

**DYNAMICS OF ZINC IN TYPIC KANDIUSTULTS WITH SPECIAL  
REFERENCE TO NUTRITION IN FODDER MAIZE (*Zea mays* L.)**

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**THESIS**

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**DEPARTMENT OF SOIL SCIENCE AND AGRICULTURAL CHEMISTRY  
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VELLAYANI, THIRUVANANTHAPURAM, KERALA**

**2010**

## DECLARATION

I hereby declare that this thesis entitled **“Dynamics of Zinc in Typic Kandiustults with special reference to nutrition in fodder maize (*Zea mays* L.)”** is a bonafide record of research work done by me during the course of research and that the thesis has not previously formed the basis for the award of any degree, diploma, associate ship, fellowship or other similar title, of any other university or society.

Vellayani,  
03.02.10

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(2006 -21-115)

## **CERTIFICATE**

Certified that this thesis entitled “**Dynamics of Zinc in Typic Kandiusults with special reference to nutrition in fodder maize (*Zea mays* L.)**” is a record of research work done independently by Ms. THANKAMONY, K. (2006-21-115) under my guidance and supervision and that it has not previously formed the basis for the award of any degree, fellowship or associate ship to her.

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## LIST OF ABBREVIATIONS

Am.FeOZn-Amorphous iron oxide bound Zn

Av.-Available

Ca-Calcium

CF-Crude Fibre

cm-centimeters

cmol-centimolper kilograms

CP-Crude Protein

CPY-Crude Protein Yield

Cu-Copper

Cv.-Cultivar

DAS-Days after sowing

DFY- Dry Fodder Yield

dSm<sup>-1</sup>-desi Siemens per meter

DTPA-Diethylene Triamine Penta Acetic acid

EC-Electrical Conductivity

Fe-Iron

FAO-Food and Agricultural Organization

FYM-Farmyard manure

GFY- Green Fodder Yield

kg ha<sup>-1</sup>- Kilo grams per hectare

K-Potassium

LAI-Leaf Area Index

LSR- Leaf Stem Ratio

mg kg<sup>-1</sup>-milligram per kilogram

Mg m<sup>3</sup>-mega gram per meter cube

Mg-Magnesium

Mn-Manganese

## LIST OF ABBREVIATIONS CONTINUED

MnO.Zn-Manganese oxide bound Zn

MOP-Muriate of Potash

N-Nitrogen

OB Zn-Organically bound Zn

PDGFY-Per Day Green Fodder Yield

POP-Package of Practice Recommendation

P-Phosphorus

ppm- parts per million

Res-Zn- Residual Zn

S-Sulfur

Ws+Ex.Zn-Water soluble +Exchangeable Zn

ZnSO<sub>4</sub>.7H<sub>2</sub>O-Zinc sulfate heptahydrate

Zn-Zinc



## 1. INTRODUCTION

Zinc is an essential trace element necessary for plants (Sommer and Lipman, 1926) and animals (Prasad, 2008). Zinc is found in nearly 100 specific enzymes and is the second most abundant transition metal in organisms after iron and it is the only metal which appears in all enzyme classes (Broadley *et al.* 2007). Zn is essential for the normal healthy growth and reproduction of plants and animals and when the supply of plant available Zn is inadequate crop yields are reduced and the quality of crop products is frequently impaired.

In plants Zn plays a key role as a structural constituent or regulatory co-factor of a wide range of enzymes. It is also involved in many biochemical pathways and these are mainly concerned with carbohydrate metabolism, both in photosynthesis and in the conversion of sugars to starch, protein metabolism, auxin metabolism, pollen formation etc. When the supply of Zn to the plant is inadequate, one or more of the many important physiological functions of Zn is unable to operate normally and the growth of the plant is adversely affected. Even a reduction in yield up to 40 per cent has been reported without obvious visible symptom. Losses of yield of 40 per cent or more in many zinc deficient soils have a major economic impact on the farmer due to the reduced income as a result of yield loss (Alloway, 2004). Zn is believed to promote RNA synthesis and its role in gene expression and regulation were also reported (Das, 2007).

The Food and Agricultural Organization has estimated that 50 per cent of the world's agricultural lands are deficient in Zn (Mikko Sillanpaa, 1990). Further the WHO attributes eight lakhs deaths world wide each year to zinc deficiency and highlights that zinc deficiency in human is largely related to inadequate intake or absorption of Zn in the diet (Hambridge and Krebs, 2007)

Among the micronutrients, deficiency of Zn is more predominant in soils of India with highest rate of 57 per cent in acid soils of Kerala and Meghalaya followed

by Jharkhand, Orissa and West Bengal (23-54 per cent ) (Sarkar and Singh, 2003). The depletion of micronutrients from the soil is increasing due to the adoption of modern technologies in crop production such as the use of high analysis fertilizers and cultivation of newer and nutrient exploitative high yielding crop varieties. Occurrence of micronutrient deficiencies in Indian soil to an extent of 46, 9, 5 and 4 per cent Zn, Fe, Cu and Mn respectively. In another report, 49 per cent of samples were found to be Zn deficient in India. (Tiwari, 2008). Majority of the Kerala soils recorded a low content of available Zn (Mathew, 2006).

The use of Zn fertilizers in Zn deficient soils can increase crop productivity as well as the Zn nutritional status of crops. Proper understanding of micronutrient availability in soils and extent of their deficiencies is the pre-requisite for efficient management of micronutrient fertilizers to sustain crop productivity. The magnitude of response to micronutrient application varies widely from crop to crop and on different soils for the same crop. Since the difference between deficiency and toxicity levels is narrow, in many situations, location specific recommendation is necessary. Not only there is inherent variability in the behaviour of soils with respect to nutrient availability but also the crops grown influence the transformation of nutrients and their availability in soils.

Maize crop was found to be responsive to Zn. Fodder crops respond to micronutrients by increasing the drymatter yields and by improving their quality as a food for animal nutrition (Mikko Sillanpaa, 1990). Among the cultivated cereal forages, maize (*Zea mays* L.) is one of the most important crops grown all round the year under irrigated conditions. African tall variety of fodder maize is a quick growing, succulent, palatable, high yielding, and nutritious variety without any antinutritional qualities that increase the milk production.

Live stock is an integral component agriculture that plays a vital role in rural economy. Balanced nutrition and adequate feeding of animals are the major factors

for improved animal production on sustainable basis. The demand for milk and meat is linked to demand for fodder. Shortage of fodder is a major problem in livestock production. Since there is only limited scope for extending the acreage under forage production, increasing forage productivity is the alternate way out for meeting the demand for fodder.

The deficit in green fodder and dry fodder in the country was estimated at 61.96 and 22.08 percent respectively during 2005 and it will increase to 62.76 and 23.46 percent during 2010. With the shift in cropping pattern of Kerala, the area under rice has come down by 50 per cent over the last two decades leading to drastic reduction in the availability of straw for feeding cattle in Kerala. One of the main constraints for increasing milk production is the shortage of quality fodder. The trend over the last five years period from 2001-02 to 2005-06 showed a continuous decline in total area brought under fodder cultivation in Kerala i.e. from 4315 ha to 1600 ha (Anon., 2008). This also necessitates the increased productivity of the fodder crop. The per capita land is decreasing every year due to unabated increase in human population, so there is little scope for horizontal expansion of area under plough and the future requirement of fodder has to be met through intensification of cropping.

Nutrient content in various parts of plant varies and logging it can assist in evaluating the nutrient status of crops. The knowledge of the plant parts which accumulate the highest concentration could prove to be useful in delineating the deficiency level of nutrients from sufficiency and toxicity levels (Sharma and Bapat, 2000). Tandon (1989) reported that middle season deficiencies in crops can easily be corrected through foliar spray. Keeping all these points in view, the present study was undertaken with the following objectives:

- ❖ To elucidate the role of Zinc in nutrition of fodder maize as evidenced by its growth, yield and quality attributes when supplied to soil as well as foliar application.

- ❖ To record the pattern of periodic release of Zn by its carriers on incubation with soil and its interaction with the major soil nutrients will also be studied.
- ❖ To select of the index plant part for foliar diagnosis for fixing up the critical nutrient level in that part for maximum crop yield.

## 2. REVIEW OF LITERATURE

Among the micronutrient disorders Zn deficiency in field crops constitutes a major soil fertility problem in many parts of the World. The critical value of Zn as determined by DTPA extraction varies from soil to soil and from crop to crop (Katyal, 1973). Zn is essential for several enzyme systems that regulate various metabolic activities in plants. Zn occurs in soils in different chemical pools which differ in their solubility and availability to plants (Mandal and Mandal, 1996 and Das, 2003). The different aspects of Zn nutrition in crops and Zn dynamics in soil are reviewed here.

### 2.1 Role of Zinc in the nutrition of crops

The essentiality of Zn for higher plants was established by Sommer and Lipman (1926). Tsui a (1948) reported that Zn has an active role in the production of auxins for a range of plant species and first suggested the role of Zn in the water relations and osmotic pressure of tomato plants.

According to Kessler (1961), Zn was required for the activity of various enzymes such as dehydrogenases, aldolases, isomerases, transphosphorylases, and RNA and DNA polymerases.

Jyung *et al.* (1975) have reported that Zn was involved in the carbohydrate and protein metabolism.

Carbonic anhydrase, alcohol dehydrogenase, Cu – Zn Super oxide dismutase and RNA polymerase contain bound Zn (Marschner, 1986).

Suge *et al.* (1986) studied that the giberellic acid metabolism was impaired by Zn deficiency in plants.

Cakmak *et al.* (1989) stated that Zn was required for the synthesis of tryptophan, a precursor for the synthesis of IAA.

Cakmak and Marschner (1990) have reported that Zn appears to be involved in all membrane proteins probably due to Zn preferential binding with –SH groups and the loss of membrane integrity in Zn deficient plants.

Graham and Reed (1991) established the relationship between carbonic anhydrase and the regulation of photosynthesis suggesting Zn involvement in the photosynthesis reaction of the plant.

Miller *et al.* (1991) and Frageria *et al.* (2002) reviewed Zn as an essential micronutrient for humans, animals and plants.

The role of Zn in higher plants was, as a divalent cation ( $Zn^{++}$ ) which acts either as a functional, structural or a regulatory co-factor of a large number of enzymes (Brown *et al.* 1993).

Kochaian (1993) reported that in grasses the non-protein amino acids called phytosiderophores can complex  $Zn^{2+}$  in the rhizosphere and transport Zn into the root cell.

Norvell and Welch (1993) investigated the role of Zn in regulating the uptake of nutrients, as Zn treatments produced an increase of P and Mn concentrations in barley shoots.

Welch and Norwell (1993) suggested that Zn had a protective role in preventing the oxidation of sulfi hydryl groups to disulfides in the root cell membrane.

Shuman *et al.* (1995) indicated the involvement of Zn in stomatal opening, possibly as a constituent of carbonic anhydrase needed to maintain adequate bicarbonate in the guard cell and also Zn affected the influx of K<sup>+</sup> uptake into guard cells.

Cakmak (2000) proved that Zn has a role in protecting cells by both controlling regeneration as well as detoxification of reactive oxygen species.

Zn has been identified as a yield limiting factor for maize in Ludhiyana (Anon, 2000).

Jyolsna (2005) reported that Zn influenced the growth, yield and quality of tomato.

Takkar (2006) reported, Zn as a constituent of carbonic anhydrase, alcoholic dehydrogenase, Super Oxide Dismutase, lactate dehydrogenase, aldolase, phosphatase, DNA -RNA polymerase etc.

The Zn plays a key role in auxin and protein synthesis, seed production, rate of maturity and membrane integrity. Zn is believed to promote RNA synthesis and recently its role has been reported in gene expression and regulation (Das, 2007).

Singh (2009) opined phytosiderophores in cereal crops increased Zn mobility in rhizosphere and uptake and also translocation of Zn from root to shoot. This may be involved in affecting Zn uptake efficiency of graminaceous species under Zn deficiency.

## **2.2. Micronutrient status of Kerala soil**

The status of available Fe for rice soils was high and reaches toxic limits (Pisharody, 1965). Jacob (1989) reported that the values of available Fe ranged

from 1.6-200 mg kg<sup>-1</sup> in Kozhikode and 9.2 to 5066 mg kg<sup>-1</sup> in Palakkad. This study also revealed that the mean values of available Fe in soils of lateritic alluvium, Karappadom, Kayal and coastal sandy alluvium were 307, 393, 282 and 127 mg kg<sup>-1</sup> respectively.

Rajagopalan (1969) observed that the level of total Mn was the highest in black soil (950 mg kg<sup>-1</sup>) and the lowest in sandy soil. (104 mg kg<sup>-1</sup>).

Praseedom (1970) noticed that the available Zn status of Kerala soils varied between 0.25-8.0 ppm and that available Zn status in soils decreased with depth.

Singh (1998) observed the low Cu status in Kerala soils.

Rajendran (1981) found that all rice soils of Kerala except the coastal sandy alluvium and the lateritic alluvium were well supplied with available Mn.

Sharma *et al* (1986) reported a critical limit of 0.5 mg kg<sup>-1</sup> in the soils of Kerala

According to Usha (1995), the available Fe content of wetland soils ranged from 89-455 mg kg<sup>-1</sup>. The lowest value was observed for soils of Wayanad and the highest for Pokkali soils.

Sheeja *et al.* (1996) reported that available Zn content in sandy soil ranged from 0.09-2.90 mg kg<sup>-1</sup> and in laterite soil the range was from 0.2-7.08 mg kg<sup>-1</sup>.

Thampatty (1997) observed that the total Fe content of plough layer of Kuttanadu soils ranged from 1.62 to 7.1 per cent.

Sathyanarayanan (1997) reported that the available and total Zn in major land resources of Southern districts of Kerala was in the range of 0.3 to 6.3 mg kg<sup>-1</sup> respectively.



Thampatty (1997) reported that Cu content ranged from 1.5 – 10.9 mg kg<sup>-1</sup> in Kuttanadu soils. Sathyanarayanan (1997) found out that available Cu status ranged from 0.6 – 5.8 mg kg<sup>-1</sup> in Southern districts.

Mathew (2000) reported that available Cu ranged from 0.7 – 2.5 mg kg<sup>-1</sup> in wet land and 1.11 -3.60 mg kg<sup>-1</sup> in garden land soils of Kerala.

The mean status of total Zn in the wet land soils and garden soils of Kerala were in the range of 24.1 to 78.9 and 11.5 to 43.5 mg kg<sup>-1</sup> respectively (Mathew, 1999). The available Zn ranged from 1.4 to 2.9 and 0.9 to 3.2 mg kg<sup>-1</sup> in the above soils. The available content of Zn in the surface samples of laterite soils in Kerala was found to be low (Mathew, 2006).

Nair *et al.* (2007) reported acute Zn deficiency in low land rice soils of Palakkad

According to Koshy *et al.* (1990) the available B content of Vellayani Series and Neyyattinkara Series was 1.52 and 1.26 ppm respectively

Jyolsna (2005) reported that the available B content of Vellayani Soil was 0.35 mg kg<sup>-1</sup>

The available Mo status of Vellayani Series was 0.04 mg kg<sup>-1</sup> while that of Neyyattinkara Series was 0.06 mg kg<sup>-1</sup> (Koshy *et al.*, 1990).

### **2.3 Physico-chemical properties of soil in relation to the availability of Zn**

Brawn *et al.* (1962) revealed that the acidification of soils increased the concentration of available Zn to plants.

Yoshida *et al.* (1973) observed that presence of excess organic matter tends to influence soil Zn and Zn uptake by plants.

Reddy and Perkins (1974) studied the absorption of Zn by kaolinite at different pH values and found that Zn was precipitated in clay lattice surfaces or adsorbed at surface exchange sites.

Saeed and Fox (1977) showed that Zn solubility at high soil pH was due to organic matter dispersion which can either release complexed Zn or supply ligands for metal complexation and decrease precipitation.

Kalbasi et al (1978) reported that the absorption of Zn by Al oxide was pH dependent and increased markedly with increasing pH. At equivalent pH values FeO absorbed more Zn than Al oxides.

The rate of absorption of Zn in goethite was slow at low pH but increased rapidly with increasing pH due to the high point of zero charge of goethite (Padmanabhan, 1983).

According to Barber (1984) and Morgan (1984) the Zn concentration in plants may decrease by 3-4 times for each one unit increase in soil pH.

Jahiruddin *et al.* (1986) revealed that at high soil pH, Zn is more strongly adsorbed on to the surface of silicate clays and oxides and hence the availability of Zn to plants is diminished. An increase in soil pH, decreased Zn availability for plant uptake. Moisture status in soil was found to have significant effect on Zn availability and it was more in air dry soil than in soil at field capacity and full saturation (Jyolsna, 2005).

Brummer *et al.* (1988) postulated that the diffusion of Zn into goethite is one possible reason for declining availability of Zn with time. The internal binding sites of goethite increased with time, temperature and Zn concentration.

Mortvedt and Kellose (1988) noticed that placing Zn with acid forming fertilizer increased the availability of applied Zn to plants.

Rattan *et al.* (1999) reported the pH determined the availability of Zn in the soils.

Clemente *et al.* (2003) suggested that soil acidification due to oxidation of metallic sulfides in the soil increases bioavailability of Zn.

Vasegi *et al.* (2003) reported that Zn concentration in maize tissues increased with decrease in pH.

Studies conducted by Lombanes and Singh (2004) revealed that Manganese deficient soil conditions induced elevated concentration of Zn in the shoots of barley plants.

Reactions of applied Zn with soil constituents appear to be the major cause of the reduction of the Zn availability with time, since capacity of soils to absorb Zn greatly exceeds the usual amount of Zn applied (Brennan, 2005).

#### **2.4 Influence of organic matter on Zn**

According to Himes and Barber (1957) Zn complexed with insoluble humus complexes is a significant component of the specifically absorbed Zn while additional Zn is non-specifically absorbed on exchange sites of organic matter.

The interactions between humic substances and Zn, fulvic acids form chelates with Zn ions over a wide pH range on increasing the solubility and mobility of Zn (Mortenson, 1963; Mc Bride, 1989; and Norvell, 1991).

It was observed that 60 to 75 per cent of soluble Zinc was present as soluble organic complexes [Hodgsen *et al* (1966) : Geering and Hodgson (1969)].

Schnitzer and Skinner (1966) and Stevenson and Adrakani (1972) determined the stability constant ( $\log K$ ) and Zn fulvate as 1.7 at pH 3.5 and 2.3 at pH 5.0.

A range of organic compounds such as humic substances, carbohydrates, proteins, peptides, amino acids, lipids, waxes, polycyclic aromatic hydrocarbons and lignin fragments were contributed by soil organic matter (Stevenson, 1991).

Stevenson and Adrakani (1972) divided the organic compounds that form stable complexes in soil into 2 groups first, simple compounds such as organic acid, polyphenols, amino acids, peptides, proteins and poly saccharides and second complex polymers formed by secondary synthesis reactions such as humic and fulvic acids. This study also revealed that soluble Zn complexes are mostly from group 1 and these maintain Zn in solution at soil pH values and resulted in insoluble precipitation of Zn compounds while, reactions of Zn with second group tend to form insoluble complexes that are unavailable to plants, although humic and fulvic acid may also form soluble complexes. They have also reported the relative significance of these compounds was difficult to determine because of analytical problems in measuring the small quantities involved, their dynamic nature due to balance between synthesis and degradation and also their heterogenous distribution throughout the soil.

However, Schnitzer and Khan (1978) reported  $\log$  values as 2.3 at pH 3 and 3.6 at pH 5.0. Thus the Zn complex stability increases with increasing pH value in the acid range. Many of these compounds of soil organic matter have a strong affinity to bind Zn.

Humic acids are insoluble in acid conditions and dissolved gradually as pH increases. Humic acids are completely soluble in alkaline media and behave as colloidal system, which means that they can be flocculated by Mg and Ca. Soil organic matter is an important factor affecting the behaviour of Zn in soils. The fulvic acids and low molecular weight organic acids mainly form soluble complexes and chelates with Zn, thus increasing its mobility in the soil (Alloway, 1990).

Stevenson (1991) reported that both humic and fulvic acid substances contained a large number of functional groups (OH, COOH, SH, C-O) that have a strong affinity for Zn and other micronutrient ions, Cu and Mn. Application of FYM along with nutrients had an additive effect on increasing the DTPA – Zinc (Jagtap, *et al*, 2007).

## **2.5 Interaction of Zn with other nutrients**

Hulagur and Dangarwala (1983) recorded the increased uptake of N, Ca, Mg and S in maize due to the application of Zn.

Application of higher doses or presence of higher levels of either Zn or P in soil may reduce or enhance the availability of the other nutrients (Orabi, 1985).

Zn and P were reported to interact either antagonistically or synergistically depending upon a number of physicochemical characters of the soil (Barrow, 1987).

Devarajan *et al.* (1988) observed the antagonistic effect of P and Zn where high levels of Zn recorded generally lower P content and uptake in pulse crops.

Loneragan and Webb (1993) reported negative relationship between Zn and P.

Nitrogen contents in grain and straw were increased significantly while P content decreased with increase in level of Zn. The low concentration of P might be due to antagonistic effect of Zn (Singh *et al.* 1993).

Takkar and Walker (1993) investigated that Zn deficiency in plants was associated with calcareous high pH soils, because of their low total Zn content.

Krishna (1995) reported significant positive effects of Zn treatment on dry matter, seed and straw yield of mung bean as well as crude protein percentage in the seeds.

Mishra and Singh (1996) reported that the interactive effect of N and Zn significantly influenced the Zn accumulation.

Geethakumari *et al.* (2005) reported Zn exhibit positive interaction with N and K, while negative with P. High concentration of Ca, Mg and Na inhibited the absorption and translocation of Zn. They also reported the interaction between Fe, Mn and Cu with Zn are also negative in nature.

Das *et al.* (2007) noticed synergistic interaction between Ca and Zn

Fe and Mn are reported to be antagonistic in the absorption of Zn by roots (Varshney *et al.*, 2008).

## **2.6. Critical level of Zn in soil and crops**

Cate and Nelson (1965) indicated that  $1.2 \text{ mg kg}^{-1}$  in soil and  $35.95 \text{ mg kg}^{-1}$  in rice plants as the critical limits for Zn.

Cate and Nelson (1971) noticed that the critical level of DTPA Zn in soil was  $0.5 \text{ mg kg}^{-1}$ .

The critical level for DTPA extractable Zn for red and laterite and specifically for the laterite soils of Kerala were fixed as  $0.6$  and  $1.2 \text{ mg kg}^{-1}$  respectively (Valsaji, 1972).

Aiyer *et al.* (1975) observed the critical limit of available Zn in the acid rice soils of Kuttanad as  $0.5 \text{ mg kg}^{-1}$ .

Brennan and Gartrell (1981) reported the critical value for Zn in soil for subterranean clover as  $0.25 \text{ mg kg}^{-1}$ .

According to Sakal *et al.* (1981), the critical Zn concentration in 52 days old rice and wheat were 37 and 29 ppm respectively.

The work done by Dudde (1986) revealed the critical levels for DTPA Zn in soil and plants respectively were  $0.74$  and  $23.35 \text{ mg kg}^{-1}$  for wheat and  $0.72$  and  $21.90 \text{ mg kg}^{-1}$  respectively for gram. The critical values were  $0.75$  and  $29 \text{ mg kg}^{-1}$  in mungbean and  $0.60$  and  $25.70 \text{ mg kg}^{-1}$  in groundnut.

Available Zn in the range of  $0.72$  to  $2.4 \text{ mg kg}^{-1}$  was suggested as the medium range of availability to crops (Miko Sillanappaa, 1990).

Maleswar and Dudde (1995) have developed the critical concentration for DTPA Zn in cotton (NHH – 44) and sorghum (CSH – 5) as  $0.86$  and  $0.85 \text{ mg kg}^{-1}$  in soil, while it was  $28.40$  and  $27.50 \text{ mg kg}^{-1}$  respectively for plants.

Gupta and Vyas (1999) found that that the critical limit of DTPA extractable Zn for soyabean in vertisols as  $0.5 \text{ mg kg}^{-1}$ .

Singh *et al.* (1999) observed that in the wetland rice soils of Meghalaya, DTPA extractable Zn was less than  $1.2 \text{ mg kg}^{-1}$  and Zn concentration below  $35.90 \text{ mg kg}^{-1}$  in rice plants, as the critical limit.

Karche *et al.* (2003) established the critical levels of DTPA Zn in soil and plants as  $0.41$  and  $117.0 \text{ mg kg}^{-1}$  for Sorghum (CSH – 5);  $0.88$  and  $40 \text{ mg kg}^{-1}$  for wheat (HD 2189);  $0.72$  and  $21.90 \text{ mg kg}^{-1}$  in Grams (Vijay). This study also

developed the critical values for DTPA Zn using groundnut (JL – 24) and Tomato (Roopali). For soil it was 0.62 and 0.72 mg kg<sup>-1</sup>, where as the values for plant was 25.70 and 20.50 mg kg<sup>-1</sup> respectively.

Pattil and Bharambe (2003) reported that the critical limits of DTPA Zn in rice and cotton growing soils was 0.62 to 0.64 mg kg<sup>-1</sup> respectively while that of plants was 32 and 21 mg kg<sup>-1</sup> dry matter.

Available Zn in clay bean soils of Kandiustults of Jammu region ranged from 0.28 – 2.4 ppm (Deepak *et al.* 2004).

Mehra *et al.* (2005) established a ranged value of critical limits for DTPA Zn for maize growing soil and the same varied from 0.30 to 4.12 mg kg<sup>-1</sup>, while for plant, it ranged between 10.21 to 47.64 mg kg<sup>-1</sup>.

According to Mehra *et al.* (2005), the critical limit for DTPA Zn was 0.95 mg kg<sup>-1</sup> in soil and 32.7 mg kg<sup>-1</sup> in plant.

John (2006) reported that the critical limit for Zn in Vellayani Soils was 0.60 mg kg<sup>-1</sup>.

Chaudhary and Raina (2008) reported the critical range of Zn in apple plants as 16.0 – 20.0 ppm.

The critical Zn concentration in cereal crops below which Zn deficiency is likely to occur varied from 12 to 24 mg kg<sup>-1</sup>. The leaf blade 3 from the top is the most sensitive in relating relative yields of maize shoots to Zn concentration in the tissue (Singh, 2009)



The critical limit of 2.10-3.74 mg kg<sup>-1</sup> for DTPA Zn in soil and 27.0-53.8 mg kg<sup>-1</sup> Zn for leaf for maximum yield of ginger in an Ustic Humitropept (Sreenivasan *et al.* 2009).

## **2.7 Availability of Zinc to crop plants**

Halvorson and Lindsay (1977) studied the availability of Zn to maize plants and stated that the availability depended on its form in solution and speciation of Zn as an important factor controlling Zn reactions in soil and its uptake by plants.

Singh and Sekhon (1977) opined that the speciation of Zn and other micronutrients was controlled by the reactions that occur in soil solution while the concentration of Zn in solution is controlled by adsorption to soil phases.

Bould *et al.* (1983) stated that concentration of Zn in wheat and barley for sufficiency was 40 and 35 ppm respectively.

Shuman (1985) reported that the availability of Zn to plants depended on several soil factors such as the concentration of Zn in solution, ion speciation and the interaction of Zn with other macro and micronutrient elements.

## **2.8. Effect of Zinc on the yield and quality of crops**

Usage of ZnSO<sub>4</sub> in tea plantations during high cropping season increased the bush ability to utilize major nutrients (Anon, 1975).

Das (1986) reported that application of ZnSO<sub>4</sub> · H<sub>2</sub>O at 20 kg ha<sup>-1</sup> gave the highest average yield of 4.12 t ha<sup>-1</sup> for rice on an acid lateritic soil.

Application of  $\text{ZnSO}_4$  @ 5-10 kg ha<sup>-1</sup> or foliar application of  $\text{ZnSO}_4$  @ 1.25 per cent increased tuber yield and decreased HCN content in cassava in laterite soils of Kerala (Nair and Abrol, 1989).

Maize was found to be responsive to Zn. Fodder crops respond to micronutrients by increasing the dry matter yields and by changing their quality as a food or animal nutrition (Mikko Sillanpaa, 1990).

Singh *et al.* (1992) revealed that mean grain and straw yield in barley increased significantly up to Zn 5 kg ha<sup>-1</sup>. The significant response of barley to Zn upto  $\text{ZnSO}_4$  5 kg ha<sup>-1</sup> was attributed to the low status of Zn availability in the soil and fulfilled by optimum requirement.

The increase in crude protein content with Zn addition was due to the involvement of Zn in the nitrogen metabolism (Frescenko and Lozek, 1998).

Channakesava *et al.* (1999) inferred that soil application of  $\text{Zn SO}_4$  @ 10 kg ha<sup>-1</sup> resulted in increased seed yield and yield components in African Tall fodder maize.

The favourable influence of  $\text{ZnSO}_4$  on the higher yield of fodder maize may be attributed to its role in various enzymatic reactions, growth processes, hormone, production and protein synthesis (Mehla, 1999).

Singh *et al.* (1999) observed that the use of Zn fertilizer increased the yield of rice in wetland soils of Meghalaya.

Meerabai (2001) reported increase in ginger yield by the application of Zn @ 4 kg ha<sup>-1</sup> increase in turmeric yield, oil content and curcumin content by applying 10 kg ha<sup>-1</sup>.

Reddy and Ahlawat (2002) reported that application of Zn along with P increased grain yield in lentil.

The optimum level of Zn in different crops ranged between 5-10 kg ha<sup>-1</sup> and response of crops to Zn application varied widely because of the differences in soil characteristics, available Zn status of soil and crop varietal characteristics (Sakal, 2001).

Singh et al. (2001) observed that the use of Zn fertilizer increased the yield of rice in wetland soils of Meghalaya.

Umamaheswarai and Singh (2002) showed that application of ZnSO<sub>4</sub> @ 5 kg ha<sup>-1</sup> increased significantly the yield and yield attributes such as number of pods plant<sup>-1</sup>, number of seeds pod<sup>-1</sup> and 100 seed weight of rajmash.

The application of Zn 10 kg ha<sup>-1</sup> significantly increased plant height and green fodder yield in pearl millet (Dadwhich and Gupta, 2003).

Devi and Rani (2003) reported that application of 25 kg ha<sup>-1</sup> of ZnSO<sub>4</sub> improved significantly the grain yield by increasing the number of panicles hill<sup>-1</sup>, number of spikelets panicle<sup>-1</sup>, number of filled grains panicles<sup>-1</sup> and test weight in various rice hybrids. The increased yield was found to be 48.2 per cent higher than that of control.

Dewal and Pareek (2004) indicated that Zn fertilization @ 10 kg ha<sup>-1</sup> significantly enhanced all the yield attributes and grain, straw and biological yields in wheat.

According to Ravikiran and Reddy (2004), foliar spray of ZnSO<sub>4</sub> (0.5 per cent) increased the total number of tillers per hill, number of productive tillers per hill, number of filled grains per panicle, 1000 grain weight and grain yield in the rice cv MT4 100.

Verma *et al.* (2004) reported that foliar application of Zn (0.5 per cent ZnSO<sub>4</sub>) increased plant height, number of leaves and branches per plant in pigeon pea.

The application of Zn 5.0 kg ha<sup>-1</sup> significantly increased growth of wheat plants which was on par with 6.25 and 7.25 kg ha<sup>-1</sup> (Singh, 2004).

Jyolsna (2005) reported that application of Zn as ZnSO<sub>4</sub> @ 5 kg ha<sup>-1</sup> was ideal for economic yield in tomato.

Application Zn 5 kg ha<sup>-1</sup> along with NPK fertilizers were found suitable for achieving greater productivity in plants (Shivoy and Kumar, 2005).

When Zn was applied @ 4.4 kg ha<sup>-1</sup> along with N, P, K and S, gave the highest number of nodules per plant, nodule N, leghaemoglobin contents, pod yield and highest protein content in groundnut (Tiwari *et al.*, 2006).

Maximum Zn uptake was obtained for maize when Zn chelates were used @ 10 mg kg<sup>-1</sup> (Alvarez, 2007).

Application of ZnSO<sub>4</sub> 10 kg ha<sup>-1</sup> resulted in increased stem diameter, more number of seed rows, and increased seed yield in maize (Channakesava *et al.* 2007).

Chaube *et al.* (2007) recorded that application of 1 per cent ZnSO<sub>4</sub> significantly increased the plant height, number of leaves, leaf area and yield in pearl millet.

Haris *et al.* (2007) application of 2.75 kg Zn ha<sup>-1</sup> significantly increased mean maize grain yield by 720 kg ha<sup>-1</sup> (25 per cent) total dry matter, number of cobs and cob weight.

## 2.9. Total Zinc and Fractions of Zinc in soil

The behaviour of Zn ions in soils and their uptake cannot be explained by the total concentration of Zn in the soil. Total Zn concentration of soils largely dependent on the composition of the parent rock material (Graham, 1953; Swaine, 1955 ;Swaine and Mitechell, 1960; Tiller, 1963. Mikko Sillanappa, 1972; Kabata Pendias and Pendias, 1984). Total Zn concentration was seldom used as a test for evaluating plant availability of Zn as the total Zn soil often incorporate Zn in unweathered minerals that is unavailable for plant growth.

The total concentration of Zn for soils ranged from 10-300 mgkg<sup>-1</sup> (Goldschmidt, 1954).

Soils with a low total Zn concentration are often Zn deficient for crop production. Since quartz contains negligible Zn, sandy soils are inherently low in total Zn concentration and frequently deficient in available Zn. Zn deficiencies of plants grown on acid soils are generally associated with low total soil Zn concentration (Lucas and Davis, 1961).

Viets (1962) and Hodgson (1963) reported that the successive pools of Zn form ions in the solution to Zn in crystal lattices and they are considered to represent pools of decreasing availability for plant uptake.

Total Zn concentration in calcareous soils are often similar to or higher than those of non-calcareous soils ,but Zn deficiency was repored for calcareous soils (Yoshida and Tanaka,1969). Though the total soil concentration of Zn in calcareous was reported to be higher, but it was not available to plants due to the effects of soil pH.

The mean Zn concentration ranged from 40 mg kg<sup>-1</sup> in acid rocks (granites) to 100 mg kg<sup>-1</sup> in basaltic rock (Krauskopf, 1972).

Lindsay (1972) reported that distribution of total Zn down the soil profile reported to be uniform, while extractable Zn decreased with depth.

The average concentration of Zn in basaltic rock was usually higher than the Zn concentration in granitic rocks. Wedepohl (1973) investigated and found magnetite as the most important Zn carrier in basaltic rocks, while biotite was the main source of Zn in granitic rocks.

Norrish (1975) reported that in igneous rock, low concentration of Zn was present in sulfides, but higher concentration of Zn in the more abundant silicates.

According to Takkar (1982) the total Zn content of Indian soils was in the range of 0 -1019 mgkg<sup>-1</sup>.

Kabata Pendias and Pendias (1984) investigated that in the sedimentary rocks, the highest Zn concentrations were found in shales and clayey sediments (80-120 mg kg<sup>-1</sup>) while sand stones, lime stones and dolomites generally have lower concentration (10-30 mg kg<sup>-1</sup>).

Shuman (1985) reported the different Zn solutions as water soluble and exchangeable, organically bound, MnO bound, Amorphous Fe O bound, Crystalline FeO bound, Residual and total. Most procedure of chemical fractionation showed that the residual Zn is the greatest component of Soil Zn.

Barrow (1986) opined that the concentration of Zn OH<sup>+</sup> explained Zn adsorption on soil surfaces better than the total concentration.

It was inferred that total Zn content is an unsatisfactory measure of Zn availability to crops (Katyal and Rattan, 1993; Sureshkumar *et al.*, 2004).

Neilson *et al.* (1986) determined the distribution of Zn in soil fractions. The total Zn was measured as 51.5-226.3 mg kg<sup>-1</sup> with an average of 89.7 mgkg<sup>-1</sup>. The exchangeable Zn consisted of 0.3-23.2 per cent, 0.5-29.7 per cent associated with organic matter, 1.3-15 per cent was in Fe and Al oxides. The MnO bound fraction ranged from 6.4-24 per cent while the residual Zn varied from 45.6-92 percent

Yaduvanshi and Sharma (1988) studied the transformation of Zn on an acid soil (Mollisol). The fractions of Zn for surface soil (0-15-cm) was found as 1.0 – 3.8ppm, 26.4 – 400ppm, 18.0 – 34.0 ppm, 10.9 – 18.0 ppm and 56.0 – 68.0 ppm for water soluble + exchangeable, specifically sorbed at inorganic sites, organically bound Zn, free oxides bound Zn and residual Zn respectively.

Hazra *et al.* (1993) studied the distribution of Zn fractions in red and lateritic soils of Birbhum (West Bengal). The total Zn in the surface layer varied from 24 - 85 mg kg<sup>-1</sup>. The water soluble + Exch. Zn (0.40 - 3.30 mg kg<sup>-1</sup>), organically bound Zn (3.8 per cent), MnO-Zn (1.7 per cent), AmFeO-Zn (23 per cent), CryFeO-Zn (0.84 per cent). For the subsurface (0.15 – 0.3 cm) layer, the various fractions ranged from 20 – 53 mg kg<sup>-1</sup> for total Zn, 0.2 – 2.2 mg kg<sup>-1</sup> for Ws + Ex – Zn and 0.2 – 3.5 mg kg<sup>-1</sup> for organically bound Zn. The MnO-Zn, AmFeO-Zn and CryFeO-Zn were in the range of 0.3 – 1.8, 8.5 – 13.7 and 2.3 – 12.3 mgkg<sup>-1</sup> respectively.

Randhawa and Singh (1995) reported that exchangeable Zn contributed to the pool of available Zn and thus played a significant role in the Zn nutrition of maize.

Prasad *et al.* (1996) have studied the transformation of applied Zn in the rice growing OH alluvial soils of Bihar plain under submerged condition. The amount of Zn recovered in Ws + Ex, OB and CryFeO-Zn forms recorded a progressive

decrease with increase in residual forms on submergence. But the complexed and amorphous Zn was not influenced by the submergence.

According to Sahu *et al.* (1996) the relative amount of different fractions of Zn were in the order of residual (22.1 mgkg<sup>-1</sup>) > complexed (6.68 mgkg<sup>-1</sup>) > organically bound (4.5 mgkg<sup>-1</sup>) > CBD extractable (3.03 mgkg<sup>-1</sup>) > water soluble + exchangeable (2.03 mgkg<sup>-1</sup>) > HCl soluble Zn (0.59 mgkg<sup>-1</sup>).

Sharma *et al.* (1996) reported that water soluble, exchangeable, complexed, organically bound, occluded and residual fractions constituted on an average of 0.10, 0.18, 0.55, 1.53, 2.06 and 90.7 per cent of the total Zn content of the soil respectively. This study also reported that complexed, occluded and residual Zn were the principal forms contributing to the pools of available Zn in Vertisols.

Deb (1997) has reported a wide variation in the total Zn content of Indian soils and that it might be due to the varied pedological conditions on the formation of soils. This study also reported that the total Zn content varied between 7.0 to 1000 mgkg<sup>-1</sup>

The different forms of Zn were usually measured as water soluble, carbonate bound, organic matter bound, Fe and Al oxide bound and residual fractions (Ma and Uran (1997). This study also found that wetting, drying and elevated soil temperature enhanced the transformation of recently added Zn to Fe, Al and Mn oxide fractions.

Nayyar (1999) reported wide variations in the total micronutrient content in Indian soils due to the dominant influence of pedological conditions on soil development.

Singh *et al.* (1999) reported the distribution of various fractions of Zn in soil. Total Zn ranged from 66.5 – 72.9 mgkg<sup>-1</sup>. The highest fraction of 65.6 – 76.6



percent of total Zn was remained as residual, while 1.12 – 2.73, 2.17 – 3.93, 3.08 – 5.31, 4.07 – 6.64, 12.30 – 19.5 per cent remained in Ws + Ex-Zn., OB, MnO-Zn, AmFeO-Zn and CryFeO-Zn form respectively.

Novillo *et al.* (2002) reported that The percentage of Zn in labile or exchangeable form is small, but it would appear that roots of plants obtain Zn from these pools and no Zn was detectable in the water soluble plus exchangeable fractions in a calcareous soil, while some Zn remained in the labile fractions of acidic and neutral soils.

According to Suresh Kumar *et al.* (2004) the Zn fractions were in the order of residual > amorphous FeO>, crystalline FeO and water soluble.

Chaudhary and Raina (2008) have reported that the amount of various fractions of Zn followed the order of WS+exch (4.0 per cent) < CBD extractable (7.7 per cent) < complexed (9.2 per cent) < organically bound (11.4 per cent) < acid soluble (13.4 per cent).

Mishra *et al.* (2009) reported that the total Zn ranged from 78.1 to 107.4 mgkg<sup>-1</sup> the ammonium acetate extractable Zn ranged from 0.02 – 0.84 mgkg<sup>-1</sup> and copper acetate extractable Zn from 0.34 – 0.55 mgkg<sup>-1</sup>. The pyrophosphate extractable, HCl extractable, CBD extractable were in the range of 2.22 – 3.06, 0.72 – 1.59 and 13.2 – 15.3 mgkg<sup>-1</sup> respectively.

## **2.10 Profile characteristics of Ultisol**

Bhattacharya *et al.* (1994) classified the soils of Meghalaya as acidic, low base status, rich in organic matter and the surface horizons with less than 35 percent base saturation and revealed the presence of argillic/kandic horizons. These soils were included in Ultisols in Soil Taxonomy.

Sen *et al.* (1994) studied the Kandi soils of Manipur. Based on the physico-chemical and morphological properties, the soils were classified as Ultisol as they contain Kandic horizon with the base saturation less than 35 percent.

Krishnan *et al.* (2000) studied the morphological characteristics and classification of low activity clay soils of Kerala. The soils are very deep, well drained, fairly rich in organic matter, acidic, poor in bases and have low CEC with kandic or argillic horizon and the soils are classified under the order Ultisol.

Sathisha *et al.* (2002) reported the characteristics and limitations of rubber growing soils of Northern Mizoram as very deep soils, well drained, fine loamy texture with rich organic carbon, low CEC and low base saturation, acidic and included under the Ultisols.

Anon. (2007), classified Vellayani soils as a member of clayey, kaolinitic, isohyperthermic, Typic Kandiustults. The soils were identified as dark reddish grey to yellowish red, slightly acidic, sandy clay loam to sandy clay A horizon and reddish brown to brown, very strongly acidic to extremely acidic, sandy clay loam to clay B horizon.

### 2.11 Foliar diagnosis in crops

Velesco and Nevero (1951) suggested the recently matured first two blades from the top at the time of flowering (when heads are almost completely emerged) as the index part in rice.

Riceman and Jones (1958) reported the highest content of Zn was observed in the uppermost leaf in clover.

Shorrocks (1962) reported the four basal leaves from each whorl (for trees older than 4 years) to be the index part in rubber.

Romney *et al.* (1965) reported that the 11<sup>th</sup> and 15<sup>th</sup> leaf of coconut to be the index part of coconut

According to Johnson (1978), 5<sup>th</sup> to 12<sup>th</sup> leaves when the plants are 3-4 month old as the index tissue in ginger.

The first fully mature leaf from fruit bearing laterals proved to be the index part in pepper (Sushama, 1980).

According to Asher *et al.* (1980), youngest fully expanded leaf, petiole or leaf blade at 3-4 month stage turned out to be the best indicator of nutritional status in cassava.

Saifudeen (1981) reported that the 3<sup>rd</sup> leaf of tillers when the plants are 3-4 months old could be selected as the index tissue in turmeric.

Gopi (1983) reported that the middle leaflets of 10<sup>th</sup> frond proved to be the best indicator of nutritional status of coconut.

Gnanadas (1989) standardized the leaves of 3<sup>rd</sup> whorl of bottom region as the index leaf in clove for Zn status.

George (1994) reported that petiole of the third leaf of banana, (*Musa*, AAB, Nendran) upto shooting stage, flag leaf thereafter as the index part for K status.

In sugarcane crop tillering phase is identified as the best stage of leaf sampling for nutrient analysis. Regarding the leaf position, the 5<sup>th</sup> leaf from top without sheath is ideal for diagnostic purpose in relation to N while, the above leaf with sheath is ideal for P and K (Ramesh, 1994).

According to Singh and Nayar (1997), youngest fully expanded leaf was selected as the diagnostic tissue in soyabean and green gram. The critical deficiency level in youngest fully expanded leaf associated with a 10 percent reduction in the yields of soyabean and green gram and the critical level had been established as Zn 22.1 and 22.3 mg kg<sup>-1</sup> respectively.

Nutrient content in various parts of plant can assist in evaluating the nutrient status of crops. The knowledge of the plant parts which accumulate the highest concentration should prove to be useful criterion in delineating the sufficiency and toxicity levels (Sharma and Bapat, 2000)

Jose (2005) reported that petiole from the middle 1/3<sup>rd</sup> of the total leaves could be selected as the index part in tapioca at four and a half month after planting

Application of Zn significantly increased its concentration in index tissue of rice and wheat. The flag leaf was selected as the index tissue and the zinc concentration ranged from 15.6 – 19.2 mgkg<sup>-1</sup> and 23.4 to 25.1 mgkg<sup>-1</sup> respectively for rice and wheat (Varshney *et al*, 2008).

### **3. MATERIALS AND METHODS**

The study entitled 'Dynamics of Zinc in Typic Kandiusults with special reference to nutrition in fodder maize (*Zea mays* L.) was conducted at College of Agriculture, Vellayani during 2006-09. The research work consisted of two parts; a laboratory incubation study and a field study.

#### **3.1 Incubation Study**

The incubation study was conducted to determine the release pattern of Zn from soil, treated with  $ZnSO_4 \cdot 7H_2O$  and incubated with and without FYM for a period of 75 days.

##### **3.1.1 Materials**

The soil for the incubation study was collected from the experimental site, at the Instructional Farm, College of Agriculture, Vellayani. The soil belonged to Typic Kandiusults (Red Soil) under Vellayani Series. One kilogram each of surface soil was taken in plastic containers of uniform size and incubated with  $ZnSO_4 \cdot 7H_2O$  as per treatments for 75 days by maintaining the moisture level at field capacity. The  $ZnSO_4 \cdot 7H_2O$  used contained 21% of Zn. The moisture level in each treatment was maintained at field capacity throughout the period of study replenishing the moisture lost by evaporation. The details of experiment carried out are presented below.

##### **3.1.2 Design and Layout of the experiment**

Design : Completely Randomized Design

Treatments : 10

Replications : 3

### 3.1.3 Treatments

- T<sub>1</sub> : Soil alone  
 T<sub>2</sub> : Soil + Zn @ 0 kg ha<sup>-1</sup> + FYM @ 10 t ha<sup>-1</sup>  
 T<sub>3</sub> : Soil + Zn @ 5 kg ha<sup>-1</sup>  
 T<sub>4</sub> : Soil + Zn @ 5 kg ha<sup>-1</sup> + FYM @ 10 t ha<sup>-1</sup>  
 T<sub>5</sub> : Soil + Zn @ 10 kg ha<sup>-1</sup>  
 T<sub>6</sub> : Soil + Zn @ 10 kg ha<sup>-1</sup> + FYM 10 t ha<sup>-1</sup>  
 T<sub>7</sub> : Soil + Zn @ 15 kg ha<sup>-1</sup>  
 T<sub>8</sub> : Soil + Zn @ 15 kg ha<sup>-1</sup> + FYM @ 10 t ha<sup>-1</sup>  
 T<sub>9</sub> : Soil + Zn @ 20 kg ha<sup>-1</sup>  
 T<sub>10</sub> : Soil + Zn @ 20 kg ha<sup>-1</sup> + FYM @ 10 t ha<sup>-1</sup>

Fig. 1 Layout of the incubation study

The soil samples were collected at an interval of 15 days for a period of 75 days and analyzed for available Zn by DTPA method. (Lindsay and Norvel, 1978) using Atomic Absorption Spectrophotometer. View of Incubation study is given in the Plate. 1.

## 3.2 Field Study

### 3.2.1

A field experiment was conducted to study the effect of Zn on the growth and yield of fodder maize (*Zea mays* L.) and the crop period extends from October, 2007 to September, 2008. Two experimental crops and two residual crops after each experimental crop were raised.

### 3.2.2 Experimental Site

The experiment was carried out at the Instructional Farm, College of Agriculture, Vellayani which is located at 8 ° 30' N latitude and 76 ° 54' E longitudes and at an altitude of 29 m above mean sea level.

### 3.2.3 Weather parameters

The major weather parameters during the crop period were monitored. The maximum temperature, relative humidity and total rainfall during the period ranged

Replications

R1

R2

R3

T4	T1	T10
T9	T7	T5
T6	T4	T8
T10	T9	T3
T2	T8	T7
T5	T2	T1
T1	T6	T4
T8	T3	T2
T3	T10	T6
T7	T5	T9

**Fig. 1. Layout of the incubation study**



**Plate 1. Incubation Experiment**



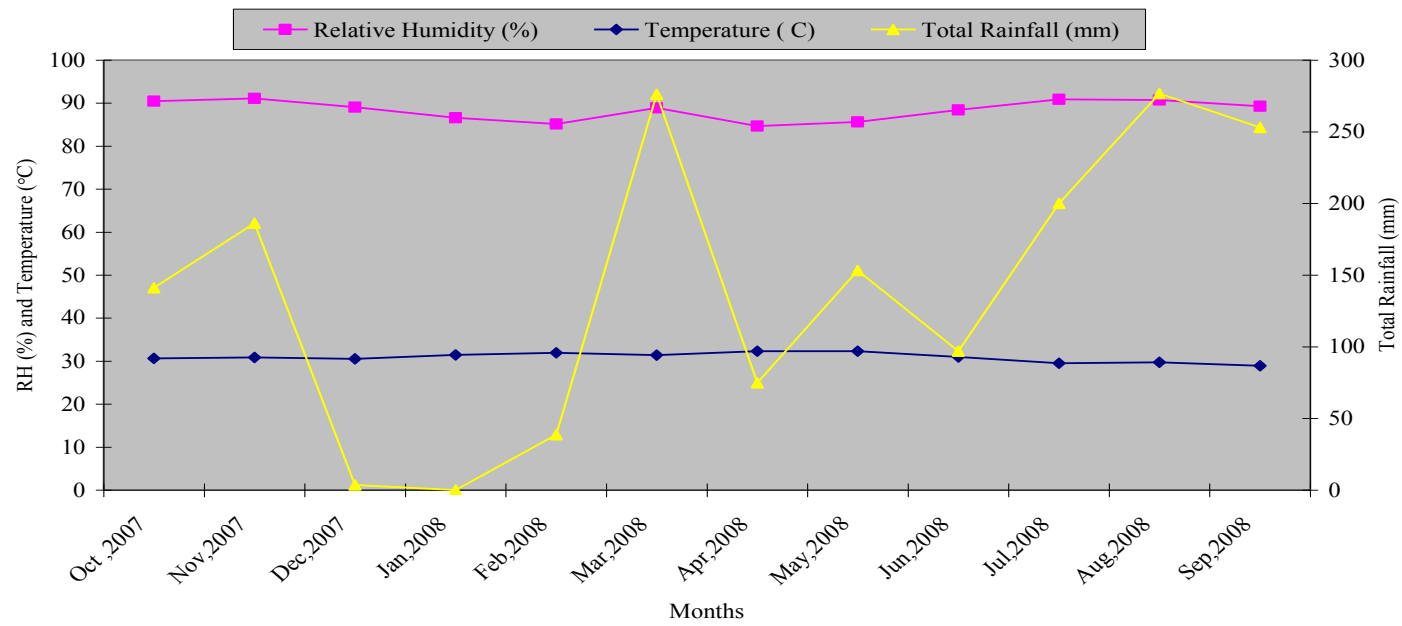
from 30.55°C to 31.45°C, 86.59% to 91.12%, 0.0 to 186.2 mm respectively. The graphical representation of the data is presented in Fig.2.

### 3.2.4 Soil

The soil of the experimental site was Clayey, Kaolinitic, Isohyperthermic Typic Kandustults belonging to Vellayani Series.

Table.1 Initial soil analysis of the experimental site

<b>Physico-chemical properties</b>	<b>Surface (0-15 cm)</b>	<b>Subsurface (15-30cm)</b>
Sand (%)	73.90	69.50
Silt (%)	10.00	12
Clay (%)	15.10	18.30
Particle density (Mg m <sup>-3</sup> )	2.36	2.38
Bulk (density) (Mg m <sup>-3</sup> )	1.28	1.30
WHC (%)	38	40.20
Texture	sandy loam	Sandy loam
<b>Chemical properties</b>		
p <sup>H</sup>	5.10	5.20
EC (dS m <sup>-1</sup> )	0.12	0.16
CEC (c mol (P+) kg <sup>-1</sup> )	3.59	2.43
Organic carbon (%)	0.71	0.69
Available Nitrogen (kg ha <sup>-1</sup> )	271.00	227.50
Available Phosphorous (kg ha <sup>-1</sup> )	22.64	24.3
Available Potassium (kg ha <sup>-1</sup> )	88.92	102.7
Exch. Ca (c mol kg <sup>-1</sup> )	1.42	1.01
Exch. Mg (c mol kg <sup>-1</sup> )	0.45	0.32
Available S (kg ha <sup>-1</sup> )	12.21	10.64
DTPA Fe (mg kg <sup>-1</sup> )	22.60	16.59
DTPA Mn (mg kg <sup>-1</sup> )	8.64	6.80
DTPA Zn (mg kg <sup>-1</sup> )	0.48	0.42
DTPA Cu (mg kg <sup>-1</sup> )	0.84	0.80
<b>Fractions of Zn (mg kg<sup>-1</sup>)</b>		
Ws+Ex-Zn	0.31	0.30
OB-Zn	1.82	0.60
MnO-Zn	0.80	1.01
Am-FeO-Zn	14.10	12.69
CryFeO-Zn	28.94	24.99
Res-Zn	43.63	43.18
Total Zn	89.60	82.50



**Fig. 2 Weather data during the cropping period (October,2007 to September ,2008)**

### 3.2.5. Soil Profile Study

A vertical section of the soil adjacent to the experimental site has been taken upto a depth of 2m and studied the morphological characteristics and physico-chemical properties of the soil (Plate. 2).

### 3.2.6. Soil sample collection for initial analysis of soil

Composite samples of Surface soil (0-15 cm) and sub surface soil (15-30 cm) were collected from the experimental field before the layout of the crop and analyzed for various physico – chemical parameters and nutrient elements.

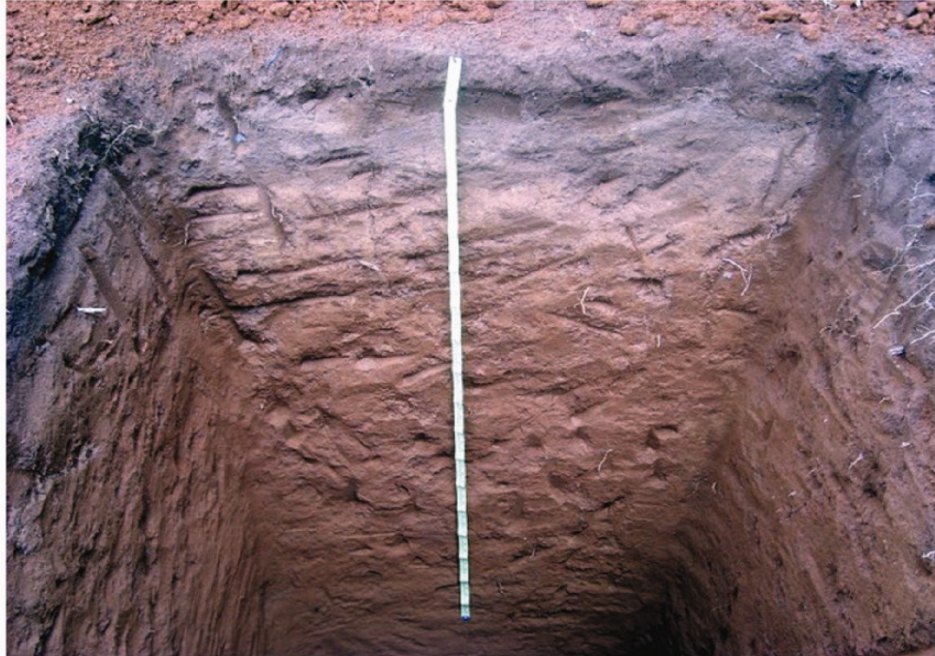
### 3.2.7. Design and Layout of the experiment

Design : Randomized Block Design  
 Plot size : 2.1 m x 2.1 m  
 Spacing : 30 x 15 cm  
 Treatments : 10  
 Variety : African Tall variety of fodder maize  
 Replications : 3

### 3.2.8. Treatments

T1 : NPK @ 120:60:40 kg ha<sup>-1</sup> without FYM  
 T2 : NPK@120:60:40kg ha<sup>-1</sup> +FYM @ 10 t ha<sup>-1</sup> (POP recommendation)  
 T3 : POP + 5 kg Zn ha<sup>-1</sup> as Zn SO<sub>4</sub> (Soil application)  
 T4 : POP + 10 kg Zn ha<sup>-1</sup> as Zn SO<sub>4</sub> (Soil application)  
 T5 : POP + 15 kg Zn ha<sup>-1</sup> as Zn SO<sub>4</sub> (Soil application)  
 T6 : POP + 20 kg Zn ha<sup>-1</sup> as Zn SO<sub>4</sub> (Soil application)  
 T7 : POP + 0.25% Zn as Zn SO<sub>4</sub> (Foliar application)  
 T8 : POP + 0.5% Zn as Zn SO<sub>4</sub> (Foliar application)  
 T9 : POP + 0.75% Zn as Zn SO<sub>4</sub> (Foliar application)  
 T10 : Control

Fig.3. Layout of the field study



**Plate 2. Soil profile**

View of Experimental and residual crops given in Plate 3 & 4.

### **3.2.9. After cultivation practices**

#### **3.2.9.1. Irrigation**

During summer months the field was irrigated twice a week and good drainage was provided to prevent water logging during rainy days.

#### **3.2.9.2 Weed control**

Before sowing, the entire field was cleared and made weed free using spade. During the crop growth period, hand weeding was resorted to as and when required.

#### **3.2.9.3 Fertilizer application**

FYM@ 10t ha<sup>-1</sup> was applied to the plots as per the treatments. Urea, rock phosphate and MOP were given @ 120:60:40 NPK kg ha<sup>-1</sup> ten days after the application of FYM. Soil application of ZnSO<sub>4</sub> .7H<sub>2</sub>O was given as per the treatments. Foliar spray of ZnSO<sub>4</sub> .7H<sub>2</sub>O was given to the treatment plots of T<sub>7</sub>, T<sub>8</sub> and T<sub>9</sub> on the 4<sup>th</sup> leaf stage.

#### **3.2.9.4 Sampling details of foliar diagnosis**

The 3<sup>rd</sup>, 4<sup>th</sup> and 5<sup>th</sup> leaf and petiole from the tip were collected on the 15<sup>th</sup>, 25<sup>th</sup>, 35<sup>th</sup> and 45<sup>th</sup> day after sowing and analyzed for Zn content for indexing of plant part.

### **3.2.10. Growth characters**

Five plants were selected from the middle row, recorded the growth parameters and the mean values were taken and tabulated.

Replications

R <sub>1</sub>	R <sub>2</sub>	R <sub>3</sub>
T <sub>7</sub>	T <sub>5</sub>	T <sub>8</sub>
T <sub>2</sub>	T <sub>7</sub>	T <sub>4</sub>
T <sub>5</sub>	T <sub>10</sub>	T <sub>7</sub>
T <sub>8</sub>	T <sub>9</sub>	T <sub>10</sub>
T <sub>9</sub>	T <sub>3</sub>	T <sub>9</sub>
T <sub>10</sub>	T <sub>2</sub>	T <sub>3</sub>
T <sub>3</sub>	T <sub>7</sub>	T <sub>5</sub>
T <sub>4</sub>	T <sub>8</sub>	T <sub>6</sub>
T <sub>1</sub>	T <sub>6</sub>	T <sub>2</sub>
T <sub>6</sub>	T <sub>4</sub>	T <sub>7</sub>

**Fig.3 Layout of the field study**



**Plate 3. Field study-Experimental crop**



**Plate 4. Field study- residual crop**



### **3.2.10.1 Plant Height (cm)**

Height of the plant was measured from the base of the plant to the topmost leaf bud on the 15<sup>th</sup>, 25<sup>th</sup>, 35<sup>th</sup> and 45<sup>th</sup> day after sowing and at the time of harvest and recorded.

### **3.2.10.2 Number of leaves**

Total numbers of leaves were recorded on the 15<sup>th</sup>, 25<sup>th</sup>, 35<sup>th</sup> and 45<sup>th</sup> day after sowing and at the time of harvest and mean number of leaves per plant were recorded.

### **3.2.10.3. Leaf Area Index (LAI)**

Leaf Area Index was computed from the values of the total leaf area of a plant and the geographical area occupied by it using the formulae

$$\text{LAI} = \frac{\text{Total leaf area of a plant}}{\text{Geographical area occupied by it}}$$

### **3.2.10.4 Leaf Stem Ratio**

Leaf of observational plants were cut at the base. The leaves and stem were separated and oven dried for three days till constant weight was obtained. The dry weight of leaves and stem of individual plants were recorded and the ratio computed by dividing leaf dry weight by stem dry weight.

## **3.2.11. Yield and Quality attributes**

### **3.2.11.1. Green fodder yield**

The green fodder yield of fodder maize was recorded per plot and calculated and presented as q ha<sup>-1</sup>.

### **3.2.11.2. Dry matter yield (q ha<sup>-1</sup>)**

Dry matter yield was calculated as qha<sup>-1</sup>

### **3.2.11.3. NPK content (%)**

NPK contents were determined according to standard procedures and calculated.

### **3.2.11.4. Crude protein (%) (CP)**

CP was calculated by multiplying the percentage content of N by a factor 6.25 (Strepson et al, 1965).

### **3.2.11.5. Crude Protein Yield (kg ha<sup>-1</sup>)**

CPY was calculated by multiplying CP content by ha<sup>-1</sup> dry matter production and expressed in kg ha<sup>-1</sup>.

### **3.2.11.6. Crude fibre content (%)**

The dried plant samples collected at the time of harvest were utilized for the estimation of crude fibre by gravimetric method (Sadasivam and Manickam, 1992).

### **3.2.11.7. Per day green fodder yield (q ha<sup>-1</sup>)**

Total green fodder yield divided by duration of the crop gave the Per day green fodder yield in q ha<sup>-1</sup>.

## **3.3. Soil Sample, Collection after the crop growth and Processing of soil**

The soil samples were collected from the individual plots at two depths viz, surface (0-15 cm) and subsurface (15-30cm). The samples were air dried, processed

and passed through a 2mm sieve and stored in polythene bags for analysis. The soil samples were analyzed for various physico-chemical parameters following the standard analytical procedures as detailed below (Table-2).

Table.2. Standard analytical procedures followed in soil analysis

Sl. No.	Parameters	Methods	Reference:
<b>I. Physical Analysis</b>			
1.	Mechanical Analysis	International Pipette Method	Piper (1966)
2.	Bulk density	Core sampling Method	Gupta and Dakshinamurthy (1980)
3.	Particle density	Pycnometer method	Gupta and Dakshinamurthy (1980)
4.	Field capacity	Core sampling Method	Black (1965)
5.	Water holding capacity	Core sampling method	Black (1965)
<b>II. Chemical Analysis</b>			
1.	pH	Direct reading using pH meter in 1:2.5 soil : water suspension	Jackson (1973)
2.	EC	Direct reading using Conductivity meter	Jackson (1973)
3.	CEC	Ammonium saturation by IN Neutral Normal Ammonium Acetate	Jackson (1973)
4.	Organic Carbon	Walkley and Black rapid titration method	Walkley and Black (1934)
5.	Available N	Alkaline Permanganate method	Subbaiah and Asija (1956)
6.	Available P	Bray No.1 Extraction and Spectrophotometry	Jackson (1973)
7.	Available K	NN Ammonium acetate extraction and flame photometry	Jackson (1973)
8.	Exch. Ca and Mg	Versenate Titration method	Hesse (1971)
9.	Available S	Turbidimetry – Barium chloride method	Chesnin and Yien (1950)
10.	Available Micronutrients	DTPA extraction and AAS	Lindsay and Norvell (1978)
11.	Fractions of Zn	Extraction using appropriate reagents and AAS	Shuman (1985)

### 3.4. Plant sample collection and processing

Plant samples collected at harvest were washed free of soil and dust particles and were dried to constant weight at 60-70<sup>0</sup> C in hot air oven. The dried plant samples were powdered in a Willey mill and wrapped in clean butter paper cover and stored for analysis. The plant samples were analyzed for their nutrient contents. The methods employed in the analyses are given in the table-3.

Table.3. Standard analytical methods followed in plant analysis

Sl. No.	Parameters	Methods	Reference:
1.	N	Microkjeldahl digestion with H <sub>2</sub> SO <sub>4</sub> and distillation	Jackson (1973)
2.	P	Diacid digestion using Nitric acid – Perchloric acid (9:4) and colorimetry using vanado molybdate yellow colour method	Jackson (1973)
3.	K	Diacid digestion using Nitric acid – Perchloric acid (9:4) and flame photometry	Jackson (1973)
4.	Ca and Mg	Diacid digestion using Nitric acid – Perchloric acid (9:4) and versenate titration	Piper (1966)
5.	S	Turbidimetry – Barium chloride method	Chesnin and Yien (1950)
6.	Fe, Mn, Zn and Cu	Diacid digestion and Atomic Absorption Spectrophotometry	Lindsay and Norvel (1978)

### **3.4.1. Nutrient uptake**

Uptake of each nutrient by the crop was computed from the dry matter yield and the per cent plant nutrient content.

## **3.5 Foliar diagnosis**

### **3.5.1. Indexing of plant part for fodder maize for zinc**

For identifying the best part and stage for sampling for Zn content, destructive sampling was done on the 15<sup>th</sup>, 25<sup>th</sup>, 35<sup>th</sup> and 45<sup>th</sup> DAS. The leaf and petiole samples from the 3<sup>rd</sup>, 4<sup>th</sup> and 5<sup>th</sup> from the tip were sampled and separated. The samples were dried, powdered and analyzed for Zn content. Correlations were worked out between the Zn content in each part at each stage to yield. The part which showed the maximum correlation as evidenced by the highest coefficient of correlation was selected as the best part and its stage of sampling as the best stage for sampling in fodder maize.

### **3.5.2. Critical level of Zn in the fodder maize for maximum yield**

The scatter diagram technique developed by Cate and Nelson (1965) was adopted to determine the critical Zn concentration graphically. Yield was plotted against Zn concentrations. The points were distributed into four quadrants. Parallel lines were drawn each to X and Y axis. The intersecting points of the line drawn parallel to Y-axis on X-axis were taken as the critical level of leaf Zn for maximum yield.

## **3.6. Economic analysis**

The economics of fertilizer application has been worked out for fodder maize under various treatments, using the cost of cultivation and BC ratio calculated.

### **3.7. Statistical Analysis**

The data from the experiments were subjected to various statistical analyses. The data collected from the incubation study and field experiments were analyzed using analysis of variance techniques (ANOVA) in CRD and RBD. Wherever significant differences between treatments were detected through ANOVA critical differences (CD) are provided for effective comparison of treatments. Correlations with different factors were also done. Path analysis was adopted to study the direct and indirect effects of different fractions of Zn on yield (Singh and Choudhary, 1985).

## 4. RESULTS

The increasing crop requirements and use of pure forms of high analysis fertilizers have brought micronutrients to the fore front of soil fertility management. Though the total reserve of micronutrients in Kerala soils are considered high, available nutrient status is often very low. Among the micronutrients, Zn deficiency is found widespread in Kerala Soils. Zn is essential for the normal healthy growth and reproduction of plants and animals and when the supply of plant available Zn is inadequate, crop yields are reduced and the quality of crop products is frequently impaired.

In this context, a research programme was initiated to find out the optimum requirement of Zn for fodder maize as evidenced by its yield and quality when supplied to soil as well as foliar application, to observe the pattern of periodic release of Zn from Zn SO<sub>4</sub> and to identify the index plant part for foliar diagnosis in fodder maize cv. African Tall. The results of the study are presented below.

### 4.1. Incubation study

The incubation experiment was conducted to study the nutrient release pattern of Zn from zinc sulphate under the laboratory conditions. The treatments involved 5 levels of Zn varying from 0-20 kg ha<sup>-1</sup> with and without FYM. The level of FYM applied was 10 t ha<sup>-1</sup>. The periodic release of DTPA extractable Zn from ZnSO<sub>4</sub>.7H<sub>2</sub>O is given in Table-4.

Table. 4. Effect of treatments and incubation periods on the availability of Zn ( $\text{mg kg}^{-1}$ )

Treatments	Days of incubation					
	1	15	30	45	60	75
T <sub>1</sub>	0.55	0.77	0.32	0.70	0.64	0.41
T <sub>2</sub>	0.64	0.89	0.52	0.90	0.87	0.79
T <sub>3</sub>	0.79	1.02	0.93	1.00	0.70	0.80
T <sub>4</sub>	1.27	1.73	1.79	0.96	1.29	0.86
T <sub>5</sub>	1.74	1.17	0.58	1.27	1.56	1.28
T <sub>6</sub>	1.33	1.24	1.59	1.76	2.00	0.81
T <sub>7</sub>	1.41	1.44	0.65	1.37	1.74	0.85
T <sub>8</sub>	1.44	1.43	1.24	1.11	1.03	1.94
T <sub>9</sub>	1.06	1.26	1.12	1.13	1.95	2.68
T <sub>10</sub>	1.47	1.07	1.80	1.37	1.78	1.75
CD	0.27	0.30	0.25	NS	0.17	0.29

On the first day of incubation the highest mean value of Zn  $1.74\text{mg kg}^{-1}$  was recorded in T<sub>5</sub> (ie. application of Zn @  $10\text{ kg ha}^{-1}$ ) and it was on par with T<sub>10</sub>. T<sub>1</sub>, the control plot released the lowest Zn content.

As the incubation completes 15 days, the highest mean value of Zn  $1.73\text{ mgkg}^{-1}$  was released from T<sub>4</sub> (Zn  $5\text{ kg ha}^{-1}$  + FYM) and it was on par with T<sub>8</sub>, The lowest was from T<sub>1</sub>.

The highest mean value of Zn ( $1.80\text{ mg kg}^{-1}$ ) was recorded in T<sub>10</sub> on the 30<sup>th</sup> day of incubation study. But this was closely followed by T<sub>4</sub> (Zn  $5\text{ kg ha}^{-1}$  + FYM). While the lowest was still observed in the control plot.



On the 45<sup>th</sup> day of incubation no significant variation in Zn release was observed among the different treatments.

As the incubation entered on the 60<sup>th</sup> day, the treatment that received Zn@ 10 kg ha<sup>-1</sup> + FYM (T<sub>6</sub>) gave the highest value of 2.00 mg kg<sup>-1</sup> Zn release and this was on par with T<sub>9</sub> ie, 20 kg Zn applied plot without FYM.

On the 75<sup>th</sup> day of incubation, the highest value of 2.68 mg kg<sup>-1</sup> was recorded in T<sub>9</sub> ie, application of Zn @ 20 kg ha<sup>-1</sup> without FYM, while the lowest value was recorded in the control plot.

#### **4.2. Soil Profile Study**

A soil profile has been taken adjacent to the experimental site in Instructional Farm, College of Agriculture, Vellayani, to study the morphological as well as physico-chemical characteristics of the soil.

The representative soil profile was taken up to 2 m depth. Various horizons were identified for the morphological descriptions of the field. Soil samples were collected from the profile based on approved guide lines. Soil colour was recorded under the field moisture conditions using Munsell Soil Colour Chart (Soil survey Staff, USDA, 1998). Soils were tentatively classified in the field itself as per keys to Soil Taxonomy (Soil survey Staff, USDA, 1998)

The soil samples from the different horizons of the soil profile were collected in polythene bags and labelled. They were analysed for physico-chemical and chemical parameters according to standard procedures.

The general description of the soil profile site is given below.

**Soil Profile – site descriptions**

Location	: Instructional Farm, College of Agriculture, Vellayani
Latitude	: 8 <sup>0</sup> 35' N; Longitude : 70 <sup>0</sup> 59'E
Altitude	: 25 m above MSL
Village	: Kalliyoor
Tehsil	: Nemom
District	: Thiruvananthapuram
State	: Kerala
Local name	: Red loam (Vellayani Series)
USDA	: Clayey, Isohyperthermic, Typic Kandiusults)
Physiographic unit	: Mid slope
Geology	: Charnockite
Parent	: Weathered charnockite
Topography	: Slopy
Slope:East West	: Gradient : 1-3%; Length:0-50 m
Erosion	: Moderate
Drainage	: Prominent
Ground water depth	: >10.0 (m)
Flooding	: Nil
Salt Alkali	: Nil
p <sup>H</sup>	: 4.5 – 5.5
EC	: <2.0 d Sm <sup>-1</sup>
Stone size	: <2.5 cm
Stoniness	: <3.0 %
Rockout crops	: Nil
Natural vegetation	: Cultivated
Crops	: Fodder
Land use	: Coconut based mixed farming

#### 4.2.1. Morphological description of soil profile

The morphological characteristics of soil profile are presented in Table. 5

Table. 5 Morphological characteristics of soil profile

Horizon	Depth(cm)	Description
Ap	0-9.5	5 YR 3/4 , sandy loam, few (<1) non-sticky fine to very fine (0-1) non-plastic, crumb massive, Dry friable, many fine pores, Argillian cutans present, fine-medium nodules, fine-to coarse roots, abrupt wavy boundary.
Ac	9.5-25	5 YR 3/4Sandy loam, medium angular blocky, non-sticky, non-plastic medium nodules and gradual smooth boundary.
B <sub>1</sub>	25-52	5 YR 4/8, sandy loam, fine to medium, granular, non-sticky, non-plastic friable few pores, gradual smooth boundary few fine roots.
B <sub>2</sub>	52-98	5 YR 4/8, sandy loam, fine to medium, pores granular, non-sticky, non-plastic, friable fine to medium roots, sub angular blocky gradual smooth boundary.
Bt <sub>2</sub>	98-123	5 YR 4/8, sandy clay, fine to medium pores, slightly sticky slightly plastic, friable, many roots, subangular blocky, gradual smooth boundary.
C <sub>1</sub>	123-150+	5 YR 4/8, sandy clay, fine to medium, pores, granular, sub angular blocky, slightly sticky, slightly plastic fine to medium roots

Six horizons viz. Ap 0-9.5 cm; Ac 9.5-25 cm; B, 25-52 cm; B2 52-98 cm Bt2 98-123 cm; and C, 123-150 + cm were identified.

The upper most 2 horizons viz. Ap and Ac had a soil colour of 5 YR 3/4, but as the depth increases the colour varied to 5 YR 4/8 till the C<sub>1</sub>, layer. The morphological examination revealed the presence of clay at a depth of 98 cm and below. The presence of clay in these horizons made the consistency slightly sticky and slightly plastic in these horizons.

The view of the soil profile taken is presented in Fig.

#### 4.2.2. Physico-chemical parameters of the soil profile

Soil samples were taken from each horizon and analyzed for various physico-chemical properties. The analytical Data is presented in Table-6

Table. 6. Physico- chemical parameters and nutrient content of the soil profile

Depth(cm)	Ap (0-95)	Ac (95-25)	B1 (25-52)	B2 (52-98)	Bt <sub>2</sub> (98-123)	C <sub>1</sub> (123-150 +)
Sand %	79.40	68.80	65.40	66.70	48.90	51.60
Silt %	11.5	16.20	15.90	14.50	10.70	9.01
Clay %	11.6	15.00	18.70	18.80	34.52	36.20
Particle density Mgm <sup>-3</sup>	2.32	2.28	2.26	2.24	2.31	2.30
Bulk density Mg m <sup>-3</sup>	1.26	1.24	1.3	1.26	1.27	1.35
Texture	SL	SL	SL	SL	SCL	SCL
pH	5.40	5.30	5.10	4.90	4.70	4.80
EC(d Sm <sup>-1</sup> )	0.12	0.15	0.13	0.12	0.12	0.12
Organic Carbon(%)	0.79	0.68	0.61	0.53	0.43	0.35
Avail. N (kg ha <sup>-1</sup> )	297.11	257.32	261.5	228.4	196.5	164.67
Avail. P (kg ha <sup>-1</sup> )	22.40	18.65	15.33	11.47	9.65	5.34
Avail. K (kg ha <sup>-1</sup> )	84.50	76.70	73.21	76.5	82.85	56.44
Exch. Ca (c mol kg <sup>-1</sup> )	1.42	0.71	0.69	0.62	0.66	0.67
Exch.Mg (c mol kg <sup>-1</sup> )	0.42	0.36	0.32	0.30	0.30	0.28
Avail. S (kg ha <sup>-1</sup> )	11.25	9.42	6.78	6.13	6.03	5.48
DTPA Fe (mg kg <sup>-1</sup> )	20.34	22.36	18.34	27.64	21.59	20.76
DTPA Mn (mg kg <sup>-1</sup> )	9.70	10.20	11.30	14.50	16.80	18.94
DTPA Cu (mg kg <sup>-1</sup> )	0.89	0.92	0.93	0.85	0.88	0.90
DTPA Zn (mg kg <sup>-1</sup> )	0.56	0.52	0.46	0.34	0.57	0.55
Fractions of Zn in the Profile (mg kg <sup>-1</sup> )						
Ws+Ex. - Zn	0.47	0.32	0.05	0.15	0.07	0.01
OB-Zn	1.50	1.30	1.27	1.18	1.15	1.12
MnO-Zn	1.30	2.10	2.00	0.70	0.50	0.23
Am-Fe-O-Zn	23.50	17.40	13.40	13.50	11.90	10.60
Cry-Fe-O-Zn	34.20	24.00	23.10	16.95	14.20	19.60
Res-Zn	24.03	35.88	37.18	39.52	38.18	30.44
Total Zn	85.00	81.00	77.00	72.00	66.00	62.00

The sand percentage decreased from 79.40 in the Ap horizon to 51.60 percentage in the C<sub>1</sub> horizon. The silt content varied from 16.20 (Ac) per cent in the upper horizon to 9.01 per cent in the C<sub>1</sub>. The clay content recorded a steady increase from 11.60 per cent in the Ap horizon to 36.20 per cent in the C<sub>1</sub> horizon and the textural class was sandy loam for the upper horizons and sandy clay loam for the lower horizons. The pH decreased as depth increased and the EC recorded a steady rate.

#### **4.2.3. Nutrient content of soil profile**

The data on the Table. 5 revealed that the Organic Carbon and other nutrients viz, Avail. N, P, K, Ca, Mg, and S decreased from Ap to C<sub>1</sub> horizon. But the available micronutrients showed a different pattern of distribution. The Mn content showed a steady increase of 9.7 to 18.94 mg kg<sup>-1</sup> from the upper to lower horizon while Zn was decreased from Ap to B<sub>2</sub> horizon.

Various fractions of the Zn were estimated as Ws+Ex.-Zn, OB- Zn, MnO-Zn, AmFeO- Zn , Cry FeO-Zn , Res--Zn and Total Zn were estimated from different profile samples. All the fractions except MnO- Zn and Res-Zn showed a decrease from upper to lower horizons. The Ws+Ex.-Zn was maximum in the Ap horizon with 0.47 mgkg<sup>-1</sup> followed by Ac with 0.32 mg kg<sup>-1</sup>. The highest values for OB-Zn was 1.5 mg kg<sup>-1</sup>. The highest values for FeO bound fractions were 23.50 mg kg<sup>-1</sup> (AmFeO-Zn) and 34.20 mg kg<sup>-1</sup>(Cry FeO-Zn) respectively. The RS-Zn was maximum in B<sub>1</sub> horizon. The highest value for total Zn was recorded in Ap horizon. The MnO-Zn was 1.30 mg kg<sup>-1</sup> in the Ap horizon and increased to 2.10 mg kg<sup>-1</sup> in the Ac horizon, thereafter from B<sub>1</sub> to C<sub>1</sub> it showed a steady decrease in content and the C<sub>1</sub> horizon recorded only 0.23 mg kg<sup>-1</sup>.

#### **4.3. Initial soil analysis of the experimental site**

Surface (0-15 cm) and subsurface (15-30cm) composite soil samples were collected from the experimental field before crop growth. Soil samples were

collected and analysed for different physical and chemical parameters and the results are given in Table 6.

Surface and sub-surface samples showed that the experimental area comes under sandy loam texture. The different physico-chemical parameters were decreased in the subsurface soil, except for available P and K. The available P and K were higher in the subsurface soil with the values of 24.3 and 102.7 kg ha<sup>-1</sup> respectively than that of 22.64 and 88.92 kg ha<sup>-1</sup> of surface soil.

The various Zn fractions were analyzed as Ws+Ex-Zn, OB-Zn, MnO-Zn, AmFeO-Zn, Cry FeO-Zn, Res-Zn and Total Zn. All the fractions, except MnO bound Zn fraction, were recorded a higher value in the surface soil. But the MnO bound fraction of Zn showed a higher value in the subsurface soil (1.01 mg kg<sup>-1</sup>) than that of the lower content of 0.80 mg kg<sup>-1</sup> in the surface soil.

#### **4.4. Field study**

The results of the two experimental crops raised as per the treatments outlined in the earlier and the residual crops that have been taken after each experimental crop are presented below.

##### **4.4.1. First field crop**

###### **4.4.1.1. Biometric observations**

Different Biometric observations viz. Height of the plant, Number of leaves plant<sup>-1</sup> etc. were recorded, on the 15<sup>th</sup>, 25<sup>th</sup>, 35<sup>th</sup>, 45<sup>th</sup> Days after sowing and at harvest.

The height in (cm) of fodder maize at different stages of growth is given in Table. 7.

Table 7. Height (cm) of fodder maize at different stages of growth – first crop

Treatments	Stages of sampling				
	15DAS	25DAS	35DAS	45DAS	HVST
T1	13.70	33.37	54.33	78.70	163.73
T2	13.23	21.30	31.70	62.23	129.02
T3	14.00	27.63	46.73	67.86	138.69
T4	16.73	41.80	59.67	87.76	180.90
T5	15.53	32.53	49.60	90.55	152.17
T <sub>6</sub>	13.53	27.28	46.37	62.70	158.48
T <sub>7</sub>	12.47	30.73	47.00	63.76	149.42
T <sub>8</sub>	13.97	35.37	51.37	72.66	151.97
T <sub>9</sub>	13.90	29.53	47.56	67.36	156.50
T <sub>10</sub>	9.53	17.97	24.56	33.46	56.13
CD	NS	3.486	10.588	16.836	22.994

The treatment means for height of the plants were not significant at 15DAS. At 25 DAS, the highest value of 41.80cm was noticed in T<sub>4</sub> and it was found superior to all other values. The lowest value was recorded in T<sub>10</sub> (17.97cm). At 35DAS the highest value was again recorded in T<sub>4</sub> (59.67cm) and T<sub>1</sub>, T<sub>8</sub> and T<sub>5</sub> were found to be on par with values of 54.33, 51.37 and 49.6cm respectively. T<sub>10</sub> recorded the lowest value of 24.56 cm. In the case of 45DAS, the highest and lowest values were T<sub>5</sub> (90.55cm) and T<sub>10</sub> (33.46cm). The treatment means of T<sub>4</sub> (87.76cm) and T<sub>1</sub> (78.7cm) were on par to T<sub>5</sub>. At the time of harvest, the highest and lowest values were found to be 180.90 cm (T<sub>4</sub>) and 56.13 cm for T<sub>10</sub>. T<sub>1</sub> and T<sub>6</sub> were found on par with T<sub>4</sub>.

The mean number of leaves recorded per plant on the 15<sup>th</sup>, 25<sup>th</sup>, 35<sup>th</sup>, 45<sup>th</sup> DAS and at the time of harvest are given in Table. 8.

Table 8 Mean number of leaves plant<sup>-1</sup> of fodder maize at different stages of growth – first crop.

Treatments	Number of leaves				
	15DAS	25DAS	35DAS	45DAS	HVST
T <sub>1</sub>	4.03	7.10	7.80	10.83	14.44
T <sub>2</sub>	4.65	6.72	7.40	11.53	12.86
T <sub>3</sub>	5.23	7.70	8.30	10.53	13.76
T <sub>4</sub>	4.41	6.50	7.73	11.07	12.72
T <sub>5</sub>	4.94	6.83	7.73	10.17	13.83
T <sub>6</sub>	4.30	7.23	7.54	11.10	13.90
T <sub>7</sub>	4.53	6.75	7.82	9.93	14.07
T <sub>8</sub>	4.23	7.23	8.26	10.37	13.2
T <sub>9</sub>	4.43	7.23	8.14	9.63	15.13
T <sub>10</sub>	3.44	4.79	5.17	8.37	9.37
CD	0.705	0.673	0.847	NS	2.846

The highest and lowest mean values (Table. 8) for the number of leaves at 15DAS were recorded as 5.23 (T<sub>3</sub>) and 3.44 (T<sub>10</sub>). T<sub>5</sub>, T<sub>2</sub> and T<sub>7</sub> were found on par with T<sub>3</sub>. For 25 DAS, the maximum number of leaves were noticed in T<sub>3</sub> (7.70) and the treatments T<sub>1</sub>, T<sub>6</sub>, T<sub>8</sub> and T<sub>9</sub> were on par with T<sub>3</sub>. The highest and lowest mean values obtained for 35DAS were 8.3 and 5.17 in T<sub>3</sub> and T<sub>10</sub> respectively, while the other treatment means except T<sub>2</sub> and T<sub>10</sub> were on par. At harvest, the maximum number of leaves was observed in T<sub>9</sub> (15.13) and the lowest in T<sub>10</sub> (9.37), all other values were found to be on par with T<sub>9</sub>.

#### 4.4.1.2. Yield and growth attributes of fodder maize

The various yields and growth attributes viz. green fodder yield, dry fodder yield, dry matter percentage, per day green fodder yield, leaf area index, Leaf stem ratio etc. are recorded in Table. 9.



Table.9. Yield and growth attributes of fodder maize of first crop

Treatments	Green fodder (qha <sup>-1</sup> )	Dry fodder (qha <sup>-1</sup> )	DM (%)	PDGFY (qha <sup>-1</sup> )	LAI	LSR
T <sub>1</sub>	408.10	83.02	20.33	6.79	10.63	2.54
T <sub>2</sub>	318.30	65.15	20.40	5.30	9.73	1.58
T <sub>3</sub>	457.10	89.76	19.63	7.62	11.62	2.61
T <sub>4</sub>	512.50	103.92	20.30	8.54	12.97	2.63
T <sub>5</sub>	418.40	81.81	19.57	6.97	11.09	2.58
T <sub>6</sub>	301.40	59.71	19.83	5.02	9.69	1.58
T <sub>7</sub>	366.10	72.86	19.90	6.10	7.51	2.40
T <sub>8</sub>	389.30	77.02	19.77	6.48	9.33	2.29
T <sub>9</sub>	395.10	77.77	19.63	6.58	10.09	2.48
T <sub>10</sub>	66.40	12.61	19.00	1.10	7.17	1.48
CD	21.31	5.56	NS	0.37	NS	0.47

As seen from the Table.9, the yield and growth attributes were significant for various treatments. The highest GFY and DFY were recorded for T<sub>4</sub>, i.e. application of Zn at 10 kg ha<sup>-1</sup> in addition to POP recommendation and was found superior to all other treatments. The yield T<sub>4</sub> treatment recorded a green fodder of (512.5 q ha<sup>-1</sup>) and dry fodder yield of 103.92 q ha<sup>-1</sup>. The lowest value was recorded in T<sub>10</sub> with 66.4 q ha<sup>-1</sup> and 12.61 q ha<sup>-1</sup> respectively for GF and DF yield.

The per day green fodder yield (PDGFY) recorded the highest value of 8.54 q ha<sup>-1</sup> and lowest value of 1.10 q ha<sup>-1</sup> respectively for T<sub>4</sub> and T<sub>10</sub>. T<sub>4</sub> was found superior to all. The treatment means for LAI was not found significant. Even though not significant, the maximum LAI of (12.97) obtained treatment i.e., T<sub>4</sub> recorded the maximum green fodder yield.

The highest LSR was noted in T<sub>4</sub> (2.63) and the lowest value of (1.48) in T<sub>10</sub>. T<sub>4</sub> was found superior to all other treatments. All other values except T<sub>2</sub>, T<sub>6</sub> and T<sub>10</sub> were found on par.

#### 4.4.1.3. Quality attributes of fodder maize of first crop

The quality attributes viz. crude protein per cent, crude protein yield and crude fibre per cent are given below.

Table. 10. Quality parameters of fodder maize – first crop

Treatments	CP (%)	CPY ( kg ha <sup>-1</sup> )	CF (%)
T <sub>1</sub>	9.89	821	23.80
T <sub>2</sub>	9.94	648	24.33
T <sub>3</sub>	10.16	911	24.40
T <sub>4</sub>	10.93	1136	23.70
T <sub>5</sub>	10.19	833	23.76
T <sub>6</sub>	9.23	551	26.63
T <sub>7</sub>	8.78	639	25.96
T <sub>8</sub>	9.00	693	25.63
T <sub>9</sub>	9.16	712	23.10
T <sub>10</sub>	6.57	216	29.40
CD	0.61	147	1.00

Regarding the quality aspects ( Table.10), maximum CP content and CPY were recorded in T<sub>4</sub> with values of 10.93 percent and 1136 kg ha<sup>-1</sup> and the lowest crude fibre was obtained in T<sub>9</sub> (23.1 percent). Even though T<sub>9</sub> showed the lowest crude fibre percentage of 23.1 per cent, it was on par with T<sub>4</sub>. The lowest CP per cent CPY and maximum CF per cent were recorded in T<sub>10</sub> with respective values of 6.57 per cent, 216 kg ha<sup>-1</sup> and 29.4 per cent respectively.

The next higher values for CP was recorded for T<sub>5</sub> (10.19 percent), T<sub>5</sub> recorded a comparable crude fibre content with T<sub>4</sub> i.e., 23.76 per cent for T<sub>5</sub> and 23.70 per cent for T<sub>4</sub>. This showed that T<sub>4</sub> (POP+Zn @10 kg ha<sup>-1</sup>) was the best treatment followed by T<sub>5</sub> (POP+Zn @10 kg ha<sup>-1</sup>).

#### 4.4.1.4. Nutrient content in fodder maize.

After the harvest of the 1<sup>st</sup> crop the plant samples collected were processed and analyzed for the nutrient concentration and the results are given in Table. 11.

Table 11 Nutrient concentration in the fodder maize –first crop

Treatments	N (%)	P (%)	K (%)	Ca (%)	Mg (%)	S (%)	Fe (mg kg <sup>-1</sup> )	Mn (mg kg <sup>-1</sup> )	Zn (mg kg <sup>-1</sup> )	Cu (mg kg <sup>-1</sup> )
T <sub>1</sub>	1.58	0.14	1.33	0.19	0.12	0.12	89.30	23.33	41.67	13.33
T <sub>2</sub>	1.59	0.17	1.56	0.22	0.12	0.13	73.00	28.00	45.67	13.00
T <sub>3</sub>	1.63	0.27	1.77	0.27	0.12	0.15	113.30	33.00	63.67	16.33
T <sub>4</sub>	1.75	0.26	2.16	0.29	0.15	0.15	116.00	35.00	68.33	17.00
T <sub>5</sub>	1.57	0.20	1.90	0.34	0.16	0.14	119.70	34.67	63.00	17.00
T <sub>6</sub>	1.48	0.15	1.86	0.26	0.13	0.17	103.70	32.33	61.33	15.00
T <sub>7</sub>	1.41	0.18	1.70	0.22	0.13	0.13	89.00	30.67	49.33	14.33
T <sub>8</sub>	1.44	0.19	1.76	0.24	0.12	0.13	84.00	29.67	52.33	13.65
T <sub>9</sub>	1.47	0.19	1.77	0.21	0.11	0.13	85.00	28.00	56.00	16.00
T <sub>10</sub>	1.05	0.10	0.99	0.10	0.07	0.11	90.70	17.67	19.00	11.67
CD	0.064	0.031	0.99	0.024	0.02	0.01	9.68	3.748	4.006	1.805

Data presented in the Table .11 revealed that the nutrient concentrations in fodder maize for different treatments were found significant. The highest value for N was obtained in T<sub>4</sub> with 1.75 per cent and it was found superior to all other treatment mean values, while the lowest was recorded in T<sub>10</sub> (1.05 per cent). For the P content, maximum mean value of 0.27 percent was observed in T<sub>3</sub> and T<sub>4</sub> (0.26 per cent) was found to be on par with T<sub>3</sub>. The lowest was noted in T<sub>10</sub> with a men value of 0.10 per cent. The highest value of 2.16 per cent K was observed in T<sub>4</sub> and all other values were on par except T<sub>10</sub> (0.99 per cent) and T<sub>1</sub> (1.23 per cent).

Maximum content of Ca was 0.34 per cent in T<sub>5</sub> while the lowest value was observed in T<sub>10</sub>. For Mg content, T<sub>5</sub> recorded the highest value of 0.16 per cent and T<sub>4</sub> (0.15 per cent) was on par while lowest value was noticed in T<sub>10</sub> (0.07 per cent). The highest S concentration was 0.17 per cent in T<sub>6</sub> which was found to be the superior to all other treatment values. T<sub>10</sub> recorded the lowest (0.11 per cent) value.

The Fe content was highest in T<sub>5</sub> (119.7 mg kg<sup>-1</sup>) and T<sub>4</sub> (116.0 mg kg<sup>-1</sup>) and T<sub>3</sub> (113.3 mg kg<sup>-1</sup>) was found to be on par with T<sub>5</sub>. The lowest value was recorded in T<sub>2</sub> (73 mg kg<sup>-1</sup>). For Mn, T<sub>4</sub> (35 mg kg<sup>-1</sup>) recorded the highest mean value and lowest value of (17.67 mgkg<sup>-1</sup>) was observed in T<sub>10</sub>. T<sub>5</sub>, T<sub>3</sub> and T<sub>6</sub> were found to be on par with T<sub>5</sub> with values of 34.67, 33.00, 32.33 mg kg<sup>-1</sup> respectively.

The maximum content was observed in T<sub>4</sub> (68.33 mg kg<sup>-1</sup>), and it was found significantly different from other mean values. The lowest value was recorded in T<sub>10</sub> (19.00 mg kg<sup>-1</sup>). The highest value for Cu was observed in T<sub>4</sub> and T<sub>5</sub> with a mean value of 17.00 mgkg<sup>-1</sup>, but T<sub>3</sub> and T<sub>9</sub> were found to be on par with T<sub>4</sub> and the lowest value of 11.67 mgkg<sup>-1</sup> was given by T<sub>10</sub>.

#### 4.4.15. Nutrient uptake

Table.12. Nutrient uptake studies of the first crop (kgha<sup>-1</sup>)

Treatments	N	P	K	Ca	Mg	S	Fe	Mn	Zn	Cu
T <sub>1</sub>	131.50	11.37	113.4	15.75	9.94	9.81	0.73	0.19	0.35	0.11
T <sub>2</sub>	103.70	10.91	103.8	14.06	7.60	8.34	0.48	0.18	0.30	0.09
T <sub>3</sub>	146.00	23.80	142.2	24.26	11.05	13.88	1.02	0.28	0.57	0.14
T <sub>4</sub>	181.80	24.91	224.7	31.15	15.61	14.64	1.20	0.36	0.71	0.18
T <sub>5</sub>	128.40	16.36	155.5	30.14	13.09	11.8	0.98	0.28	0.52	0.13
T <sub>6</sub>	88.20	12.54	111.2	10.21	8.23	9.91	0.70	0.19	0.37	0.10
T <sub>7</sub>	102.40	13.09	123.6	16.26	9.71	10.89	0.65	0.21	0.36	0.10
T <sub>8</sub>	77.50	14.34	135.7	18.23	9.25	10.14	0.67	0.23	0.40	0.10
T <sub>9</sub>	114.00	14.78	137.3	15.99	8.80	10.44	0.66	0.22	0.44	0.12
T <sub>10</sub>	13.20	1.26	12.5	1.26	0.92	1.35	0.11	0.02	0.02	0.01
CD	30.78	3.407	15.97	3.063	1.962	1.568	0.125	0.025	0.043	0.021

Data in the Table.12 revealed that the uptake of different nutrients were significant. For the uptake of N, maximum value was obtained in T<sub>4</sub> (181.80 kg ha<sup>-1</sup>) which was found to be superior and the lowest value of 13.20 (kg ha<sup>-1</sup>) was observed in T<sub>10</sub>. For the P uptake, highest value was observed in T<sub>4</sub> 24.91(kg ha<sup>-1</sup>) and T<sub>3</sub> (23.80 kgha<sup>-1</sup>) was found to be on par. The lowest value of 1.26 kg ha<sup>-1</sup> was noted in T<sub>10</sub>. Maximum value for K uptake was noticed in T<sub>4</sub> 224.7(kg ha<sup>-1</sup>) and found significantly superior to all other treatments, while the lowest value (12.5 kgha<sup>-1</sup>) was noticed in T<sub>10</sub>.

For the uptake of Ca, the highest value of 31.15 kg ha<sup>-1</sup> was observed in T<sub>4</sub>, while T<sub>5</sub> (30.14 kgha<sup>-1</sup>) was found on par with T<sub>4</sub> and T<sub>10</sub> recorded the lowest value (0.92 kgha<sup>-1</sup>). For Mg uptake, T<sub>4</sub> recorded highest (15.61kgha<sup>-1</sup>) and was superior when compared with other treatment means. T<sub>10</sub> recorded the lowest value of (1.35 kgha<sup>-1</sup>). The highest value for Mg uptake was found in T<sub>4</sub> (14.64 kgha<sup>-1</sup>) and found superior to all other treatments. The lowest value was in T<sub>10</sub> (1.35kgha<sup>-1</sup>). For the S uptake, maximum value was recorded in T<sub>4</sub> (14.64 kgha<sup>-1</sup>) and it was on par with T<sub>3</sub> with a value of 13.88 kgha<sup>-1</sup>. The control plot recorded the lowest value.

The highest value of Fe uptake was noticed in T<sub>4</sub> (1.20 kgha<sup>-1</sup>) while it was on par with T<sub>3</sub> (1.02 kgha<sup>-1</sup>). For the uptake of Mn, there were significant differences among the treatments. T<sub>4</sub> recorded the highest value of 0.36 kg ha<sup>-1</sup> and was found to be superior. Maximum uptake of Zn and Cu was obtained in T<sub>4</sub> with values of 0.71 kgha<sup>-1</sup> and 0.18 kgha<sup>-1</sup> respectively and found significantly different from all other mean values. T<sub>10</sub> recorded the lowest value for all micronutrient uptake.

#### **4.4.1.6. Physico-chemical properties and primary nutrient concentration in the surface soil after the first crop**

Surface soil samples (0-15 cm) were collected from each plot after the 1<sup>st</sup> crop growth and analyzed for different physico-chemical parameters and primary nutrient elements and the data are presented in Table-13.

Table 13 Physico-chemical parameters and primary nutrients content of subsurface soil after the first crop

Treatments	pH	EC (dSm <sup>1</sup> )	Organic Carbon (%)	Av .N kg <sub>ha</sub> <sup>-1</sup>	Av. P kg <sub>ha</sub> <sup>-1</sup>	Av.K kg ha <sup>-1</sup>
T <sub>1</sub>	5.0	0.11	0.75	187.6	14.09	145.67
T <sub>2</sub>	4.8	0.14	1.06	221	12.02	133
T <sub>3</sub>	5.0	0.12	0.79	218.4	16.57	183.73
T <sub>4</sub>	4.9	0.10	1.12	224.60	13.29	204.3
T <sub>5</sub>	5.0	0.11	0.7	208.82	12.35	172.67
T <sub>6</sub>	4.9	0.11	0.82	216.60	10.83	108.93
T <sub>7</sub>	5.0	0.12	0.76	200.77	12.13	177.46
T <sub>8</sub>	5.0	0.14	0.84	214.73	12.32	179.77
T <sub>9</sub>	4.9	0.10	0.98	205.03	13.81	186.84
T <sub>10</sub>	4.8	0.12	0.58	89.4	4.95	69.00
CD	NS	NS	NS	21.66	1.661	9.132

Data presented in the Table 13 showed that treatment effects were significant except for, P<sup>H</sup>, EC and organic carbon. The highest value of available N was observed in T<sub>4</sub> with 224.60 kg ha<sup>-1</sup> which was on par with T<sub>2</sub> (221.0 kg ha<sup>-1</sup>), T<sub>3</sub> (218.4 kg ha<sup>-1</sup>) T<sub>6</sub> (216.60 kg ha<sup>-1</sup>), T<sub>8</sub> (214.73 kg ha<sup>-1</sup>), T<sub>5</sub> (208.82 kg ha<sup>-1</sup>) and T<sub>9</sub> (205.03 kg ha<sup>-1</sup>) respectively. The lowest value was noticed in T<sub>10</sub> (89.4 kg ha<sup>-1</sup>). For available P, maximum was noted in T<sub>3</sub> (16.57 kg ha<sup>-1</sup>) which was found to be superior and T<sub>10</sub> recorded the lowest (4.95 kg ha<sup>-1</sup>). The highest value of available K was 204.3 kg ha<sup>-1</sup> (T<sub>4</sub>) and was found superior to all other treatments, while the lowest value of 69.00 kg ha<sup>-1</sup> was recorded in T<sub>10</sub>.

#### 4.4.1.7. Secondary and micro nutrient content in the surface soil after the first crop

The surface soil (0-15 cm) samples collected from each plot, after the harvest of the 1<sup>st</sup> crop were analyzed for the secondary and micronutrient contents and the data is presented in Table 14.

Table -14 Secondary and micronutrient contents of subsurface soil after the first

Treatments	Ca (c mol kg <sup>-1</sup> )	Ex Mg (c mol kg <sup>-1</sup> )	Av. S (kg ha <sup>-1</sup> )	DTPA Fe (mg kg <sup>-1</sup> )	DTPA Mn (mg kg <sup>-1</sup> )	DTPA Zn (mg kg <sup>-1</sup> )	DTPA Cu (mg kg <sup>-1</sup> )
T <sub>1</sub>	1.29	0.48	9.26	18.03	3.29	2.92	0.82
T <sub>2</sub>	1.3	0.46	11.11	14.62	2.3	2.32	0.5
T <sub>3</sub>	1.28	0.51	11.88	22.37	4.53	3.69	0.61
T <sub>4</sub>	1.27	0.52	12.1	19.80	3.61	4.89	0.65
T <sub>5</sub>	1.26	0.48	12.88	18.85	3.86	4.66	0.61
T <sub>6</sub>	1.15	0.43	9.78	14	2.66	2.38	0.32
T <sub>7</sub>	1.25	0.38	11.35	18	4.06	2.72	0.46
T <sub>8</sub>	1.22	0.37	11.58	17.20	3.93	2.64	0.47
T <sub>9</sub>	1.23	0.38	11.54	18.51	4.94	2.59	0.49
T <sub>10</sub>	1.07	0.35	6.05	15.80	0.77	0.77	0.21
CD	0.070	0.034	2.106	5.03	0.544	0.194	0.08

The maximum and minimum value for Exch. Ca was in T<sub>2</sub> (1.30 c mol kg<sup>-1</sup>) and T<sub>10</sub> (1.07 c mol kg<sup>-1</sup>). While T<sub>1</sub>, T<sub>3</sub>, T<sub>4</sub>, T<sub>5</sub>, T<sub>7</sub> and T<sub>9</sub> were on par with T<sub>2</sub>. The value of 1.07 c mol kg<sup>-1</sup> was noted as the lowest (T<sub>10</sub>). The highest value of Exch. Mg was noted in T<sub>4</sub> (0.52 cmol kg<sup>-1</sup>) followed by T<sub>3</sub> (0.51 c mol kg<sup>-1</sup>) and the lowest value of was recorded in T<sub>10</sub> (0.35 c mol kg<sup>-1</sup>). The available S was maximum in T<sub>5</sub> (12.88 kg ha<sup>-1</sup>) and T<sub>4</sub> (12.1 kg ha<sup>-1</sup>), T<sub>3</sub> (11.88 kg ha<sup>-1</sup>), T<sub>8</sub> (11.58 kg ha<sup>-1</sup>), T<sub>9</sub> (11.54 kg ha<sup>-1</sup>), T<sub>7</sub> (11.35 kg ha<sup>-1</sup>), T<sub>2</sub> (11.11 kg ha<sup>-1</sup>) were found to be on par with T<sub>5</sub>. But lowest was 6.05 kg ha<sup>-1</sup> in T<sub>10</sub>.

For the DTPA extractable Fe, the highest value of 22.37 mg kg<sup>-1</sup> (T<sub>3</sub>) was superior to all other treatments except T<sub>10</sub> and T<sub>2</sub> and T<sub>6</sub>. The highest value for DTPA extractable Mn was 4.94 mg kg<sup>-1</sup> (T<sub>9</sub>) and it was found on par with T<sub>3</sub>. The highest and lowest for DTPA extractable Zn was in T<sub>4</sub> and T<sub>10</sub> and the corresponding values were 4.89 mg kg<sup>-1</sup> and 0.77 mg kg<sup>-1</sup> respectively. The highest mean value of 0.82 mg kg<sup>-1</sup> was noticed in T<sub>1</sub> for DTPA extractable Cu, while the lowest was found in T<sub>10</sub> with 0.21 mg kg<sup>-1</sup>. T<sub>1</sub> was found superior to all other values.

#### 4.4.1.8. Zn fraction of surface soil after the first crop

The surface soil samples collected from each plot were analyzed for different fractions of Zn and the data is given in Table-15.

Table 15. Fraction of Zinc ( $\text{mg kg}^{-1}$ ) in the surface soil after the first crop

Treatments	Ws+Ex.	OB Zn	Mn O Zn	Am FeO Zn	Cry FeO Zn	Res Zn	Total
T <sub>1</sub>	3.75 (3.34)	2.3 (2.05)	0.88 (0.78)	21.82 (19.48)	42.64 (38.07)	40.61 (36.28)	112
T <sub>2</sub>	1.34 (1.14)	2.92 (2.50)	0.68 (0.58)	45.00 (38.46)	37.85 (32.40)	29.21 (24.92)	117
T <sub>3</sub>	4.31 (2.89)	2.11 (1.42)	0.96 (0.64)	25.27 (16.95)	75.94 (50.96)	40.41 (27.14)	149
T <sub>4</sub>	4.36 (3.57)	2.41 (1.98)	1.01 (0.83)	27.01 (22.14)	55.61 (45.58)	31.35 (23.58)	122
T <sub>5</sub>	5.43 (4.08)	2.21 (1.66)	0.61 (0.46)	23.40 (17.59)	70.00 (52.63)	31.35 (23.58)	133
T <sub>6</sub>	1.54 (1.39)	3.84 (3.46)	1.14 (0.55)	26.14 (23.55)	24.51 (22.08)	53.83 (48.49)	111
T <sub>7</sub>	2.76 (2.67)	2.52 (2.44)	0.43 (0.42)	27.00 (26.21)	21.54 (20.91)	48.75 (47.33)	103
T <sub>8</sub>	2.87 (2.73)	2.14 (2.04)	0.44 (0.42)	40.91 (38.96)	35.38 (33.70)	23.26 (22.15)	105
T <sub>9</sub>	5.91 (4.02)	2.01 (1.37)	0.36 (0.25)	36.75 (25.00)	90.33 (61.45)	11.64 (7.92)	147
T <sub>10</sub>	0.30 (0.34)	3.38 (3.89)	0.10 (0.11)	16.64 (19.13)	43.91 (50.47)	22.67 (26.06)	87
CD	0.204	0.564	0.082	4.19	10.58	6.67	14.19

The values in parenthesis are the percentage content of the individual fractions to Total Zinc.

The Ws+Ex-Zn was maximum in T<sub>9</sub> ( $5.91 \text{ mg kg}^{-1}$ ) and it was found significantly different from other values. The lowest value was obtained in the control ( $0.3 \text{ mg kg}^{-1}$ ). The highest value for OB fraction of Zn was in T<sub>6</sub> ( $3.84 \text{ mg kg}^{-1}$ ), but it was on par with T<sub>10</sub> ( $3.38 \text{ mg kg}^{-1}$ ) and the lowest was recorded in T<sub>9</sub> ( $2.01 \text{ mg kg}^{-1}$ ). For the Am FeO bound fraction, the highest value was noted in



T<sub>2</sub>(45 mg kg<sup>-1</sup>) and it was on par with T<sub>8</sub>(40.91 mg kg<sup>-1</sup>) while the control recorded the lowest. The highest and lowest values for CryFeO content of Zn was T<sub>9</sub> and T<sub>7</sub> with mean values of 90.33 and 21.54 mg kg<sup>-1</sup> respectively and also T<sub>9</sub> was found superior. The residual Zn was highest in T<sub>6</sub>(53.83 mg kg<sup>-1</sup>) and was on par with T<sub>7</sub>. With a mean value of 48.75 mg kg<sup>-1</sup>. The highest value for total Zn was seen in T<sub>3</sub> (149 mgkg<sup>-1</sup>) and T<sub>9</sub> (147 mg kg<sup>-1</sup>) was on par with T<sub>3</sub>. The control denoted the lowest value of 87 mg kg<sup>-1</sup>.

#### 4.4.1.9 . Physico-chemical properties and primary nutrient concentration in the subsurface soil after the crop growth in first crop

Soil surface soil samples (15-30 cm) were collected from each plot after the first crop period and analyzed for different physico-chemical parameters and primary nutrient elements and the data is presented in Table-16.

Table 16. Physico-chemical properties and nutrient content in the surface soil after the first crop

Treatments	pH	EC (dSm <sup>-1</sup> )	Organic carbon (%)	Av. N kgha <sup>-1</sup>	Av .P kgha <sup>-1</sup>	Av . K kgha <sup>-1</sup>
T <sub>1</sub>	5.3	0.11	0.4	149.06	5.45	150.66
T <sub>2</sub>	5.4	0.12	0.78	108.97	5.71	192.26
T <sub>3</sub>	5.5	0.11	0.58	175.07	2.56	182.16
T <sub>4</sub>	5.3	0.11	0.91	213.43	2.78	174.53
T <sub>5</sub>	5.3	0.09	0.66	141.37	2.26	160.13
T <sub>6</sub>	5.5	0.08	0.52	191.77	5.82	165.1
T <sub>7</sub>	5.6	0.07	0.38	159.23	5.25	182.03
T <sub>8</sub>	5.5	0.11	0.62	197.83	4.27	165.5
T <sub>9</sub>	5.3	0.12	0.57	158.87	3.21	103.23
T <sub>10</sub>	5.4	0.11	0.37	75.17	1.55	57.17
CD	0.305	NS	NS	48.565	2.510	18.802

The data (Table. 16) on pH, EC and OC per cent were found to be non-significant. The highest and lowest value of available N was observed in T<sub>4</sub> (213.43 kg ha<sup>-1</sup>) and T<sub>10</sub> (75.17 kg ha<sup>-1</sup>). The values of T<sub>8</sub> (197.83 kg ha<sup>-1</sup>), T<sub>6</sub> (191.77 kg ha<sup>-1</sup>) and T<sub>3</sub> (175.07 kg ha<sup>-1</sup>) were found on par with T<sub>4</sub>. The maximum value of available P was recorded in T<sub>6</sub> with 5.82 kg ha<sup>-1</sup> and was found on par with T<sub>2</sub> (5.71 kg ha<sup>-1</sup>), T<sub>8</sub> (4.27 kg ha<sup>-1</sup>) and T<sub>1</sub>. The lowest was found in T<sub>10</sub> (1.55 kg ha<sup>-1</sup>). in T<sub>3</sub> and it was found to be superior to all other treatment means and the lowest was found in T<sub>10</sub>. The highest value of 192.26 kg ha<sup>-1</sup> for available K was found in T<sub>2</sub> and it was superior to all others.

#### 4.4.1.10. Secondary and micronutrient contents in the subsurface soil after crop growth first crop

The sub-surface (15-30 cm) soil samples collected from each plot after the harvest of the first crop were analyzed for the secondary and micronutrient content and the data is presented in Table 17.

Table. 17 Secondary and micronutrient contents of subsurface soil after the first crop

Treatments	Ex. Ca (c mol kg <sup>-1</sup> )	Ex .Mg (c mol kg <sup>-1</sup> )	Avail. S (kgha <sup>-1</sup> )	DTPA Fe (mgkg <sup>-1</sup> )	DTPA Mn (mgkg <sup>-1</sup> )	DTPA Zn (mgkg <sup>-1</sup> )	DTPA Cu (mgkg <sup>-1</sup> )
T <sub>1</sub>	0.97	0.38	11.22	15.50	2.17	2.35	0.71
T <sub>2</sub>	0.92	0.41	7.43	14.47	2.79	2.27	0.71
T <sub>3</sub>	1.07	0.35	11.52	12.30	3.24	3.22	0.56
T <sub>4</sub>	1.02	0.42	11.61	15.83	2.44	3.88	0.71
T <sub>5</sub>	1.01	0.46	11.98	15.40	3.01	3.28	0.76
T <sub>6</sub>	0.70	0.43	10.26	13.13	2.82	2.18	0.41
T <sub>7</sub>	0.89	0.39	10.33	12.60	2.71	2.33	0.39
T <sub>8</sub>	0.77	0.35	10.42	5.00	2.87	2.14	0.48
T <sub>9</sub>	0.77	0.38	11.07	8.37	3.58	2.26	0.47
T <sub>10</sub>	0.64	0.33	3.72	12.37	3.01	0.61	0.44
CD	0.128	0.037	1.767	4.759	NS	0.252	0.139

The highest and lowest value of exchangeable Ca was noticed in T<sub>3</sub> with the values of 1.07 c mol kg<sup>-1</sup> and 0.640 c mol kg<sup>-1</sup> in T<sub>10</sub> and T<sub>4</sub> and T<sub>5</sub> were found to be on par with T<sub>3</sub>. The highest mean value for exchangeable Mg was found to be in T<sub>5</sub> (0.46c mol kg<sup>-1</sup>) while it was on par with T<sub>6</sub> and T<sub>4</sub> with the respective values of 0.43 and 0.42 c mol kg<sup>-1</sup> respectively. T<sub>10</sub> (0.33 c mol kg<sup>-1</sup>) recorded the lowest. The maximum value for available S was noted in T<sub>5</sub> with a value of 11.98 kg ha<sup>-1</sup> and T<sub>4</sub>, T<sub>3</sub>, T<sub>1</sub>, T<sub>9</sub>, T<sub>8</sub>, T<sub>7</sub> and T<sub>6</sub> were found on par with T<sub>5</sub>.

The highest values of DTPA Fe was found in T<sub>4</sub> (15.83mg kg<sup>-1</sup>) and T<sub>1</sub> (15.50 mg kg<sup>-1</sup>), T<sub>5</sub>(15.40mg kg<sup>-1</sup>), T<sub>2</sub>(14.47mg kg<sup>-1</sup>), T<sub>6</sub>(13.13mg kg<sup>-1</sup>),T<sub>7</sub>(12.60mg kg<sup>-1</sup>)and T<sub>10</sub>(12.37mg kg<sup>-1</sup>) were found on par with T<sub>4</sub>. The lowest was obtained in T<sub>8</sub> (5.mg kg<sup>-1</sup>).

For the DTPA extractable Zn, the highest and lowest values were 3.30 and 0.61mg kg<sup>-1</sup> respectively for T<sub>4</sub> and T<sub>10</sub>. However the values of 3.28 (T<sub>5</sub>), 3.22(T<sub>3</sub>) and 3.00 mg kg<sup>-1</sup> in T<sub>9</sub> found to be on par. The highest value for Cu was recorded in T<sub>5</sub> (0.76 mg kg<sup>-1</sup>) and it was found on par withT<sub>1</sub>, T<sub>2</sub> and T<sub>4</sub> while the lowest was observed in T<sub>7</sub> (0.39 mgkg<sup>-1</sup>).

#### **4.4.1.11. Zn fractionation of subsurface soil after the first crop**

The sub-surface soil samples collected from each plot were analyzed for different fractions of Zn and the data is presented in Table. 18. The values in parentheses are the percentage content of the individual fraction to Total Zn.

Table. 18 Different fractions of Zn in the sub surface soil after the first crop (mg kg<sup>-1</sup>)

Treatments	WS+EX.- Zn	OB- Zn	MnO- Zn	Am Fe-Zn	Cry Fe-Zn	Res- Zn	Total- Zn
T1	0.72 (2.23)	0.61 (2.25)	0.92 (0.98)	26.4 (13.58)	34.21 (57.63)	35.14 (23.33)	98
T2	1.01 (1.35)	1.83 (2.36)	0.43 (0.98)	32.73 (17.81)	40.33 (25.51)	14.67 (52)	91
T3	0.85 (0.69)	1.44 (2.09)	1.01 (1.47)	14.52 (21.16)	36.56 (29.72)	68.62 (55.78)	123
T4	1.17 (0.83)	1.84 (1.16)	1.47 (1.02)	25.8 (20.69)	33.66 (37.50)	39.06 (38.79)	103
T5	2.1 (1.13)	0.79 (0.80)	1.18 (1.03)	20.5 (28.72)	69.63 (33.34)	32.8 (34.98)	127
T6	1.24 (2.59)	2.18 (1.39)	0.61 (1.43)	34.5 (24.34)	33.43 (45.95)	24.04 (24.39)	96
T7	1.83 (1.88)	1.35 (1.39)	0.87 (0.89)	21.43 (22.09)	43.65 (45.00)	27.87 (28.73)	97
T8	1.55 (2.30)	1.44 (1.86)	0.64 (1.70)	17.95 (19.50)	30.7 (58.69)	41.72 (19.19)	94
T9	1.73 (1.58)	1.22 (0.99)	0.43 (1.07)	16.17 (30.24)	35.36 (36.78)	57.09 (29.34)	112
T10	0.073 (0.11)	0.24 (0.36)	0.28 (0.42)	10.88 (16.49)	31.81 (48.19)	22.72 (34.40)	66
CD	0.134	0.161	0.112	5.106	2.454	4.75	20.3

The values in parenthesis are the percentage content of the individual fractions to Total Zinc.

The highest value obtained for the Ws+Ex. content of Zn, was in T<sub>5</sub> (2.1 mg kg<sup>-1</sup>) and it was found superior to all other treatments, while the lowest mean was recorded in T<sub>10</sub>. The highest value for OB fraction was noticed in T<sub>6</sub> with a mean value of 2.18 mg kg<sup>-1</sup> and the lowest was noted in the control (0.24 mg kg<sup>-1</sup>). For the

Mn O bound Zn was highest in T<sub>4</sub>(1.47 mg kg<sup>-1</sup>) while the lowest was obtained in the control. The Am Feo was highest in T<sub>6</sub> and T<sub>2</sub> was found to be on par. The lowest value of 10.88 mg kg<sup>-1</sup> was obtained in T<sub>10</sub>. The highest mean value of Cry Feo-Zn was obtained T<sub>5</sub> (69.63 mg kg<sup>-1</sup>) and it was found superior to all others. the control recorded the lowest. The Res-Zn content was found to be maximum in T<sub>3</sub> (68.62 mg kg<sup>-1</sup>) and it was significantly different from other treatment means, while T<sub>2</sub> (14.67 mg kg<sup>-1</sup>) recorded the lowest mean value. T<sub>5</sub> recorded the highest value for total Zn (127 mg kg<sup>-1</sup>) and T<sub>3</sub> and T<sub>9</sub> were on par to T<sub>5</sub> with the values of 123 and 112 mg kg<sup>-1</sup> respectively. The lowest mean value was noted in T<sub>10</sub> (66 mg kg<sup>-1</sup>)

#### 4.4.2. First residual crop

A residual crop has been raised after the 1<sup>st</sup> experimental crop without adding any of the manures or fertilizers and harvested. The green fodder yield and dry fodder yield of the crop is given in Table.19.

Table 19. Green fodder and Dry fodder yield of the first residual crop

Treatments	GFY(q ha <sup>-1</sup> )	DFY(q ha <sup>-1</sup> )
T1	10.32	2.06
T2	14.96	2.89
T3	15.67	3.06
T4	10.32	2.05
T5	34.65	6.86
T6	13.21	2.59
T7	26.62	4.08
T8	20.68	4.24
T9	10.54	2.16
T10	7.61	1.78
CD	6.618	0.457

The residual crop was raised after the harvest of first crop without applying any manures or fertilizer to know the residual impact of nutrients supplied in the first crop season to the yield. The data showed that T<sub>5</sub> (Zn applied at 15 kgha<sup>-1</sup>) along with POP recommendation of 120: 60:40 NPK kgha<sup>-1</sup> and FYM 10tha<sup>-1</sup> gave

the maximum yield of 34.65 qha<sup>-1</sup> of GFY, while the application of the highest doze of Zn(20 kgha<sup>-1</sup>) along with POP recorded only a lower value for GFY (13.21qha<sup>-1</sup>).

#### 4.4.2..1 Total and DTPA extractable Zn after the crop growth first residual crop

Soil samples both surface and subsurface, were collected from each experimental plot after the harvest of the first residual crop and analyzed for Total and DTPH extractable Zn and presented in Table 20.

Table.20. Total and DTPA extractable Zn in the surface and sub surface soil after first residual crop (mg kg<sup>-1</sup>)

Treatments	DTPA Zn surface (mg kg <sup>-1</sup> )	DTPA Zn Subsurface (mg kg <sup>-1</sup> )	Total Zn Surface (mg kg <sup>-1</sup> )	Total Zn Subsurface (mg kg <sup>-1</sup> )
T1	1.03	0.69	41.29	31.9
T2	0.93	0.87	50.73	25.3
T3	1.33	0.79	54.2	27.93
T4	2.06	1.12	48.83	26.07
T5	1.71	1.31	48.13	33.78
T6	1.17	0.98	45.93	43.25
T7	0.97	0.63	37.33	31.07
T8	0.99	0.82	36.67	33.23
T9	1.07	0.84	33.57	28.00
T10	0.56	0.51	38.3	28.83
CD	0.13	0.109	13.07	NS

The treatment means were significantly differ among themselves except for total Zn in the subsurface soil. For the DTPA extractable Zn in the surface and subsurface soil .T<sub>4</sub> and T<sub>5</sub> recorded the highest value of 2.06 and 1.31 mg kg<sup>-1</sup> respectively. In the surface soil the highest value for total Zn was in T<sub>3</sub> (54.2 mg kg<sup>-1</sup>) while T<sub>2</sub>, T<sub>4</sub>, T<sub>5</sub> and T<sub>6</sub> were found to be on par. The highest and lowest value for total Zn was 43.25 (T<sub>6</sub>) and 25.3 mgkg<sup>-1</sup> (T<sub>2</sub>) for subsurface soil.

#### 4.4.3 Second field crop

##### 4.4.3.1 Biometric observations

Different biometric observations viz. Height of the plant no. of leaves per plant etc. were recorded, on the 15<sup>th</sup>, 25<sup>th</sup>, 35<sup>th</sup>, 45<sup>th</sup> days after sowing and at harvest.

Table. 21 . Height (cm) of fodder maize at different stages of growth – second crop

Treatment	Height(cm)				
	15DAS	25DAS	35DAS	45DAS	HVST
T1	4.90	20.17	46.40	114.50	133.40
T2	5.34	19.57	41.50	104.90	119.00
T3	6.00	21.30	50.27	113.90	133.80
T4	5.53	17.40	33.60	123.70	139.20
T5	4.56	16.83	34.20	94.20	137.90
T6	5.47	16.73	36.20	119.70	136.30
T7	5.24	19.78	36.70	93.70	108.80
T8	4.18	15.10	32.90	104.70	117.80
T9	4.20	16.94	27.50	93.00	111.20
T10	1.90	9.00	18.20	26.20	37.40
CD	1.386	2.767	4.435	14.684	12.280

Data presented in Table.21 showed that the height of fodder maize at different stages of growth were significant. At 15 DAS, the highest mean value was observed in T<sub>3</sub> (6 cm) while T<sub>4</sub>, T<sub>2</sub> and T<sub>1</sub> were on par. The lowest value was in T<sub>10</sub>. At 25 DAS, maximum height was in T<sub>3</sub> (21.30 cm) while T<sub>1</sub> (20.17 cm) T<sub>7</sub> (19.78 cm) and T<sub>2</sub> (19.57 cm) were found to be on par with T<sub>3</sub>. T<sub>10</sub> recorded the lowest

value of 9 cm at 25 DAS. At 35 DAS, highest mean value of 50.27 cm was found in T<sub>3</sub> and it was on par with T<sub>1</sub> (46.40 cm). However T<sub>10</sub> recorded the lowest. In the case of 45 DAS, the highest mean value of 123.70 cm was noticed in T<sub>4</sub> and it was on par with T<sub>6</sub>, T<sub>1</sub> and T<sub>3</sub>. While the lowest was noted in T<sub>10</sub> (26.20 cm). At the time of harvest, T<sub>4</sub> recorded the highest value of 139.20 cm, and T<sub>5</sub>, T<sub>6</sub>, T<sub>3</sub> and T<sub>1</sub> were found to be on par and lowest value was recorded in T<sub>10</sub>.

The mean number of leaves recorded plant<sup>-1</sup> on the 15<sup>th</sup>, 25<sup>th</sup>, 35<sup>th</sup>, 45<sup>th</sup> DAS and at the time of harvest are given in Table-22.

Table-22 Mean number of leaves plant<sup>-1</sup> of fodder maize at different stages of growth

Treatments	Number of leaves				
	15DAS	25DAS	35DAS	45DAS	HVST
T1	4.50	8.15	8.80	10.86	11.30
T2	4.97	7.40	8.63	10.33	11.93
T3	4.93	7.20	8.47	11.60	12.63
T4	5.14	6.80	8.50	10.53	12.13
T5	4.59	6.70	8.43	10.76	11.87
T6	4.37	6.50	8.30	9.51	11.53
T7	4.73	7.20	7.90	10.70	11.36
T8	4.20	5.97	7.13	11.30	11.00
T9	4.04	7.07	8.20	11.30	10.63
T10	2.95	4.17	7.60	8.0	10.37
CD	0.742	1.124	NS	1.716	NS

The data presented in Table.22 showed that the number of leaves at different growth stages except 35 DAS and at harvest were found to be significant. At 15 DAS, the highest mean value of 5.14 was recorded in T<sub>4</sub> and T<sub>2</sub>, T<sub>3</sub>, T<sub>7</sub>, T<sub>6</sub> and T<sub>1</sub> were found on par with T<sub>4</sub>. The observation taken after 25 DAS showed T<sub>1</sub> as the maximum leaf produced treatment. The number of leaves 35 DAS and at harvest were found to be non significant. At 45 DAS T<sub>8</sub> and T<sub>9</sub> ie., the Zn foliar application received treatments on the 13<sup>th</sup> DAS, produced the maximum number of leaves, next to T<sub>3</sub>.



**3.4.3.2.** Yield and yield attributes of fodder maize attributes viz. green fodder yield, dry fodder yield, dry matter percentage, per day green fodder yield, leaf area index, leaf stem ratio etc. were recorded in Table-23.

Table-23 Yield and yield attributes of fodder maize - second crop

Treatments	Green fodder (qha <sup>-1</sup> )	Dry Fodder (qha <sup>-1</sup> )	DM (%)	PDGF (qha <sup>-1</sup> )	LAI	LSR
T1	170.40	45.47	18.43	3.97	7.97	1.79
T2	374.10	74.45	19.87	6.79	8.03	1.86
T3	348.70	74.82	21.47	6.33	7.73	1.79
T4	416.70	83.78	20.80	7.57	8.21	1.94
T5	519.30	108.05	20.77	9.44	8.31	2.48
T6	465.40	93.67	20.10	8.46	8.23	2.31
T7	385.00	77.09	20.03	7.00	7.68	1.92
T8	418.70	82.61	19.70	7.61	7.71	1.90
T9	441.60	86.97	19.67	8.03	8.22	1.94
T10	142.50	22.59	15.87	2.59	5.01	1.49
CD	69.850	5.795	1.221	0.671	0.665	NS

Data presented in Table-23 revealed that the yield and yield attributes of second crop were significant except for LSR. The GFY was maximum in T<sub>5</sub> (soil application of Zn @ 15 kg ha<sup>-1</sup> +POP) with a value of 519.3 qha<sup>-1</sup> and T<sub>6</sub> was found on par with an yield of 465.4 qha<sup>-1</sup>. The lowest GFY was recorded in T<sub>10</sub> with 142.5 q ha<sup>-1</sup>. In the case of DFY, T<sub>5</sub> recorded the highest value of 108.05 and T<sub>10</sub> observed the lowest (22.59 qha<sup>-1</sup>). T<sub>5</sub> was found to be superior over other mean values.

T<sub>3</sub> recorded the maximum dry matter content which was on par with T<sub>4</sub> and T<sub>5</sub>. The values were 21.47, 20.80 and 20.77 per cent respectively. While the lowest was recorded in T<sub>10</sub> (15.87 per cent). For the PDGFY, T<sub>5</sub> recorded the highest (9.44 qha<sup>-1</sup>) while T<sub>10</sub> observed the lowest value of 2.59 qha<sup>-1</sup>. T<sub>5</sub> found significantly superior to all other values. For LAI the highest value was found to be in T<sub>5</sub> (8.31) and all other treatment means except T<sub>10</sub> were found to be on par. As in the case of

1<sup>st</sup> crop for the 2<sup>nd</sup> crop too, the maximum LAI recorded treatment had given the maximum green fodder yield. The leaf stem ratio was found to be non-significant among treatments.

#### 4.4.3.3 Quality attributes of fodder maize of second crop

The quality attributes viz. crude protein per cent, crude protein yield and crude fibre per cent are given in Table-24

Table-24 Quality attributes of fodder maize of second crop

Treatments	CP (%)	CPY (kg ha <sup>-1</sup> )	CF (%)
T1	9.52	433	27.94
T2	10.06	749	23.43
T3	10.23	766	26.1
T4	9.78	818	25.46
T5	10.08	1090	24.23
T6	10.14	949	23.93
T7	9.46	730	25.6
T8	9.52	787	24.3
T9	9.67	841	24.1
T10	6.50	147	29.73
CD	0.452	826	0.846

The data in Table.24 revealed that the highest CP content was observed in T<sub>3</sub> (10.23 per cent) and T<sub>6</sub>, T<sub>5</sub>, T<sub>2</sub>, and T<sub>4</sub> were found to be on par with values of 10.14, 10.08, 10.06 and 9.78 respectively. In the case of CPY, it was maximum in T<sub>5</sub> (1090 kg ha<sup>-1</sup>) and it was found significantly different from all other mean values. The lowest CF was obtained in T<sub>2</sub> (23.43 per cent).

#### 4.4.3.4. Nutrient content in fodder maize – Second crop

After the harvest of the 2<sup>nd</sup> crop the plant samples collected were processed and analyzed for the nutrient concentration and the results are given in Table-25.

Table.25. Nutrient concentration in the fodder maize – second crop

Treatments	N (%)	P (%)	K (%)	Ca (%)	Mg (%)	S (%)	Fe (mgkg <sup>-1</sup> )	Mn (mgkg <sup>-1</sup> )	Zn (mgkg <sup>-1</sup> )	Cu (mgkg <sup>-1</sup> )
T1	1.52	0.12	1.33	0.26	0.14	0.13	109.0	41.67	56.33	12.33
T2	1.61	0.21	1.6	0.32	0.19	0.17	128.3	45.67	61.67	15.00
T3	1.67	0.18	1.76	0.24	0.13	0.16	108.7	39.33	66.00	14.00
T4	1.56	0.25	1.83	0.31	0.21	0.16	96.0	36.00	75.00	13.00
T5	1.61	0.21	2.18	0.35	0.17	0.15	97.3	34.67	89.00	12.67
T6	1.62	0.20	1.50	0.41	0.18	0.14	98.0	32.67	84.00	12.67
T7	1.51	0.18	0.84	0.25	0.14	0.15	119.3	34.00	85.67	12.67
T8	1.52	0.19	0.87	0.28	0.16	0.15	116.0	32.67	86.33	13.00
T9	1.55	0.20	0.90	0.29	0.17	0.16	117.7	35.00	90.33	13.33
T10	1.04	0.08	0.76	0.10	0.09	0.11	102.7	28.67	29.33	10.33
CD	0.072	0.020	0.170	0.241	0.019	0.004	15.480	5.353	5.220	1.502

The data from the Table. 25 revealed that the nutrient concentration in fodder maize among the different treatments were significant. Maximum content of N was noticed in T<sub>3</sub> (1.67 per cent) and T<sub>6</sub> (1.62 per cent), T<sub>5</sub> and T<sub>2</sub> (1.61 percent each) were found to be on par with T<sub>3</sub>. The lowest was noticed in T<sub>10</sub> (1.04 per cent). The highest value for P was in T<sub>4</sub> (0.25 per cent) and was found superior to other treatments. The lowest was seen in T<sub>10</sub> (0.08 per cent). The highest value for K (2.18 per cent) was showed in T<sub>5</sub> and this treatment was found to be superior, while the lowest was recorded by T<sub>10</sub> (0.76 per cent).

The Ca content was maximum in T<sub>6</sub> (0.41 per cent). All other treatments were found on par with T<sub>6</sub> except T<sub>10</sub> which recorded the lowest value of 0.1 per cent. The highest value for Mg was observed in T<sub>4</sub> (0.21 per cent) and T<sub>2</sub> was at par with T<sub>4</sub>, while the lowest value was noted by T<sub>10</sub> (0.09 per cent). In the case of S, T<sub>2</sub> recorded the maximum content of 0.17 per cent and was superior to all other mean values. But lowest was found in T<sub>10</sub> (0.11 per cent).

In the case of Fe content, T<sub>2</sub> recorded the maximum value of 128.3 mg kg<sup>-1</sup> which was on par with T<sub>7</sub>, T<sub>9</sub> and T<sub>8</sub>. The lowest value was noticed in T<sub>4</sub> (96.0 mg kg<sup>-1</sup>). For the Mn concentration, T<sub>2</sub> recorded the highest value of 45.67 mg kg<sup>-1</sup> and T<sub>1</sub> (41.67 mg kg<sup>-1</sup>) was on par. The lowest value of 49 mg kg<sup>-1</sup> was noted in T<sub>10</sub>. The Zn concentration was highest (90.33 mg kg<sup>-1</sup>) on T<sub>9</sub> and it was found on par with T<sub>5</sub>, T<sub>7</sub> and T<sub>8</sub>. T<sub>10</sub> (29.33 mg kg<sup>-1</sup>) recorded the lowest. Maximum Cu content was observed in T<sub>2</sub> and T<sub>3</sub> was found at par with T<sub>2</sub> and the values were 15 mg kg<sup>-1</sup> and 14 mg kg<sup>-1</sup> respectively. T<sub>10</sub> recorded the lowest value of 10.33 mg kg<sup>-1</sup>.

#### 4.4.3.5. Nutrient uptake Table 26

Nutrient uptake studies are given in the Table.26

Table.26 Nutrient uptake studies of second crop in kg ha<sup>-1</sup>

Treatments	N	P	K	Ca	Mg	S	Fe	Mn	Zn	Cu
T1	69.26	5.46	60.63	11.2	6.35	9.81	0.49	0.18	0.25	0.06
T2	119.91	15.37	119.29	23.83	13.92	8.34	0.95	0.34	0.46	0.11
T3	122.55	12.82	131.76	17.7	12.54	13.88	0.90	0.30	0.49	0.10
T4	130.95	20.94	153.97	25.96	17.60	14.64	0.80	0.30	0.59	0.13
T5	174.52	23.01	235.27	37.42	30.87	11.80	1.06	0.37	0.96	0.12
T6	151.90	18.50	157.76	38.43	17.78	9.91	0.90	0.31	0.79	0.12
T7	116.72	13.89	64.67	19.82	10.56	10.89	0.92	0.26	0.70	0.10
T8	125.86	15.42	71.85	23.10	13.24	10.14	0.95	0.27	0.71	0.11
T9	134.55	17.13	77.90	25.24	14.83	10.44	1.02	0.30	0.79	0.12
T10	23.54	1.89	17.12	2.23	2.19	1.35	0.23	0.06	0.07	0.07
CD	13.083	2.994	13.075	3.258	6.852	1.568	0.148	0.038	0.068	0.019

Uptake studies (Table 26) revealed that the different treatment means were significant. The highest N uptake was observed in T<sub>5</sub> and was found superior to 174.52 kg ha<sup>-1</sup>, while the lowest value of 23.54 kg ha<sup>-1</sup> was noticed in T<sub>10</sub>. The P uptake was maximum in T<sub>5</sub> (23.01 kg ha<sup>-1</sup>) and T<sub>4</sub> (20.94 kg ha<sup>-1</sup>) was on par with T<sub>5</sub> and T<sub>10</sub> (1.89 kg ha<sup>-1</sup>) recorded the lowest. The uptake of K was maximum in T<sub>5</sub> (235.27 kg ha<sup>-1</sup>) and was significantly superior. The lowest value of 17.21 kg ha<sup>-1</sup> was observed in T<sub>10</sub>.

For Ca uptake, T<sub>6</sub> (38.43 kg ha<sup>-1</sup>) recorded the highest and was superior to all treatment means and the lowest was observed in 2.23 kg ha<sup>-1</sup>. T<sub>5</sub> with a value of 37.42 kg ha<sup>-1</sup> on par with T<sub>6</sub>. For the uptake of Mg, the highest value was observed in T<sub>5</sub> (30.87) and significantly superior, but lowest value was obtained for T<sub>10</sub> (2.19 kg ha<sup>-1</sup>). In the case of S, T<sub>4</sub> was noticed the highest uptake of 14.64 and T<sub>3</sub> (13.88 kg ha<sup>-1</sup>) found to be on par with T<sub>4</sub>, while the value of 1.35 kg ha<sup>-1</sup> observed as lowest (T<sub>10</sub>).

For the highest uptake of Fe was in T<sub>5</sub> (1.06 kg ha<sup>-1</sup>), while T<sub>9</sub> (1.02 kg ha<sup>-1</sup>), T<sub>8</sub> and T<sub>2</sub> (0.95 kg ha<sup>-1</sup>) and T<sub>7</sub> (0.92 kg ha<sup>-1</sup>) were found to be on par with T<sub>5</sub>. But T<sub>10</sub> showed the lowest value of 0.23 kg ha<sup>-1</sup>. The highest value of Mn uptake (0.37 kg ha<sup>-1</sup>) was observed in T<sub>5</sub> and T<sub>2</sub> was found to be on par (0.34 kg ha<sup>-1</sup>). T<sub>10</sub> (0.04 kg ha<sup>-1</sup>) observed the lowest value. In the case of Zn, T<sub>5</sub> (0.96 kg ha<sup>-1</sup>) was noticed the maximum uptake value and lowest was showed in T<sub>10</sub> (0.069 kg ha<sup>-1</sup>). But T<sub>5</sub> observed as significantly superior. Highest uptake value for Cu was noticed in T<sub>4</sub> (0.13 kg ha<sup>-1</sup>), but T<sub>5</sub>, T<sub>6</sub> and T<sub>9</sub> (0.12 kg ha<sup>-1</sup>), T<sub>2</sub> and T<sub>8</sub> (0.11 kg ha<sup>-1</sup>) was on par with T<sub>4</sub>. T<sub>1</sub> recorded the lowest (0.06 kg ha<sup>-1</sup>).

#### **4.4.3.6. Physico-chemical properties and primary nutrient concentration in the surface soil after the crop growth**

Surface soil samples (0-15 cm) were collected from each plot after the second crop growth and analyzed for different physico-chemical parameters and primary nutrient elements and the data is presented in Table-27.

Table.27 Physico-chemical parameters and primary nutrient content of surface soil after the second crop

Treatments	pH	EC (dSm <sup>-1</sup> )	Organic Carbon (%)	Avail.N (kg ha <sup>-1</sup> )	Avail.P (kg ha <sup>-1</sup> )	Avail.K <sup>-</sup> (kg ha <sup>-1</sup> )
T <sub>1</sub>	5.1	0.13	0.79	178.2	12.13	109.9
T <sub>2</sub>	5.2	0.13	0.99	215.9	12.32	188.2
T <sub>3</sub>	5.1	0.13	0.73	185.9	13.81	156.7
T <sub>4</sub>	5.1	0.15	0.80	211.1	14.12	144.0
T <sub>5</sub>	5.4	0.14	0.70	254.2	13.32	86.2
T <sub>6</sub>	5.3	0.12	0.80	207.5	12.88	104.2
T <sub>7</sub>	5.3	0.13	1.11	189.7	9.56	150.2
T <sub>8</sub>	5.2	0.16	0.97	205.4	10.81	115.3
T <sub>9</sub>	5.2	0.13	1.01	223.5	11.84	140.8
T <sub>10</sub>	5.3	0.12	0.78	73.0	4.95	75.8
CD	0.45	NS	NS	21.5	1.661	15.97

The treatment means were significant for the physico-chemical parameters and primary nutrient content of the surface soil after the second crop except for EC and organic carbon (Table 25). The pH was maximum in T<sub>5</sub>. The highest content of available N was with T<sub>5</sub> (254.20 kg ha<sup>-1</sup>) while the lowest value was recorded in T<sub>10</sub> (73.0 kg ha<sup>-1</sup>). Available P was highest in T<sub>4</sub> (14.12 kg ha<sup>-1</sup>) and was on par with T<sub>3</sub>, T<sub>5</sub> and T<sub>6</sub> while the lowest was recorded in T<sub>10</sub> 2.85 kg ha<sup>-1</sup>). The highest and lowest value of K was noticed in T<sub>2</sub> and T<sub>10</sub> with values of 188.20 and 75.80 kg ha<sup>-1</sup> respectively.

#### 4.4.3.7 Secondary and micronutrient content in the surface soil after the second crop

The surface soil (0-15 cm) samples collected from each plot, after the harvest of the second crop were analyzed for the secondary and micronutrient content and the data is presented in Table-28

Table 28 Secondary and micronutrient content of surface soil after crop growth –  
Second crop

Treatments	Exch. Ca (cmol kg <sup>-1</sup> )	Exch. Mg (cmol kg <sup>-1</sup> )	Avail.S (kg ha <sup>-1</sup> )	DTPA Fe (mg kg <sup>-1</sup> )	DTPA Mn (mg kg <sup>-1</sup> )	DTPA Zn (mg kg <sup>-1</sup> )	DTPA Cu (mg kg <sup>-1</sup> )
T <sub>1</sub>	1.30	0.54	11.50	16.33	7.79	2.5	0.75
T <sub>2</sub>	1.30	0.46	17.50	15.33	6.20	3.74	0.86
T <sub>3</sub>	1.26	0.55	12.83	20.67	5.60	4.63	0.62
T <sub>4</sub>	1.15	0.46	13.60	20.00	7.57	6.15	0.60
T <sub>5</sub>	1.48	0.51	15.18	15.00	2.38	5.93	0.40
T <sub>6</sub>	1.27	0.46	15.74	17.00	8.19	5.99	0.52
T <sub>7</sub>	1.22	0.60	12.00	16.00	6.11	3.55	0.32
T <sub>8</sub>	1.23	0.44	13.73	17.33	8.15	3.56	0.37
T <sub>9</sub>	1.25	0.42	14.50	13.00	5.59	3.68	0.45
T <sub>10</sub>	1.07	0.40	7.60	17.00	1.14	0.97	0.27
CD	0.068	0.102	1.158	2.932	1.317	0.229	0.28

The highest value for exch. Ca was 1.48 cmol kg<sup>-1</sup> and it was found superior to all other treatments, while the lowest was observed in T<sub>10</sub>(1.07 cmol kg<sup>-1</sup>). For exch. Mg, the highest mean value of 0.55 c mol kg<sup>-1</sup> was observed in T<sub>3</sub> while T<sub>1</sub> was found on par with a value of 0.54 cmol kg<sup>-1</sup> and T<sub>10</sub> recorded the lowest. Available S was highest in T<sub>2</sub> with a value of 17.50 kg ha<sup>-1</sup> which was found superior and lowest value of 7.60 kg ha<sup>-1</sup> was observed in T<sub>10</sub>.

Maximum value for DTPA Fe was observed in T<sub>3</sub> (20.67 mg kg<sup>-1</sup>) while T<sub>4</sub> and T<sub>8</sub> were found on par with values of 20 and 17.33 mg kg<sup>-1</sup>. T<sub>9</sub> recorded the lowest value of 13 mg kg<sup>-1</sup>. For DTPA Mn, maximum value of 8.19 mg kg<sup>-1</sup> was observed in T<sub>6</sub> while T<sub>8</sub>, T<sub>4</sub> and T<sub>1</sub> were found to be on par. T<sub>10</sub> recorded the lowest of 1.14 mgkg<sup>-1</sup>. The highest value of DTPA Zn was observed in T<sub>4</sub> (6.15 mgkg<sup>-1</sup>), while T<sub>6</sub> (5.99 mgkg<sup>-1</sup>) and T<sub>5</sub> (5.93 mgkg<sup>-1</sup>) were found on par. The lowest value was 0.97 mg kg<sup>-1</sup> in T<sub>10</sub>. Maximum value for DTPA Cu was observed in T<sub>2</sub> (0.86 mgkg<sup>-1</sup>) and was on par with T<sub>1</sub> (0.75 mgkg<sup>-1</sup>), T<sub>3</sub> (0.62 mgkg<sup>-1</sup>) and T<sub>4</sub> (0.60 mg kg<sup>-1</sup>). The lowest value was in recorded in T<sub>10</sub> (0.27 mgkg<sup>-1</sup>).

#### 4.4.3.8. Zn fractionation of surface soil after the second crop

The surface soil samples collected from each plot were analyzed for different fractions of Zn and the data is given in Table-29.

Table 29 Fractions of Zn ( $\text{mg kg}^{-1}$ ) in the surface soil after the second crop

Treatments	Ws+Ex-Zn	OB-Zn	MnO-Zn	AmFeO-Zn	Cry FeO-Zn	Res-Zn	Total Zn
T1	2.34 (1.97)	4.47 (3.76)	1.09 (1.12)	27.89 (43.56)	33.30 (27.91)	29.91 (25.13)	119.00
T2	1.65 (1.70)	1.92 (1.98)	0.61 (0.51)	30.53 (31.47)	43.00 (44.33)	19.29 (19.80)	97.00
T3	3.68 (3.07)	3.93 (3.28)	1.26 (1.05)	27.82 (23.18)	46.00 (38.33)	37.31 (31.09)	120.00
T4	4.66 (3.17)	3.22 (2.19)	1.46 (0.99)	27.65 (18.81)	86.00 (58.50)	24.01 (16.33)	147.00
T5	5.06 (3.89)	3.28 (2.19)	1.22 (0.94)	23.28 (17.91)	75.00 (57.69)	22.16 (17.04)	130.00
T6	4.74 (4.83)	3.16 (2.52)	0.84 (0.85)	26.95 (27.50)	39.00 (39.80)	23.31 (23.79)	98.00
T7	3.32 (2.20)	2.25 (3.22)	0.47 (0.31)	23.93 (15.85)	76.00 (50.33)	45.03 (29.82)	151.00
T8	2.28 (1.65)	2.76 (1.49)	0.66 (0.49)	13.82 (10.01)	91.00 (66.00)	27.48 (19.91)	138.00
T9	2.16 (1.38)	3.09 (2.00)	0.73 (0.46)	22.11 (14.08)	90.00 (57.32)	18.91 (12.00)	157.00
T10	0.59 (0.72)	0.91 (1.97)	0.31 (0.38)	22.00 (26.83)	33.00 (40.24)	25.19 (30.72)	82.00
CD	0.194	0.186	0.093	3.69	10.05	5.07	30.85

The values in parentheses are the percentage content of the individual fractions to total Zn.

The data from the Table. 29 indicated that the different fractions of Zn in the surface soil after the second crop was found significant among treatments. The highest content of Ws+Ex-Zn was of  $5.06 \text{ mg kg}^{-1}$  was in T<sub>5</sub> significantly superior and the lowest was in the control ( $0.59 \text{ mg kg}^{-1}$ ). The OB fraction, the highest and the lowest values were in T<sub>1</sub> ( $4.47 \text{ mg kg}^{-1}$ ) and T<sub>10</sub> ( $0.91 \text{ mg kg}^{-1}$ ), For the Mn O



bound content, the maximum value of  $1.46 \text{ mg kg}^{-1}$  was recorded in  $T_4$ . Maximum value of AmFeO-Zn ( $30.53 \text{ mg kg}^{-1}$ ) was noticed in  $T_2$  and the lowest value ( $13.82 \text{ mg kg}^{-1}$ ) was noted in  $T_8$ . The Cry FeO-Zn content was highest in  $T_8$  ( $91 \text{ mg kg}^{-1}$ ) but it was closely followed by  $T_9$  and  $T_4$ , with values of 90 and  $86 \text{ mg kg}^{-1}$ . The lowest was in the control. The Res- Zn was highest in  $T_7$  ( $45.03 \text{ mg kg}^{-1}$ ), The lowest value was in  $T_9$ . For the total Zn, maximum value was found to be  $157 \text{ mg kg}^{-1}$  ( $T_9$ ) while  $T_7$ ,  $T_4$ ,  $T_8$  and  $T_5$  were on par with mean values of 151, 147, 138 and  $130 \text{ mg kg}^{-1}$  respectively and the lowest value was in  $T_{10}$  ( $82 \text{ mg kg}^{-1}$ ).

#### 3.4.3.9 Physico-chemical properties and primary nutrient concentration in the subsurface soil after the crop growth

Subsurface soil samples (15-30cm) were collected from each plot after the second crop growth and analyzed for different physico-chemical parameters and primary nutrients and the data is presented in the Table .30

Table.30. Physico-chemical parameters of the subsurface soil after the second crop

Treatments	pH	EC ( $\text{dSm}^{-1}$ )	Organic carbon (%)	Av N ( $\text{kg ha}^{-1}$ )	Av. P ( $\text{kg ha}^{-1}$ )	Av.K ( $\text{kg ha}^{-1}$ )
$T_1$	5.01	0.11	0.44	143.8	3.21	245.67
$T_2$	4.8	0.14	0.46	155.3	3.42	133.00
$T_3$	5.0	0.12	0.40	164.4	9.15	183.73
$T_4$	4.9	0.10	0.30	171.1	6.10	204.3
$T_5$	5.1	0.11	0.20	219.2	3.29	172.67
$T_6$	4.9	0.12	0.82	166.9	3.87	108.93
$T_7$	5.0	0.12	0.76	157.8	3.58	177.46
$T_8$	5.03	0.15	0.84	193.9	4.63	179.77
$T_9$	4.97	0.10	0.98	138.7	3.81	186.84
$T_{10}$	4.89	0.12	0.58	98.0	2.85	69.00
CD	NS	NS	NS	32.41	3.504	9.132

The pH, EC and OC contents were not significant among treatments. The maximum content of available N was observed in  $T_5$ , available P in  $T_3$  and available K in  $T_1$ .

#### 4.4.3.10. Secondary and micronutrient contents in the subsurface soil after the crop growth

The sub-surface (15-30 cm) soil samples collected from each plot after the harvest of the second crop were analysed for the secondary and micronutrient contents and the data is presented in Table-31.

Table 31. Secondary and micro nutrient contents of subsurface soil after the second crop

Treatments	Ex ca (c mol kg <sup>-1</sup> )	Ex Mg (c mol kg <sup>-1</sup> )	Avail.S (kg ha <sup>-1</sup> )	DTPA Fe (mg kg <sup>-1</sup> )	DTPA Mn (mg kg <sup>-1</sup> )	DTPA Zn (mg kg <sup>-1</sup> )	DTPA Cu (mg kg <sup>-1</sup> )
T <sub>1</sub>	1.29	0.48	9.26	18.03	3.29	2.17	0.82
T <sub>2</sub>	1.3	0.46	11.11	14.62	2.3	2.44	0.50
T <sub>3</sub>	1.28	0.51	11.88	22.37	4.53	3.25	0.61
T <sub>4</sub>	1.27	0.52	12.1	19.80	3.61	3.28	0.65
T <sub>5</sub>	1.26	0.48	12.88	18.85	3.86	3.20	0.61
T <sub>6</sub>	1.15	0.43	9.78	14	2.66	2.72	0.32
T <sub>7</sub>	1.25	0.38	11.35	18	4.06	1.75	0.46
T <sub>8</sub>	1.22	0.37	11.58	17.20	3.93	2.16	0.47
T <sub>9</sub>	1.23	0.38	11.54	18.51	4.94	2.12	0.49
T <sub>10</sub>	1.07	0.35	6.05	15.80	0.77	0.85	0.21
CD	0.070	0.034	2.106	5.03	0.544	0.35	0.08

The exch.Ca and DTPA C u were higher in T<sub>1</sub> and T<sub>4</sub> recorded the highest exch.Mg and DTPA Zn .The avail. S ,DTPA Fe and DTPA Mn recorded higher values in T<sub>5</sub>, T<sub>3</sub> and T<sub>9</sub> respectively.

#### 4.4.3.11 Zn fractionation of sub-surface soil after the Second crop

The sub-surface soil samples collected from each plot were analysed for different fractions of Zn and the data is presented in Table-32.

Table. 32 Fractions of Zn in the subsurface soil (mgkg<sup>1</sup>)

Treatments	WS+EX.	OB	Mn O	Am Fe	Cry Fe	Res	Total
T <sub>1</sub>	1.46 (1.70)	1.18 (1.37)	1.56 (1.81)	20.65 (24.01)	33.5 (38.95)	27.65 (32.15)	86
T <sub>2</sub>	1.28 (1.35)	2.24 (2.36)	0.93 (0.98)	16.92 (17.81)	39.33 (41.4)	34.3 (36.10)	95
T <sub>3</sub>	2.63 (2.22)	2.65 (2.25)	1.16 (0.98)	16.03 (13.58)	68 (57.63)	27.53 (23.33)	118
T <sub>4</sub>	1.57 (1.22)	2.08 (1.61)	1.53 (1.18)	48.97 (37.96)	52 (40.31)	22.85 (17.71)	129
T <sub>5</sub>	1.21 (1.44)	0.86 (1.02)	1.1 (1.02)	30.73 (36.58)	35.67 (42.46)	14.43 (17.18)	84
T <sub>6</sub>	2.2 (2.59)	1.18 (1.39)	1.18 (1.39)	20.69 (24.34)	39.06 (46.0)	20.69 (24.34)	85
T <sub>7</sub>	0.7 (0.83)	0.98 (1.17)	0.86 (1.02)	17.38 (20.69)	31.5 (38)	32.5 (38.69)	84
T <sub>8</sub>	3.01 (2.30)	1.34 (1.02)	2.2 (1.68)	25.54 (19.50)	76.89 (59)	22.02 (16.81)	131
T <sub>9</sub>	1.8 (1.58)	1.13 (0.99)	1.22 (1.07)	34.37 (30.14)	41.93 (37)	33.55 (29.43)	114
T <sub>10</sub>	0.15 (0.20)	0.66 (0.86)	0.74 (0.96)	21.24 (27.58)	27.54 (36)	26.67 (34.63)	77
CD	0.163	0.124	0.17	3.734	10.64	4.66	9.678

The values in parentheses are the percentage content of the individual fractions to Total Zn.

The data presented in the Table. 32 were significant for various treatment means of Zn fractions in the subsurface soil. The highest value of  $W_s+Ex$ ,  $MnO$  bound,  $Cry Fe O$  and total Zn was observed in  $T_8$  and these values were found significantly different when compared to other mean values. The highest and lowest values for  $Am FeO - Zn$  were recorded for  $T_4(48.97 \text{ mgkg}^{-1})$  and  $T_3(16.03 \text{ mgkg}^{-1})$  respectively. For residual content of Zn, though  $T_2 (34.3 \text{ mgkg}^{-1})$  recorded maximum,  $T_9$  and  $T_7$  were found so par with  $T_2$ .

#### 4.4. 4 Second Residual Crop after the second experimental crop

A second residual crop has been raised after the second experimental crop without adding any of the manures or fertilizers and harvested. The green fodder and dry fodder yield of the second residual crop are given in Table-33.

Table.33. Green fodder and Dry fodder yield of the second residual crop

Treatments	Green fodder Yield ( $qh a^{-1}$ )	Dry fodder Yield ( $qha^{-1}$ )
$T_1$	21.54	4.34
$T_2$	30.93	6.3
$T_3$	31.94	6.32
$T_4$	26.87	5.3
$T_5$	26.00	5.19
$T_6$	39.49	7.77
$T_7$	32.16	6.39
$T_8$	23.17	4.59
$T_9$	34.29	6.93
$T_{10}$	18.37	3.02
CD	3.381	0.964

The residual crop was raised after the harvest of the second crop without applying any manures or fertilizers to know the residual effect of nutrients applied earlier.

The data shown in the Table. 33 of the second residual crop were significant among treatments. The highest mean value for green fodder as well as dry fodder was observed in T<sub>6</sub> with the values of 39.49 qha<sup>-1</sup> and 7.77 qha<sup>-1</sup>) which was found superior while the lowest was noticed in the control.

#### 4.4.4.1 Total and DTPA extractable Zn after the crop growth – Second residual crop

Soil samples both surface and sub surface were collected from each experimental plot after the harvest of the second residual crop and analysed for Total and DTPA extractable Zn and presented in Table-34.

Table 34 DTPA extractable and total Zn in the surface and sub surface soil after second residual crop (mg kg<sup>-1</sup>)

Treatments	DTPA Zn surface	DTPA Zn subsurface	Total Zn Surface	Total Zn subsurface
T <sub>1</sub>	1.05	0.77	41.27	35.63
T <sub>2</sub>	0.98	0.63	50.73	33.17
T <sub>3</sub>	1.89	0.87	54.2	43.87
T <sub>4</sub>	2.1	0.72	57.53	38.03
T <sub>5</sub>	1.62	1.09	45.5	27.42
T <sub>6</sub>	1.28	0.99	52.53	50.47
T <sub>7</sub>	1.17	0.59	36.83	33.63
T <sub>8</sub>	1.41	0.65	36.67	37.37
T <sub>9</sub>	1.49	0.52	35.97	33.83
T <sub>10</sub>	0.54	0.49	33.6	31.53
CD	0.076	0.11	13.81	5.084

The data presented in the Table. 34 revealed that the treatment means for the DTPA extractable and total Zn in the surface and subsurface soil for the second residual crop were significant. The highest mean value for the DTPA extractable Zn in the surface soil was observed in T<sub>4</sub> (2.1 mgkg<sup>-1</sup>) and it was found

superior to all other values. The lowest value of 0.54 (mgkg<sup>-1</sup>) was noted in the control. For the subsurface, the highest was 1.09 mg kg<sup>-1</sup> which was found on par with T<sub>6</sub> and lowest was noted in T<sub>10</sub>. The maximum value for total Zn in the surface soil was noted in T<sub>4</sub> (57.53 mgkg<sup>-1</sup>) while it was on par to T<sub>3</sub>, T<sub>6</sub>, T<sub>2</sub> and T<sub>5</sub> while the lowest value was in T<sub>10</sub>. For the subsurface soil, T<sub>6</sub> recorded the highest value (50.47 mg kg<sup>-1</sup>) and it was found significantly differ from all other values. The lowest value was in the control plot.

#### 4.4.5. Pooled analysis – Yield

The yield data generated out of the two field experiments were pooled and analysed statistically. The pooled data on yield is given in Table 35.

Table. 35 . Pooled analysis data on yield

Treatment	Green fodder yield (q ha <sup>-1</sup> )	Dry fodder yield ( q ha <sup>-1</sup> )
T <sub>1</sub>	327.60	64.20
T <sub>2</sub>	346.10	69.70
T <sub>3</sub>	402.80	82.20
T <sub>4</sub>	464.50	93.80
T <sub>5</sub>	468.80	94.90
T <sub>6</sub>	383.30	76.60
T <sub>7</sub>	375.50	74.90
T <sub>8</sub>	403.90	79.80
T <sub>9</sub>	418.30	82.30
T <sub>10</sub>	104.30	17.60
CD	66.30	6.67

The data presented in the Table-35 revealed that the pooled analysis data for the yield was found significant. The highest yield was recorded for T<sub>5</sub> with a mean value of 468.8 q ha<sup>-1</sup>, but it was on par with T<sub>4</sub>, T<sub>9</sub> and T<sub>8</sub>. The highest dry fodder yield was in T<sub>5</sub> which was on par to T<sub>4</sub> with the values of 94.9 and 93.8 q ha<sup>-1</sup> respectively.

#### 4.4.6 Foliar diagnosis

To identify the best part and best stage of sampling the 3<sup>rd</sup>, 4<sup>th</sup> and 5<sup>th</sup> leaf lamina and petiole were analysed for Zn at different stages of crop growth viz, 15, 25,35 and 45 DAS.

##### 4.4.6.1 Zinc concentration (mgkg<sup>-1</sup>) of leaf and petiole of fodder maize at 15 DAS

Table. 36. Zinc concentration (mgkg<sup>-1</sup>) of leaf lamina and petiole of fodder maize at 15 DAS

Treatments	Leaf lamina			Petiole		
	3 <sup>rd</sup>	4 <sup>th</sup>	5 <sup>th</sup>	3 <sup>rd</sup>	4 <sup>th</sup>	5 <sup>th</sup>
T <sub>1</sub>	4.60	5.22	4.67	6.08	6.63	7.10
T <sub>2</sub>	4.63	6.10	4.70	5.37	5.83	6.50
T <sub>3</sub>	5.33	5.20	5.70	6.60	7.03	7.93
T <sub>4</sub>	6.57	6.47	6.73	7.73	8.10	7.77
T <sub>5</sub>	9.97	9.83	10.03	11.93	11.53	11.33
T <sub>6</sub>	8.07	8.03	8.27	10.47	9.77	10.13
T <sub>7</sub>	8.82	8.67	8.90	11.33	9.70	10.32
T <sub>8</sub>	9.27	9.47	9.57	12.13	11.90	12.10
T <sub>9</sub>	9.63	9.93	10.05	12.00	11.27	11.00
T <sub>10</sub>	4.13	4.83	4.50	5.42	4.90	5.75
CD	0.794	1.495	0.713	1.664	1.691	1.490

Data presented in the Table. 36 showed that the treatment means were significant at 15 DAS. For the third leaf lamina, the highest mean value obtained for T<sub>5</sub> (9.97 mgkg<sup>-1</sup>) and was found significantly superior. The lowest mean value 4.13 mgkg<sup>-1</sup> for T<sub>10</sub>. The treatments T<sub>9</sub> and T<sub>8</sub> were found to be on par with T<sub>5</sub>. For the 4<sup>th</sup> leaf lamina the maximum value was recorded 9.93 mgkg<sup>-1</sup> for T<sub>9</sub> and it was on par with T<sub>5</sub>, T<sub>8</sub> and T<sub>7</sub> and the lowest value was recorded for T<sub>10</sub> with 4.83 mgkg<sup>-1</sup>. While in the 5<sup>th</sup> leaf lamina highest mean value was noticed 10.05 mgkg<sup>-1</sup> for T<sub>9</sub> and it was on par with T<sub>5</sub>, and T<sub>8</sub> and the lowest value was noticed in T<sub>10</sub> with 4.50 mgkg<sup>-1</sup>.

The concentration of Zn in the petiole was in the range of 4.90 mg kg<sup>-1</sup> to 12.13 mgkg<sup>-1</sup>. For the 3<sup>rd</sup> petiole, the highest value of 12.13 mgkg<sup>-1</sup> was recorded for T<sub>8</sub> and it was on par with T<sub>9</sub>, T<sub>5</sub> T<sub>7</sub> and T<sub>6</sub> with the values 12.00 ,11.93, 11.33 and 10.47 mgkg<sup>-1</sup> respectively while the lowest value was observed for T<sub>10</sub> (5.42 mg kg<sup>-1</sup>). For the 4<sup>th</sup> petiole, the highest value obtained for T<sub>8</sub> (11.90 mgkg<sup>-1</sup>) and it was on par to T<sub>5</sub>(11.53 mg kg<sup>-1</sup>), T<sub>9</sub>(11.27 mg kg<sup>-1</sup>) respectively and the lowest value of 4.90 mg kg<sup>-1</sup> was recorded for T<sub>10</sub>. In the case of 5<sup>th</sup> petiole, the highest value of 12.10 mg kg<sup>-1</sup> was noticed in T<sub>8</sub> which was found to be on par with T<sub>5</sub> and T<sub>9</sub> with values of 11.33 and 11.00 mgkg<sup>-1</sup>. The lowest value (5.75 mg kg<sup>-1</sup>) was recorded for T<sub>10</sub>.

#### 4.4.6.2 Zinc concentration (mgkg<sup>-1</sup>) of leaf and petiole of fodder maize at 25DAS

Table.37. Zinc concentration (mgkg<sup>-1</sup>) of leaf lamina and petiole of fodder maize at 25DAS

Treatments	Leaf lamina			Petiole		
	3 <sup>rd</sup>	4 <sup>th</sup>	5 <sup>th</sup>	3 <sup>rd</sup>	4 <sup>th</sup>	5 <sup>th</sup>
T <sub>1</sub>	12.23	13.38	13.00	10.45	16.10	9.73
T <sub>2</sub>	15.83	11.70	15.67	11.80	17.60	11.21
T <sub>3</sub>	18.22	21.03	18.90	16.15	22.20	17.57
T <sub>4</sub>	19.83	21.80	19.70	20.09	26.20	17.00
T <sub>5</sub>	23.00	25.26	15.67	24.64	14.20	23.42
T <sub>6</sub>	20.85	18.93	19.47	15.26	21.40	17.85
T <sub>7</sub>	15.67	18.91	15.47	18.99	17.00	19.13
T <sub>8</sub>	17.17	19.99	17.30	19.06	18.60	20.32
T <sub>9</sub>	17.93	18.88	17.17	20.07	28.80	22.13
T <sub>10</sub>	8.45	8.96	10.64	8.61	10.64	8.67
CD	5.199	1.996	3.191	2.15	2.01	3.608

Zinc concentration (mg kg<sup>-1</sup>) of leaf and petiole of fodder maize at 25DAS (Tab.35) showed that the different treatment means were found significant. For the 3<sup>rd</sup> leaf, T<sub>5</sub> recorded the highest value of 23.00 mgkg<sup>-1</sup> and was found to be on par



with T<sub>6</sub>, T<sub>4</sub>, T<sub>3</sub> and T<sub>9</sub> with values of 20.85, 19.83, 18.22 and 17.93 mgkg<sup>-1</sup> respectively. The lowest value was 8.45 mgkg<sup>-1</sup> (T<sub>10</sub>). For the 4<sup>th</sup> leaf, the highest value of 25.26 mgkg<sup>-1</sup> (T<sub>5</sub>) was found significantly superior. The lowest value was found in T<sub>10</sub> (8.96 mgkg<sup>-1</sup>). For the 5<sup>th</sup> petiole, T<sub>4</sub> recorded the highest value (19.70 mgkg<sup>-1</sup>). T<sub>6</sub>, T<sub>3</sub>, T<sub>8</sub> and T<sub>9</sub> were found to be on par while T<sub>10</sub> (10.64 mgkg<sup>-1</sup>) was found to be the lowest.

For the 3<sup>rd</sup> petiole, T<sub>5</sub> was recorded the highest and the T<sub>10</sub> recorded the lowest values with corresponding values of 24.64 and 8.61 mgkg<sup>-1</sup> and T<sub>5</sub> was found to be superior. For the 4<sup>th</sup> petiole, highest value of 28.88 mgkg<sup>-1</sup> was noticed in T<sub>9</sub> and was found significant difference over other treatment means while T<sub>10</sub> (10.64 mgkg<sup>-1</sup>) recorded the lowest value. The maximum value recorded for the 5<sup>th</sup> petiole was in T<sub>5</sub> (23.42 mgkg<sup>-1</sup>) while T<sub>9</sub> and T<sub>8</sub> were found on par with values of 22.13 and 20.32 mgkg<sup>-1</sup> respectively. T<sub>10</sub> recorded the lowest value of 8.67 mgkg<sup>-1</sup>.

#### 4.4.6.3 Zinc concentration (mg kg<sup>-1</sup>) of leaf lamina and petiole of fodder maize at 35DAS

Table.38. Zinc concentration (mgkg<sup>-1</sup>) of leaf and petiole of fodder maize at 35DAS

Treatments	Leaf lamina			Petiole		
	3 <sup>rd</sup>	4 <sup>th</sup>	5 <sup>th</sup>	3 <sup>rd</sup>	4 <sup>th</sup>	5 <sup>th</sup>
T <sub>1</sub>	17.17	17.90	20.07	19.40	19.91	16.38
T <sub>2</sub>	19.89	20.20	23.67	41.40	21.86	17.46
T <sub>3</sub>	22.50	29.80	32.16	24.30	33.20	17.20
T <sub>4</sub>	28.53	37.25	40.25	32.00	31.90	23.81
T <sub>5</sub>	33.33	38.20	41.51	44.30	41.50	32.00
T <sub>6</sub>	29.33	31.60	33.27	35.70	34.65	26.83
T <sub>7</sub>	28.04	26.00	31.00	21.00	24.80	16.48
T <sub>8</sub>	25.00	26.50	33.83	14.70	33.67	17.45
T <sub>9</sub>	28.40	35.00	35.77	46.20	28.08	18.21
T <sub>10</sub>	16.94	14.52	13.70	14.90	15.85	12.57
CD	5.177	4.13	8.819	NS	7.846	1.202

As seen from the Table. 38 the treatment means were found significant at 35 DAS except for 3<sup>rd</sup> petiole. T<sub>5</sub> and T<sub>10</sub> recorded highest and lowest mean values in all cases. For the third leaf, T<sub>5</sub> with the maximum value of 33.33 mgkg<sup>-1</sup> and T<sub>10</sub> recorded the lowest value (16.94 mgkg<sup>-1</sup>). While T<sub>6</sub>, T<sub>4</sub> and T<sub>9</sub> were found to be on par with values of 29.33, 28.53, and 28.40 mgkg<sup>-1</sup> respectively. In the 4<sup>th</sup> leaf, the highest value of 38.20 mgkg<sup>-1</sup> was noticed by T<sub>5</sub> while the lowest value was observed in T<sub>10</sub> (14.52 mgkg<sup>-1</sup>). T<sub>4</sub> and T<sub>9</sub> were found on par with T<sub>5</sub>. For the 5<sup>th</sup> leaf, T<sub>5</sub> with highest value of 41.51 mgkg<sup>-1</sup> and it was on par to T<sub>4</sub> (40.25 mgkg<sup>-1</sup>), T<sub>9</sub> (35.77 mgkg<sup>-1</sup>) and T<sub>8</sub> (33.83 mgkg<sup>-1</sup>) and the control plot recorded the lowest value (13.70 mgkg<sup>-1</sup>). The Zn content of 3<sup>rd</sup> petiole was not significant for various mean values while the highest and lowest mean values were 46.20 mgkg<sup>-1</sup> (T<sub>9</sub>) and 14.90 mgkg<sup>-1</sup> (T<sub>10</sub>) respectively. The maximum value for Zn content in the 4<sup>th</sup> petiole was 41.50 mgkg<sup>-1</sup> which was on par with T<sub>6</sub> (34.65 mg kg<sup>-1</sup>). T<sub>10</sub> recorded the lowest value of 15.85 mgkg<sup>-1</sup>. For the 5<sup>th</sup> petiole, T<sub>5</sub> was found to be significantly different with a value of 32.00 mgkg<sup>-1</sup> and control with the lowest value of 12.57 mgkg<sup>-1</sup>.

#### 4.4.6.4. Zinc concentration (mg kg<sup>-1</sup>) of leaf lamina and petiole of fodder maize at 45DAS

Table.39. Zinc concentration (mgkg<sup>-1</sup>) of leaf and petiole of fodder maize at 45DAS

Treatments	Leaf lamina			Petiole		
	3 <sup>rd</sup>	4 <sup>th</sup>	5 <sup>th</sup>	3 <sup>rd</sup>	4 <sup>th</sup>	5 <sup>th</sup>
T <sub>1</sub>	28.50	27.10	29.33	32.57	25.70	24.80
T <sub>2</sub>	31.20	29.27	32.27	28.90	29.00	25.30
T <sub>3</sub>	41.00	35.00	43.76	35.24	34.59	39.20
T <sub>4</sub>	45.50	38.50	45.46	41.60	47.20	39.90
T <sub>5</sub>	46.54	43.67	49.54	39.00	39.76	40.10
T <sub>6</sub>	45.30	36.64	43.87	40.80	45.70	36.20
T <sub>7</sub>	42.50	32.40	44.33	36.00	32.54	35.60
T <sub>8</sub>	41.76	34.25	45.57	38.50	42.74	33.20
T <sub>9</sub>	47.40	32.63	47.00	45.90	42.30	34.30
T <sub>10</sub>	15.78	17.93	21.53	15.20	18.80	16.10
CD	11.29	6.981	7.693	9.76	11.384	18.446

The data presented in the Table.39. The 3<sup>rd</sup>,4<sup>th</sup> and 5<sup>th</sup> leaf follows the same pattern for the Zn concentration . The highest and lowest values of 47.40 and 15.78 mgkg<sup>-1</sup>, 43.67 and 17.93 mgkg<sup>-1</sup>, 49.54 and 21.53 mgkg<sup>-1</sup> respectively for 3<sup>rd</sup> ,4<sup>th</sup> and 5<sup>th</sup> petiole and all other mean values except T<sub>1</sub>, T<sub>2</sub> and T<sub>10</sub> respectively found on par with T<sub>5</sub> .

The highest mean value of the 3<sup>rd</sup> petiole was recorded in T<sub>9</sub> (45.90 mgkg<sup>-1</sup>) and it was on par with T<sub>4</sub>(41.60 mgkg<sup>-1</sup>) ,T<sub>6</sub>(40.80 mgkg<sup>-1</sup>) T<sub>5</sub>(39.00 mgkg<sup>-1</sup>) and T<sub>8</sub>(38.50 mgkg<sup>-1</sup>)while the lowest was recorded in T<sub>10</sub>(15.20 mgkg<sup>-1</sup>). T<sub>4</sub> and T<sub>10</sub> recorded the highest and the lowest mean value o 47.20 and 18.80 mg kg<sup>-1</sup> respectively in the 4<sup>th</sup> petiole. But T<sub>6</sub>, T<sub>8</sub> ,T<sub>9</sub> and T<sub>5</sub> were found to be on par with T<sub>4</sub> and values were 45.70, 42.74 ,42.30 and 39.76 mgkg<sup>-1</sup> respectively. For the 5<sup>th</sup> petiole, maximum value (40.10 mgkg<sup>-1</sup>) has been observed in T<sub>5</sub> and all other treatment means except T<sub>1</sub>, T<sub>2</sub> and , T<sub>10</sub>. The lowest was in T<sub>10</sub> (16.10 mgkg<sup>-1</sup>).

## 5. DISCUSSION

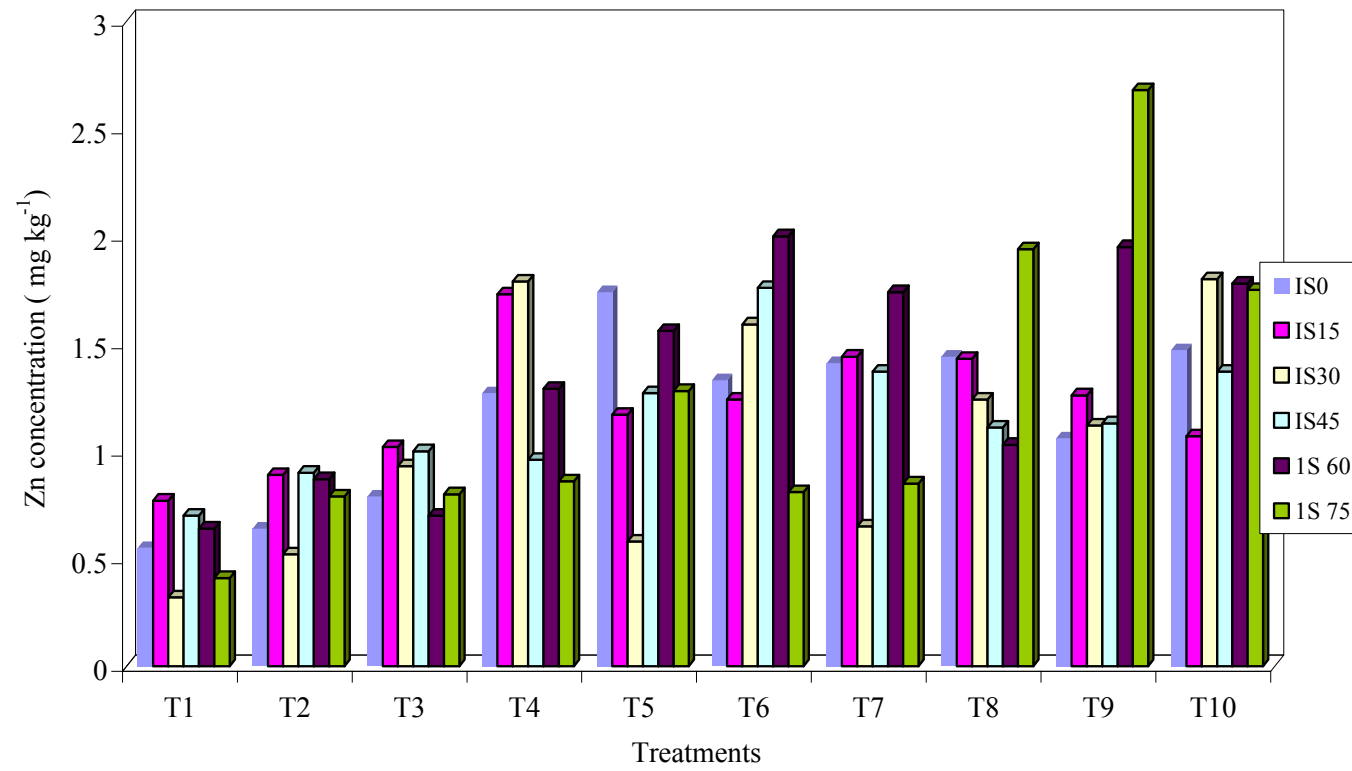
A study has been undertaken on the dynamics of Zinc in Typic Kandiusults with special reference to nutrition in fodder maize (*Zea mays* L.). This comprised of an incubation study to determine the release pattern of Zn from ZnSO<sub>4</sub> · 7H<sub>2</sub>O, a preliminary investigation on the Physico-chemical parameters especially Zn dynamics in soil profile and the surface and sub-surface soil of the experimental site, and a field study to find out the optimum requirement of Zn by the fodder maize cv. African Tall through soil and foliar application. The study also aimed to identify the index plant part of fodder maize for foliar diagnosis. The results of the experiments conducted are discussed below.

### 5.1. Study on the release pattern of Zn from ZnSO<sub>4</sub> · 7H<sub>2</sub>O on incubation with and without FYM

The medium range of available Zn proposed by FAO (Mikko Sillanpaa, 1990) was 0.7-2.4 mg kg<sup>-1</sup>. Data given in Table. 3 ie. the release status of Zn from Zinc sulphate indicate that when Zn was applied @ 5, 10 and 15 kg ha<sup>-1</sup>, the availability of Zn was in the medium range during the period of incubation and it has exceeded 2.4 mg kg<sup>-1</sup> on the 75<sup>th</sup> day of incubation only, when Zn was applied @ 20 kg ha<sup>-1</sup>.

The release pattern of Zn from zinc sulphate is graphically presented in Fig. 4.

On examining the data on the Zn release status during incubation, the Zn release was more at 5 and 10 kg ha<sup>-1</sup> of Zn application along with FYM. The chelation of inorganic sources of Zn with the organic compounds in farm yard manure might have increased the Zn availability. But as the levels of Zn application increased to 15 and 20 kg ha<sup>-1</sup>, the effect of FYM could not be much evidenced on the Zn release. This showed that at a higher level of Zn application the complexing of Zn with organic matter is immaterial (Shover *et al.* 2007).



**Fig. 4** Effect of different treatments with and without FYM on the availability of Zn

The incorporation of organic manures in soil is quite effective to mitigate Zn deficiency in crops. Organic manures may serve as a source of Zn and their decomposition products give rise to natural complexing agents which mobilize the native Zn already present in soil through chelation (Sakal, 2001).

## **5.2. Study of the soil profile**

The photograph of the soil profile examined is given in plate-1.

### **5.2.1. Morphological characteristics of the soil profile**

The solum was very deep (10.123 cm). Five horizons from Ap to Bt<sub>2</sub> (Table-4) were identified in the solum. The soil colour varied from 5 YR 3/4 in the Ap horizon to 5 YR 4/8 in the Bt<sub>2</sub> horizon. The soil colour appears to be the function of different chemical and mineralogical composition as well as textural make up of soil and conditioned by topography and moisture regime (Walia and Rao, 1997).

The texture varied vertically from sandy loam to sandy clay loam. The textural variation might be due to variation in the parent material, weathering and translocation of clay. The structure of the upper horizon was crumb and this might be due to continuous addition of organic matter through vegetation. The lower horizons had a subangular blocky structure and this was attributed to the presence of higher quantities of clay fraction (Sharma *et al.* 2004).

The consistence of the soil were dry, friable, non-sticky and non-plastic in the upper horizons and as the depth increases, at the B2t and C<sub>1</sub> horizons it was slightly sticky and slightly plastic. The consistence of the soils viz. non-sticky to slightly sticky and non-plastic to slightly plastic might be due to negligible or very small amount of expanding clay minerals (Thangasamy *et al.* 2004).

The horizon boundaries were abrupt wavy to gradual smooth in topography. The roots in different horizons of the pedons were fine to coarse in size and more in surface layer and decreased with depth.

### **5.2.2. Physico-chemical characteristics of soil profile**

The detailed physico-chemical characteristics of the soil profile are presented in Table 5. Particle size analysis of the different horizons from Ap, Ac, B<sub>1</sub>, B<sub>2</sub>, Bt<sub>2</sub> and C<sub>1</sub> was carried out. In the Ap horizon the sand content was 79.4 per cent and silt and clay contents were 11.5 and 11.6 per cent respectively and classified as sandy loam. The Ac, B<sub>1</sub> and B<sub>2</sub> horizons also could be classified under sandy loam, as the sand content varied from 50-80, silt 0-50 and clay 0-20 per cent. For the Bt<sub>2</sub> silt and clay content estimated were 48.9, 10.7 and 34.52 per cent. The C<sub>1</sub> horizon was observed with a sand, silt and clay contents of 51.6, 9.01 and 36.2 per cent respectively and classified under the textural class sandy clay loam. The Bt<sub>2</sub> and C<sub>1</sub> horizons contained 34.5 and 36.2 percentage of clay and the increase in clay content could be attributed to vertical migration or translocation of clay (Sarkar et al. 2002).

The bulk density of the C<sub>1</sub> horizon (1.35 Mgm<sup>-3</sup>) was the highest, which might be due to more compaction, low organic matter and less aggregation. Similar findings were reported by Singh and Agrawal (2005).

The pH of the soil varied from 5.4 in the upper horizon to 4.8 in the lower horizon and the EC did not varied much among horizons.

### **5.2.3. Nutrient content of soil profile (Table. 5)**

The organic carbon percentage varied from 0.79 in the Ap horizon to 0.35 in the C<sub>1</sub> horizon. This decreased trend in organic carbon content with depth could be attributed to the addition of plant residues and farm yard manure to surface horizons.

The nutrient content of N, P, K, Ca, Mg and S steadily decreased from the Ap to C<sub>1</sub> horizon.

The higher availability of Nitrogen and Phosphorus and Sulphur in the surface horizons and steady decrease with depth may be due to decreasing trend of organic carbon with depth, cultivation of crops which is mainly confined to the surface horizon only and also due to supplementation of depleted nitrogen and phosphorus content by the external addition of fertilizers during crop cultivation (Prasuna Rani *et al.* 1992).

The highest available potassium content in the surface horizon might be attributed to more intense weathering, release of labile K from organic residues, application of K fertilizers etc. Available K had a significant and positive correlation with organic carbon (Thangasamy *et al.* 2005). But the micronutrients viz. Fe, Mn, Zn and Cu expressed a different pattern. The vertical distribution of Fe, Zn and Cu contents exhibited little variation from Ap to C<sub>1</sub> horizon. Distribution of total Zn with depth down the soil profile has been reported to be uniform (Lindsay, 1972).

The Mn content increased from 9.7 mg kg<sup>-1</sup> in Ap horizon to 18.94 mg kg<sup>-1</sup> in the C<sub>1</sub> horizon.

The water soluble + exchangeable form of Zn was maximum i.e., 0.41 mg kg<sup>-1</sup> in the Ap horizon (0-9.5 cm) followed by 0.32 mg kg<sup>-1</sup> in the Ac horizon (9.5 – 25 cm). According to Hazra and Mandal (1996), the availability of added zinc is directly related to the amount of water soluble + exchangeable fractions of Zn in soil. The added organic manures and fertilizers to the surface soil might have contributed to the highest content of the W<sub>s</sub>+Ex-Zn.



### 5.3. Field study – Fodder maize cv. African Tall

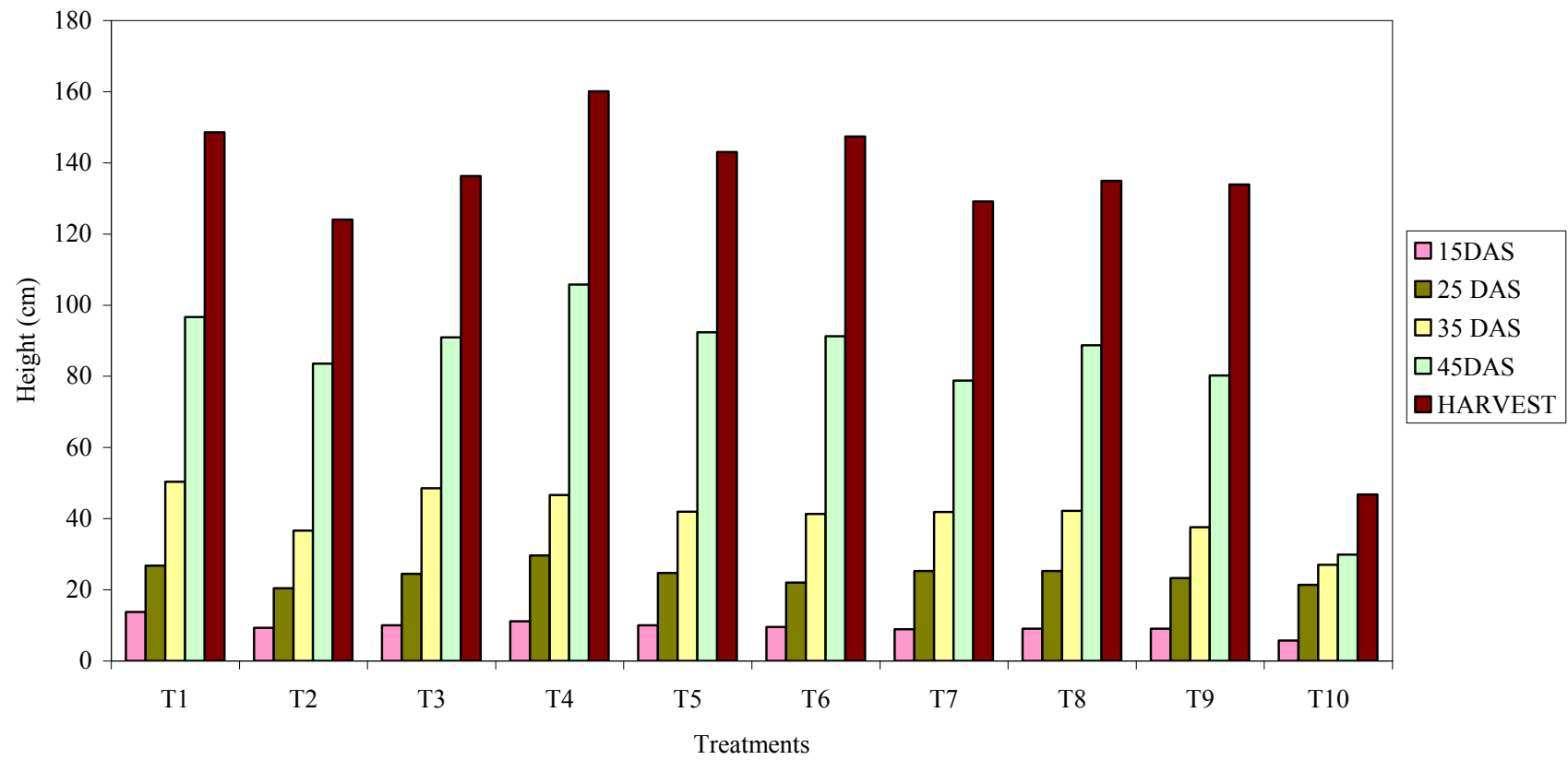
#### 5.3.1. Height (cm) and different stages of crop growth

The data presented in Table-7 , 20 and Fig. 5 revealed that at the different stages of observation viz. 25 and 35 DAS and at harvest the T<sub>4</sub> (FYM 10 t ha<sup>-1</sup> 120:60:40 NPK kg ha<sup>-1</sup> and 10 kg Zn ha<sup>-1</sup> as ZnSO<sub>4</sub> 7H<sub>2</sub>O) recorded the maximum height of the plant. At 45 DAS, even though T<sub>5</sub> gave the maximum height which was on par with T<sub>4</sub>. Tsui (b) (1948) investigated that Zn have an active role in the production of auxins for a plant species. The Zn plays a pivotal role in regulating the auxin concentration in plant and might have contributed to the increased height. Similar findings were reported by Dewal and Parek (2004) and Jain and Dahama (2006).

For the 2<sup>nd</sup> crop also, the T<sub>4</sub> treatment recorded the maximum height on the 45<sup>th</sup> DAS and at harvest. For the 1<sup>st</sup> crop 45 DAS and for the 2<sup>nd</sup> crop 35 DAS, a boost on height was noticed. The favourable climatic condition viz. rainfall at the active vegetative growth of the crop have influenced this enhanced height. The first experimental crop has been sown during the last week of October, 2007 and the total rainfall in mm recorded for the month of November, 2007 was 186.20. Like that the second experimental crop was raised during the first week of May, 2008 and the rainfall observed was 153.20 mm. This evidently established the impact of rainfall during the active vegetative growth of the crop and enhancement in height. The meteorological data is given in Fig.-2.

#### 5.3.2. Mean number of leaves at different stages of crop growth

At the time of harvest the first crop recorded maximum production of leaves in T<sub>9</sub> (POP+0.75% Zn given as foliar) and it was on par with T<sub>3</sub>. (Table-8). As zinc plays an important role in chlorophyll synthesis, the photosynthetic rate might have increased, resulted in the maximum production of leaves. Similar findings on foliar spray of mustard crop with 1% ZnSO<sub>4</sub> (Sushama *et al.* 2003) and on pigeon pea with

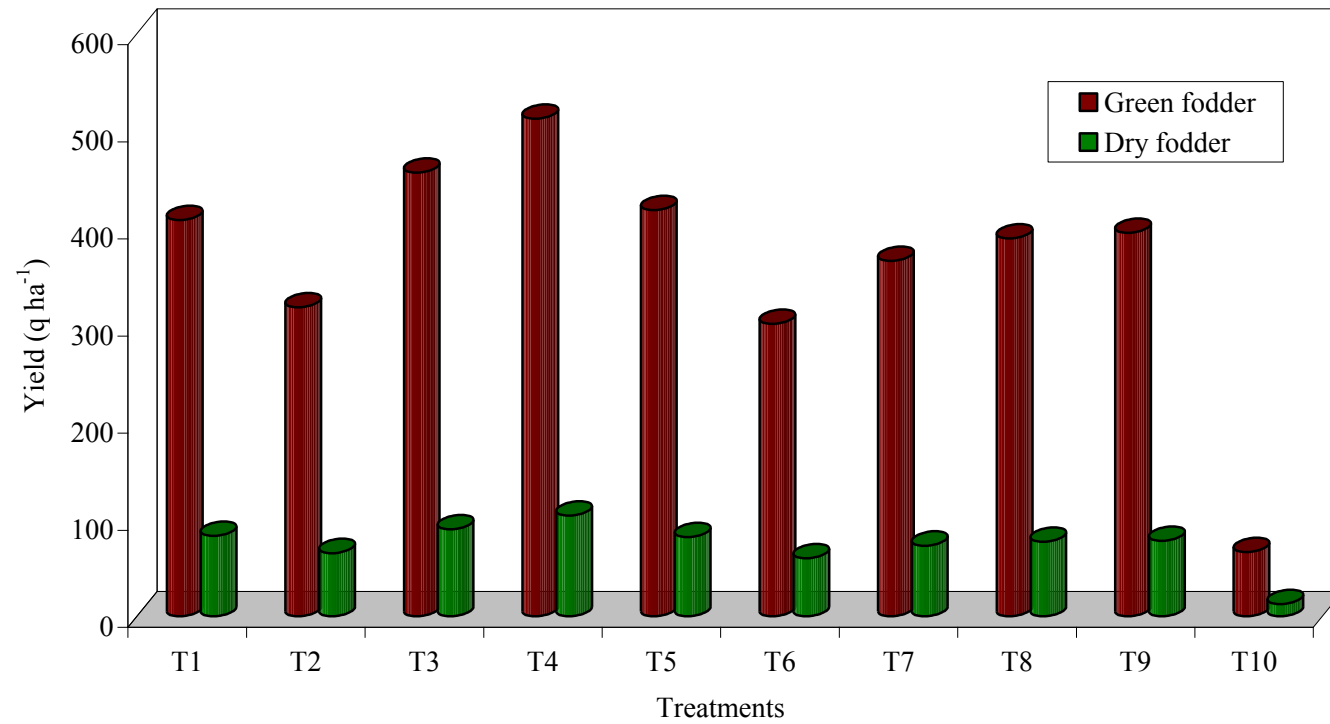


**Fig. 5. Mean values of height (cm) at different stages**

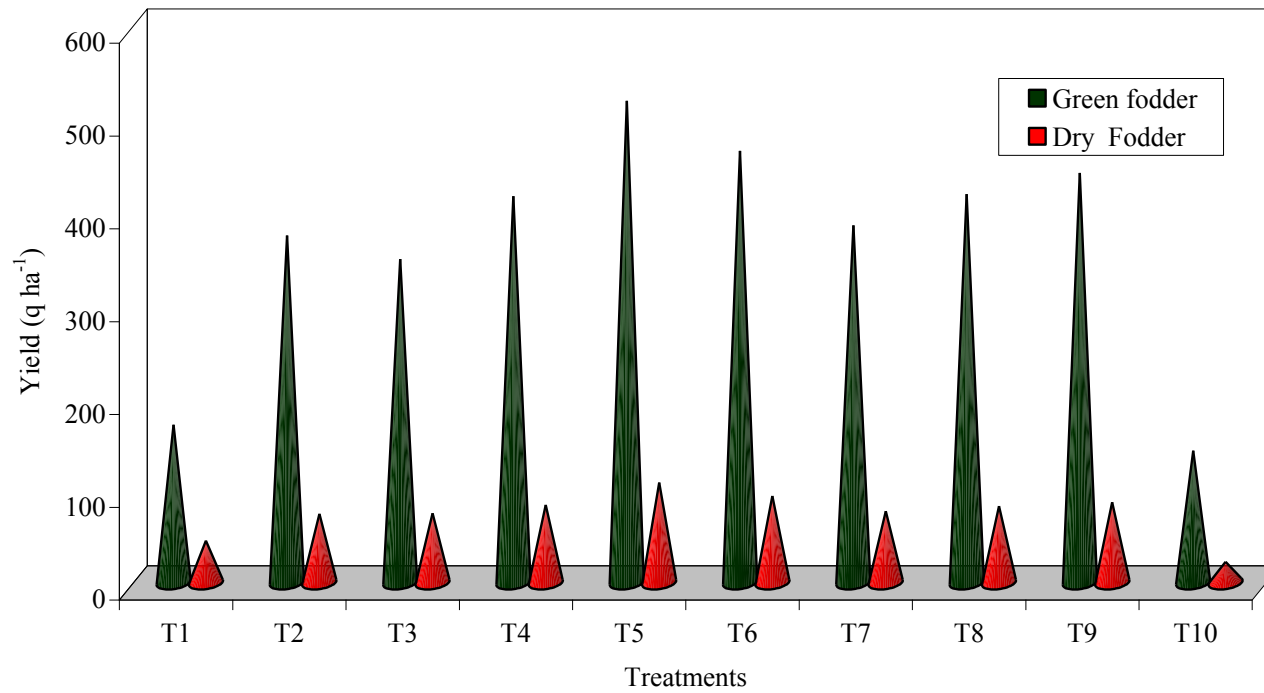
0.5% ZnSO<sub>4</sub> (Dube *et al.* 2001; Verma *et al.* 2004) were reported. At the time of harvest the number of leaves produced among treatments were not significant for the 2<sup>nd</sup> crop. . At 45 DAS T<sub>8</sub> and T<sub>9</sub> ie, Zn foliar applied treatments on the 4<sup>th</sup> leaf stage produced the maximum number of leaves, next to T<sub>3</sub>, the maximum leaf produced treatment (Table-21).

### 5.3.3. Yield and yield attributes of fodder maize

The highest green fodder yield (512.5 qha<sup>-1</sup>) was recorded in T<sub>4</sub> (POP + Zn@10 kg ha<sup>-1</sup>) and was found superior to all other treatments. The per day green fodder yield and LAI were also highest in T<sub>4</sub>. (Table-9 and Fig. 6&7 ). Not only there is inherent variability in the behaviour of soils with respect to nutrient availability but also the crops grown influence the transformation of nutrients and their availability. Sakal 2001 also reported that Zn application varied widely because of the differences in soil characteristics, available Zn status of soils and crop varietal characteristics. All these factors might have influenced the fodder production and the present study revealed that an application rate of organic and inorganic nutrients @10t FYM + 120:60:40 NPK kg ha<sup>-1</sup> + 10 kg Zn ha<sup>-1</sup> as soil application was the best treatment for fodder maize cv. African Tall. The optimum level of Zn for different crops ranged between 5-10 kg ha<sup>-1</sup>. The favourable influence of ZnSO<sub>4</sub> on the higher yield of fodder may be attributed to its role in various enzymatic reactions, growth processes, hormone production and protein synthesis (Mehla, 1999). Data presented in Table-22 revealed that the yield and yield attributes of second crop were significant except for LSR. The green fodder yield was maximum in T<sub>5</sub> (Soil application of Zn @ 15 kg ha<sup>-1</sup> + POP) with a value of 519.3 q ha<sup>-1</sup> and T<sub>6</sub> was found on par with an yield of 465.4 q ha<sup>-1</sup>. As in the case of first crop for the second crop too, the maximum LAI recorded treatment had given the maximum green fodder yield. For the first crop, the highest LSR was noted in T<sub>4</sub>, but for the second crop, the leaf stem ratio was found to be non-significant among treatment.



**Fig.6 Green and dry fodder yield ( $q\ ha^{-1}$ ) of fodder maize of first crop**



**Fig. 7. Green and dry fodder yield ( $q\ ha^{-1}$ ) of the second crop**

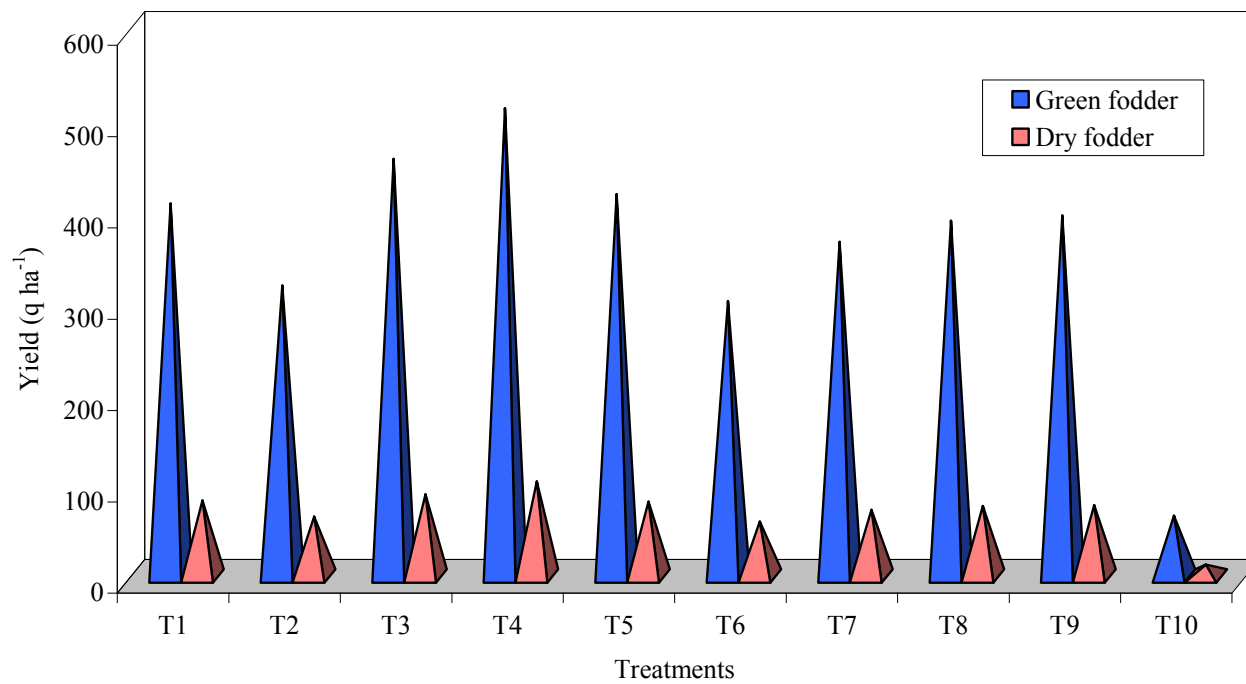
#### 5.3.4. Effect of Zn application on the yield of fodder maize

On examining the yield data of the 1<sup>st</sup> experimental crop, the T<sub>4</sub> treatment i.e. POP+10 kg Zn ha<sup>-1</sup> as soil application recorded the highest yield of 512.5 q ha<sup>-1</sup> (Table-9). For the 1<sup>st</sup> residual crop the treatment that received Zn @ 15 kg Zn ha<sup>-1</sup> showed the highest yield (Table-19).

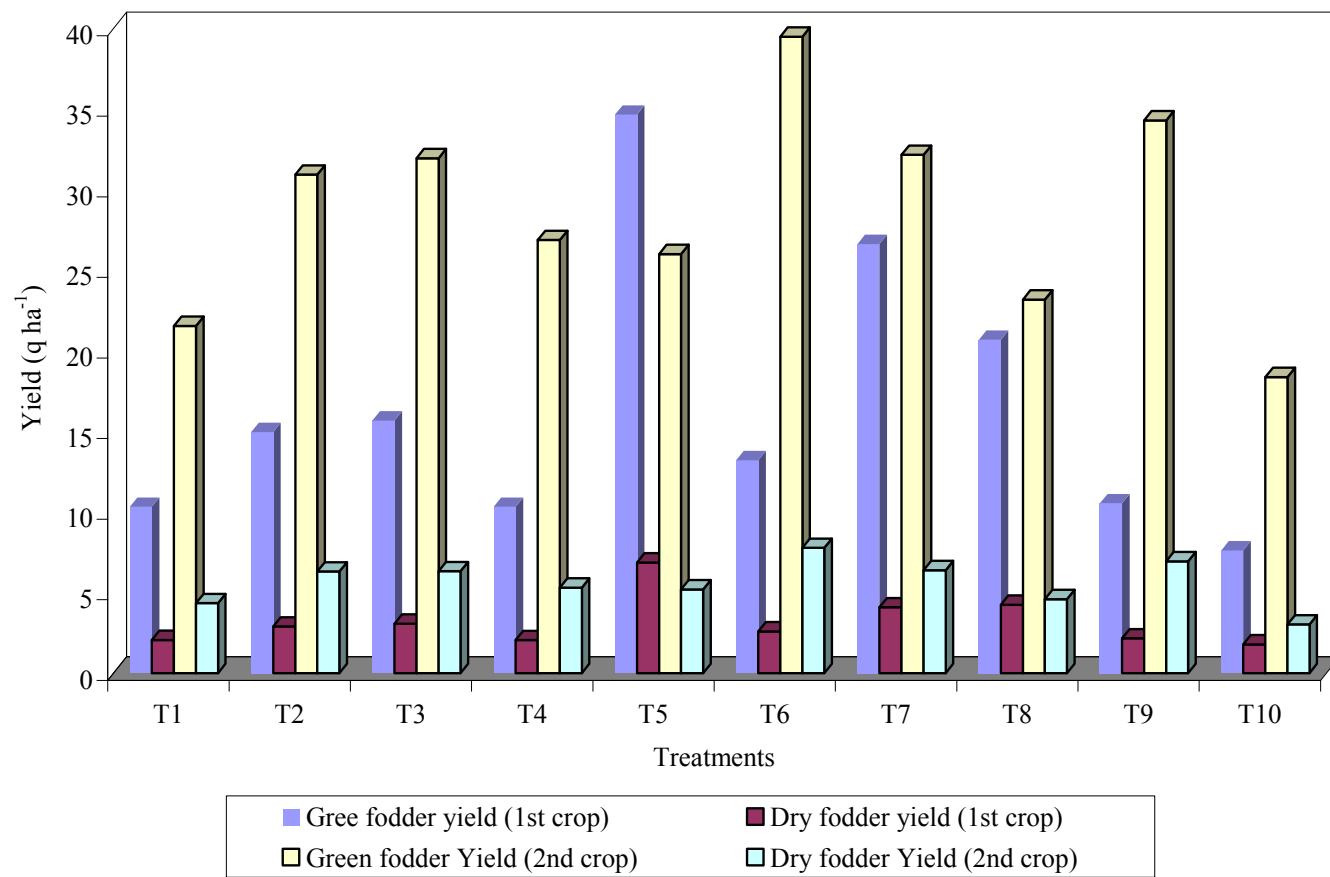
In the case of 2<sup>nd</sup> experimental crop the T<sub>5</sub> treatment that received 15 kg Zn ha<sup>-1</sup> recorded the highest yield (519.30 q ha<sup>-1</sup>) and it was on par with T<sub>6</sub>. The yield of the fodder maize due to the foliar application of Zn leaf at 4<sup>th</sup> leaf stage showed a steady increase from the lowest to highest level of foliar spray given and the trend was same for both the experimental crops raised (Table 9 and 23).

The yield data of the 1<sup>st</sup> and 2<sup>nd</sup> experimental crop were pooled and analyzed statistically ( Fig. 8). The pooled analysis data for the yield was found significant for the Zn applied treatments. Katyal *et al.* (1999) emphasized the role of Zn in increasing yield of pearl millet. For the pooled data, the T<sub>5</sub> with a mean value of 468.8 q ha<sup>-1</sup> showed the highest yield and it was on par with T<sub>4</sub>, T<sub>8</sub> and T<sub>9</sub> which recorded a yield of 465.5 q ha<sup>-1</sup>, 403.90 ha<sup>-1</sup> and 418.30 q ha<sup>-1</sup> respectively. The T<sub>8</sub> and T<sub>9</sub> were the foliar applied plots. It is seen that the yield difference between T<sub>4</sub> and T<sub>5</sub> was only 4.3 q ha<sup>-1</sup> a meager difference only (Table . 35).

For the of 1<sup>st</sup> residual crop, the plot that received 15 kg Zn ha<sup>-1</sup> and for the 2<sup>nd</sup> residual crop, the plot that received 20 kg Zn ha<sup>-1</sup> recorded the maximum yield (Fig. 9). But for the experimental crops the highest yields were at 10 kg Zn ha<sup>-1</sup> and 15 kg Zn ha<sup>-1</sup> for the 1<sup>st</sup> and 2<sup>nd</sup> crop respectively. From this it is presumed that the plants haven't utilized fully the added Zn fertilizer at these higher doses during the year the fertilizer was applied. This may be due to the tendency of the soil organic matter to chelate the added zinc or may be due to the slow rate of movement of this element to plant roots in the soil.



**Fig. 8** Pooled analysis data on green and dry fodder yield ( $q\ ha^{-1}$ )



**Fig.9 . Green and dry fodder yield (q ha<sup>-1</sup>) of residual crops**



Continuing application of Zn fertilizers in time tends to increase the level of this nutrient in the soil and particularly its level in the labile forms which can release Zn in the soil solution. Thus even though much of the Zn added in fertilizers was not used during the year of application it may provide an important source in future years.

From the above results the following conclusion could be drawn.

For raising a single crop of fodder maize var. African Tall, soil application of Zinc sulphate @10kg Znha<sup>-1</sup> along with pop ie. 10t ha<sup>-1</sup> FYM and NPK 120:60:40 kg ha<sup>-1</sup> is being recommended. The increase in yield caused by FYM with Zn could be attributed partly to the presence of Zn in FYM itself and also to the chelation effect of organic acids formed during decomposition on both native and applied Zn. One common characteristic of all the micronutrients is that they are required in very small amounts. Also, they are all harmful when the available forms are present in the soil in larger amounts that can be tolerated by plants or by animals consuming the plants. Thus the range of concentration of these elements in which plants will grow satisfactorily must not be too great. Irrespective of the source, grain yield and zinc uptake significantly increased upto 10kg Zn ha<sup>-1</sup> (Prasad and Umar,1993). Application of 15kg ZnSo<sub>4</sub>ha<sup>-1</sup> as soil application along with POP for the 1<sup>st</sup> crop will help to skip the Zn application of the succeeding crop. The profitability of Zn application in conjunction with FYM could be higher if the carry over benefits of a single application to succeeding crops are taken into account. Residual carry over of available zinc varies from slight to moderate, increasingly as soils become less alkaline.

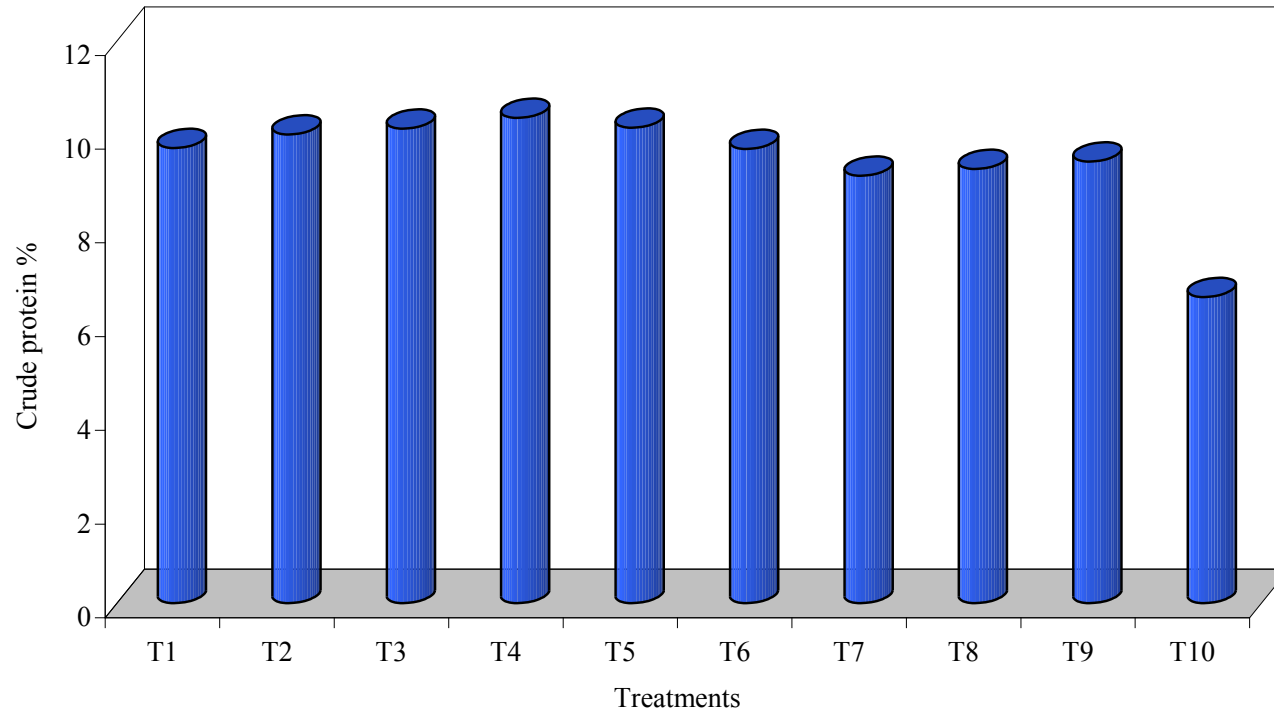
The foliar application @0.75% ZnSo<sub>4</sub> is recommended at 15 DAS. The 0.75% ZnSO<sub>4</sub> as foliar spray at 15 DAS recorded the highest yield among foliar application treatments and it was on par with T<sub>4</sub>. Foliar spray of zinc sulphate was an effective emergency method but in highly zinc deficient soils it did not compare well with soil application of zinc. (Katyal and Rattan, 2003)

### 5.3.5. Quality attributes of fodder maize

Regarding the quality aspects of first crop maximum crude protein percentage and crude protein yield was also recorded for T<sub>4</sub>, where maximum fodder yield has been obtained. The lowest crude fibre was obtained in T<sub>9</sub> (23.1%), but it was on par with T<sub>4</sub> (23.7%) (Table-10 and Fig. 10). For the second crop (Table-23) the highest crude protein was for T<sub>3</sub> but it was on par with T<sub>4</sub>. Considering the quality aspects viz. increased crude protein and reduced fibre, the T<sub>4</sub> treatment could be identified as the best treatment. Forage with less fibre and more protein was relished by the cattle. Crude fibre consists largely of cellulose and lignin (97%) plus some mineral matter. The crude fibre content is commonly used and measure of the nutritive value of livestock feeds. In addition to the nutrition of the crop, the genetic make up and potentiality of the African Tall variety also might have influenced the better performance of the variety. The increase in protein content due to zinc addition might be due to its involvement in the N metabolism of the plant (Frescenko and Lozek, 1998). Zn activates enzymes in protein synthesis. The higher crude protein was due to the high nitrogen content (Evans and Sagar, 1966).

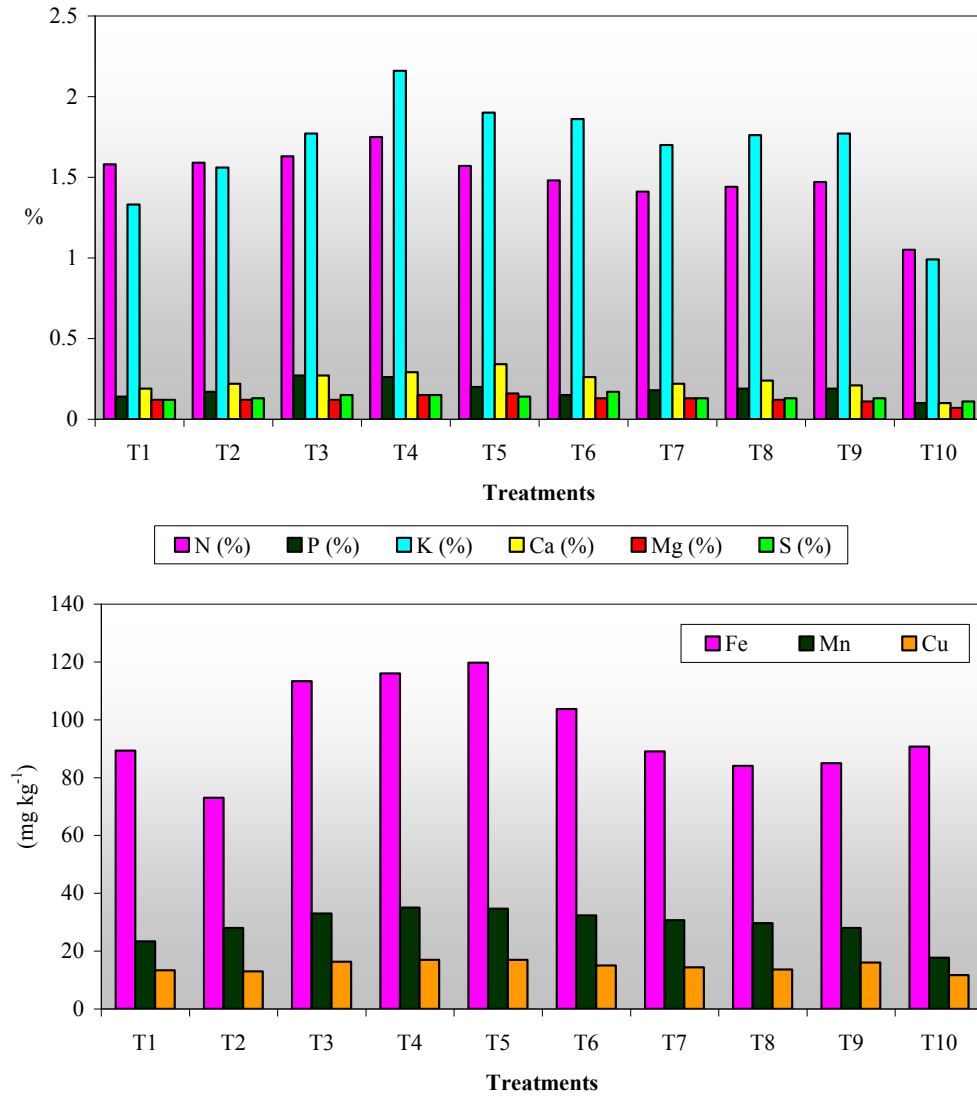
### 5.3.6. Nutrient concentration in fodder maize

The N, P, K, Ca, Mg, S and other micronutrient contents showed significant variation among treatments. In the case of N,P,K, Fe, Mn, Zn and Cu, T<sub>4</sub> showed the highest or statistically on par with the highest, nutrient concentration in the plant (Table-11 and Fig.11 & 12). Increased uptake of nutrients due to the application of Zn to maize was reported earlier (Hulagur and Dangarwal, 1983). But Ca and Mg recorded the highest concentration in T<sub>5</sub> but in the case of Mg it was on par with T<sub>4</sub>. At the highest level of Zn application ie at 20 kg Znha<sup>-1</sup>, the Cu content was found to be decreased. Plant roots appear to absorb Zn and Cu by the same mechanism. This might have caused interference in the uptake of Cu, when Zn was in excess.

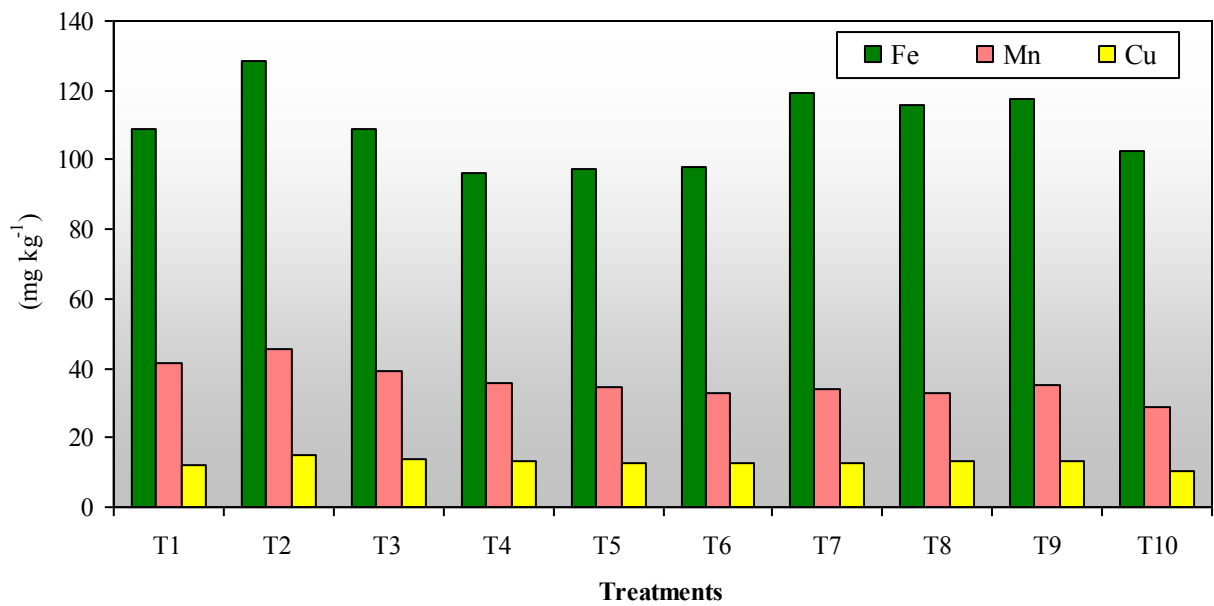
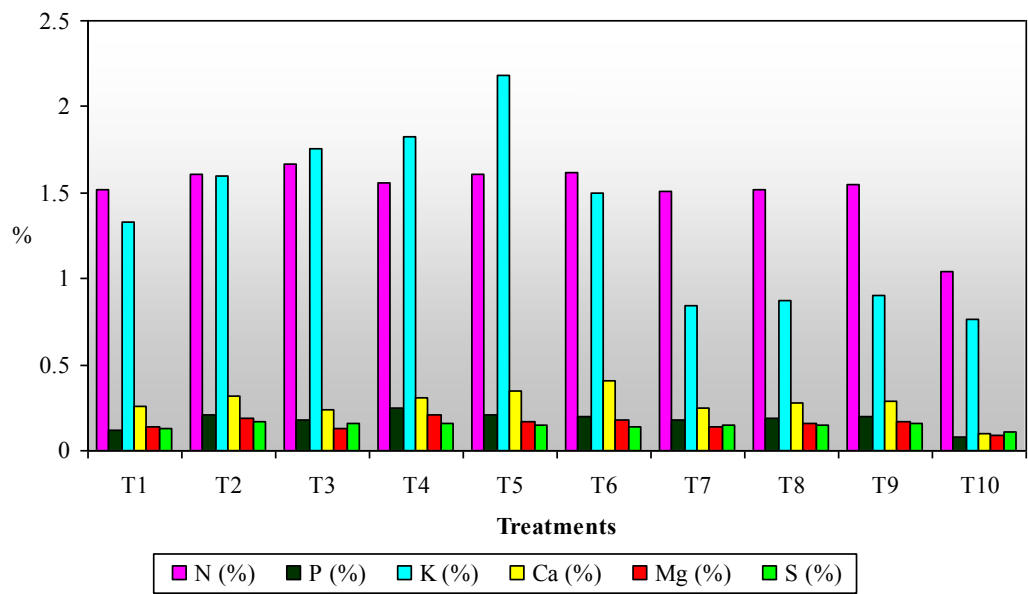


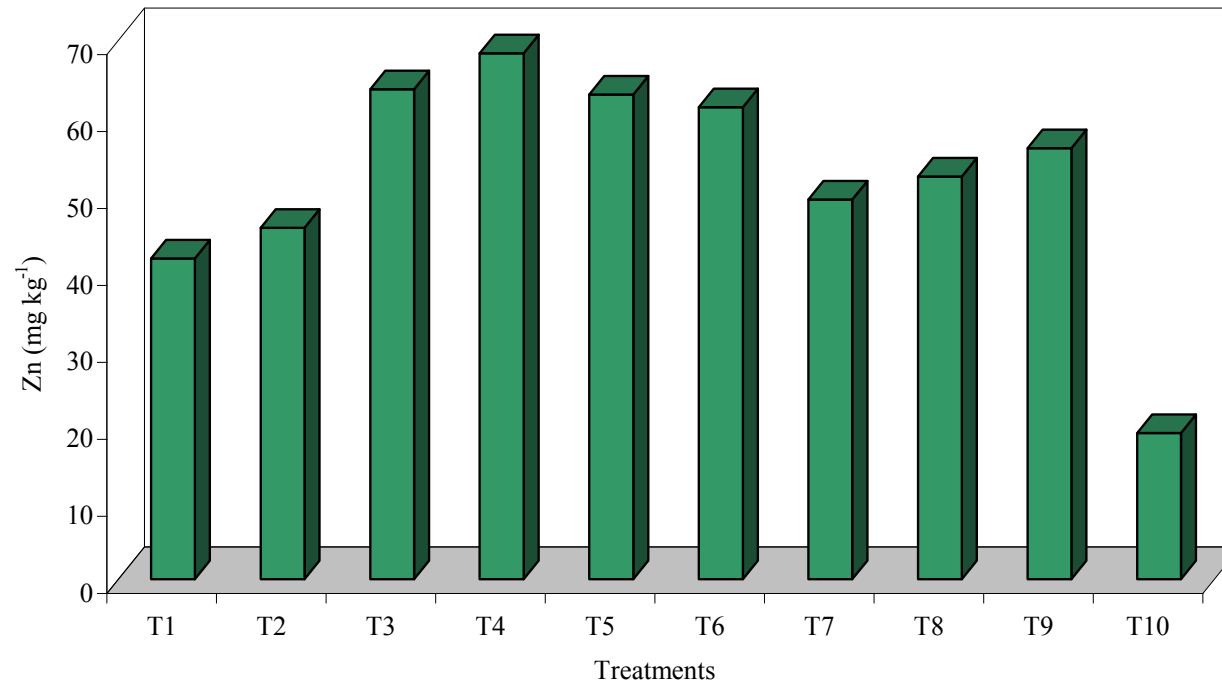
**Fig. 10 Mean value of crude protein content of 1<sup>st</sup> and 2<sup>nd</sup> crop**

**First crop**



**Fig. 11. Total nutrient concentration in the fodder maize**





**Fig. 12 Total Zn concentration in the plant**

For the second crop also (Table-24) the Zn applied treatments (soil application) recorded the maximum and comparable N content in plants.

In the control plot, the concentration of nutrients in the plants, were minimum, the low N availability might have decreased the vigour of plants to an extent that it affects the uptake of other nutrients.

### **5.3.7. Physico-chemical properties and nutrient content of the soil after the crop growth**

After the crop growth (Table-12 & 26, and Fig. 13 & 14), a reduction in the Nitrogen and phosphorus content was noticed in the soil. But the potassium content has been increased from the initial level. The release of non-exchangeable K might have contributed to this increased content. The crop requirements were partly met from the released K and both the applied K and released K caused available K build up in the soil. The differential release pattern of non-exchangeable K from the soil reserve besides variation in K uptake by the crop was also responsible for differences in the available K status in soil (Yaduvanshi et al. 1985). The experimental area represents more of coarse fractions of sand and that also might have influenced the K availability. Pasricha (2002) reported that coarse fractions of sand and soil in many soils can be an important source of K for crop plants. Studies on the release of K from sand fractions showed comparable or slightly higher release of K in soils.

In the case of available P, maximum content of  $16.57 \text{ kg ha}^{-1}$  after the 1<sup>st</sup> crop was noticed for the treatment where Zn @  $5 \text{ kg ha}^{-1}$  has been applied. As the level of Zn application increased the P content was found to be decreased. The antagonistic relationship of P and Zn was reported by many scientists (Orabi *et al.* 1985 :Barrow, 1987; Lonaragan and Webb ,1993). Much variation after the crop growth has not been showed for the nutrients viz. Ca, Mg, S, Fe and Cu (Table 14

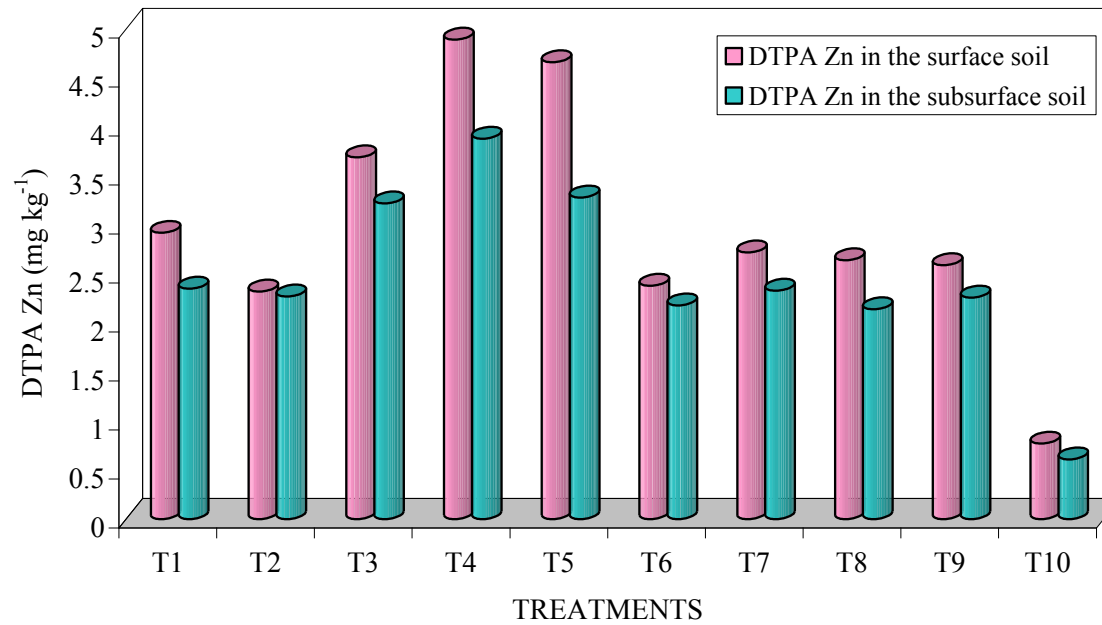
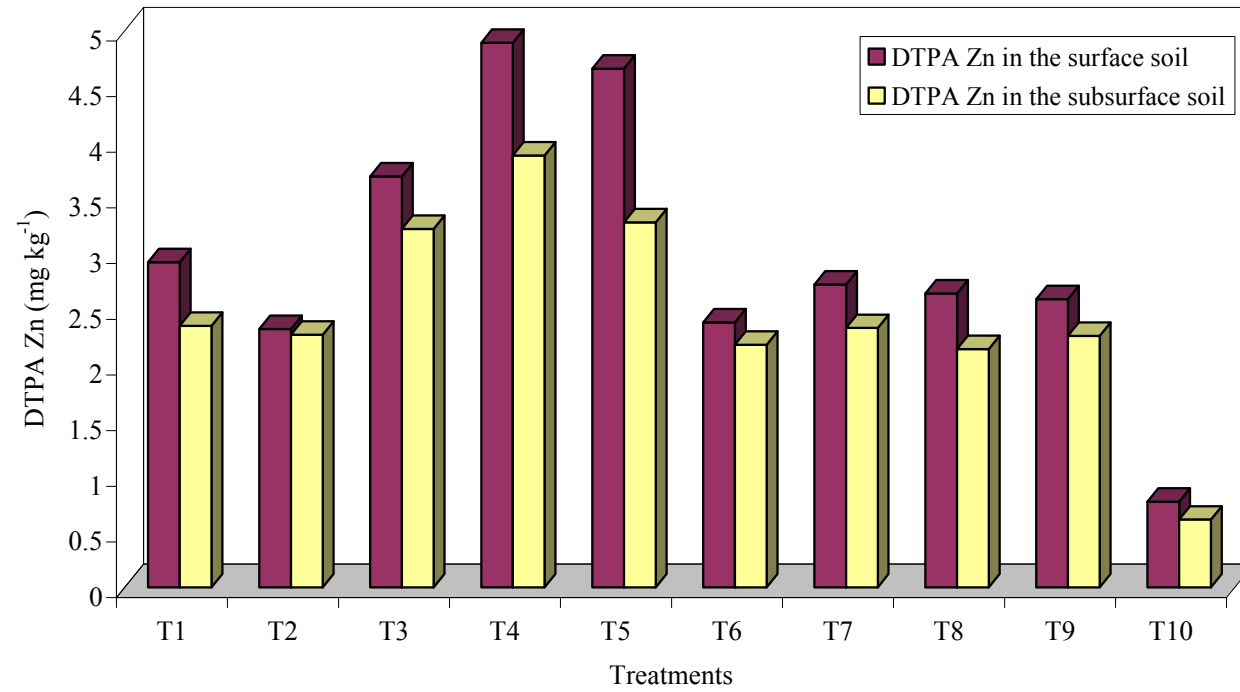


Fig.13 DTPA Zn in the surface and subsurface soil after the first crop





**Fig.14. Concentration of Zn in the surface and subsurface soil after the second crop ( mg kg<sup>-1</sup>)**

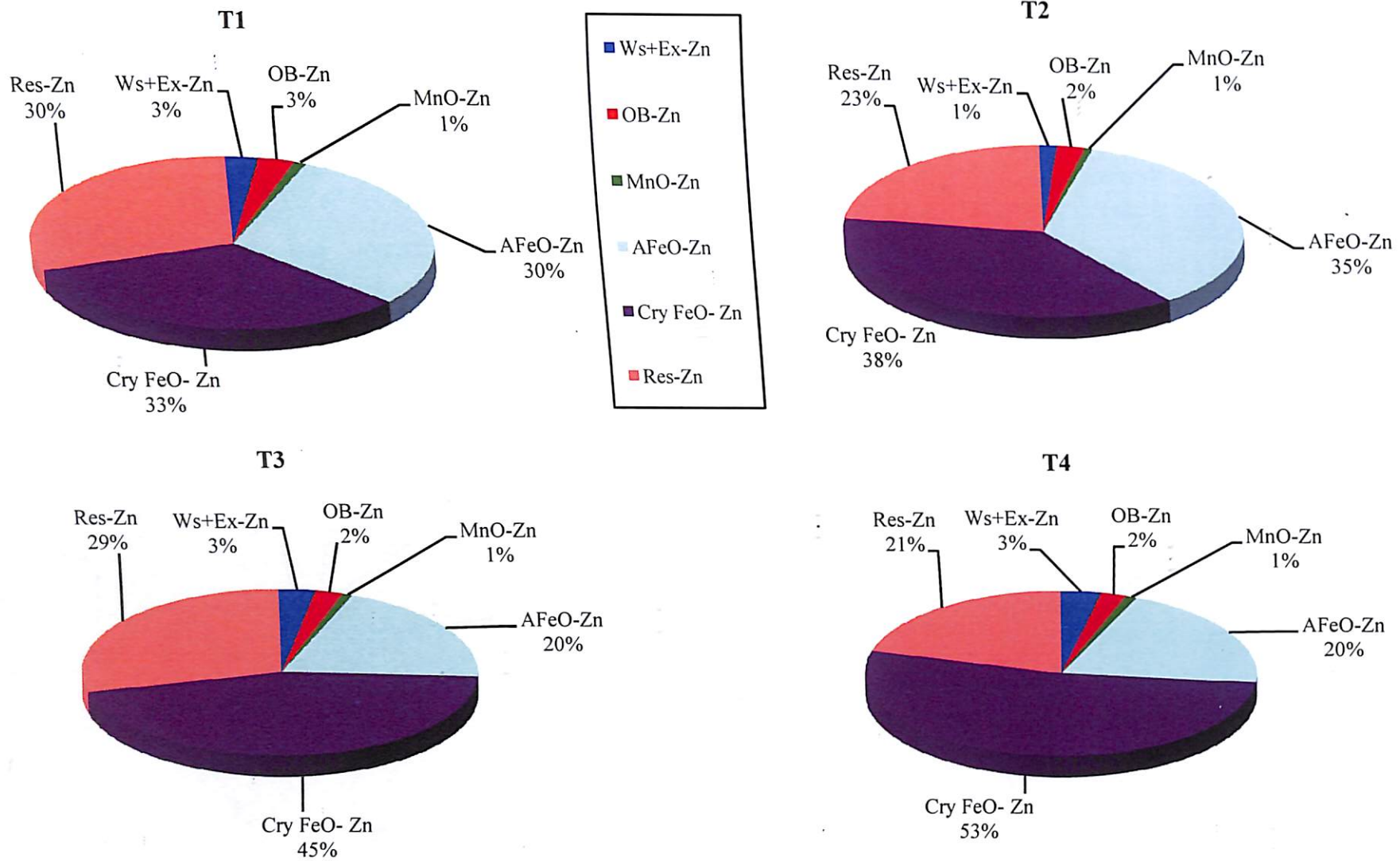
and 28). Zn content got increased in the Zinc applied as well as farm yard manure applied plots. The applied Zn nutrient as well as the chelated Zn in the organic matter might have contributed to this. A reduction in the Mn content was noticed after the crop growth. Similar findings were reported by Lombanaes and Singh (2004).

### **5.3.8. Zinc fractions of soil after the crop growth**

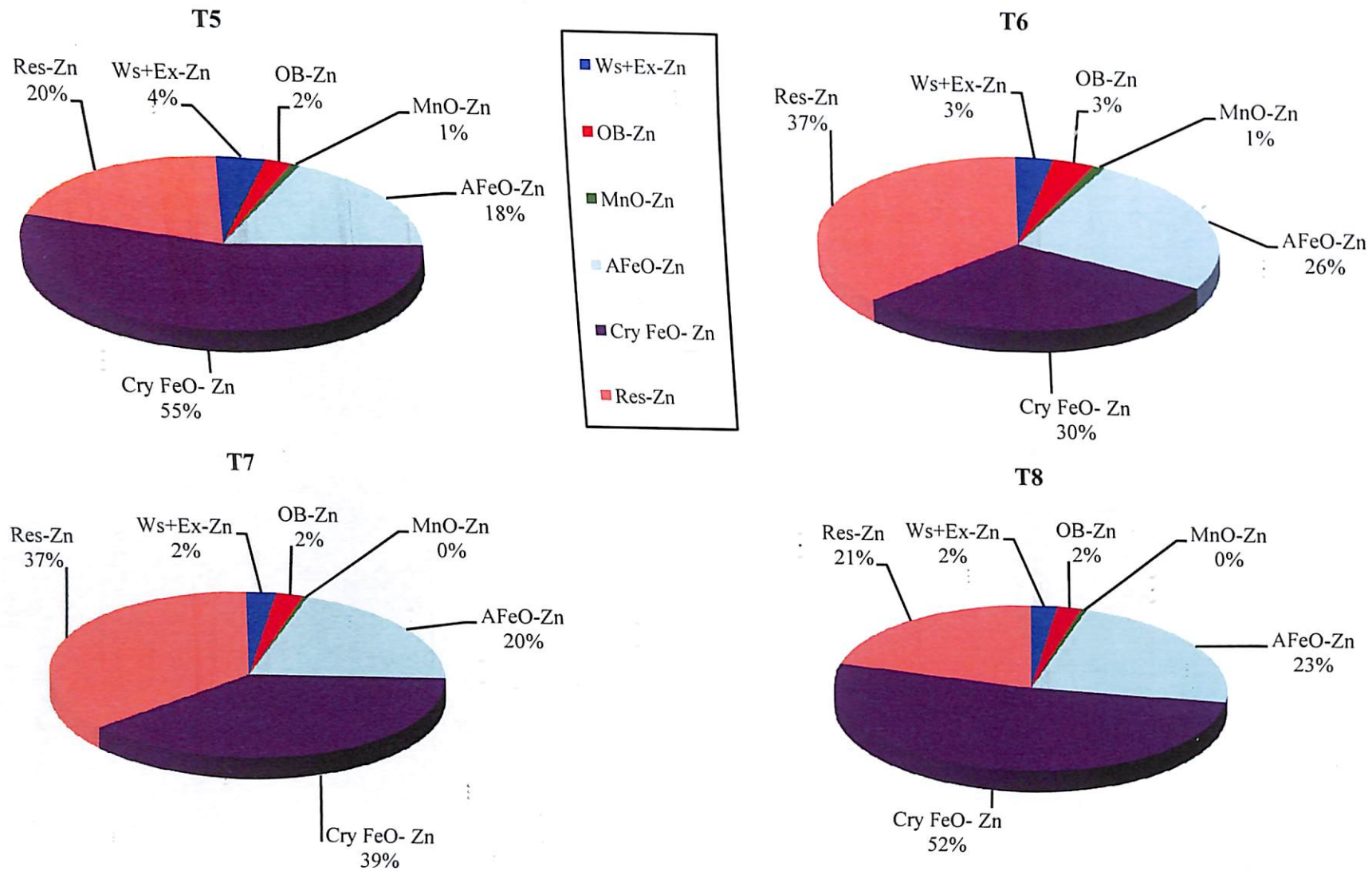
The data on the different fractions of Zn in the surface soil (Table 15 & 29 and Fig. 15 & 16) revealed that the Ws+Exch. Zn was maximum in T<sub>9</sub> and it was found significantly superior to other treatments. Actually these Ws+Exch-Zn. Fractions contributes to the available Zn and that might not have been utilized by the plant and reflected in the significantly lower yield. In T<sub>4</sub>, where the highest yield (512.5 qha<sup>-1</sup>) obtained plot recorded a significantly lower content of Ws + Exch. Fraction of Zn. Randhawa and Singh (1995) reported that exchangeable Zn contributed to the pool of available Zn and thus played a significant role in the Zn nutrition of maize.

The percentage contribution of different fraction of Zn to the total Zn in the surface soil, after the growth of first crop varied from 0.34 to 4.08 Ws+Ex- Zn, 1.37 to 3.89 OB- Zn, 0.11 to 0.83 MnO-Zn, 16.95 to 38.96 Am FeO-Zn, 20.91 to 61.45 Cry Feo-Zn, 7.92 to 48.49 Res- Zn and for the subsurface soil, it varied from 0.11 to 2.59 Ws+Ex- Zn, 0.36 to 2.36 OB- Zn, 0.42 to 1.70 MnO-Zn, 13.58 to 30.24 Am FeO-Zn, 25.51 to 58.69 Cry Feo-Zn, 19.19 to 55.78 Res- Zn. The range values of different fractions showed that all the fractions, except Mn O-Zn were lower in the surface soil.

After the growth of the second crop also, the soil samples were collected from the surface and subsurface and analyzed the different Zn fractions. The percentage contribution of the different fractions to the total Zn varied from 0.72 to 4.83 Ws+Ex- Zn, 1.49 to 3.76 OB- Zn, 0.31 to 1.12 MnO-Zn, 10.01 to 43.56 Am

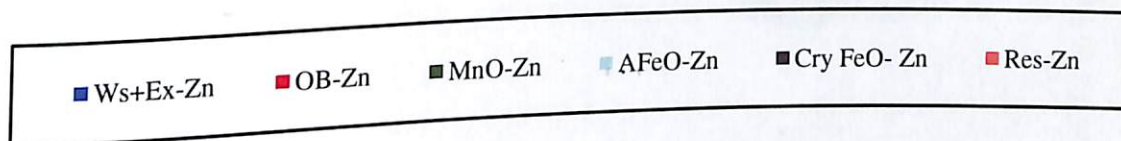
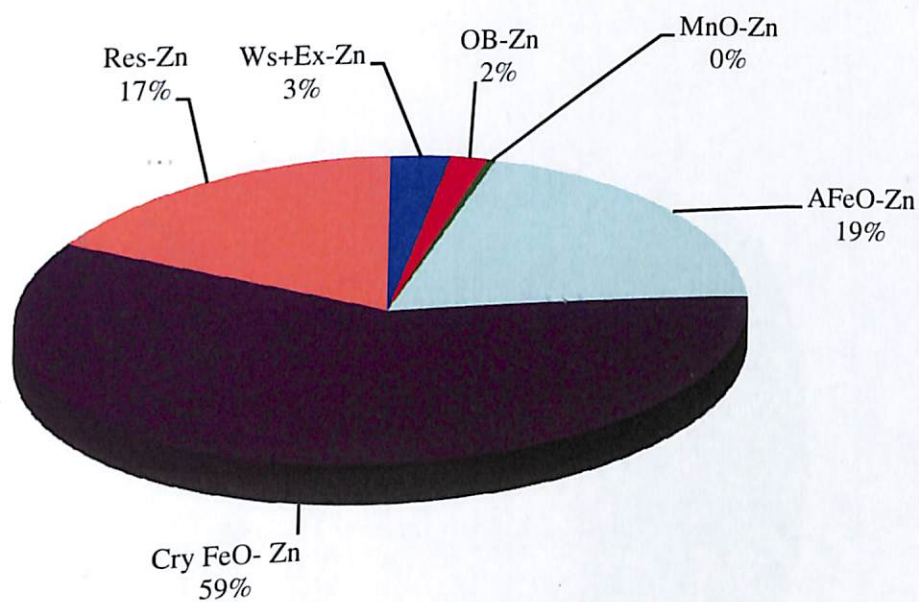


**Fig. 9 Mean values of different fractions (%) of Zn to the total Zn in the surface soil**



**Fig. 9 Mean values of different fractions (%) of Zn to the total Zn in the surface soil (Continued)**

T9



T10

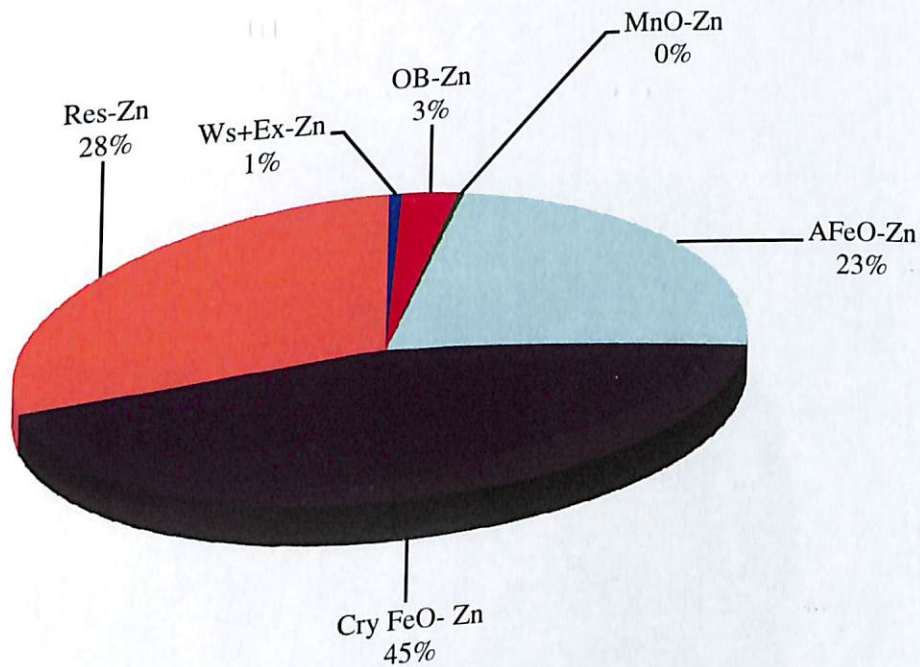
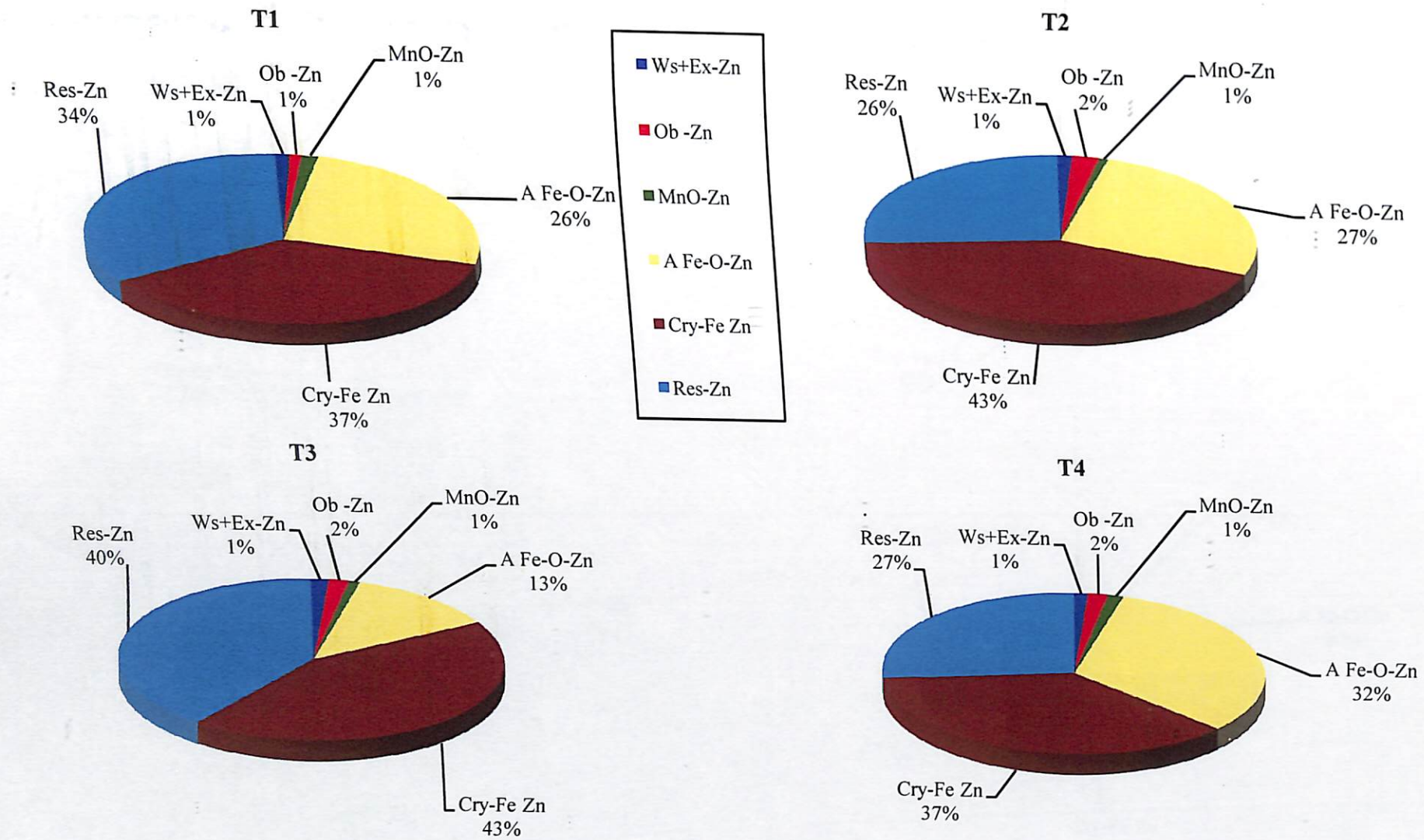


Fig. 9 Mean values of different fractions (%) of Zn to the total Zn in the surface soil (Continued)



**Fig. 10** Mean values of different fractions (%) of Zn to the total Zn in the sub surface soil

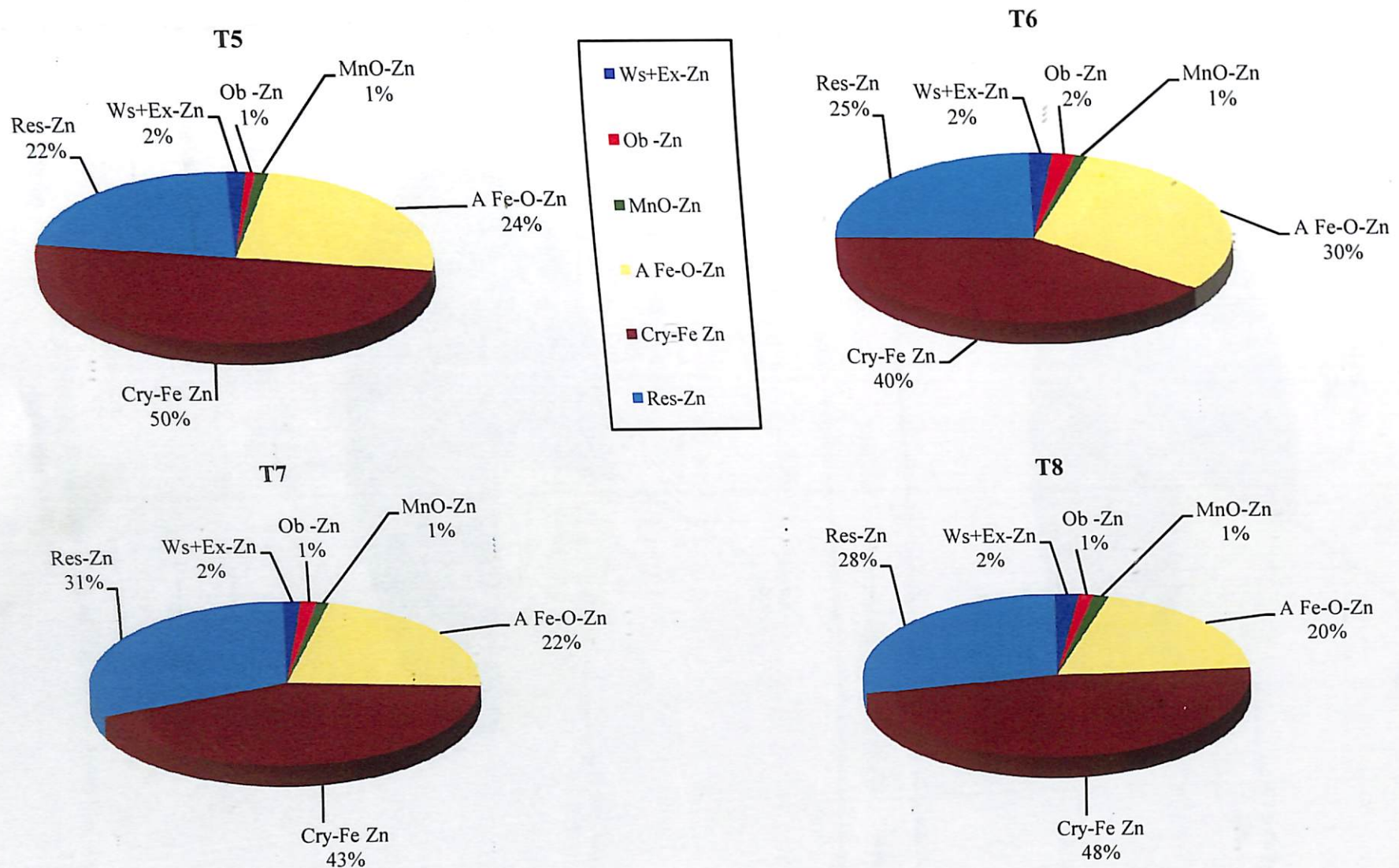
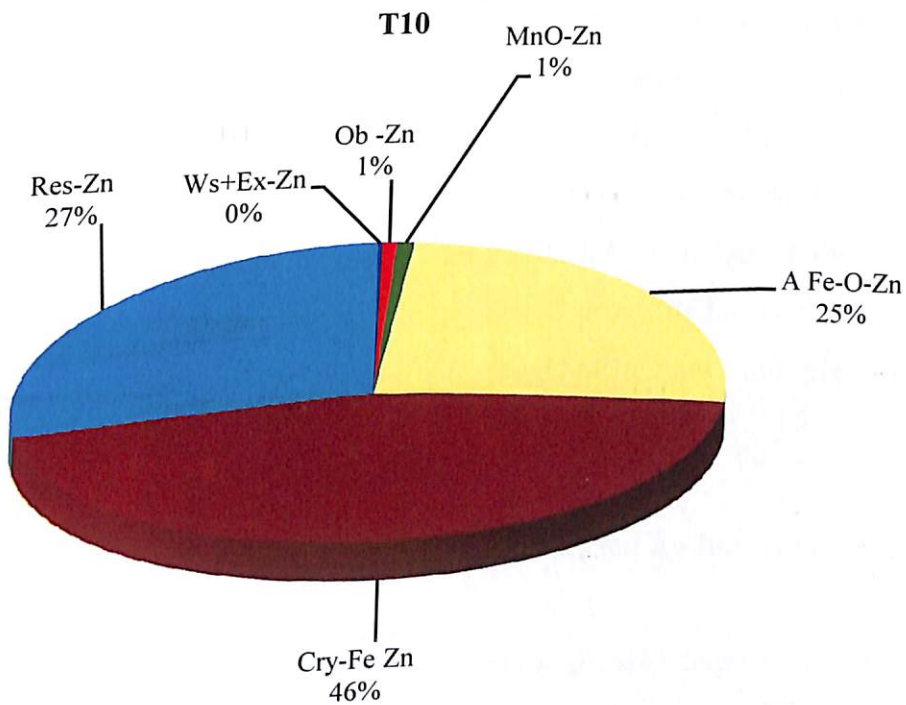
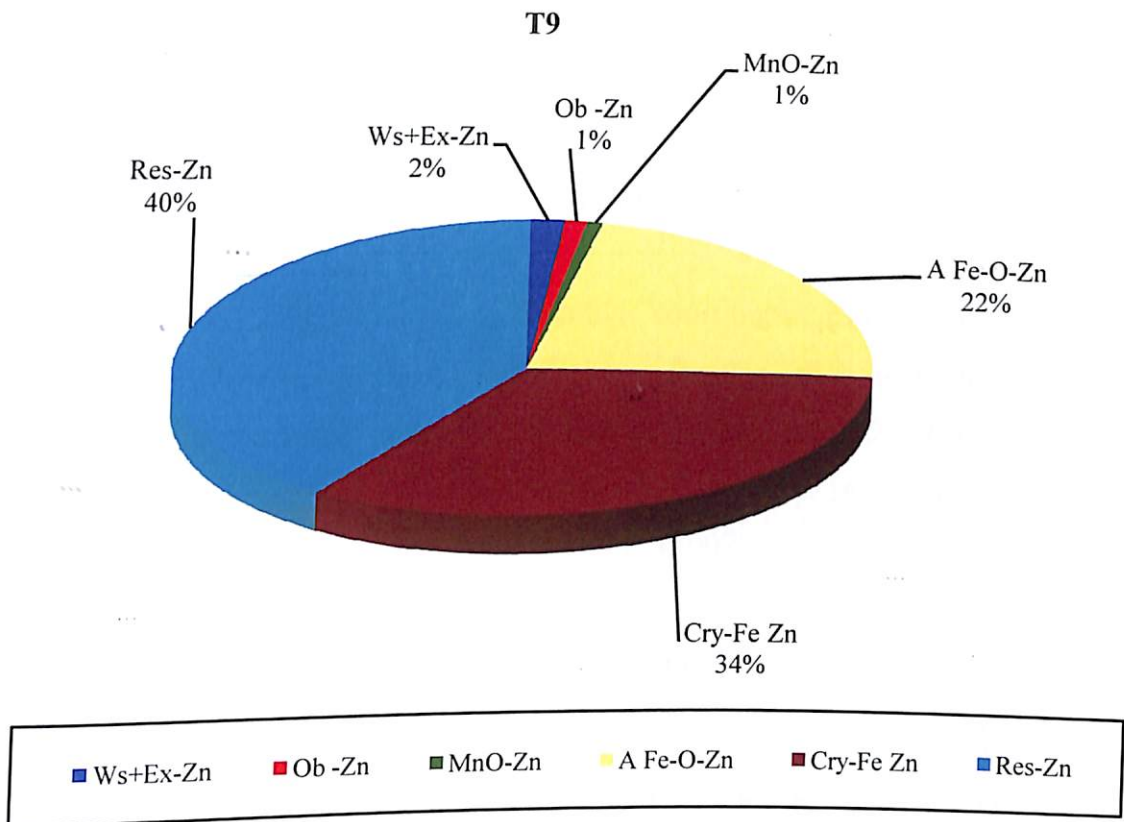


Fig. 10 Mean values of different fractions (%) of Zn to the total Zn in the sub surface soil



**Fig. 10 Mean values of different fractions (%) of Zn to the total Zn in the sub surface soil**



FeO-Zn, 27.91 to 66.0 Cry Feo-Zn, 16.33 to 31.09 Res- Zn for the surface soil and 0.20 to 2.59 Ws+Ex- Zn, 0.86 to 2.36 OB- Zn, 0.86 to 1.81 MnO-Zn, 13.58 to 37.96 Am FeO-Zn, 36 to 59 Cry Feo-Zn, 16.81 to 38.69 Res- Zn for the subsurface soil. As in the MnO-Zn fractions recorded a higher content in the sub-surface layer.

The perusal of the data on the percentage contribution of each fraction to the total Zn showed that the highest contribution was for the Am Feo-Zn and Cry Feo-Zn. In both the surface and subsurface soils after the crop growth. The crystalline iron oxides present in the soils of the experimental area might have influenced this. Similar results are reported by Arundino *et al.* (2007).

### **5.3.9 Indexing of leaf part in fodder maize for Zn status**

Leaf analysis as a diagnostic tool was first recognized by Weinhold (1862). Lundegardh (1935) defined index plant part as that part of a plant which gives the highest predictability on yield. This is based on the assumption that concentration of the nutrients in the leaf at the specific growth stage is related to the performance of the crop (Bould *et al.* 1984). Nutrient content of various plant parts of plant can assist in evaluating the nutrient status of crops. The knowledge of the plant part which accumulate the highest concentration should prove to be useful criterion in delineating the deficiency level of nutrients from sufficiency and toxicity levels (Sharma and Bapat, 2000).

The leaf lamina and petiole samples were analysed for the Zn content.

The values of coefficient of correlation (r) obtained at each stage of the crop are given below.

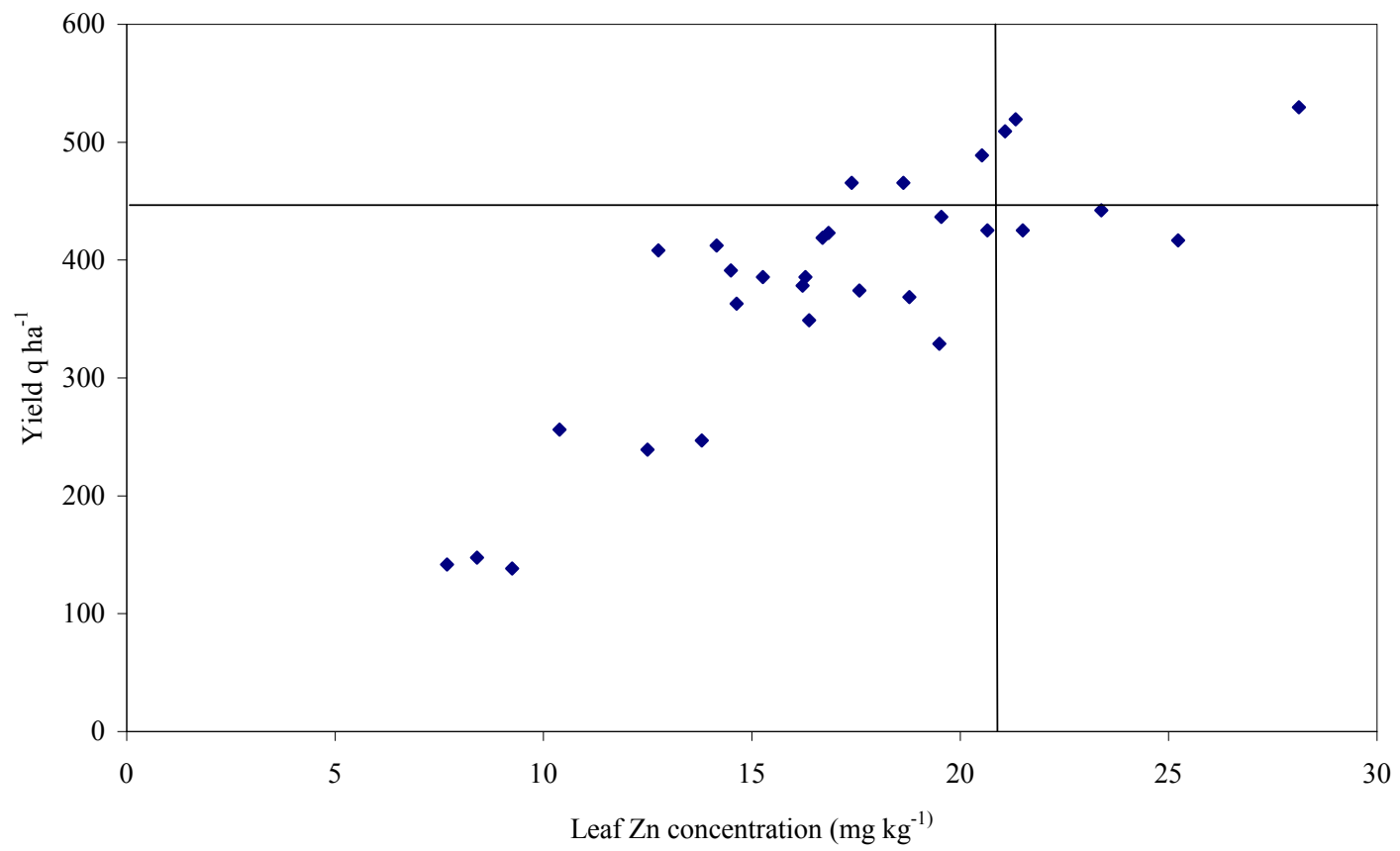
Table .40 Coefficient of Correlation of Zn content and yield at different stages of sampling

Stages of sampling	3 <sup>rd</sup> leaf lamina	3 <sup>rd</sup> petiole	4 <sup>th</sup> leaf lamina	4 <sup>th</sup> petiole	5 <sup>th</sup> leaf lamina	5 <sup>th</sup> petiole
15	0.844*	0.839*	0.786*	0.806*	0.780*	0.751*
25	0.933*	0.804*	0.746*	0.848*	0.673*	0.868*
35	0.906*	0.847*	0.868*	0.660*	0.847*	0.743*
45	0.907*	0.884*	0.903*	0.788*	0.851*	0.722*

\* Significant at 5% level

The 3<sup>rd</sup> leaf lamina at the 25<sup>th</sup> DAS recorded the maximum relationship with green fodder yield as evidenced by the highest value for coefficient of correlation (0.933) and it was selected as the best index part for foliar diagnosis as a close relationship exists between the concentration in that part and yield. Young leaves are particularly suitable for determining Zn status of the plants because symptoms of Zn deficiency usually develop in young leaves and also Zn content of the young leaves is usually more stable than that of older leaves (Rossel and Ulrich,1964).

For determining the critical Zn content in the selected index part (third leaf lamina at 25 DAS) for maximum yield the Scatter diagram technique outlined by Cate and Nelson (1965) was employed (Fig.17). Graph was plotted relating the Zn content in that part at 25 DAS (X- axis) to yield (Y-axis). The points were distributed into four quadrants using plastic overlays maximizing the points into the first and third quadrant. The intersecting point of the line on X-axis drawn parallel to



**Fig. 17. Scatter diagram for critical level of Zn for maximum yield**

Y-axis was taken as the plant critical Zn level in the third leaf lamina at 25 DAS was computed as 21 mgkg<sup>-1</sup> for maximum yield in fodder maize (Fig.17.). This is in accordance with the report by Alvarez *et al.* (1996).

### 5.3.10 Path Analysis of various fractions of Zn with Yield

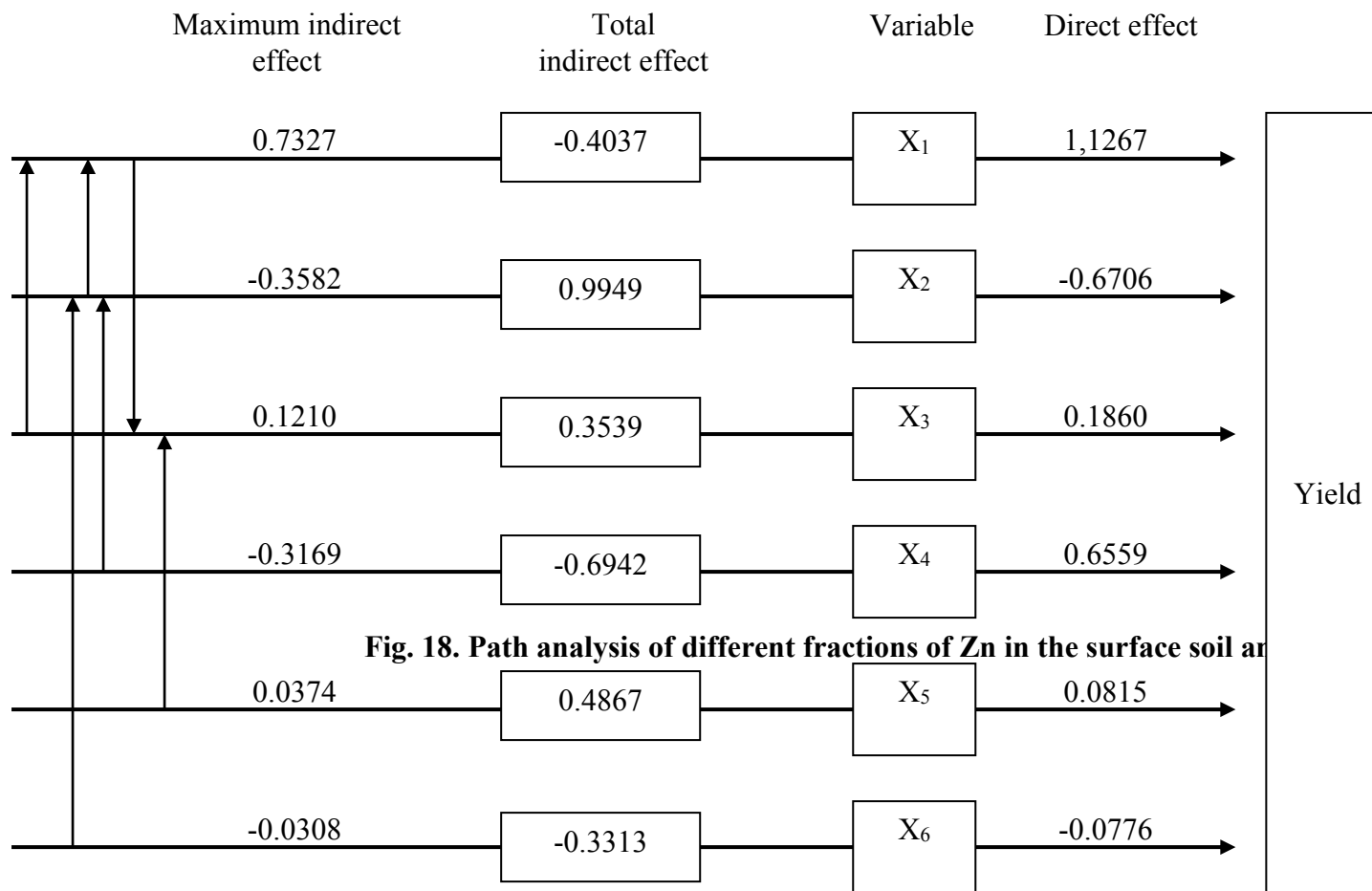
Different fractions of Zn in the surface soil were selected for studying their direct and indirect effect on yield. The different fractions considered were Ws+Ex.-Zn(X<sub>1</sub>), OB-Zn (X<sub>2</sub>), MnO-Zn (X<sub>3</sub>), Am FeO-Zn (X<sub>4</sub>), Cry-FeO-Zn(X<sub>5</sub>) and Total – Zn (X<sub>6</sub>). Path coefficient analysis technique was employed to study the cause and effect relationship of these factors with yield. The results are presented in Table. 41 and illustrated in the Fig. 18

Table . 41 Coefficient of Correlation of Zn fractions with Yield

Parameters	Direct effect	Total indirect effect	Total correlation	Maximum indirect effect
Ws+Ex.-Zn(X <sub>1</sub> )	1.1267	-0.4037	0.7230*	0.7327
OB-Zn (X <sub>2</sub> )	-0.6706	0.9949	0.3243	-0.3582
MnO-Zn (X <sub>3</sub> )	0.1860	0.3539	0.5399*	0.1210
Am FeO-Zn (X <sub>4</sub> )	0.6559	0.6942	-0.0384	0.3169
Cry-FeO-Zn(X <sub>5</sub> )	0.0815	0.4867	0.5682*	0.0374
Total –Zn (X <sub>6</sub> )	-0.0776	-0.3313	0.4414	-0.0308

\*Significant at 5% level

From the above it can be observed that the fraction X<sub>1</sub> (Ws+Ex.-Zn) had maximum direct effect on yield (1.1267). The influence of X<sub>4</sub> (Am FeO-Zn) was also positive and direct (0.6559). But the effect of OB-Zn (X<sub>2</sub>), even though direct, was negatively significant (-0.6706).



**Fig. 18. Path analysis of different fractions of Zn in the surface soil ar**

The indirect effect of  $X_1$  was maximum (0.7327) through  $X_3$ . The indirect effect of  $X_2$  was through  $X_1$  (-0.3582). The indirect effect of  $X_3$  was through  $X_1$  (0.1210) even though it was not significant. The indirect effect of  $X_4$  was through  $X_2$  (-0.3169) while that of  $X_5$  was through  $X_3$  (-0.0308) and  $X_6$  through  $X_2$  (-0.3582).

The results suggested that the depletion of Zn from fractions that are readily available to plants will be compensated by replenishment from other fractions of soil agreeing with reports by Viets (1962). The dominance of  $W_s+Ex$ -Zn fraction through directly contributed to yield is quite evident. Hydrous oxides have a strong affinity for Zn, but they do not fix it in an available form (Mandal and Mandal, 1996). So the  $W_s+Ex$ -Zn fraction exert direct effect on yield. Similar findings were reported by Sharma *et al.* (1986) also.

The effect of total Zn is negative and significant. It indicates that the total Zn content is an unsatisfactory or unreliable measure of Zn availability to crops. Similar conclusions have been drawn by Katyal and Rattan, (1993) and Sureshkumar *et al.* (2004)

Table. 42 Economics of cultivation

Treatments	Yield ( q ha <sup>-1</sup> )	Total returns (Rs.) x	Total cost of cultivation (Rs.) y	BC ratio (x/y)
T <sub>1</sub>	408.10	12243	7805	1.56
T <sub>2</sub>	318.30	9549	7990	1.15
T <sub>3</sub>	457.10	13713	8755	1.57
T <sub>4</sub>	512.50	15375	9505	1.61
T <sub>5</sub>	418.40	12552	10255	1.22
T <sub>6</sub>	301.40	9042	11005	0.82
T <sub>7</sub>	366.10	7322	7998	0.91
T <sub>8</sub>	389.30	7786	8006	0.97
T <sub>9</sub>	395.10	7902	8014	0.99
T <sub>10</sub>	66.40	1328	4375	0.30

Economics of fodder maize cultivation presented in the Table.42 showed that T<sub>4</sub> (NPK@120:60:40 kg ha<sup>-1</sup> + FYM 10 t ha<sup>-1</sup>+Zn@ 10 kg ha<sup>-1</sup> as ZnSO<sub>4</sub>.7H<sub>2</sub>O) gave the maximum net returns (Rs.15375/-) and BC ratio of 1.61, followed by T<sub>3</sub> (NPK@120:60:40 kg ha<sup>-1</sup> + FYM 10 t ha<sup>-1</sup>+Zn@ 5 kg ha<sup>-1</sup> as ZnSO<sub>4</sub>) with a BC ratio of 1.57 and net returns of Rs.13713/-. Net returns and BC ratio were considerably lower for other treatments. So application of Zn @ 10 kg ha<sup>-1</sup> as ZnSO<sub>4</sub>.7H<sub>2</sub>O along with NPK @120:60:40 kg ha<sup>-1</sup> + FYM 10 t ha<sup>-1</sup> is more remunerative.

## 6. SUMMARY

Zinc is an essential micronutrient for plants and animals and when the supply of plant available Zn is inadequate, yield and quality of crop products get reduced. Evaluation of total micronutrient content in Indian soils revealed Zn as the most limiting nutrient. Maize is classified as a very sensitive crop to low levels of available Zn. In this context, the present study on the dynamics of Zn in soils and Zn nutrition of fodder maize was carried out and the salient findings of the study are summarized below.

- The results of the incubation study showed that the availability of Zn from the  $\text{ZnSO}_4 \cdot 7\text{H}_2\text{O}$  was in the medium range ( $0.4\text{-}2.4 \text{ mgkg}^{-1}$ ) on application of Zn @ 5, 10 and  $15 \text{ kg ha}^{-1}$ , up to 60 days of incubation.
- On the 75<sup>th</sup> day of incubation the available Zn release exceeded the medium range of  $2.4 \text{ mgkg}^{-1}$  at the highest level of Zn application ie,  $20 \text{ kg Zn ha}^{-1}$ .
- The soil profile study identified six horizons viz. Ap (0-9.5 cm), Ac (9.5-25 cm), B<sub>1</sub> (25-52 cm), B<sub>2</sub> (52-98 cm), Bt<sub>2</sub> (98-123 cm) and C<sub>1</sub> (123-150 cm).
- Regarding the soil morphological studies, the colour varied from 5YR 3/4 to 5YR4/8, soil structure from crumb to subangular blocky, and soil consistence from non sticky- non plastic to slightly sticky and slightly plastic, as the depth of soil profile increased.
- The particle size analysis of the soil samples from different horizons showed that the percentage of sand decreased from 79.4 in the Ap horizon to 51.60 in C<sub>1</sub> horizon. The silt content varied from 16.20 in the Ac to 9.01 percent in the C<sub>1</sub> horizon. The clay content recorded a steady increase from 11.60 percent (Ap horizon) to 36.20 percent in the C<sub>1</sub> horizon. The textural classes were sandy loam for the upper horizons and sandy clay loam for the lower ones.



- The availability of primary and secondary nutrients showed a gradual decrease with depth of soil profile, while available micronutrients followed an irregular pattern of distribution throughout the profile. The Mn content showed a steady increase from 9.7 to 18.94 mg kg<sup>-1</sup> from the upper to lower horizon. Zn content decreased from Ap to B<sub>2</sub>.
- Regarding the different fractions of Zn, W<sub>s</sub> + Ex. – Zn, OB – Zn, Am FeO-Zn, CrY FeO-Zn decreased from upper to lower horizons.
- Initial analysis of surface (0-15 cm) and subsurface soil samples (15-30 cm) were carried out. Among the physico-chemical properties silt percent, Particle density, Bulk Density, WHC etc. showed a slight increase in the subsurface soil. The pH, Available P and K showed a slight increase in the subsurface soil, while all other nutrients recorded higher values in the surface soil.
- All the Zn fractions, except MnO-Zn recorded a higher value in the surface soil.
- At the time of harvest maximum height was observed for T<sub>4</sub>, NPK @ 120:60:40 kg ha<sup>-1</sup> + FYM 10 t ha<sup>-1</sup> + Zn @ 10 kgha<sup>-1</sup> with a value of 180.90 cm while T<sub>1</sub> (NPK @ 120:60:40 kgha<sup>-1</sup>) and T<sub>6</sub> ( NPK @ 120:60:40 kgha<sup>-1</sup> + FYM 10 t ha<sup>-1</sup> + Zn @ 20 kgha<sup>-1</sup>) were found to be par with values of 163.73 and 158.48 cm respectively.
- The maximum number of leaves at the time of harvest for the first crop recorded for T<sub>9</sub>(NPK @120:60:40 kgha<sup>-1</sup> + FYM 10 t ha<sup>-1</sup> + foliar application of 0.75 percent ZnSO<sub>4</sub> 7H<sub>2</sub>O) with a mean value of 15.13.

- The T<sub>4</sub> (NPK @ 120:60:40 kg ha<sup>-1</sup> + FYM 10 t ha<sup>-1</sup> and soil application of ZnSO<sub>4</sub> · 7H<sub>2</sub>O @ 10 kg ha<sup>-1</sup>) recorded the highest green fodder yield of 512.5 q ha<sup>-1</sup>, dry fodder yield of 103.92 q ha<sup>-1</sup> and these values were significantly superior to all others treatments in the first experimental crop. The PDGFY was also found to be maximum in T<sub>4</sub> (8.54 q ha<sup>-1</sup>).
- The highest mean value for LSR was recorded for T<sub>4</sub> (POP + Soil application of Zn 10 kg ha<sup>-1</sup>) found to be on par with T<sub>3</sub>, T<sub>5</sub>, T<sub>1</sub>, T<sub>9</sub>, T<sub>7</sub> and T<sub>8</sub>.
- Regarding the quality aspects, T<sub>4</sub> recorded the highest value of CP and CPY with values of 10.93 and 1136 kg ha<sup>-1</sup> respectively. While the lowest value of 23.16 percent for CF was observed for POP + foliar application of 0.75 percent ZnSO<sub>4</sub> · 7H<sub>2</sub>O. But T<sub>4</sub> also recorded a lower value of 23.70 percent of crude fibre and it was on par with T<sub>9</sub>.
- The nutrient concentrations of the fodder maize, samples of the first experimental crop revealed the following. T<sub>4</sub> (POP + Soil application of Zn @ 10 kg ha<sup>-1</sup>) recorded the highest value of N, K, Mn, Zn and Cu while the highest value for P was obtained in T<sub>3</sub> (0.27 percent). The T<sub>5</sub> treatment recorded the maximum Ca and Mg content. S concentration (0.17 percent) was maximum for the highest dose of Zn with 20 kg ha<sup>-1</sup>. The highest value of Zn was obtained for soil application of Zn @ 10 kg ha<sup>-1</sup> with a mean value of 68.33 mg kg<sup>-1</sup>.
- Physico-chemical parameters of surface soil showed that pH, EC and organic carbon were not significant. The available Nitrogen and Potassium were maximum in T<sub>4</sub> with values of 224.60 kg ha<sup>-1</sup> and 204.3 kg ha<sup>-1</sup>.
- The Ex-Ca and Mg were maximum in T<sub>2</sub> and T<sub>4</sub> with values of 1.30 and 0.52 c mol kg respectively. T<sub>5</sub> recorded the highest value of available S (12.88 kg ha<sup>-1</sup>). For the micronutrients, T<sub>3</sub> (POP + Zn @ 5 kg ha<sup>-1</sup>) recorded

the highest value for DTPA Fe with ( $22.37 \text{ mgkg}^{-1}$ ) which was on par with  $T_4$ ,  $T_5$ ,  $T_9$  and  $T_1$ . While DTPA Mn was maximum in  $T_9$  ( $4.94 \text{ mgkg}^{-1}$ ). The highest content of DTPA Zn was in  $T_4$  with a value of  $4.89 \text{ mgkg}^{-1}$ .

- For the first residual crop  $T_5$  (POP + Zn @  $15 \text{ kg ha}^{-1}$  as  $\text{ZnSO}_4 \cdot 7\text{H}_2\text{O}$ ) recorded the maximum green and dry fodder yield of  $34.65$  and  $6.86 \text{ q ha}^{-1}$  respectively.
- The highest value for DTPA extractable Zn in the surface soil was observed in  $T_4$  (POP + Zn @  $10 \text{ kg ha}^{-1}$  as  $\text{ZnSO}_4 \cdot 7\text{H}_2\text{O}$ ) as  $2.06 \text{ mgkg}^{-1}$  and lowest was in the control ( $0.56 \text{ mgkg}^{-1}$ ).
- For the second experimental crop the  $T_4$  (POP+Zn@  $10 \text{ kg ha}^{-1}$  as Zn  $\text{SO}_4$ ) showed the maximum height at 45 DAS and at harvest with the values of  $123.70$  and  $139.2 \text{ cm}$  respectively.
- Considering the yield and yield attributes the highest yield as well as per day green fodder yield was obtained in  $T_5$  (POP+Zn  $15 \text{ kg ha}^{-1}$ ) with  $519.3 \text{ q ha}^{-1}$  and  $9.44 \text{ q ha}^{-1}$  respectively, for the 2<sup>nd</sup> experimental crop The highest value of crude protein was  $10.23$  per cent in  $T_3$  and  $T_6$ ,  $T_5$ ,  $T_2$  and  $T_4$  were found to be on par with this. The crude protein yield was maximum in  $T_5$  with  $1090 \text{ kg ha}^{-1}$ .
- After the crop growth, a reduction in the Nitrogen and phosphorus contents and enhancement of potassium was noticed. Among Zn applied plots maximum P content was noticed where minimum quantity of Zn has been applied.
- The highest yielded plot showed a significantly lower content of  $W_s+Ex\text{-Zn}$  fractions after the crop growth, revealing its contribution to the available pool

- The Am Feo-Zn fractions contributed more to the total Zn in both surface and subsurface soil..
- For the second residual crop the highest green fodder yield of 39.49 q ha<sup>-1</sup> and dry fodder yield 67.77 q ha<sup>-1</sup> was obtained in T<sub>6</sub> and it was significantly different from other treatments.
- For raising a single crop of fodder maize Variety African Tall, soil application of Zinc Sulphate @Zn 10kg/ha along with FYM @10tha<sup>-1</sup> and NPK 120:60:40 kgha<sup>-1</sup> is recommended. T<sub>4</sub> treatment ie. (POP+Zn @ 10 kg ha<sup>-1</sup> as Zn SO<sub>4</sub>) showed the highest BC ratio of 1.61 and was found economical for maximum yield , Zn application could be skipped for the second crop of fodder maize, if . Zn @15 kg ha<sup>-1</sup> for the first crop have been applied.
- The pooled analysis data on yield showed that the highest yield was observed in T<sub>5</sub> (468.80 q ha<sup>-1</sup>), but T<sub>4</sub>, T<sub>9</sub> and T<sub>8</sub> were found to be on par. The highest dry fodder yield was also in T<sub>5</sub> (94.90 q ha<sup>-1</sup>) which was on par with T<sub>4</sub> (93.80 q ha<sup>-1</sup>).
- The 3<sup>rd</sup> leaf lamina at 25<sup>th</sup> DAS recorded the maximum coefficient of correlation (r values 0.933) with yield and it was selected as the index part and stage for foliar diagnosis.
- For determining the critical level of Zn content in the index plant part (3<sup>rd</sup> leaf lamina at 25 DAS) for maximum yield, the scatter diagram technique of Caté and Nelson was employed and the critical Zn level was computed as 21 mg/kg for maximum yield in fodder maize.
- A path coefficient analysis of different fractions of Zn in the surface soil was selected for studying their direct and indirect effect on yield. The W<sub>s</sub>+Ex-Zn fraction directly contributed to the yield while the total Zn is negative and significant.

## 7. REFERENCES

- Aiyer, R.S., Rajagopal, C.K. and Money, N. S. 1975. Available Zn, Cu, Fe and Mn status of the rice soils of Kuttanadu. *Agric.Res. J. Kerala* 13:15-19
- Alloway, B. J. 2004. Zinc in soils and crop nutrition. International Zinc Association Communications .IZA Publication, Brussel Paris, France
- Alloway, B.J. 1990. Heavy Metals in Soils .Blackie, Glasglow, P.440
- Alvarez, J. M. 2007. Influence of soil type on the mobility and bioavailability of chelated zinc. *J. agric . Food Chem.* 55(9): 3568-3576
- Alvarez, J.M. Orabar, A. and Rico, M.I. 1996. Effects of chelated Zn, soluble and coated fertilizers on soil Zn status and Zn nutrition of maize .*Commn. Soil Sci. Plant Anal.* 27:7-19
- Anonymous. 1975. Annual report ,UPASI, Mysore
- Anonymous. 2000. Three decades of AICRP on long term fertilizer experiments to study the changes in soil quality, crop productivity and sustainability, IISS, Bhopal, p.59
- Anonymous. 2007. Benchmark soils of Kerala. (ed. Premachandran, P. N), Soil Survey Organization, Government of Kerala, Thiruvananthapuram, P.623
- Anonymous. 2008. Economic Review, 2007. State Planning Board, Government of Kerala, Thiruvananthapuram P.485
- Arundino, E., Barberies, E., Ajnore Marson, F., Zaini, E. and Franchini, M. 2007. Iron oxides and clay minerals within profiles as indicators of soil age in Northern Haly *Geoderma* 136: 37: 45-55
- Asher, C.J., Edwards, D.G. and Howeler, R. H. 1980. Nutritional disorders of cassava (*Manihot esculenta*, Crantz), University of Queensland, St. Lucia, Australia, P.48

- Barber, S. A. 1984. Soil nutrient bioavailability: A mechanistic approach. John Wiley and Sons, New York, P.532
- Barrow, N. J. 1987. Reactions with variable charges in soils. Nijhoff, Dordrecht, The Netherlands, p.212
- Barrow, N. J. 1986. The effects of time and temperature on the reaction of Zn with a soil. *J. Soil. Sci.* 37: 295-302
- Bhattacharya, T., Sen, T. K., Singh, R. S., Nayak, and Seghal, J. L. 1994. Morphology and classification of Ultisols with Kandic horizon in North Eastern Region. *Indian Soc. Soil Sci.* 42(2); 301-306
- Black, C.A. 1965. Methods of Soil Analysis. Part D. Physical and mineralogical properties. Agronomy series No.9, A.S.A., SSA, Madison, Wisconsin, GSA.
- Bould, C., Hewitt, E. J. and Needham, P. 1983. Diagnosis of mineral disorders in plants. Vol.1. Principles. HMSO, London, p.501
- Brawn, A.L., Krantz, B.A. and Martin, P.E. 1962. Plant uptake and fate of soil applied Zn. *Soil.Sci.Soc.Am.Proc.* 26, 167-170
- Brown, P.H., Cakmak, I. and Zhang, Q. 1993. Forms and functions of Zinc in plants. (ed. Robson, A.D.). In: *Zinc in Soils and Plants*, Kluwer Academic, Publications, Dordrecht, The Netherlands, p. 93-106
- Brennan, R. F. 2005. Zinc application and its availability to plants. Ph.D. thesis, Murdoch University P. 252
- Brennan, R. F. and Gartrell, J. W. 1981. Zinc requirement of crops and pastures. *Our Land*, 13:10-13
- Broadley, M. R., White, P.J. Hammond, J.P; Zelko, S and Lux, A. 2007. Zinc in plants. *New Phytologist*, 173(4): 677-702
- Brawn, A.L., Krantz, B.A. and Martin, P.E. 1962. Plant uptake and fate of soil applied Zn. *Soil.Sci.Soc.Am.Proc.* 26, 167-170

- Brown, P.H., Cakmak, I. and Zhang, Q. 1993. Forms and Functions of Zinc in plants (Ed. Robson, A.D.). In Zinc in Soils and Plants Kluwer Academic, Publications, Dordrecht, The Netherlands, p. 93-106
- Brummer, G.W., Gerth, J. and Tiller, K.G. 1988. Reaction kinetics of the adsorption of nickel, Zn, and Cadmium by goethite. I. Adsorption and diffusion of metals. J. Soil Sci. 39: 37-52
- Cakmak, I. and Marschner, H. 1990. Decrease in nitrate uptake and increase in proton release in Zinc deficient cotton, sunflower and buckwheat plants. Plant Soil. 129, 261-268
- Cakmak, I., Marschner, H. and Baugerth, P. 1989. Effect of zinc nutrition status on growth, protein metabolism and level of indole 3 acetic acid and other phytohormones in bean (*Phaseolus vulgaris* L.). J. Exp. Bot. 40, 405-415
- Cakmak, I. 2000. Possible roles of zinc in protecting cells from damage by reactive oxygen species. New Phytol. 146, 185-205
- Cate, R. D. and Nelson, L. A. 1965. A rapid method for correlation of soil test analyses with plant response data, North Carolina State University Interpretation Soil Testing Series Tech. Bull. 1. North Carolina Agric. Res. Station, Raleigh, USA
- Cate, R. D. and Nelson, L. A. 1971. A simple statistical procedure for portioning of soil test correlation data into two classes. Soil.Sci. Am.Proc. 35:651-660
- Channakesava, B.C., Prasanna, K.P.R. and Ramachandrappa, B.K. 1999. Effect of plant growth regulators and micronutrients on seed yield and yield components in African Tall fodder maize (*Zea mays* L.). Mysore J. Agric. Sci. 33: 111-114

- Channakesava, B.C., Ramaprasanna, K.P. and Ramachandrappa, B.K. 2007. Effect of plant growth regulators and micronutrients on growth components and seed yield in African Tall fodder maize (*Zea mays* L.). *Agric. Sci. Dig.* 27(1): 38-40
- Chaube, A.K., Rubella, R., Rajachakraborty, Ganwar, M.S., Srivasthava, P.C. and Singh, A.K. 2007. Management of Zinc fertilizer under pearl millet wheat cropping system in a typic ustipsamment. *J. Indian Soc. Soil Sci.* 55(2): 196-202
- Chaudhary, S.K. and Raina, J.N. 2008. Zinc transformation and its critical limits in apple orchards of Himachal Pradesh. *J. Indian Soc. Soil Sci.* 56(4):430-435
- Chesnin ,L. and Yien,C.H. 1950. Turbidimetric determination of available sulfates. *Proc.Soil . Sci. Soc. Am.* 15:149-151
- Clemente, R., Walkar, D.J, Roig, A. and Bernal, M.P. 2003. Heavy metal bioavailability in a soil ,affected by mineral sulfides contamination following the mine spillage Aznalcollar ( Spain). *Biodegrad.*14(3): 199-205
- Dadwich, L.K. and Gupta, A.K. 2003. Productivity and economics of pearl millet fodder as influenced by S1Zn and planting pattern. *Forage Res.* 28(4): 207-209.
- Das, D.K. 1986. Effect of Zn in rice (*Oryza Sativa*) in the laterite soils . *Maharashtra agric.University*,11: 120-122
- Das, D.K. 2003. Secondary and micronutrient management increasing crop production in acid soil..Winter School on Characterization and Sustainable Management of Acid Soil ,OUAT,Bhuvanewar,2003.
- Das, D.K. 2007. Micronutrients, their behaviour in soils and plants. Kalyani Publishers, New Delhi. P. 362
- Deb,D. L.1997. Micronutrient research and crop production in India. *J. Indian Soc. Soil Sci.*45 (40) :675-692



- Deepak, K., Khajuria.B.M. Jalai, V.K. and Sharma,R. 2004. Distribution of micronutrient cations in soils of Kandi belt. J. Indian Soc. Soil Sci. 3(1):97-103
- Devarajan, R., Kumaresan, K.P., Ramanathan, G.and Panchanathan, R. M. 1988. Response of sunflower to micronutrients. Madras agric. J. 75: 401-404
- Devi, P.N. and Rani, Y.A. 2003. Effect of different levels of zinc on yield and yield components of different rice hybrids. The Andhra Agric J. 50(384): 283-286
- Dewal, G.S. and Pareek, R.G. 2004. Effect of phosphorus, sulfur and zinc on growth, yield and nutrient uptake of wheat (*Triticum aestivum*). Indian J. Agron. 49(3): 160-162
- Dube, B.K., Sharma, C.P. and Chatterjee, C. 2001. Response of pigeonpea to applied Zn in listifluent soils of Western Uttar Pradesh. Indian Soc. Soil Sci. 49(3): 471-475
- Dudde, K.B. 1986. Ph.D. thesis, Marathwada Agricultural University.p.215
- Evans, H.J. and Sargar, G.J. 1966. Role of mineral elements with emphasis on univalent cations. Ann. Rev. Plant Physiol 35 : 223-242
- Frageria, N.K., Baligar, V.C. and Clark, R.B. 2002. Micronutrients in crop production. Adv. Agron. 77. 185-268
- Frecenko, J. and Lozek.O. 1998. Rostelimma-vyrob, 44, 15
- Geering, H. R. and Hodgson, J. F. 1969. Micronutrient cation complexes in soil solution.111. Characteristics of soil solution ligands and their complexes with Zn<sup>2+</sup> and Cu<sup>2+</sup>. Soil Sci. Soc. Am.Proc.33: 54-59
- Geethakumari,V, L.,Mathew,U. and Varghese,S.S. 2005. Proceedings of the National Workshop on soil resource management (eds.Kunnungal,M. C. and Maghi.E. K), 22-24, December.2005. Soil Survey Organization, Trivandrum, p. 310-323

- George,S.1994. Standardization of plant part as an index of potassium status in banana, MUSA (AAB group). Ph.D thesis, Kerala Agricultural University, Thrissur, p.185
- Gnadas.1989. Standardization of index plant part in clove (*Myristica fragrans*) in relation with micronutrients. M.Sc. thesis, Kerala Agricultural University, Thrissur p.105
- Goldschmidt. M. 1954. Geochemistry, Oxford University Press, London, U.K
- Gopi, C.S. 1983. Foliar diagnosis in coconut in relation to NPK. M.Sc.Thesis, Kerala Agricultural University, Thrissur, P.97
- Graham, R.D. and Reed, M.L. 1991. Carbonic unhydrase and the regulation of photosynthesis. Nature New Biol. 231, 81-83
- Graham, E.R. 1953. Soil mineralogy as an index to the trace element status of some Australian soils. Soil Sci. 75:335-345
- Gupta, R.P. and Dekshinamoorthi, C. 1980. Procedures for Physical Analysis of soil and collection of Agro-Meteorological dates, Indian Meterological Department, Pune.
- Gupta,P.K. and Vyas,K.K. 1999. Critical level of DTPA Zn for soybean in a vertisol. J.Indian Soc.Soil. Sci. 47(3):564-566
- Halvorson, A. D. and Lindsay, W. L. 1977. The critical Zn<sup>2+</sup> concentration for corn and the non absorption of chelated Zn. Soil.Sci. Soc.Am.J. 41: 531-534
- Hambridge, K. M. and Krebs. F. 2007. Zinc deficiency: a special challenge .J. Nutr. 137(4): 1101-1105
- Harris,D ., Rashid, A .,Miraj,G., Arif,M. and Shah,H. 2007. Field Crops Res. 102(2): 119-127
- Hazra, G.C. and Mandal, B. 1996. Desorption of adsorbed Zn in soils in relation to soil properties. J. Indian Soc. Soil Sci. 44 : 233-237

- Hazra, G.C., Saha, J.K., Mete, P.K. and Mandal, B. 1993. Distribution of Zn fractions in red and laterite soils of Birbhum, West Bengal. *J. Indian Soc. Soil Sci.* 41 (3):472-476
- Hesse, P.R. 1971. A text book of Soil Chemical Analysis. William cloures and Sons, London, 513 p.
- Himes, F.L. and Barber, S.A. 1957. Chelating ability of soil organic matter. *Soil Sci. Soc.Am.Proc.* 21, 368-373
- Hodgson, J. F., Lindsay, W. L. and Trierweiler, J. F. 1966. Micronutrient cation complexing in soil solution. II Complexing of Zn and Cu in displaced solution from calcareous soils. *Soil Sci.Soc.Am. Proc.* 30, 7233-726
- Hodgson, J.F. 1963. Chemistry of micronutrient elements in solid. *Adv. Agron.* 15, 119-159
- Hulagur, B. F. and Dungurwala, R. T. 1983. Effect of Zn, Cu, and P fertilizers on the content and uptake and secondary nutrients by hybrid maize. *Madras agric.J.* 70(2): 88-89
- Jacob, M. 1989. The occurrence and distribution of the micronutrient elements in the rice soils of South Kerala. M.Sc.(Ag.) thesis, Kerala Agricultural University, Thrissur, P.86
- Jackson, M.L. 1973. Soil Chemical Analysis. Prentice Hall of India Pvt. Ltd., New Delhi, 498 p.
- Jagtap, P.B. , Patil J.D., Nimbalkar, C.A., and Kadlag, A.D. 2007 Influence of Integrated Nutrient Management on Soil properties and release of nutrients in a saline – sodic soil. *J. Indian Soc. Soil Sci.* 55 (2): 147-156
- Jahiruddin, M., Chambers, B.T., Livesey, N.T. and Cresser, M.S. 1986. Observations on the effect of soil  $p^H$  upon soil absorption by soils. *Commn.Soil Sci. Plant Anal.* 16, 909-922

- Jain ,N.K.and Dahama,A.K.2006. Direct and residual effects of P and Zn fertilization on productivity of wheat (*Triticum aestivum* ) - pearl millet (*Pennisetum glaucum*) cropping system .Indian J.Agron. 51 (3):165-169
- John, S. K. 2006. Yield maximization in cassava (*Manihot esculenta* Crantz) through Hunters systematic approach in fertilizer use. Ph. D thesis, Kerala Agricultural University, Trichur, P.225
- Johnson, P.T. 1978. Foliar diagnosis, yield and quality of ginger in relation to NPK. M.Sc.Thesis, Kerala Agricultural University, Thrissur, P.92
- Jose, A.I. 2005. Soil Fertility Evaluation. Proc.Soil and Plant analysis, (ed.John, S.), 12-17, December, 2005, CTCRI, Trivandrum, 38-42
- Jyolsna, V.K. 2005. Zinc and boron availability in soils and impact of carriers on crop productivity. M.Sc.thesis, Kerala Agricultural University, Thrissur, P.100
- Jyung, W.H., Ehmann, A., Schlender, K.K. and Sclar, J. 1975. Zinc nutrition and starch metabolism in *Phaseolus vulgaris*. Plant Physiol. 55, 414-420
- Kabata Pendias , A. and Pendias ,H. 1984. Trace elements in Soils and Plants. CRC Press, Boca-Raton, Florida, p.235
- Kalbasi, M. Racz, G.J. and Lowever – Rudger, L.A. 1978. Mechanism of Zinc adsorption by Fe and Al oxides. Soil Sci. 125, 146-150
- Karche, V.K., Pharamde, A.L. and Patil, J.D. 2003. 2003. Micronutrient in soil and plants. An overview for western Maharashtra. Interface session on micronutrient, MCEAR, Pune.
- Katyal, J.C. 1973. Annual report 1972-73. All India Co-ordinated Scheme on micro nutrients in soils and plants. Indian Council of Agricultural Research New Delhi, p.101
- Katyal, J.C. and Rattan, R.K. 1993. Distribution of Zn in Indian soils .Fert. News. 38(2):15-26

- Katyal, J.C. and Rattan, R.K. 2003. Secondary and micronutrients .Research gaps and future needs. *Fert.News*,48,9-20
- Kessler, B. 1961. Ribonucleases as a guide for the determination of Zinc deficiency in orchard trees (ed. Reuther, M.). In *plant Analysis and fertilizer Problems* American Inst. Biol. Sci., Washington, USA Pp. 314-322
- Kochaian ,L.V. 1993. Zinc absorption from hydroponic solutions by plant roots .In *Applied Soil Trace Elements* (ed. Robson. A. D), Kluwer Accademic Publications, Dodrecht, The Netherlands, P.45-57
- Koshy, M.M., Korah, P.A., Usha, P.B. and Sudharmayidevi, C.R.1990. *Quinquennial Report of Centre of Excellence of Tropical Soils*. Kerala Agricultural University, Vellanikkara, p.51
- Krauskopf, K. B. 1972. Geochemistry of micronutrients . In: *Micronutrients in Agriculture*. (ed. Mortvedt,J.J., Giordano, P. M . and Lindsay,W.L.) *Soil Sci. Soc.Am. Wisconsin, USA*,p.7-40
- Krishna, S., 1995. Effect of sulphur and zinc and application on yield, S and Zn uptake and protein 24. Yang, J., J. Zhang, Z. Wang, Q. Zhu and L. Liu,content of mung (green gram). *Legume Res.*, 18: 2001. 89-92.
- Krishnan, P., Venugopal, K.R. and Nair, K.M. 2000. Morphology, characteristics and classification of low activity clay soils of Kerala. *J. Indian So. Soil Sci.* 48(4): 819-823
- Lindsay, W.L. 1972. Zinc in Soils and Plant Nutrition, *Adv. Agron.*, 24, 147 - 186.
- Lindsay, W.L. and Norvell, W.A. 1978. Development of DTPA Soil test for Zn, Fe, Mn and Cu. *J. Soil Sci. Soc. Am.* 42: 421-428
- Lombanaes, P. and Singh,B.R. 2004. Effects of true Mn activity on the yield and uptake of micronutrient cation by barley and oat grown in chelator buffered nutrient solution .*Field Crops Abst.* 57(4): 349

- Loneragan, J. F. and Webb, M. J. 1993. Interaction between zinc and other nutrients affecting the growth of plants. In: Zinc in Soils and Plants. (ed. Robson, A. D), Kluwer Acad. Publ. Dordrecht, The Netherlands, p.119-133
- Lucas, R.E. and Davis, J. F. 1961. Relationship between  $p^H$  values of organic soils and availability of twelve plant nutrients . Soil Sci. 92 ,177-182
- Lundegardh, H. 1935. The influence of the soil upon the growth of the plant .Soil. Sci.40: 89-101
- Ma, Y. B and Uren N. C 1997. Application of new fractionation scheme for heavy metals in soils. Commn. Soil Sci. and Plant Analy. 26,3291–3303.
- Malewar, G.U. and Dudde, K.B. 1995. Evaluation of critical limits of DTPA Zinc for sorghum and cotton in chromusterts of Parbani (Maharashtra). J. Maharashtra agric. University. 30, 336-338.
- Mandal ,L.N. and Mandal,B. 1996. Zinc fractions in soils in relation to zinc nutrition of lowland rice. Soil Sci. 142,141-148
- Marschner, H. 1986. Mineral nutrition of higher plants. Academic Press. London, U.K. P.325
- Mathew,U. 1999. Status and impact of heavy metals in selected soils and crops of Kerala, . Ph.D thesis,Kerala Agricultural University, Thrissur,P.208
- Mathew,U.2000. Effect of application of Zn and Cu on the yield of rice.J.Trop.Agric. 38:100-101
- Mathew,U.2006. Assessment of micronutrients in the laterite soils of Kerala. XXV1 ZERAC meeting ,15-16,December, 2006. Kerala Agricultural University, p.144
- MC Bride, Bride, 1989. Reactions controlling heavy metal solubility in soils. Adv.Soil.Sci. 10, 1-54

- Meerabai, M. 2001. Effect of major and minor micronutrients on growth ,yield and quality of turmeric intercropped in coconut garden. Proc.11<sup>th</sup> Swadeshi Science Congress, KFRI, Thrissur.
- Mehla,D.S.1999. Effect of frequency of Zn application on yield of rice (*Oryza sativa*) and wheat (*Triticum aestivum*) in rice-wheat sequence. Indian J. Agron.44, 463-466
- Mehra, R.K., Sharma, K.M., Jat, J.R. and Dadheech, R.C. 2005. Critical limits of Zn and Fe in soil and plants for maize in Haplustalfs of sub humid Southern plain and Aravalli Hills of Rajasthan. Indian Soc. Soil Sci. 53(2): 227-231
- Mikko Sillanpaa, 1972. Trace elements in Soils and Agriculture. Soils Bull. 17, FAO, Rome
- Mikko Sillanpaa, 1990. Micronutrient assessment at the country level: An international study. FAO Soils Bull.63. FAO-UN in co-operation with FINNIDA, 178p.
- Miller, E.R., Lei, X. and Ullrey, D.E. 1991. Trace element in Animal nutrition In Micronutrients in Agriculture (Eds. Mortvedt, J.J., Fox, F.R. Shuman, L.M. and Walch, R.M.). Second edn. Soil Sci. Soc. America, Madison, USA, pp. 593-662
- Mishra and Singh 1996. In Agrl. N use and its environmental implications By Abrol I.P and Reghuraman.N. [www.google.com/books](http://www.google.com/books) google.co.in/books%3Fop%3Dlibrary ht:en.
- Mishra, P., Singh, R., Srivastava, P.C. and Ram, B. 2009. Effect of continuous cropping and fertilization on Zinc fractions and their contribution to plant uptake under rice -wheat system. Indian J. Soil Sci. 57(2): 167-171
- Morgan, J, T. 1984. Differential response to five species to phosphorous and zinc fertilizers .Commn. Soil. Sci. Plant Anal. 15, 437- 447

- Mortensen, J. J. 1963. Complexing of metals by organic matter. Soc.Am.Proc.27, 179- 186
- Mortvedt, J, J. and Kelose, J, J. 1988. Grain sorghum response to banded acid type fertilizers in iron deficient soil. J. Plant Nut. 11: 1297-1310
- Nair, K. M. and Abrol, I. P. 1989. Long term fertilizer experiments in India. Fert. News. 34(4)
- Nair, K.M., Sureshkumar, Kutty, M.C.N., Prasanna, R., Balachandran, P.V., Premachandran, P.N. and Vadivelu, S. 2007. Evaluation of soil fertility in Palakkad. Proceedings of the National Workshop on fertility evaluation for soil Health Management (Ed. Jayakumar, K) p. 111-124
- Nayyar, V. K. 1999. Micronutrient management for sustainable intensive agriculture. Indian Soc. Soil Sci. 47 (4):666-680
- Neilsen, D., Hoyt, P.B. and Mackenzie, A.F. 1986. Distribution of soil Zinc fractions in British Columbia interior orchard soils. Canadian J. Soil Sci. 66: 445-454
- Norrish, K. 1975. The geochemistry and mineralogy of trace elements. In: Trace elements in soil- plant animal systems. (eds. Nicholas , D.J.D. and Ergon,A.R.), Accad. Press, New York, p.181-198
- Norvell, W.A. 1991. Reaction of metal chelates in soils and nutrient solutions . In: Micronutrients in Agriculture. (eds. Mortvedt, J, J., Cox, F. R., Shuman , L. M. and Welch, R. M ) . 2<sup>nd</sup> edn. Soil Sci. Soc. Am, Madison, USA,p. 187-228
- Norvell, W.A. and Welch, R.M. 1993. Growth and nutrient uptake by barley (*Hordeum vulgare* L. cv. Herta). Studies using an N (2-hydroxyethyl) ethylene dinitriilotriacetic acid buffer nutrient solution technique I. Zinc ion requirement. Plant Physiol. 101, 619-625



- Novillo, J., Obrador, A., Lopez-Valdivia, L. M. and Alvarez, J. M. 2002. Mobility and distribution of Zn forms in columns of an acid, a neutral and a calcareous soil treated with three organic Zn complexes under laboratory conditions. *Aust. J. Soil Res.* 40:791-803
- Orabi, A., El-Kobble, T. and Fathi, A.I. 1985. *Plant and Soil* 83: 317-321
- Padmanabhan, M. 1983. Comparative study of the adsorption-desorption of Cu (111), Zn (11), Co (11) and Pb (11) at the goethite solution interface. *Aust. J. Soil Res.* 21, 515-525
- Pasricha, N. S. 2002. Potassium dynamics in soils in relation to crop production. *J. Indian Soc. Soil. Sci.* 41:67-69
- Pattil, D.B. and Bharambe, P.R. 2003. Ongoing micronutrient research work under Dr.P.D.K.V. Akola, Interface on micronutrients, MCEAR, Pune.
- Pisharody, P.N. 1965. Forms and distribution of Fe and Mn in rice soils of Kerala. M.Sc.(Ag.) thesis, University of Kerala, Thiruvananthapuram, P.72
- Piper, C.S. 1966. *Soil and Plant Analysis*. Asia Publishing House, Bombay. 358 p.
- Praseedom, R. 1970. Distribution of Cu and Zn in the soils of Kerala. M.Sc(Ag.) thesis Kerala Agricultural University, Thrissur, p.71
- Prasad, A.S. 2008. Zinc in human health; effect of zinc on immune cells. *Mol. Med.* 14 (5-6):353-7
- Prasad, B. and Umar, S.M. 1993. Direct and residual effect of soil application of Zn sulfate on yields and Zn uptake in rice wheat rotation. *J. Indian Soc. of Soil Sci.* 41: 192-194
- Prasad, R., Prasad, B.L. and Sakal, R. 1996. Effect of submergence on the transformation of Zinc forms in old alluvial soils growing rice as related to soil properties II. Transformation of applied Zn. *J. Indian Soc. Soil Sci.* 44(1): 74-76

- Prasuna rani,P.,Pillai,R,. N., Bhanuprasad,V.and Subbaiah, G.V.1992. Nutrient status of some red and associated soils of Nellore district under Somasila project in Andhra Pradesh. *The Andhra agric.J.* 39:1-5
- Rajagopalan.V.1969. Distribution of manganese and molybdenum in the soils of Kerala and the effect of molybdenum on the growth of cowpea.M.Sc.(Ag.) thesis, University of Kerala Thiruvananthapuram,p.75
- Rajendran,P. 1981. Manganese and zinc status of rice soils of Kerala. M.Sc.(Ag.) thesis, Kerala Agricultural University, Thrissur p.88
- Ramesh, V. 1994. Foliar diagnosis and yield production in sugarcane in relation to NPK. M.Sc.(Ag.) thesis, Kerala Agricultural University,Thrissur,P.110
- Randhawa, H.S. and Singh, S.P. 1995. Zinc fractions in soils and their availability to maize. *J. Indian Soc. Soil Sci.* 43(2): 293-94
- Rattan,R.K.,Sahran,N.and Datta,S.P.1999. Micronutrient depletion in Indian Soils:extent,causes and remedies .*Fert.News* : 44(2):43-50
- Ravikiran, S. and Reddy, G. L. 2004. In vivo fortification of Zinc in rice (*Oryza sativa*. L.). *The Andhra Agric. J.* 51(182): 184-187
- Reddy, M.R. and Perkins, H.F. 1974. Fixation of Zn by clay minerals. *Soil Sci. Soc. Am. Proc.* 38, 229-230
- Reddy, B. and Ahlawat,Y.S. 2002. DARE/ICAR report on lentil. <http://www.icar.org.in/files/icar.2001.pdf>
- Ricemann,D. S. and Jones,G. B.1958. Distribution of Zn in Subterranean clover (*Trifolium subterranean* L.) grown to maturity in a culture solution containing Zn labelled with radioactive isotope Zn <sup>65</sup>. *Aust. J. Agric. Res.* 9,730-744

- Romney et al. 1965. Soil Fertility Evaluation. Jose, A.I. 2005. Proc. Soil and Plant analysis, (ed. John, S.), 12-17, December, 2005, CTCRI, Trivandrum, 38-42
- Rosell, R. A. and Ulrich, A. 1964. Critical zinc concentrations and leaf minerals of sugar beet leaves. *Soil Sci.* 97, 152-67.
- Sadasivam, S. and Manickam, A. 1992. Biochemical methods for agricultural sciences. Wiley International Ltd. New Delhi, p.246
- Saeed, M. and Fox R.L. 1973. Relations between suspension pH and Zn solubility in acid and calcareous soils. *Soil Sci.* 124, 199-204
- Sahu, S.K., Mitra, G.N. and Pani, S.C. 1996. Effect of Zn application on uptake of nutrients by rice on an Inceptisol. *J. Indian Soc. Soil. Sci.* 44(3): 564-566
- Saifudeen, N. 1981. Foliar diagnosis, yield and quality of turmeric in relation to NPK. M.Sc. Thesis, Kerala Agricultural University, Thrissur, P.92
- Sakal, R. 2001. Efficient management of micronutrient for sustainable crop production. *J. Indian Soc. Soil Sci.* 49(4) : 593-608
- Sakal, R., Singh, A.P., Sinha, H. and Takkar, P.N. 1981. Evaluation of critical concentration of Zn in rice and wheat grown in Tarai Soils. *J. Indian Soc. Soil. Sci.* 29(1): 107-109
- Sarkar, A.K. and Singh, S. 2003. Crop response of secondary and micronutrients in acidic soils of Inida. *Fert. New* 48, 47-54
- Sarkar, I., Baruah, U., Gangopadhyay, S. K., Sahoo, A. K. and Velayudham, M. 2002. Characteristics and classification of soils of Loktak Command Area of Manipur for sustainable land use planning. *J. Indian Soc. Soil. Sci.* 50:196:204
- Sathyanarayanan, R. 1997. Distribution of extractable micronutrients in soils and selected major land resources of Kerala. M.Sc. Thesis, Kerala Agricultural University, Thrissur, P.98

- Satisha, G.C., Karthikakuttyamma, M., Paul, T.K., Dey, S.K. and Varghese, Y.A. 2002. Rubber growing soils of Northern Mizoram their characteristics of and limitations. *Indian J. Soil Sci.* 50:123-125
- Schnitzer, M. and Khan, S.U. 1978. *Soil Organic Matter*. Elsevier, Amsterdam. P.115
- Schnitzer, M. and Skinner, S.I.M. 1966. Organo metallic interaction in soils. 5. Stability constants of  $\text{Cu}^{++}$ ,  $\text{Fe}^{++}$  and  $\text{Zn}^{++}$  fulvic acid complexes. *Soil Sci.* 102, 361-365
- Sen, T.K., Dubey, P.N., Maji, A.K. and Chamvam, G.S. 1994. Status of micronutrients in some dominate soils of Manipur. *J. Indian Soc. Soil. Sci.* 45: 388-390
- Sharma, B.L and Bapat, P.N. 2000. Levels of micronutrient cations in various wheat parts of wheat as influenced by Zn and P application. *J. Indian Soc. Soil Sci.* 48 (1):130-134
- Sharma, B.L., Bhadoria, A.K.S., Rathore, G.S. and Bapat, P.N. 1996. Evaluation of Extractants for available Zn and its forms in vertisols of Madhya Pradesh. *J. Indian Soil Sci.* 44(4): 701-704
- Sharma, B.L., Rathosa, S.B., Dubey, S.B., Khambaria, R.S and Singh, S.B. 1986. Response of rice to Zn and evaluation of soil test methods for Zn. *J. Indian Soil Sci.* 3: 106-10
- Sharma, S.S., Totawat, K.L. and Shyampura, R.L. 2004. Characterization and classification of salt affected soils of Southern Rajasthan. *J. Indian Soc. Soil Sci.* 52, 209-213
- Sheeja et al. (1996). In: micronutrient management for sustaining crop productivity with special reference to Kerala. Geethakumari, V, L., Mathew, U. and Varghese, S.S. 2005. Proceedings of the National Workshop on soil resource management (eds. Kunnungal, M. C. and Maghi, E. K), 22-24, December. 2005. Soil Survey Organization, Trivandrum, p. 310-323

- Shivoy, Y.S. and Kumar, D. 2005. Effect of P and Zn fertilization on the productivity of transplanted aromatic rice. Proceedings of an International Workshop on micronutrients in South and South East Asia held in Kathamandu, Nepal, 8-11 Sept. 2004
- Shorrocks, P.1962: Soil Fertility Evaluation. Jose, A.I. 2005. Proc.Soil and Plant analysis, (ed.John, S.), 12-17, December, 2005, CTCRI, Trivandrum, 38-42
- Shover, T.M., Westfall, D.G. and Ronaghi, M. 2007. Zinc fertilizer solubility and its effects on zinc bioavailability over time. J. Plant Nutr. 30: 123-133
- Shuman, L. M . 1985. Fractionation methods for soil microelements. Soil Sci.140, 11-22.
- Shuman, L. M . 1995. Effect of ionic strength and anions on zinc adsorption by two soils. Soil Sci. Am. J.50:1438-1442
- Shuman, L. M., White,R.J.and Drwsen.E. 1995. Zinc in Plants. (ed. Nriagu,V),John Wiley & Sons,London.p.125
- Singh,M. V. 1998. Progress Report,1996, AICRP on secondary and micronutrients and pollutant elements in soils and plants.IISS,Bhopal.
- Singh. S., Singh,B.and Pande,R,1992. Response of barley to the application of Zn SO<sub>4</sub> .agric. Res. 5:17-18
- Singh, A.K., Khan, S.K. and Nongkynrih, P. 1999. Transformation of Zn in wetland rice soils in relation to nutrition of rice crop. J. Indian Soc. Soil Sci. 47(2): 248-253
- Singh, A.K., Nongkynrih, P., Sachdev, P. and Sachdev, M.S. 2001. Utilization of applied Zn by rice crop in wetland acidic soils. J. Nuclear Agrl. Biol. 30 (3&4): 148-151
- Singh, B. 2009. Phytosiderphores improve Zn deficiency of cereals. ICAR News letter, 15(2)-April –June, 2009

- Singh, B. and Sekhon, G.S. 1977. The effect of soil properties on adsorption and desorption of zinc by alkaline soils. *Soil Sci.* 124:366-369
- Singh, B., Vaidya, P.K. and Lal, R. 1993. Response of barley to Zn application as influenced by genotypic variability. *Soil Plant* 42(5):7-8
- Singh, R.K and Choudhary, B. D. 1985. Biometrical methods in quantitative genetic analysis, Kalyani Publishers, New Delhi
- Singh, S.P. and Nayar, V.K. 1993. Critical limit of Zn for cotton on a Typic ustrochrept. *J. Indian Soc. Soil Sci.* 43(3): 479-481
- Singh, S.P. and Nayar, V.K. 1997. Critical deficiency level of Zn in soyabean and green gram. *J. Indian Soc. Soil Sci.* 45(1): 201-202
- Singh, I. S. and Agrawal, H. P. 2005. Characterization, genetic and classification of rice soils of Eastern Region of Varanasi, Uttar Pradesh, *Agropedology*, 15: 29-38
- Singh, Y. P. 2004. Effect of N and Zn on wheat irrigated with alkali water. *Ann. Agric. Res. News service.* 25 (2): 233-236
- Soil Survey Staff. 1998. *Keys to Soil Taxonomy*. Eighth Edition Natural Resource conservation service, USDA, Blacksburg, Virginia
- Sommer, A.L. and Lipman, C.B. 1926. Evidence on the indispensable nature of zinc and boron for higher green plants. *Plant Physiol.* 231-249
- Sreenivasan, V., Hamza, S. and Dinesh, R. 2009. Critical limits of Zn in soil and plant for increased productivity of ginger (*Zingier officinale* Rosc.). *J. Indian Soc. Soil Sci.* 57(2): 191-195
- Stevenson, F.J. 1991. Organic matter macronutrient reactions in soil. In *macronutrients in Agriculture*. (eds. Mortvedt, J.J., Cox, F.R., Shuman, L.M. and Welch, R.M). *Soil Sci. Soc. Am.* Madison, U.S.A P. 145-186

- Stevenson, F.J. and Adrakani, M.S. 1972. Organic matter reactions involving micronutrients in soils. (Eds. Mortvedt, J.J., Giordano, P.M. and Lindsay, W.L) Soil Sci.Soc.Am. Madison, U.S.A P. 79-114
- Subbaiah, B.V. and Asija, G.L. 1956. A rapid procedure for estimation of available nitrogen in soils. Curr. Sci. 25: 259-260.
- Suge, H., Takahshi, H., Arida, S. and Takaki, H. 1986. Gibberellin relationship in Zinc deficient plants. Plant Cell Physiol. 27 : 1010-1012
- Sureshkumar, P., Rattan, R.K. and Singh, A.K. 2004. Chemical forms of Zn in soils and their contribution to available pool. J. Indian Soc. Soil Sci. 52(4): 421-425
- Sushama, P.K. 1980. Foliar diagnosis, yield and quality of pepper in relation to NPK. M.Sc.Thesis, Kerala Agricultural University, Thrissur, P.95
- Sushama,M.M.,Deotale,R. D.,Wandile,R. M. and Dighe,R. S. 2003. Influence of zinc sulfate on growth and yield of mustard (*Brassica juncea*) .J. Soils Crops 13(2): 397-399
- Swaine ,D. J. 1955. The trace element content of soils .Commonw. Bur. Soil Sci. Tech. Commn. No. 48. Commonw. Agric. Bur. Farnham Royal,U.K
- Swaine ,D. J. and Mitchell,R. L. 1960. Trace element distribution in soil profiles. J.Soil Sci.11,347-368
- Takkar, P.N. 1982. In review of soil research in India,part1.p.361.
- Takkar ,P.N. 2006. Zinc deficiency in soil and plants. J.agric. Res.4(3): 28-32
- Takkar ,P.N. and Walkar, C.D.1993. The distribution and correction of Zn deficiency .In: Zinc in Soils and Plants (ed.Robson, A. D.), Kluwer Accad. Publ.Dordrecht,The Netherlands, p.151-165

- Tandon,H.L.S. 1989. Nutrient status of soils and crops. In: Secondary and micronutrient recommendation for crops-A Guide book, Fertilizer Development and Consultation Organization ,New Delhi,p.18-32
- Thampatty,K. C. M. 1997. Morphological, physical and chemical characterization of soils of North Kuttanadu. Ph. D thesis,Kerala Agricultural University, Thrissur,P.210
- Thangasamy,A., Naidu,M. V. S. and Ramavatharam,N. 2004. Clay mineralogy of soils in the Sivagiri microwatershed of Chittoor district,Andhra Pradesh. J.Indian Soc.Soil. Sci . 52: 454-461
- Thangasamy,A. and Reddy, R.C. 2005. Characterization , classification and evaluation of soil resources in the Sivagiri microwatershed of Chittoor district,Andhra Pradesh for sustainable land use planning . J.Indian Soc.Soil. Sci . 53: 11-21
- Tiller,K. G. 1963. Micronutrients. In: Soils: an Australian view point,CSIRO, Melbourne/ Accad.Press , London ,pp. 368-387
- Tiwari, K.N. 2008. The Professor J.N.M. Future Plant Nutrition Research in India. J.Indian Soc.Soil Soc.56 (4): 327-336
- Tiwari, V. N., Tripathi, S. K., Tiwari, K. N and Pandey, R. K. 2006. Nutritional studies on yield and quality of some genotypes of groundnut in alluvial soils of Uttar Pradesh. Farm Sci. J. 15(1): 9-12
- Tsui, C. 1948a. The role of Zn in auxin synthesis in the tomato plant. American J. Bot. 35, 172-179
- Tsui, C. 1948b. The effect of Zinc on water relation and osmotic pressure of the tomato plant. American J. Bot. 35, 309-311
- Umamaheswari.P.and Singh,C.P.2002. Influence of irrigation levels and micronutrients (Fe and Zn)on yield and yield attributes of rajmash (*Phaseolus vulgaris*).www.crida.ernet.in/AICRPDA/AR02-03.pdf



- Usha,P. B. 1995. Evaluation of acidity parameters in wet land soils of Kerala in relation to nutrient availability . Ph. D thesis,Kerala Agricultural University, Thrissur,p.200
- Valsaji,K. 1972. Studies on the status and distribution of Cu and Zn in two soil series of Trivandrum district. M.Sc. (Ag.) thesis, University of Kerala,Trivandrum P.75
- Varshney, P., Singh, S.K. and Srivasthava, P.c. 2008. Frequency and rates of zinc application under hybrid rice wheat sequence in a mollisol of Uttarakhand. Indian Soc. Soil Sci. 56(1): 92-98
- Vaseghi, S. Afuni, M. Shariamadari. H. and Mobli. M. 2003. Effect of sewage sludge and soil pH on micronutrient and heavy metal availability. J. Sci. Technol. agric Natural Res. 7(3): 96-106
- Velesco and Nevero. 1951. In: Soil Fertility Evaluation. Jose, A.I. 2005. Proc.Soil and Plant analysis, (ed. John, S.), 12-17, December, 2005, CTCRI, Trivandrum, 38-42
- Verma,C.B., Lallu,S. and Yadav,.R. S. 2004. Effect of boron and zinc application on growth and yield of pigeon pea. Indian .J. Pulses Res. 17(2): 149-151
- Viets, F. G. 1962. Chemistry and availability of micronutrients in soils. Agric. Food Chem, 10: 174-178
- Walia, C.S. and Rao, Y.S. 1997. Characteristics and classification of some soils of Trans-Yamuna Plain. J. Indian Soc. Soil Sci. 45: 156-162
- Walkley, A. and Black, A.A. 1934. An examination of the Degjareff method for determining soil organic matter and a proposed modification of the chromic acid titration method. Soil Sci. 37: 29-34
- Wedepohl, K. H.1972. Zinc. In; Handbook of Geochemistry.Vol.11& 111,Springer-Verlag,NewYork, p,125

Weinhold, A. 1862. Landw vers Sta. 4, 188

Welch, R.M. and Norvell, W.A. 1993. Growth and nutrient uptake by barley (*Hordeum vulgare* L. cv. Herta). Studies using an N- (2 hydroxy ethyl) ethylene dinitrotri-acetic acid buffer nutrient solution technique II. Role of zinc in the uptake and root leakage of mineral nutrients. Plant Physiol. 101, 627-631

Yaduvanshi, H.S. and Sharma, K.L. 1988. Zinc fractionation in acid alfisol under long term manuring and sequential cropping system. J. Indian Soc. Soil Sci. 36:173-175

Yaduvanshi, H.S., Tripathi, B.R. and Kanwar, B.S. 1985. Effect of continuous manuring on some soil properties of an Alfisol. J. Indian Soc. Soil Sci. 33 : 700-703

Yoshida, S. and Tanaka, A. 1969. Zinc deficiency of rice plants in calcareous soils. Soil Sci. Pl. Nutr. 19 (12): 81-83

Yoshida, S. Ahri, J.S. and Formo, D.A. 1973. Occurrence, diagnosis and correction of Zn deficiency of low land rice. Soil Sci. Pl. Nutr. 19 (12): 83-93.

## APPENDIX I

### Weather data during the cropping period

(October, 2007 to September, 2008)

Months	Relative Humidity (%)	Temperature (° C)	Total Rainfall ( mm)
Oct ,2007	90.46	30.61	141.16
Nov,2007	91.12	30.84	186.20
Dec,2007	89.07	30.55	3.60
Jan,2008	86.59	31.45	0.00
Feb,2008	85.13	31.94	38.60
Mar,2008	88.88	31.37	276.21
Apr,2008	84.65	32.32	74.80
May,2008	85.62	32.31	153.20
Jun,2008	88.40	30.98	96.90
Jul,2008	90.90	29.50	200.00
Aug,2008	90.72	29.72	276.60
Sep,2008	89.25	28.91	253.00

## APPENDIX II

### Nutrient content of the manure and fertilizers used for the experiment

Manure/Fertilizer	Nutrient content
FYM	
N (%)	0.55
P (%)	0.24
K (%)	0.62
Ca (%)	0.65
Mg (%)	0.12
S	010
Fe (ppm)	138
Mn (ppm)	57
Zn(ppm)	60
Cu(ppm)	2.3
Urea	46 % (N) : 4% (Zn)
RP	18 % (P)
Muriate of Potash	60% (K)
ZnSO <sub>4</sub> .7H <sub>2</sub> O	21% (Zn),10% (S)

### APPENDIX III

#### Mean values of different fractions of Zinc in the surface soil

Treatment	Ws+Ex-Zn	Ob - Zn	MnO-Zn	AFe -O-Zn	Cry Fe-O-Zn	Res- Zn	Total Zn
T1	3.05 (2.66)	3.39 (2.91)	0.99 (0.95)	34.86 (30.18)	37.97 (32.99)	35.24 (30.31)	115.50
T2	1.49 (1.39)	2.42 (2.26)	0.65 (0.61)	37.77 (35.30)	40.42 (37.78)	24.25 (22.66)	107
T3	3.99 (2.97)	3.02 (2.24)	1.11 (0.83)	26.54 (19.73)	60.97 (45.33)	38.87 (28.90)	134.50
T4	4.51 (3.35)	2.81 (2.09)	1.24 (0.92)	27.33 (20.32)	70.80 (52.64)	27.81 (20.68)	134.50
T5	5.25 (3.99)	2.75 (2.09)	0.92 (0.70)	23.34 (17.75)	72.50 (55.13)	26.74 (20.34)	131.50
T6	3.14 (3.00)	3.5 (3.34)	0.99 (0.95)	26.55 (25.40)	31.76 (30.40)	38.55 (36.91)	104.50
T7	3.04 (2.39)	2.89 (2.28)	0.45 (0.35)	25.47 (20.05)	48.77 (38.40)	46.38 (36.53)	127.00
T8	2.58 (2.12)	2.45 (2.02)	0.55 (0.45)	27.37 (22.52)	63.19 (52.01)	25.36 (20.88)	121.50
T9	4.04 (2.66)	2.55 (1.68)	0.55 (0.36)	29.43 (19.36)	90.17 (59.32)	25.26 (16.62)	152.00
T10	0.45 (0.53)	2.15 (2.54)	0.21 (0.25)	19.32 (22.86)	38.46 (45.51)	23.91 (28.31)	84.50

Values in the paranthesis shows the % share of individual fractions to total.

## APPENDIX IV

### Mean values of different fractions of Zinc in the sub surface soil.

Treatment	Ws+Ex-Zn	Ob- Zn	MnO-Zn	A Fe-O-Zn	Cry-Fe-O-Zn	Res- Zn	Total Zn
T1	1.09 (1.18)	0.90 (0.98)	1.24 (1.35)	23.53 (25.57)	33.86 (36.80)	31.40 (34.12)	92.00
T2	1.14 (1.22)	2.03 (2.18)	0.68 (0.73)	24.82 (26.69)	39.83 (42.83)	24.50 (26.35)	93.00
T3	1.74 (1.48)	2.04 (1.69)	1.09 (0.90)	15.20 (12.67)	52.28 (43.39)	48.15 (39.87)	12.50
T4	1.37 (1.18)	1.96 (1.69)	1.50 (1.29)	37.38 (32.22)	42.83 (36.92)	30.96 (26.68)	116.00
T5	1.66 (1.57)	0.83 (0.79)	1.14 (1.08)	25.61 (24.27)	52.65 (49.90)	23.61 (22.39)	105.50
T6	1.72 (1.90)	1.68 (1.86)	0.89 (0.98)	27.60 (30.49)	36.24 (40.04)	22.37 (24.73)	90.50
T7	1.26 (1.39)	1.16 (1.28)	0.86 (0.95)	19.40 (21.44)	37.58 (41.54)	30.19 (30.24)	90.50
T8	2.28 (2.02)	1.39 (1.24)	1.42 (1.26)	21.75 (19.34)	53.80 (47.82)	31.86 (28.32)	112.5
T9	1.77 (1.57)	1.17 (1.04)	0.82 (0.72)	25.27 (22.30)	38.65 (34.30)	45.32 (40.07)	113
T10	0.11 (0.15)	0.45 (0.63)	0.51 (0.71)	16.06 (22.46)	29.67 (41.50)	24.69 (24.70)	71.5

Values in the paranthesis shows the % share of individual fractions to total.

## **APPENDIX V**

### **Input Cost and market price of produce**

1kg urea- Rs.5.50/-;

1 kg RP- Rs.5.50/-;

1kg MOP-Rs.5/-

1kg ZnSO<sub>4</sub>.7H<sub>2</sub>O –Rs.30/- ;

1kg seed-Rs.15/-

1kg fodder-Rs.3/-;

Labour charge –Rs.175/-

**DYNAMICS OF ZINC IN TYPIC KANDIUSTULTS WITH SPECIAL  
REFERENCE TO NUTRITION IN FODDER MAIZE (*Zea mays* L.)**

**THANKAMONY, K.**

**ABSTRACT OF THE THESIS**

**Submitted in partial fulfillment of the requirement  
for the degree**

**DOCTOR OF PHILOSOPHY  
Faculty of Agriculture  
Kerala Agricultural University**

**DEPARTMENT OF SOIL SCIENCE AND AGRICULTURAL CHEMISTRY  
COLLEGE OF AGRICULTURE  
VELLAYANI, THIRUVANANTHAPURAM, KERALA**

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## ABSTRACT

Zinc is an essential mineral nutrient with exceptional biological significance in plant and animal nutrition. It is associated with several enzyme systems that regulate various metabolic activities in plants.

In Indian Scenario more than 45% of soils are Zn deficient. Majority of the Kerala soils also recorded a low content of available Zn. Proper understanding of micronutrient availability in soil and extent of their deficiencies is the pre-requisite for efficient management of micronutrient fertilizer to sustain crop productivity.

The response of Zn application varies widely from crop to crop and soil to soil. Maize crop was found to be responsive to Zn. Fodder crops respond to micronutrients by increasing the dry matter yields and by changing their quality as a feed for animal nutrition.

In this context the present study on the Dynamics of zinc in Typic Kandiusults with special reference to nutrition in fodder maize (*Zea mays*.L) was carried out to satisfy the objectives viz. understanding the release pattern of Zinc from Zinc sulphate with and without FYM under the laboratory conditions, to elucidate the role of Zn in the growth, yield and quality parameters of fodder maize. when applied to soil as well as foliar application. The selection of the index plant part for foliar diagnosis and fixing up of the critical nutrient level in that plant part was also included in the experiment.

The incubation study on the release pattern of Zn from zinc sulphate was conducted in complete randomized design with 10 treatments and three replications. The treatments are given below:

T <sub>1</sub>	Soil alone:	T <sub>2</sub>	Soil + Zn @ 0 kg ha <sup>-1</sup> + FYM @ 10
tha <sup>-1</sup>			
T <sub>3</sub>	Soil + Zn @ 5 kg ha <sup>-1</sup> :	T <sub>4</sub>	Soil + Zn @ 5 kg ha <sup>-1</sup> + FYM
@ 10 tha <sup>-1</sup> :			
T <sub>5</sub>	Soil + Zn @ 10 kg ha <sup>-1</sup> :	T <sub>6</sub>	Soil + Zn @ 10 kg ha <sup>-1</sup> + FYM
@ 10 tha <sup>-1</sup> :			
T <sub>7</sub>	Soil + Zn @ 15 kg ha <sup>-1</sup> :	T <sub>8</sub>	Soil + Zn @ 15 kg ha <sup>-1</sup> + FYM
@ 10 tha <sup>-1</sup> :			
T <sub>9</sub>	Soil + Zn @ 20 kg ha <sup>-1</sup> :	T <sub>10</sub>	Soil + Zn @ 20 kg ha <sup>-1</sup> + FYM
@ 10 tha <sup>-1</sup>			

The soil samples were collected at an interval of 15 days for a period of 75 days and analyzed for available Zn by DTPA method.

The results of the incubation study showed that when Zn was applied at 5, 10, 15 kg ha<sup>-1</sup>, the availability was in the medium range during the period of incubation and it has exceeded 2.4mg kg<sup>-1</sup> on the 75<sup>th</sup> day of incubation only, when Zn was applied @ 20 kg ha<sup>-1</sup>.

Initial soil profile study and the initial analysis of the surface and subsurface soil samples of experimental site were also carried out.

The field study comprised of two experimental crops with fodder maize cv. African Tall, in the Instructional Farm, College of Agriculture, Vellayani in RBD with 10 treatments and 3 replications. The treatments are given below:

T<sub>1</sub> : NPK @ 120:60:40 kg ha<sup>-1</sup> ; T<sub>2</sub> : NPK @ 120:60:40 kg ha<sup>-1</sup> + FYM @ 10 t ha<sup>-1</sup> (POP recommendation); T<sub>3</sub> : POP + Zn 5 kg ha<sup>-1</sup> as ZnSO<sub>4</sub>(Soil application); T<sub>4</sub> : POP + Zn 10 kg ha<sup>-1</sup> as ZnSO<sub>4</sub>(Soil application); T<sub>5</sub> : POP + Zn 15 kg ha<sup>-1</sup> as ZnSO<sub>4</sub>(Soil application); T<sub>6</sub> : POP + Zn 10 kg ha<sup>-1</sup> as ZnSO<sub>4</sub>(Soil application); T<sub>7</sub> : POP + 0.25% Zn as ZnSO<sub>4</sub>(foliar application); T<sub>8</sub> : POP + 0.5% Zn as ZnSO<sub>4</sub> (foliar application); T<sub>9</sub> : POP + 0.75% Zn as ZnSO<sub>4</sub>(foliar application); T<sub>10</sub> : Control.

After each experimental crop, a residual crop was also raised without any fertilizers or manures to study the residual effect of applied zinc for the previous crop.

The yield data of the first experimental crop revealed that T<sub>4</sub> (POP + Zn 10 kg ha<sup>-1</sup> as ZnSO<sub>4</sub> as soil application), recorded the highest yield of 512 q ha<sup>-1</sup>. The higher dry fodder yield (103.92 q ha<sup>-1</sup>) per day green fodder yield (8.54 q ha<sup>-1</sup>) and leaf stem ratio (2.63) were also higher for the above treatment. The quality attributes viz. crude protein (10.93 per cent), crude protein yield were also showed for this treatment. For the first residual crop, T<sub>5</sub> (POP + Zn 15 kg ha<sup>-1</sup>) recorded the maximum yield. In the second experimental crop, the T<sub>5</sub> showed the maximum yield (519.3 q ha<sup>-1</sup>). For the second residual crop, T<sub>6</sub> (POP + Zn 20 kg ha<sup>-1</sup> as ZnSO<sub>4</sub>) of soil applied Zn gave the maximum yield.

The yield data of the first and second experimental crops were pooled and analyzed statistically. The pooled analysis data for the yield was found significant among treatments. For the pooled data, the T<sub>5</sub> (POP + Zn 15 kg ha<sup>-1</sup>) with a mean yield of 468.8 q ha<sup>-1</sup> showed the highest yield and it was on par with T<sub>4</sub>, T<sub>8</sub> and T<sub>9</sub> which recorded and yield of 464.5 q ha<sup>-1</sup>, 403.90 q ha<sup>-1</sup>, 418.30 q ha<sup>-1</sup> respectively. It is seen that the yield difference between T<sub>4</sub> and T<sub>5</sub> was only 4.3 q ha<sup>-1</sup>. For raising a single crop of fodder maize (African Tall), soil application of zinc sulphate @ Zn 10 kg ha<sup>-1</sup> along with 120:60:40 NPK kg ha<sup>-1</sup> + FYM 10 t ha<sup>-1</sup> is recommended. Foliar spray of zinc sulphate @ 0.75% Zn at the 4<sup>th</sup> leaf stage recorded the highest yield among foliar application treatments and it was on par with T<sub>4</sub>.

Application of Zn @ 15 kg ha<sup>-1</sup> as soil application along with 120:60:40 NPK kg ha<sup>-1</sup> + FYM 10 t ha<sup>-1</sup> for the first crop of fodder maize will help to skip the Zn application for the succeeding crop.

From the studies on foliar diagnosis to identify the index plant part of this crop, 3<sup>rd</sup> leaf lamina from the tip at 25 DAS was selected as the index plant part. The

critical nutrient level of Zn for maximum response to yield in plant was standardized using the graphical method proposed by Cate and Nelson (1971). The content of Zinc in the 3<sup>rd</sup> leaf lamina was found to be 21 mg kg<sup>-1</sup>. Foliar spray @ 0.75% Zn during the crop growth is recommended on identifying zinc deficiency to the crop. Zinc is an essential element for plant growth and metabolism, and exists in soil in different fractions such as Ws+Ex, OB, MnO, AmFeO, Cry FeO, and Res and all these forms are existed in a dynamic equilibrium. So the amount and rate of transformation of these forms determine the size of the labile pool of Zinc. The different fractions of Zn was also studied for the surface (0-15 cm) and subsurface soil (15-30 cm). The percentage share of different fractions in the surface soil to the total Zn was as follows i.e, Ws+exch.Zn (0.53-3.99%), OB-Zn(2.15-3.34%), MnO-Zn (0.21-0.92%), Am. FeO-Zn (17.75-35.30%) and Cry-Fe-O Zn (30.40-59.32%) and RS-Zn (16.62-36.91%).

A path coefficient analysis was also carried out with the different fractions of Zinc on the yield of fodder maize and the results revealed that direct effect of water soluble + exchangeable fraction soil Zn on the yield of fodder maize.