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## EFFECTIVENESS AND COMPETITIVE ABILITY OF THE ANTIBIOTIC RESISTATNT MUTANTS OF RHIZOBIUM SP.\*

Infectiveness and effectiveness are two genetic characters of the genus *Rhizobium* which permit the organism to enter into a symbiotic association with the host legume to form nodules for nitrogen fixation. A method used in genetic studies on *Rhizobium* for isolation of ineffective and non-ineffective mutants is the selection of strains resistant to certain antibiotics. However, the results reported so far present a controversial picture of the relationship between resistance and symbiotic nitrogen fixation of *Rhizobium* (Damary and Alexander, 1969, Zelazna–Kowalska, 1971, Levin and Montgomery, 1974).

Present study was carried out to determine the effectiveness and compettive ability of the mutants of *Rhizobium* sp. to streptomycin, erythromycin and kanamycin, Native *Rhizobium* sp. from nodulating pigeonpea (*Cajanus cajan* Mill sp.) plants was isolated and a clonal population was raised which was designated as AS<sub>1</sub>. Yeast extract mannitol (YM) was used as the growth medium. Exponentially growing cells of AS<sub>1</sub> were plated on YM plates containing 10,100 and 500 $\mu$ g/ml of the each antibiotic (streptomycin, erythromycin add kanamycin) and spontaneous antibiotic resistant mutants were isolated randomly (Vincent, 1970).

Plant nodulation test was carried out in pigeonpea cultivar Bahar in a randomised block design with nine replications of each treatment. Surface sterilized seeds of pigeonpea were germinated under aseptic conditions. The 48h old seedlings were transferred to the agar slants prepared in large glass tubes (200 x 38 mm) supplemented with Thornton's nutrient medium (Vincent, 1970) and 1 ml of exponentially growing cultures of *Rhizobium* strains under test was poured on the roots-When the seedlings attained appropriate height, the shoot portion was pushed out gently under aseptic condition. Plants were grown under fluorescent lights in a growth chamber on an 18 h/day illumination at  $27 \pm 2^{\circ}$ C and were watered regularly with Thorton's broth. After sixty days, the plants were taken out and length of the shoot, total number of nodules per plant, dry weight of the nodules, plant dry weight as well as total nirogen content per plant (Micro-kjeldahl method) were determined.

Based upon the data regarding shoot length, number of nodules per plant, nodule dry weight, plant dry weight and total nitrogen content per plant of the inoculated pigeonpea plants (Table 1), all the antibiotic resistant mutants were classified into four groups viz., '(i) Non-infective  $(kan_s)$  (ii) Partially effective  $(str_1, str_2, str_3, ery_1, ety, and kan_s)$  (iii) Effective  $(kan_1)$  and (iv) Very effective  $(ery_2)$ .

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### Research Notes

Strain kan<sub>s</sub> was non-infective whereas  $ery_2$  showed hyper efficiency in nitrogen fixation. This hyper efficiency might be due to increased activity of nitrogen fixing (*nif*) genes (Jagat Ram, 1979).

The loss of infectiveness of streptomycin resistant mutants (Maier and Brill 1978J and kanamycin resistant mutants(Damery and Alexander, 1969) was due to an altered cell wall and lipopoiysaccharide suggesting that the lipopoiysaccharide was a receptor for plant attachment. Mutants with altered cell wall and lipopolysaccharide may not form nodules (Vincent, 1980). Here also the non-infectivity of kan<sub>3</sub> might be due to the above explained reason.

Out of the nine mutants isolated, only two mutants namely  $ery_{2}$  and  $kan_{1}$  were effective in nitrogen fixation. This result indicates that these mutants may not have defect in general cell surface or function. However, in this study the rest of the mutants ( $str_{1}$ ,  $str_{2}$ ,  $str_{3}$ ,  $ery_{1}$ ,  $ery_{3}$  and  $kan_{2}$ ) were inferior to the parent strain This varied effect of nitrogen fixation might be due to different mechanisms of resistance in such cases.

These marked effective rhizobial strain can be used for the competition studies for knowing the efficiency of field inoculation.

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Treatment	Length of shoot (cm)	Total no. of nodules	Nodule dry weight (mg)	Dry weight of plant (mg)	Total N/ plant (mg)
Control	25.83	*	*	159.00	5.82
AS,	30.33	20.33	12.66	191.00	9.33
str,	29.66	19.66	9.33	180.00	6.83
str <sub>2</sub>	28.83	17.66	12.00	184.66	8.85
str.	29.66	16.00	9.60	166.33	6,81
ery,	28.00	17.66	9.66	166.66	6,06
ery <sub>2</sub>	33,66	29.66	19.00	246.33	16.20
ery <sub>3</sub>	29.83	17.00	11.00	174.66	6.82
kan,	32.00	20,66	12.73	192.66	9.30
kan <sub>2</sub>	25.50	13.00	2.00	165.00	6.45
kan <sub>s</sub>	26.66	*	*	159.80	5.85
CD (0.05)	2.18	1.05	0.45	4.28	0.066

#### Table 1

# Different plant characters of pigeonpea plants inoculated with parent and mutant strains of *Bhizobium* sp

111

No nodules

### moloano

തുവരയുടെ വേരുകളിൽ വളരുന്ന rararairoT ക്ഷത്തിൽ നിന്ന് പാകൃജനകം സംഭരി കുന്ന റൈസോബിയം എന്ന ബാക്ടീരിയയിൽ സ്റ്റെപ്ടോമൈസിൻ, എറിത്രോമൈസിൻ, കനാമൈസിൻ എന്നീ ആൻറീബയോട്ടിക്കുകയംക്കെതിരെയുള്ള ഒൻപതു തരം ബാക്ടീരിയക ളെ വേർതിരിച്ചെടുക്കുകയും, അവ എത്രമാത്രം പാക്യജനകം ബഹാർ എന്നയിനം തുവരയിൽ സംഭരിക്കുന്നു എന്ന് ഇൻസ്റ്റിറ്റ്യൂട്ട് ഓഫ് അഗ്രിക്കാച്ചറൽ സയൻസസ്, ബനാറസ് ഹിന്ദു സർവകലാശാലയിൽ പരീക്ഷണത്തിന് വിധേയമാക്കി. എറിത്രോമൈസിൻ, കനാ മൈസിൻ എന്നീ ആൻറീബയോട്ടിക്കുകയംക്കതിരെ പ്രതിരോധശക്തിയുള്ള രണ്ടുതരം റൈ സോബിയം ബാക്ടീരിയകയം സാധാരണ രീതിയിൽ പാക്യജനകംസംഭരിക്കുന്നു എന്നും, കനാ മൈസിൻ പ്രതിരോധശക്തിയുള്ള റൈസോബിയം വേരുകളിൽ മൂലാർബുദങ്ങയം ഉണ്ടാക്കു വാൻശേഷി നശിച്ചതായും ബാക്കിയുള്ളവ സാധാരണ രീതിയിൽ നിന്ന് താഴ്ന്ന നിലവാര ത്തിൽ പാക്യജനകം സംഭരിക്കുന്നതായും തെളിഞ്ഞു.

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### References

- Damery, J, T. and Alexander, M. 1969. Physical differences between effective and ineffective strains of *Rhizobium.* Soil Sci. 108: 209-215
- Jagat Ram, Grover, R, P., Riwari, R. P., and Kumar, S, 1978. Improvement in the N. fixing effectiveness of *Rhizobium leguminosarum* by incorporating genetic resistance to azide. *Indian J. Exp. Biol* 16: 1321-1322.
- Levin, R. A. and Montgomery, M. A. 1974. Symbiotic effectiveness of antibiotic resistant mutants of *Rhizobiumjaponicum*. *Pl. Soil* **41**: 669-676
- Maier, R. J. and Brill, W. J, 1978. Involvement of *Rhizobium japonicium* G antigen in soybean nodulation. *J. Bacteriol.* **133**: 1294–1299.
- Vincent, J, M. 1970, A Manual for the Practical Study of Root Nodule Bacteria. I. B. P. Hand book No. 15, Black well, Oxford and Edinburg, pp 32-36.
- Vincent, J. M. 1980. Factors controlling the legume-Rhizobium symbiosis. In Nitrogen Fixation Volume II, (W. E. Newton and W, H. Orme-Johnson, eds) University Park Press, Baltimore, pp 103-129.

Zelazna-Kowalska, I. 1971. Correlation between streptomycin resistance and infectiveness of *Rhizobium trifoli.Pl. Soil* sp. vol. 67-71.

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