# PHOSPHORUS AND BORON INTERACTIONS IN BLACK COTTON SOILS OF KERALA WITH RESPECT TO GROUNDNUT (Arachis hypogaea L.)

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#### **THESIS**

Submitted in partial fulfilment of the requirement for the degree of

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SOIL SCIENCE AND AGRICULTURAL CHEMISTRY

COLLEGE OF HORTICULTURE

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2018

#### **DECLARATION**

I, hereby declare that the thesis entitled "Phosphorus and boron interactions in black cotton soils of Kerala with respect to groundnut (Arachis hypogaea L.)" is a bonafide record of research work done by me during the course of research and the thesis has not previously formed the basis for the award to me of any degree, diploma, associateship, fellowship or other similar title, of any other university or society.

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#### **ABBREVIATIONS**

B Boron

Ca Calcium

CD Critical difference

CEC Cation exchange capacity

cm Centimeter

Cu Copper

DAP Days after planting

dS Deci seimens

EC Electrical conductivity

Fe Iron

FYM Farm yard manure

Ha Hectare

K Potassium

Kg Kilogram

LAI Leaf area index

m Meter

Mg Magnesium

mg Milligram

Mn Manganese

MOP Muriate of potash

N Nitrogen

OC Organic carbon

P Phosphorus

S Sulphur

SSP Single super phosphate

Zn Zinc

# Introduction

#### 1. INTRODUCTION

Balanced supply of nutrients is one of the most important factors determining crop yield. Sometimes the applied nutrients may not be available for plant use, as their availability depend on interaction between nutrients. When the supply of one of the nutrients affects absorption and utilization of other nutrient element, the elements are said to be in interaction. Interaction between nutrients in soil affects the overall performance of crop. Nutrient interaction may be negative or positive. When nutrients are in combination and results in a greater response than individual response, the interaction is positive (synergism). When combination results in a lesser response, then interaction is negative (antagonism).

In Kerala, black cotton soils are seen in Chittur taluk of Palakkad district occupying an area of 2000 ha (Padmaja *et al.*, 1994). These soils are dark, low in organic matter, calcareous, neutral to alkaline (pH 7.0 to 8.5), high in clay content and CEC. The texture of soil ranges from clay loam to clay. The level of total nitrogen in the soil is satisfactory but, only very small fraction of phosphorus is in available form (less than 1 per cent) due to the process of fixation under high pH and clay content.

Even though these soils are fertile, the nutrient imbalances and poor physical conditions may adversely affect the yield of the crop (Krishnakumar, 1978; Padmaja *et al.*, 1994). Except for available P and B, all other soil nutrients are present either in medium level or adequate level. The availability of P to plants for uptake and utilization is impaired in alkaline soils due to the formation of poorly soluble calcium phosphate.

The groundnut (*Arachis hypogaea* L.) belonging to the family *leguminosae*, originated in South America (southern Bolivia/north west Argentina region) and cultivation started as early as 1000 B.C. Groundnut is an important oilseed crop, about two thirds of world production is used for oil production. The nut (kernel) is a rich source of edible oil, containing 36 to 54% oil and 25 to 32% protein. Even though groundnut can grow in soils of marginal fertility, proper fertilizer application will help to achieve full yield potential of crop. Groundnut has the capacity to utilize soil nutrients that are relatively unavailable to other crops, and therefore can make good use of residual fertility (Cox *et al.*, 1982).

Phosphorus is the second limiting nutrient for crop production (Mallikarjuna et al., 2003). In majority of Indian soils available P ranges from low to medium. It stimulates setting of pods, decreases the number of unfilled pods (pops) and hastens the maturity of the crop. P enhances nitrogen use efficiency by plants. It is essential for energy storage and transfer and hence called "energy currency" of the living system.

Boron is the most limiting micronutrient for groundnut production, as it has a role in kernel quality and flavor. Boron deficiency results in "hollow-heart" in groundnut kernels. B is involved in sugar transport, cell wall synthesis, and IAA synthesis. Boron deficiency causes low pod filling, shriveled seeds and hollow darkening or off-color in the center of the seed. Plants respond to B deficiency by slowing down growth. Boron deficiency inhibits root elongation by limiting cell enlargement and cell division. Severe B deficiency, leads to the death of root tips. Low B inhibits leaf expansion and thereby reduces photosynthetic capacity of plants. The early inhibition of root growth, compared to shoot growth, increases the shoot: root ratio, which may enhance the susceptibility of plants to environmental stresses such as deficient supplies of other nutrients and water deficit in soil.

In this context present study entitled "Phosphorus and boron interactions in black cotton soils of Kerala with respect to groundnut (*Arachis hypogaea* L.)" was carried out with following objectives:

- 1. To find out the interactions of boron and phosphorus in black cotton soils of Palakkad.
- 2. To assess the treatment level of boron and phosphorus for maximizing yield.

# **Review of Literature**

#### 2. REVIEW OF LITERATURE

Literature on nutritional requirement of groundnut, nutrient uptake and interactions between nutrients are reviewed in this chapter

#### 2.1 NUTRITIONAL REQUIREMENTS OF GROUNDNUT

#### 2.1.1 Nitrogen

#### 2.1.1.1 Influence of nitrogen on growth parameters

Groundnut is a leguminous crop, which can fix atmospheric nitrogen in its root nodule at the rate of 200-260 kg N ha<sup>-1</sup>, which reduces the demand for applied N. Even though it fixes atmospheric N, it responds well to N fertilizer application (York and Colwell, 1951 and Williams, 1979). Application of N in early stages has beneficial effect on growth characters of groundnut (Reddy and Rao 1965).

Application of N increased the number of leaves, branches and height of groundnut plant (Punnoose, 1968). Nitrogen (N) is the structural constituent of the plant cell and plays an important role in plant metabolism (Mahapatra *et al.*, 1985). Singh and Ahuja (1985) opined that application of N at 25 kg ha<sup>-1</sup> increased the growth of groundnut.

#### 2.1.1.2 Influence of nitrogen on yield parameters

Reddy and Rao (1965) observed a decrease in yield of groundnut by application of 40 kg N ha<sup>-1</sup>. Nijhawan and Maini (1966) observed increased yield of groundnut at small doses of N. Puntamkar and Bathkal (1967) viewed that application of 20 kg N resulted in significant increase in weight of pods and number of pods per plant in groundnut. Saini and Tripathi (1973) observed that application of 15 kg N ha<sup>-1</sup> resulted in highest pod yield and better oil content.

Jadhar and Narkhende (1980) observed that N has significant role on number of pods per plant and number of filled pods per plant. About 86-92 per cent of N uptake by groundnut was through biological nitrogen fixation (BNF) which is equivalent to 125-178 kg N ha<sup>-1</sup> (Dart *et al.*, 1983).

Higher doses of N produced more number of flowers and pegs (Saradhi *et al.*, 1990).N is required by plants in comparatively larger amounts than other elements. As a crop of *leguminosae* family, groundnut can fix as much as 40-80 kg N ha<sup>-1</sup> yr<sup>-1</sup> (Islam and Noor, 1992).

Application of 10-30 kg N ha<sup>-1</sup> produced higher pod yield in groundnut and after that there was a decrease in pod yield (Pant and Katiyar, 1996; Patel *et al.*, 1994) whereas, Reddy *et al.* (1984) reported that application of N at the rate of 40-60 kg ha<sup>-1</sup> increased number of pods per plant.

#### 2.1.2 Phosphorus

#### 2.1.2.1 Influence of phosphorus on growth parameters

Phosphorus (P) is the second major essential nutrient element for crop growth. The most obvious effect of P is on the plant root system. The requirement of P in nodulating legumes is higher compared to non-nodulating crops as it plays a significant role in nodule formation and fixation of atmospheric nitrogen (Brady and Weil, 2002). Application of P at 132 kg ha<sup>-1</sup> increased haulm yield in groundnut (York and Colwell, 1951). Punnoose (1968) studied effect of P on growth, yield and quality of groundnut and found that P application increased the number of leaves, branches, height of plant and weight of nodules per plant.

Shelke and Khuspe (1980) reported that dry matter production and P uptake by groundnut cv. Latur No.33 was highest with the application of 17.5 kg P<sub>2</sub>O<sub>5</sub> ha<sup>-1</sup>. Basha and Rao (1980) observed decrease in shoot length and number of leaves in

groundnut under P deficiency. Change in the rate of P application from 30 to 90 kg ha<sup>-1</sup> enhanced the growth of plant (Singh and Ahuja, 1985). Sebale and Khuspe (1986) reported that plant height, number of leaves, branches and dry weight per plant were increased by application of P at the rate of 60 kg ha<sup>-1</sup>. Juan *et al.* (1986) reported that P application increased the plant height and dry matter yield.

Phosphorus is essential for energy storage and transfer. Adenosine di and tri phosphate (ADP and ATP) are known as energy currency of plants. (Tisdale *et al.*, 1993). Higher dose of P increased the number of root nodules the number and weight of nodules, nitrogenase activity, leghaemoglobin content, leaf area and dry matter production increased by enhancing P<sub>2</sub>O<sub>5</sub> rate from 0 to 30, 60 and 90 kg ha<sup>-1</sup> (Patel *et al.*, 1994). Higher levels of P significantly increased root and shoot growth (Patel *et al.*, 1994). Plants meet their P requirement by uptake of phosphate anions from soil solution. P is important for root growth, root formation and N fixation (Lakshmamma and Raj, 1997).

Kamara et al. (2011) reported an increase in biomass of groundnut after the application of phosphorus fertilizer and attributed it to the availability of soluble phosphate that enhanced extensive root development.

### 2.1.2.2 Influence of phosphorus on yield parameters

Banerjee *et al.* (1967) observed an increase in yield of groundnut by the application of P<sub>2</sub>O<sub>5</sub> up to the level of 67.2 kg ha<sup>-1</sup>. Puri (1969) reported a significant response of groundnut to superphosphate application. Choudhary (1979) viewed that pod yield of irrigated groundnut variety, TMV-2 was higher with 60 kg P<sub>2</sub>O<sub>5</sub> ha<sup>-1</sup> than with 30 kg P<sub>2</sub>O<sub>5</sub> ha<sup>-1</sup>. Nakagawa *et al.* (1981) reported that application of 40 kg P<sub>2</sub>O<sub>5</sub> ha<sup>-1</sup> increased pod yield from 1.42 to 2.5 t ha<sup>-1</sup> and seed yield from 0.91 to 1.58 t ha<sup>-1</sup>. P application increased seed size and 100 pod weight. Application of P above 60 kg ha<sup>-1</sup> had no effect on number of pods and it depended on soil fertility status (Rao

et al., 1984). Shelling per cent was increased by application of moderate to high level of P (Chauhan et al., 1987).

Agasimani and Hosmani (1989) reported that response of P could be obtained when available soil P is less than 35 kg ha<sup>-1</sup>. Balasubramanian and Palaniappan (1991) reported that higher the concentration of phosphorus, higher will be the amount of nitrogen fixed.

Total nitrogen uptake and proportion of nitrogen present in kernel is greatly influenced by phosphorus level and interaction between phosphorus and potassium had significant influence on kernel yield (Balasubramanian and Palaniappan, 1991). Samtana *et al.* (1994) observed improvement in yield attributes by addition of P. This was due to formation and proliferation of new roots and improvement in their functional activity. Ae *et al.* (1996) reported that groundnut showed superior ability to take up P from a soil with low P fertility compared with sorghum and soybean in acid soils. Root cell walls of groundnut showed higher P-solubilizing activity than those of soybean or sorghum. Response of crop to P application depends on initial available P in the soil.

There is higher requirement for P in legumes in comparison to non-legume crops because of its role in nodule formation and fixation of atmospheric nitrogen (Brady and Weil, 2002). Bala *et al.* (2011) reported that increased pod and seed index and shelling per cent of groundnut was due to early and greater availability of nitrogen and phosphorus to plants which favorably influenced the kernel development and kernel size..

#### 2.1.3 Potassium

#### 2.1.3.1 Influence of potassium on growth parameters

Dry matter production increased with higher levels of K in groundnut cv. TMV-2 (Rao, 1979). Mathew *et al.* (1983) observed that growth characters like plant height, number of branches, leaves per plant and LAI were increased with K application.

#### 2.1.3.2 Influence of potassium on yield parameters

York and Colwell (1951) observed that groundnut grew well even in K deficient soil where other crops could not. High levels of soil K in the pod zone were undesirable as they resulted in pod rot and interfered with uptake of Ca by pegs and pods, which resulted in a higher per cent of pops and Ca deficiency in the seeds (Hallock and Garren, 1968; Csinos and Gaines, 1986).

Potassium nutrition has favorable effect on photosynthesis and translocation of food reserves from leaves to developing pods (Koch and Mengal, 1977). Number of pods per plant and seed test weight increased with higher level of K in groundnut variety, TMV-2 (Rao, 1979). There was an increase in pod yield and yield contributing characters with increase in K level (Loganathan and Krisnamoorthy, 1980). Loganathan and Krishnamoorthy (1980) reported that optimum dose of K for groundnut was 52 kg ha<sup>-1</sup> for irrigated crop and 26 kg ha<sup>-1</sup> for rainfed crop. Nair *et al.* (1981) reported that application of K fertilizer @ the rate of 80 kg ha<sup>-1</sup> increased number of pods per plant. According to Ramanathan *et al.* (1982) application K fertilizer at 50 kg ha<sup>-1</sup> yielded maximum number of pods per plant and test weight of seed. Chavan and Kalra (1983) observed that dry pod yield, shelling per cent, 1000 grain weight and oil content of groundnut cv. TG-1 were higher with 50 kg ha<sup>-1</sup> than with 25 kg ha<sup>-1</sup>.

Groundnut is a heavy feeder of K and adequate supply of this nutrient must be provided to obtain a better yield (Geetalakshmi *et al.*, 1993). Yakadri *et al.* (1992) viewed that 40-60 kg K<sub>2</sub>O ha<sup>-1</sup> was the optimum dose of K for groundnut. Hadwani and Gundalia (2005) reported that yield of pod and haulm was increased with K fertilizer application.

#### 2.1.4 Secondary nutrients

#### 2.1.4.1 Influence of Ca on growth and yield parameters

In order to avoid Ca deficiency, application of soluble source of Ca at early flowering stage is advocated. The developing pods require high Ca level in surrounding soil, because Ca absorbed by root did not translocate into pods and Ca required for pod development is absorbed directly from soil solution (Skelton and Shear, 1971). Calcium deficiency resulted in lower yield, darkened plumule in the seed, empty pods and sometimes plants remained green and continued to produce flowers and pegs without pods, which might be infertile (Sulhiram *et al.*, 1974).

Calcium (Ca) is required by groundnut plants from the time pegs began to appear for fruit formation, until the pods mature (Walker, 1975). Calcium is essential for proper development of pod and for production of high quality seed (Cox *et al.*, 1982). Ca requirement varied with seed size and smaller seeded cultivars required less Ca than larger seeded type, because of larger surface to volume ratio. Soil characters like soil water content, soluble Ca, exchangeable Ca and type of soil mineral present affected Ca uptake by groundnut (Keisling *et al.*, 1983).

For production of good quality groundnut kernel, soil Ca level must be in the range of 600-800 mg kg<sup>-1</sup> (Kvien *et al.*, 1988; Sumner *et al.*, 1986). Morphological

characters of pod like pod surface area, pod volume, number of days for maturation of pods and shell thickness influenced Ca uptake by pods (Kvien *et al.*, 1988). Ca deficiency led to high per cent of aborted seeds (empty pods), improperly filled pods and caused aborted or shrivelled fruit, including darkened plumules and production of pods without seed (Singh and Oswalt, 1995). Mandal *et al.* (2005) observed that gypsum application for summer and rainy season groundnut in sandy loam soils of West Bengal @ the rate of 400 kg ha<sup>-1</sup> recorded highest plant height (65.1cm).

#### 2.1.4.2 Influence of magnesium on growth and yield parameters

Dowood (1982) indicated that application of three levels of Mg (0,120 and 240 kg MgSO<sub>4</sub>.7H<sub>2</sub>O ha<sup>-1</sup>) caused significant increase in phosphorus uptake by plant. Al-lami (1999) showed that increase in adding of MgSO<sub>4</sub>.7H<sub>2</sub>O from 0 to 80 kg ha<sup>-1</sup> caused a significant increase in available phosphorus in the soil from 0.23 to 0.25 c mol kg<sup>-1</sup>.

#### 2.1.4.3 Influence of sulphur on growth and yield parameters

Umadevi *et al.* (1999) reported that pod yield enhanced by increasing S levels from 15 to 30 and 75 kg ha<sup>-1</sup> in red loamy sandy soils of Ananthapur, Andhra Pradesh. Highest pod yield was recorded by addition of S at 75 kg ha<sup>-1</sup>. Babu *et al.* (2007) reported that application of S through gypsum @ of 40 kg ha<sup>-1</sup> recorded highest pod and haulm yield. Singh and Chaudhari (2008) observed that in calcareous soil, plants grown by addition of S had increased plant height, number of flowers, number and weight of nodules, higher dry matter, seed, haulm (leaves and stems), and oil content compared to those without S.

Rao *et al.* (2013) found that application of S at 45 kg ha<sup>-1</sup> through gypsum recorded highest plant height (71.45 cm) in sandy loam soils of Andhra Pradesh. Application of S influenced growth, yield attributing characters, yield and oil content regardless of the sources and levels of S. Addition of S at 45 kg ha<sup>-1</sup> recorded highest plant height, number of filled pods per plant, 100 pod weight, 100 kernel weight, pod yield, haulm yield and oil content of kernels (Rao *et al.*, 2013).

The S uptake by pods increased significantly with increasing levels of S and maximum uptake (10.89 kg ha<sup>-1</sup>) was noticed with application of 60 kg S ha<sup>-1</sup>. Giri *et al.* (2014) observed that number of pods per plant was highest in sandy loam soils when S was applied at 15 kg ha<sup>-1</sup> (25.52).

#### 2.1.6 Micronutrients

# 2.1.6.1 Influence of micronutrients on growth and yield parameters

Mahajan *et al.* (1994) recorded that boron at 0.5 kg ha<sup>-1</sup> applied through boronated super phosphate or borax increased dry pod yield (3200 kg ha<sup>-1</sup>) followed by boron spray of 0.5 ppm (2 sprays, first at 30 days after germination and second at the time of flowering) gave higher yield than control. Subrahmaniyan *et al.* (2001) noticed that combined application of ZnSO<sub>4</sub> at 5 kg ha<sup>-1</sup>, borax at 5 kg ha<sup>-1</sup>, ferrous sulphate at 10 kg ha<sup>-1</sup> recorded maximum number of pods per plant. Mahamoud *et al.* (2006) observed that foliar application of B at 25-50 ppm increased plant height, leaf area, total dry matter, number of pods and seed yield. Availability of micronutrients in soils is governed by soil pH, cation and anion exchange capacity and nutrient interactions. The application of B also promoted absorption of N by groundnut and increased plant height, plant dry weight and the total number of pods.

Singh *et al.* (2009) reported that soil application of boron at 1 kg ha<sup>-1</sup> as agricol, solubor and borosol increased pod yield by 23, 18 and 12 per cent, respectively, compared to the spray of 9 per cent as borax and 5 per cent as boric acid.

Barthi *et al.* (2010) reported that application of micronutrients (Fe, Zn, B) along with recommended fertilizers improved growth parameters and chlorophyll content of groundnut. They observed highest pod yield and dry matter content on application of 20 kg ha<sup>-1</sup> ZnSO<sub>4</sub> and 5 kg borax. Reddy *et al.* (2011) reported that soil application of micronutrients viz., ZnSO<sub>4</sub> at 10 kg ha<sup>-1</sup>, borax at 5 kg ha<sup>-1</sup> and copper sulphate at 5 kg ha<sup>-1</sup> enhanced number of pods per plant. Kamalakannan and Ravichandran (2013) observed that application of 100 per cent NPK, ZnSO<sub>4</sub> at 25 kg ha<sup>-1</sup>, B at 10 kg ha<sup>-1</sup> and FYM at 12.5 t ha<sup>-1</sup> recorded highest plant height at all critical stages of groundnut

Ravichandra *et al.* (2015) viewed that application of boron as foliar spray along with rhizobium at flowering and pod formation stage had positive effect on growth and yield of groundnut with increased plant height, number of branches, plant dry weight, number of pods per plant, 100 pod weight, seed index and pod yield. Excess spray of boron as foliar nutrition led to decrease in the above mentioned parameters.

# 2.1.7 Response of groundnut to combined application of N, P and K

Das (1982) reported that growth components were increased by applying NPK. Venkateswaralu and Nath (1989) reported the importance of balanced fertilizer schedule and its influence on groundnut. Combined application of NPK at the rate of 20: 40: 40 kg ha<sup>-1</sup> produced highest yield (Pradhan and Das, 1989). Yadav (1990) found that the application of NPK at 20: 60: 40 kg ha<sup>-1</sup> resulted in highest yield.

Whereas Balasubramaniam and Palaniappan (1991) reported that the application rate of 150 kg N and 50 kg K<sub>2</sub>O ha<sup>-1</sup> produced higher yield.

Patel *et al.* (1994) reported that application of 25 kg N + 50 kg P<sub>2</sub>O<sub>5</sub> ha<sup>-1</sup> increased the pod and haulm yield of groundnut *cv.* GAUG-1. Kachot *et al.* (1984) observed higher number of pegs per plant, number of pods per plant, pod weight per plant and test weight when 12.5 kg N ha<sup>-1</sup> and 50 kg P<sub>2</sub>O<sub>5</sub> ha<sup>-1</sup> was applied. Rana *et al.* (1984) reported that higher pod yield of 23.19 q ha<sup>-1</sup> was obtained at 20 kg N ha<sup>-1</sup>, 60 kg P<sub>2</sub>O<sub>5</sub> ha<sup>-1</sup> and 40 kg K<sub>2</sub>O ha<sup>-1</sup>. Application of NPK at the rate of 50: 100: 50 kg NPK ha<sup>-1</sup> the increased the number of branches per plant significantly (Dholaria *et al.*, 1972).

Ghadekar *et al.* (1993) reported that pod yield was higher at the fertilizer rate of 40 kg N, 80 kg  $P_2O_5$  and 30 kg  $K_2O$  ha<sup>-1</sup>. NPK application @ the rate of 25 kg N + 75 kg  $P_2O_5$  + 37.5 kg  $K_2O$  ha<sup>-1</sup> gave mean pod yield of 3.55, 4.10 and 4.99 t ha<sup>-1</sup> respectively (Thimmegowda, 1993).

Dahatonde (1982) revealed that combined application of inorganic fertilizer and organic manure recorded favorable effects on various growth characters and yield attributes of groundnut. They reported that application of 25:50 kg N and P<sub>2</sub>O<sub>5</sub> ha<sup>-1</sup> + 5 t FYM ha<sup>-1</sup> recorded highest plant height, plant spread, no.of branches per plant and total dry matter per plant at harvest and yield attributes *viz.*, filled pod per plant, dry pod per plant and pod and haulm yield of summer groundnut.

Kuchanwar *et al.* (1997) reported that nitrogen and phosphorus uptake was highest with combination of 25:50 kg N and P<sub>2</sub>O<sub>5</sub> ha<sup>-1</sup> respectively. Dubey (1997) reported that, application of SSP at 60 kg P<sub>2</sub>O<sub>5</sub> ha<sup>-1</sup> was highest, but was on par with 30 kg P<sub>2</sub>O<sub>5</sub> ha<sup>-1</sup> as SSP along with *Pseudomonas striata* with regard to N, P and K

uptake (straw + grain) in black or medium clay soil (Vertisol). Shipkule *et al.* (2008) reported that application of 80:60:20 kg ha<sup>-1</sup> N, P<sub>2</sub>O<sub>5</sub> and K<sub>2</sub>O respectively showed maximum nutrient content as well as highest nitrogen, phosphorus and potassium uptake by kernel and haulm of groundnut. Sanchez and Owen (1978) reported that application 150 kg P<sub>2</sub>O<sub>5</sub> ha<sup>-1</sup> along with N and K increased pod yield from 0.75 to 2.07 t ha<sup>-1</sup>.

Babu *et al.* (2007) reported that uptake of NPK by groundnut was higher in highly fertilized plots. But this higher uptake did not contribute to higher yield but enhanced vegetative growth. Dholaria *et al.* (1972) reported that pod weight, number of pods and number of branches per plant increased under high fertility condition (50: 100: 50 kg NPK ha<sup>-1</sup>).

# 2.2 NUTRIENT CONTENT AND UPTAKE

# 2.2.1 Influence of nitrogen on nutrient content and uptake

Uptake of nitrogen is more intensive during flowering and pod formation stages. During reproductive stages, there is continuous mobilization of N from leaves to the developing fruit, and this sometimes resulted in appearance of N deficiency symptoms (Kvien *et al.*, 1986). Chahal *et al.* (1983) observed that shoot N content was high at early and mid-flowering stages. Reddy *et al.* (1984) studied N uptake of groundnut and it was maximum by the application of 10 kg N as basal and 20 kg N at 30 DAP. Boote *et al.* (1985) viewed that during seed filling stage leaf N declined from 4.01 to 2.85 per cent and stem N from 1.65 to 1.13 per cent and root N from 2.19 to 1.50 per cent. Reddy and Murthy (1985) observed that N content was highest in kernel and lowest in the shell. N content decreased as crop grew older.



# 2.2.2 Influence of phosphorus on nutrient content and uptake

Basha and Rao (1980) reported that P deficiency decreased N,P,K and Ca contents in 30 day old groundnut plants. Shelke and Khuspe (1980) reported that dry matter production and P uptake by groundnut cv. Latur No. 33 was highest with 17.5 kg ha<sup>-1</sup> than with 0 or 35 kg ha<sup>-1</sup>. Nakagawa *et al.* (1981) reported that highest rate of P application significantly increased P content in seeds. Patel *et al.* (1994) reported the effect of application of 100ppm P<sub>2</sub>O<sub>5</sub> on growth and nutrient uptake by groundnut in calcareous soil. High level of P increased shoot and root growth and uptake of P by root and shoot. Higher level of P was also effective in increasing uptake of N. Chahal *et al.* (1983) observed that application of P increased N and P uptake and dry matter yield.

Application of 60 kg P<sub>2</sub>O<sub>5</sub> ha<sup>-1</sup> increased N and P uptake and content in seed (Islam and Noor, 1982). Chavan and Kalra (1983) reported that P increased the N content and NPK uptake in plants. Bell (1985) reported that the tissue P content during vegetative growth was 0.3 per cent of dry matter and declined during reproductive stage as 0.27 per cent at 60 Days After Emergence to 0.12 per cent at 100 days.

# 2.2.3 Influence of potassium on nutrient content and uptake

Rao (1979) reported that uptake of N and P increased with application of K at rate of 0, 40, 80 kg ha<sup>-1</sup> in groundnut variety, TMV-2 under irrigated condition. Reddy *et al.* (1983) reported that uptake of K in groundnut was maximum when it was applied as basal @ of 40 kg ha<sup>-1</sup>. Survase *et al.* (1986) reported that average plant N, P, K contents at flowering stage were 2.7, 0.21, and 2.28 per cent respectively.

# 2.2.4 Influence of secondary nutrients on nutrient content and uptake

Giri *et al.* (2014) reported that uptake of nutrients (N, P, K and S) by kernel, shell and haulm of groundnut and also total uptake of nutrients by groundnut were significantly influenced by levels of sulphur. Umadevi *et al.* (1999) reported that application of S at 75 kg ha<sup>-1</sup> recorded highest uptake of nitrogen (100.7 kg ha<sup>-1</sup>), phosphorus (10.40 kg ha<sup>-1</sup>), potassium (40.4 kg ha<sup>-1</sup>), sulphur (12.21 kg ha<sup>-1</sup>), calcium (34.6 kg ha<sup>-1</sup>) and magnesium (15.59 kg ha<sup>-1</sup>) by groundnut. Singh *et al.* (2005) observed that S uptake by pods of groundnut increased significantly with increased levels of S up to 60 kg ha<sup>-1</sup> (10.89 kg ha<sup>-1</sup>). Patel *et al.* (2009) viewed that successive increase in S application up to 40 kg ha<sup>-1</sup> improved NPS uptake by groundnut. The maximum uptake of nutrients was recorded at S application at 15 kg ha<sup>-1</sup>. The minimum uptake of nutrients was recorded in no S treatment.

Rao and Shaktawat (2002) reported that application of gypsum at 250 kg ha<sup>-1</sup> (half at sowing + half at 35 DAS) for groundnut crop significantly increased nitrogen, phosphorus, potassium, sulphur, calcium and magnesium uptake by 13.2, 11.0, 10.6, 10.4, 11.1 and 8.9 per cent respectively over control.

Veerabhadrappa and Yeledhalli (2005) revealed that foliar spray (N, P, K, Ca and S - commercial formulation of urea, SSP and MOP at 1 per cent level each) at 60 DAS along with application of 100 per cent RDF recorded higher levels of nitrogen (252 kg ha<sup>-1</sup>), phosphorus (28.9 kg ha<sup>-1</sup>), potassium (204 kg ha<sup>-1</sup>), calcium (74.8 kg ha<sup>-1</sup>) and S (31.4kg ha<sup>-1</sup>) uptake by groundnut.



# 2.2.5 Influence of micronutrients on nutrient content and uptake

Mahajan *et al.* (1994) reported that soil application of boron at 0.5 kg ha<sup>-1</sup> through boronated superphosphate recorded higher uptake of nitrogen (127.4 kg ha<sup>-1</sup>) and phosphorus (11.7 kg ha<sup>-1</sup>) in clayey soils. Kamalakannan and Ravichandran (2013) reported that application of 100 per cent NPK, zinc sulphate at 25 kgha<sup>-1</sup>, borox at 10 kg ha<sup>-1</sup> and FYM at 12.5 t ha<sup>-1</sup> showed highest NPK uptake at all the growth stages of crop. Nadaf and Chidanandappa (2015) observed that borax at 5 kg ha<sup>-1</sup> and zinc sulphate at 20 kg ha<sup>-1</sup> recorded highest uptake of nitrogen (95.72 kg ha<sup>-1</sup>), phosphorus (23.50 kg ha<sup>-1</sup>), potassium (92.68 kg ha<sup>-1</sup>), calcium (38.34 kg ha<sup>-1</sup>), magnesium (20.87 kg ha<sup>-1</sup>) and sulphur (28.16 kg ha<sup>-1</sup>).

#### 2.3 PROTEIN CONTENT

Nijhawan (1962) reported beneficial effect of application of nitrogen in increasing the protein content. Punnoose (1968) found that graded doses of P enhanced protein content in kernels. Basha and Rao (1980) observed that P deficiency decreased protein content in groundnut. Nair and Sadanandan (1981) found that protein content was increased with increased P application at 50-100 kg ha<sup>-1</sup>. Nair and Sadanandan (1981) found that the protein content was decreased with increased dose of K from 25 to 75 kg ha<sup>-1</sup>.

#### 2.4 INTERACTIONS BETWEEN NUTRIENTS

#### 2.4.1 P and B interaction

The effects of phosphorus deficiency like reduction in dry matter, soluble protein, DNA, activity of ribonuclease and increase in the activities of peroxidase, acid phosphatase and polyphenol oxidase were intensified by combined deficiency of boron and phosphorus in maize (Chatterjee *et al.*, 1990). The excessive application

of P reduced uptake of B in citrus (Bingham et al., 1958) and in strawberry (May and Pritts, 1993).

Gunes and Alpaslan (2000) reported that application of B resulted in increased B uptake and a decreased P uptake in all genotypes of maize. Similar way, application of P resulted in decreased B uptake and increased P uptake of the genotypes. B application caused decrease in dry weight of all genotypes where as dry weight of all genotypes was increased by application of P.

B is more toxic in the absence of P rather than the presence of P, and this toxicity could be alleviated by applications of P in the calcareous soils of semiarid areas. Pollard *et al.* (1976) reported that capacity for absorption of phosphate was shown to be reduced in *Zea mays* and *Vicia faba* suffering from boron deficiency. Tavajjoh *et al.* (2016) found that application of P and B fertilizers increased the concentration and total uptake of P and B in the seeds of rapeseed cultivars and there is a synergistic interaction between P and B. Kabir *et al.* (2013) reported that fertilizer level for P, Ca and B should be 50 kg ha<sup>-1</sup>, 110 kg ha<sup>-1</sup> and 2.5 kg ha<sup>-1</sup> to attain maximum yield.

Adams and Winsor (1974) observed that boron content of the foliage was reduced by lime addition and high phosphate content in tomato. Kaya *et al.* (2009) showed that interaction between P and B had an effect on biomass of tomato. Fruit yield, number of fruits and average fruit weight decreased in the plants grown under high boron conditions.

#### 2.4.2 Interaction of P with other nutrients

Phosphorus deficiency is a main yield limiting variable for crop production in acid and alkaline soils (Fageria *et al.*, 1983). Assessing the interaction of phosphorus with other nutrient is critical to keep up a balanced nutrient supply for enhancing crop

yield. Phosphorus has a positive interaction with nitrogen and plant development (Sumner and Farina, 1986). Sumner and Farina (1986) viewed that increased growth required more of both N and P, the mutually synergistic effects resulted in growth stimulation and enhanced uptake of N and P.

If large amount of P is supplied, P: Fe and P: Zn ratio in plant tissue is raised and led to deficiency of these nutrients (Loneragan *et al.*, 1979; Loneragan *et al.*, 1982).

High available P induced the deficiencies of Zn and Mn in maize (Adriano and Murphy, 1970) and potato (Adriano et al., 1971). P formed chemical bond with Zn, at high levels of P and P bounds large quantity of Zn resulting in P induced Zn deficiency, which led to reduced shoot growth. Friesen et al. (1980) reported increased total Zn uptake with P addition which led to increased root growth, however extreme level of P caused Zn deficiency. Saeed and Fox (1979) reported an increase in Zn sorption in Hawaiian soil due to P application. Sorption of P on the surface of Fe and Al oxides increased negative charges on them resulting in increased sorption of Zn. Gupta and Raj (1983) observed positive interaction between K and Zn on yield of wheat.

Heavy application of P resulted in Fe deficiency. P interactions with Fe form Fe- phosphates which led to Fe chlorosis in plants (Ayed, 1970). P inhibits Fe absorption by roots and Fe transport from roots to shoots and inactivates plant Fe (Elliott and Lauchli, 1985; Moraghan and Mascagni, 1991).

Interaction between Ca and P was complex; it had both synergistic and antagonistic effect. The synergistic effect was due to simultaneous uptake of Ca and P. Antagonistic effect was due to precipitation of P into less soluble calcium phosphate. Fageria (1983) reported decreased uptake of P and Ca with increased concentration of K in rice. Lundergardh (1934) reported a higher absorption of P and

Ca at lower concentration of K. Acidifying effect of S application was important in mobilization of Fe, Mn, Zn and P in calcareous soil (Soliman *et al.*, 1992).

# 2.4.3 Interaction of B with nutrients

Hosseini *et al.* (2007) showed that Zn application increased plant height and dry matter yield where as high levels of B decreased plant height and dry matter yield. In grape, increased Zn concentration decreased B toxicity Similarly, B toxicity symptoms were severe in Zn deficient orange (Swietlik, 1995).

Mahmoud *et al.* (2006) reported synergistic interaction between B and N. The foliar application of B (25-50ppm) in combination with 40 kg N ha<sup>-1</sup> has significantly increased plant height, leaf area, total dry weight, number of pods and seed yield. Daliparthy *et al.* (2008) reported that application of K led to decrease in B level and caused B deficiency.

# **Materials and Methods**

#### 3. MATERIALS AND METHODS

The present study entitled "Phosphorus and boron interactions in black cotton soils of Kerala with respect to groundnut (*Arachis hypogaea* L.)" was carried out in College of Horticulture, Vellanikkara during 2016-18. Study consists of a field experiment with groundnut in black cotton soils of Chittur, Palakkad followed by laboratory analysis of soil, plant and pod samples taken from experimental field. The materials used and methods followed in the study are described in this chapter.

# 3.1 COLLECTION AND ANALYSIS SOIL SAMPLE

Soil samples were collected from different locations of Chittur and analyzed for available P and B. Three to four samples were collected from each location from a depth of 0-15 cm. Collected samples were air dried, processed and sieved through 2mm sieve. These samples were analyzed for available P and B. Available P and B status of samples collected from different locations are given below.

Table 1: Available P and B status of soil samples collected from different locations of Chittur

Sl. no.	Place	Available P	Water soluble B
		(kg ha <sup>-1</sup> )	(mg kg <sup>-1</sup> )
1	Erimedu	32.00	0.30
2	Kochikkadu	22.67	0.48
3	Kammalathara	15.74	0.50
4	Malayampallam	25.65	0.37
5	Nellimedu	8.70	0.30



Plate 1: Initial soil sample collection from experimental field



Plate 2: View of field before experiment

The experiment was carried out where deficiencies of both P and B were noticed. The techniques followed for estimation of physico – chemical properties of soil are given below.

Table 2: Methods followed in soil analysis

Parameter	Method	Reference
Bulk density	Keen – Raczkowski cup	Piper ( 1966)
Texture	International pipette method	
pH and electrical	1:2.5 soil water suspension- pH	Jackson (1973)
conductivity	meter and conductivity meter.	
Organic carbon	Walkley and Black method	Walkley and Black
		(1934)
Available nitrogen	Alkaline permanganate mehod	Subbiah and Asija
		( 1956)
Available phosphorus	Olsen extraction (0.5 M NaHCO <sub>3</sub> at	Watanabe and Olsen
	pH 8.5) and estimation by	(1965)
	spectrophotometer	
Available potassium	Neutral normal ammonium acetate	
	extraction and estimation by flame	
	photometer	
Available calcium and	Neutral normal ammonium acetate	
magnesium	extraction and estimation by AAS	Jackson ( 1973)
Available sulphur	CaCl <sub>2</sub> extraction and estimation by	
	spectrophotometer	
Available boron	Hot water extraction and estimation	
	by spectrophotometer	
Available micronutrients	DTPA extraction and estimation by	Lindsay and Norvell
(Fe, Mn, Zn, Cu)	ICP-OES	(1978)

Table 3: Initial physico - chemical properties of soil in the experimental site

Parameter	Value
Bulk density (Mg m <sup>-3</sup> )	1.35
Texture	Sandy clay loam
Coarse sand (%)	31.85
Fine sand (%)	27.30
Silt (%)	18.60
Clay (%)	22.25
pH	7.83
Electrical conductivity (dS m <sup>-1</sup> )	0.242
Organic carbon (%)	1.40
Available nitrogen (kg ha <sup>-1</sup> )	288.50
Available phosphorus (kg ha <sup>-1</sup> )	8.70
Available potassium (kg ha <sup>-1</sup> )	384.48
Available calcium (mg kg <sup>-1</sup> )	4454.50
Available magnesium (mg kg <sup>-1</sup> )	812.54
Available sulphur (mg kg <sup>-1</sup> )	6.05
Available B (mg kg <sup>-1</sup> )	0.31
Available Fe (mg kg <sup>-1</sup> )	8.39
Available Mn (mg kg <sup>-1</sup> )	7.18
Available Zn (mg kg <sup>-1</sup> )	1.03
Available Cu (mg kg <sup>-1</sup> )	1.24

# 3.2 GENERAL DETAILS OF FIELD EXPERIMENT

# 3.2.1 Experimental site

The field experiment was conducted in black soils of Chittur, Palakkad. Geographically the field is situated at eastern side of Palakkad at  $10^0$  38' 3.88" N latitude and  $76^0$  44' 53.90" E longitude at an elevation of 129 m from mean sea level.

3.2.2 Climate and weather

The experimental site has humid tropical climate.

3.2.3 Cropping season

Experiment was conducted during May to August, 2017.

3.2.4 Cropping history of field

Maize was cultivated in 2015 and then field was left fallow for one year.

3.2.5 Crop variety

Groundnut variety, K-6 (Kadiri-6) was used for experiment. This variety was released from Agricultural Research station, Kadiri, Andhra Pradesh, which is suitable for both kharif and rabi season. It is a semi spreading, short duration variety resistant to leaf spot.

3.3 EXPERIMENTAL DETAILS

Soil samples were collected from different locations of Chittur and experiment was conducted where deficiencies of both P and B are noticed. The experimental details are given below.

Crop

: Groundnut

Variety

: K-6

Season

: May - August

Design

: RBD (factorial)

Treatments

 $: 4^2 + 1$ 

Replications: 4

Spacing

: 15cm x 15cm.

Plot size

 $: 4 \times 2 \text{ m}^2$ 

POP recommendations of groundnut- 10:75:75 N:P:K kg ha<sup>-1</sup>

Seeds were collected from National Seed Cooperation (NSC), Palakkad.

#### 3.3.1 Treatment details

Different combinations of P and B at different doses were the treatments. Treatment combinations are made with four levels of P and four levels of B and soil test based recommendation is taken as control.

### Factor A

# Levels of phosphorus (P) - 4

Po -Control

P<sub>1</sub> -60 kg ha<sup>-1</sup>

 $P_2 - 75 \text{ kg ha}^{-1}$ 

 $P_3$  -90 kg ha<sup>-1</sup>

#### Factor B

# Levels of boron (B) – 4

B<sub>0</sub> - Control

 $B_1 - 5 \text{ kg ha}^{-1}$ 

 $B_2 - 10 \text{ kg ha}^{-1}$ 

 $B_3 - 15 \text{ kg ha}^{-1}$ 





Plate 3: View of field at flowering stage of groundnut crop

### **Treatment combinations**

T<sub>1</sub>: Soil test based recommendations

Treatment	Notation	Treatment	Notation
T <sub>2</sub>	$P_0 B_0$	T <sub>10</sub>	$P_2 B_0$
T <sub>3</sub>	$P_0 B_1$	T <sub>11</sub>	$P_2 B_1$
T <sub>4</sub>	$P_0 B_2$	T <sub>12</sub>	$P_2 B_2$
T <sub>5</sub>	$P_0 B_3$	T <sub>13</sub>	$P_2 B_3$
T <sub>6</sub>	$P_1 B_0$	T <sub>14</sub>	$P_3 B_0$
T <sub>7</sub>	$P_1B_1$	T <sub>15</sub>	$P_3 B_1$
T <sub>8</sub>	$P_1 B_2$	T <sub>16</sub>	$P_3 B_2$
T <sub>9</sub>	$P_1 B_3$	T <sub>17</sub>	$P_3 B_3$

Nitrogen and potassium levels are kept same (based on POP recommendations of KAU) for all treatments except for the first treatment where soil test based recommendations were given.

# 3.4.1 Land preparation

Land was ploughed thoroughly using tractor and made into fine tilth. Gross area of field was 17 cents and net area was 13.6 cents. The selected field was divided into four blocks and each block was further divided into 17 treatment plots. Furrow of 1m width was taken in between blocks for drainage.

T3	419	T15	T4	T16	T10	T17	T14	T11	T7	T6	T13	T1	T8	T12	T5	T2
T4	T12	T8	T10	T6	T17	T1	T13	T9	T3	T14	T5	T16	T2	T15	T11	T7
	14	T15	13	T11	T10	1.0 J.	.16	114	T2	T3	F1	T5	L9	T17	T8	T12
T7									L	I	L	T	L			
T15	T8	T11	T17	T1	T12	T14	T2	41 L	TS	T13	T16	T7	T6	T4	T3	T10

4 m

4 m

4 m

4 m



Plate 4: View of field at pegging stage of groundnut crop



Plate 5: View of field at pod setting stage of groundnut crop

# 3.4.2 Application of manures and fertilizers

Fertilizers were applied as basal dose at the time of sowing. Farm yard manure was applied in all the plots equally. Urea and MOP were also applied in equal quantity based on POP except in  $T_1$  where soil test based recommendations were given. Different levels of SSP and borax were given based on treatment requirements.

Table 3: Rate of application of fertilizers in the experimental field

Treatments	Urea	SSP	МОР	Borax
	(kg ha <sup>-1</sup> )			
$T_{I}$	21.74	416.67	150	10
$T_2$	`21.74	0	150	0
$T_3$	21.74	0	150	5
$T_4$	21.74	0	150	10
$T_5$	21.74	0	150	15
$T_6$	21.74	333.33	150	0
T <sub>7</sub>	21.74	333.33	150	5
$T_8$	21.74	333.33	150	10
T <sub>9</sub>	21.74	333.33	150	15
T <sub>10</sub>	21.74	416.67	150	0
$T_{11}$	21.74	416.67	150	5
T <sub>12</sub>	21.74	416.67	150	10
T <sub>13</sub>	21.74	416.67	150	15
T <sub>14</sub>	21.74	500	150	0
T <sub>15</sub>	21.74	500	150	- 5
T <sub>16</sub>	21.74	500	150	10
T <sub>17</sub>	21.74	500	150	15

# 3.4.3 Irrigation

Furrow irrigation was done once in 7 days.

# 3.4.4 Weed management

Hand weeding was done at 15 days interval. At the time of flowering, earthing up was done along with weeding. Field was kept without any disturbance after 45 days of sowing.

# 3.4.5 Plant protection

Pest and disease incidence was very meager in the field. Peacock menace was controlled by using colored reflecting papers across the field.

# 3.4.6 Harvesting

Harvesting was done when leaves started yellowing and began to dry up. Plants were ready for harvest at 80 days after sowing. The plants were uprooted and pods were separated manually.

### 3.5 OBSERVATIONS

# 3.5.1 Biometric observations

Five plants were selected randomly and tagged for taking biometric observations. The following biometric observations were recorded from these plants at flowering, peg development, pod setting and harvesting stages and mean values were computed.

# Plant height

Plant height was recorded at flowering, peg development, pod setting and harvesting stages and mean values were recorded.

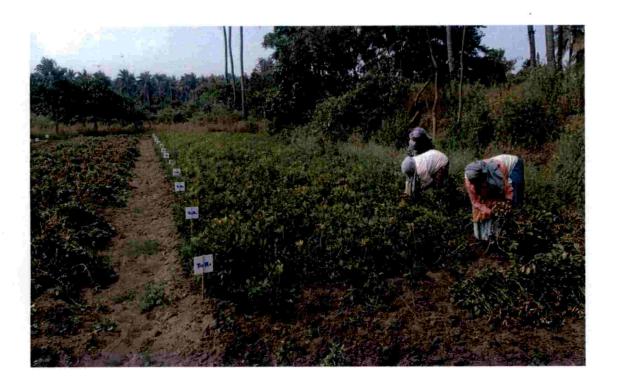




Plate 6: Harvesting of groundnut crop

### Number of leaves

Number of leaves was taken at flowering, peg development, pod setting and harvesting stages and mean values were recorded.

# No.of pods/plant

No. of pods/plant were recorded at harvesting stage and mean values were recorded.

#### Yield

Fresh weight of pods and haulm were taken at harvesting stage and mean values were noteded.

# 3.5.2 Soil analysis

Soil samples were collected from a depth of 0-15 cm and analyzed for pH, EC, OC, major nutrients (N, P, K), secondary nutrients (Ca, Mg, S) and micronutrients (Fe, Mn, Zn, Cu, B) both before and after the crop. Physical properties of soil *viz*. soil texture, bulk density and soil moisture were also analyzed. The methods employed for analysis are given in table no. 2.

#### 3.5.3 Plant analysis

# Collection of samples

Plant samples were collected at flowering, peg development, pod setting and harvesting stages. Plant samples were first washed with tap water to remove dirt and soil. These were then washed with single and double distilled water, and shade dried for a week. The shade dried samples were kept in an oven @ 60 °C and dry weight was recorded. The samples were powdered and stored in polythene bags. The content of major nutrients (N, P, and K), secondary nutrients (Ca, Mg, and S) and

micronutrients (Fe, Mn, Zn, Cu, and B) of these samples were analyzed. The methods followed to determine the nutrients are given in the table below.

Table 4: Method of plant analysis

Parameter	Method	Reference
Nitrogen	Micro kjeldahl distillation	Jackson, 1973
Phosphorus	Vanado – molybdo – phosphoric	Jackson, 1973
	(Bartons reagent) yellow color	
	method	
Potassium	Nitric acid digestion and	
	estimation by flame photometer	
Calcium and magnesium	Nitric acid digestion and	Piper,1966
	estimation by ICP- OES	
Sulphur	Nitric acid digestion and	Black, 1965
	estimation by turbidimetry	
Boron	Nitric acid digestion and	Page et al., 1982
	estimation by ICP- OES	
Micro nutrients (Fe, Mn,	Nitric acid digestion and	Piper,1966
Zn, Cu)	estimation by ICP- OES	

# 3.5.4 Nutrient uptake

Uptake of major, secondary, and micro nutrients were calculated by using the formula,

Uptake of nutrient (kg ha<sup>-1</sup>) = Nutrient concentration (%) x biomass (kg ha<sup>-1</sup>)

100



Plate 7: Harvested groundnut pods

# 3.6 Statistical analysis

Statistical analysis of experimental data was done by Fisher's method of analysis of variance as outlined by Gomez and Gomez (1984).

# Results

#### 4. RESULTS

The results of the experiment entitled "Phosphorus and boron interactions in black cotton soils of Kerala with respect to groundnut (*Arachis hypogaea* L.)" are presented in this chapter.

#### 4.1 GROWTH AND YIELD PARAMETERS OF GROUNDNUT

The effect of P and B application on growth parameters of groundnut like plant height and number of leaves, yield parameters like number of pods and yield at flowering, pegging, pod development and harvest stages under different doses of phosphorus and boron are given.

## 4.1.1 Growth parameters

# 4.1.1.1 Plant height

Plant heights at flowering, pegging, pod development and harvest stages are shown in table 6, 7, 8, 9 respectively. The treatment,  $P_3B_0$  (P, 90kg ha<sup>-1</sup> and B,0 kg ha<sup>-1</sup>) showed highest plant height at flowering (15.42 cm), pegging (24.21 cm) and pod development (32.99 cm)stage. At harvest stage, the treatment  $P_1B_1$  (P, 60 kg ha<sup>-1</sup> and B, 10 kg ha<sup>-1</sup>) showed highest plant height (37.15 cm). Plant heights at different stages were found to be significantly influenced due to main effect and interaction effect. Plant height was enhanced by increased dose of P but decreased with increased dose of B.

Table 6: Effect of P and B application on plant height at flowering stage (cm)

T<sub>1</sub>: 11.07

	$\mathbf{B}_0$	<b>B</b> <sub>1</sub>	$\mathbf{B}_2$	$\mathbf{B}_3$	Mean
$P_0$	12.44	10.65	8.88	8.55	10.13
P <sub>1</sub>	12.66	12.06	11.59	12.31	12.15
P <sub>2</sub>	14.72	13.67	11.03	9.38	12.2
P <sub>3</sub>	15.42	13.87	13.01	12.9	13.8
Mean	13.81	12.56	11.13	10.78	
CD (0.05) P; 0.104		CD (0.05) B; 0.104		CD (0.05) PxB; 0.209	
SE (m) P; 0.037		SE (m)	SE (m) B; 0.037		xB; 0.073

Table 7: Effect of P and B application on plant height at pegging stage (cm)

T<sub>1</sub>: 20.10

	$\mathbf{B}_0$	<b>B</b> <sub>1</sub>	$\mathbf{B}_2$	B <sub>3</sub>	Mean
$P_0$	21.43	19.59	17.34	16.98	18.83
P <sub>1</sub>	21.96	21.12	20.72	21.37	21.29
P <sub>2</sub>	23.11	22.57	19.93	17.72	20.83
P <sub>3</sub>	24.21	23.34	22.8	22.39	23.18
Mean	22.68	21.65	20.19	19.61	
CD (0.05) P	CD (0.05) P; 0.058		CD (0.05) B; 0.058		.05) PxB; 0.117
SE (m) P;	SE (m) P; 0.02		SE (m) B; 0.02		m) PxB; 0.041

Table 8: Effect of P and B application on plant height at pod setting stage (cm)
T<sub>1</sub>: 30.85

	$\mathbf{B_0}$	$\mathbf{B}_1$	$\mathbf{B}_2$	<b>B</b> <sub>3</sub>	Mean	
$P_0$	30.42	28.53	25.79	25.41	27.53	
P <sub>1</sub>	31.26	30.17	29.84	30.43	30.43	
P <sub>2</sub>	31.49	31.46	28.83	26.06	29.46	
P <sub>3</sub>	32.99	32.8	32.59	31.87	32.56	
Mean	31.54	30.74	29.26	28.44		
CD (0.05)	CD (0.05) P; 0.05		CD (0.05) B; 0.05		5) PxB; 0.1	
SE (m) P; 0.018		SE (m)	B; 0.018	SE (m) PxB; 0.035		

Table 9: Effect of P and B application on plant height at harvest stage (cm)

T<sub>1</sub>: 35.75

	$\mathbf{B_0}$	<b>B</b> <sub>1</sub>	$\mathbf{B}_2$	<b>B</b> <sub>3</sub>	Mean	
$\mathbf{P}_{0}$	33.6	30.93	35.4	34.68	33.65	
Pi	35.43	33.23	37.15	34.55	35.09	
P <sub>2</sub>	31.23	34	35.83	32.95	33.5	
P <sub>3</sub>	31.95	34.15	36.23	34.95	34.32	
Mean	33.05	33.08	36.15	34.28		
CD (0.05) P; 0.865		CD (0.05) B; 0.865		CD (0.05) PxB; 1.73		
SE (m) P; 0.303		SE (m) B; 0.303		SE (m) PxB; 0.606		

# 4.1.1.2 Number of leaves

Number of leaves at flowering, pegging, pod development and harvest stages are shown in the tables 10, 11, 12, 13 respectively. Both the main effect and interaction effect of treatments significantly influenced the number of leaves. The

treatment, P<sub>3</sub>B<sub>0</sub> (P, 90kg ha<sup>-1</sup> B, 0 kg ha<sup>-1</sup>) showed highest number of leaves at flowering stage (39.64), pegging stage (60.58) and pod development stage (81.53).

Table 10: Effect of P and B application on number of leaves at flowering stage T<sub>1</sub>: 29.6

	$\mathbf{B_0}$	$\mathbf{B}_1$	$\mathbf{B}_2$	$\mathbf{B}_3$	Mean	
$P_0$	34.44	29.26	28.65	25.62	29.49	
P <sub>1</sub>	34.63	30.40	30.42	30.66	31.53	
P <sub>2</sub>	39.25	36.45	29.22	28.82	33.43	
P <sub>3</sub>	39.64	39.13	35.24	34.79	37.19	
Mean	36.99	33.81	30.88	29.97		
CD (0.05) P; 0.083		CD (0.05)	В; 0.083	CD (0.05) PxB; 0.165		
SE (m) F	SE (m) P; 0.029		B; 0.029	SE (m) PxB; 0.058		

Table 12: Effect of P and B application on number of leaves at pod setting T<sub>1</sub>: 49.5

	$\mathbf{B}_0$	$\mathbf{B}_1$	$\mathbf{B_2}$	$\mathbf{B_3}$	Mean
$P_0$	52.05	48.26	43.11	53.74	49.29
$\mathbf{P_1}$	48.14	45.47	52.43	47.63	48.42
$P_2$	50.45	54.99	51.30	53.21	52.49
P <sub>3</sub>	60.58	58.6	47.79	53.79	55.19
Mean	52.81	51.83	48.66	52.09	
CD (0.05) P; 0.089		CD (0.05) B; 0.089		CD (0.05) PxB; 0.178	
SE (m) P; 0.031		SE (m)	) B; 0.031 SE (m) PxB; 0.0		B; 0.062

Table 11: Effect of application of P and B on number of leaves at pegging stage

T<sub>1</sub>: 68.00

	$\mathbf{B_0}$	$\mathbf{B}_1$	$\mathbf{B_2}$	$\mathbf{B}_3$	Mean
P <sub>0</sub>	69.65	67.25	57.57	82.02	69.12
P <sub>1</sub>	61.60	60.37	74.45	64.60	65.26
P <sub>2</sub>	61.65	73.53	73.43	77.60	71.55
P <sub>3</sub>	81.52	78.07	60.34	72.77	73.18
Mean	68.61	69.81	66.45	74.25	
CD (0.05) P; 0.126		CD (0.05) B; 0.126		CD (0.05) PxB; 0.252	
SE (m) P; 0.044		SE (m) B; 0.044		4 SE (m) PxB; 0.088	

Table 13: Effect of P and B application on number of leaves at harvest stage

T<sub>1</sub>: 55.95

	$\mathbf{B_0}$	$\mathbf{B}_1$	$\mathbf{B}_2$	B <sub>3</sub>	Mean
$\mathbf{P_0}$	59.65	54.25	47.58	62.03	55.88
P <sub>1</sub>	51.60	40.38	54.45	54.60	50.26
P <sub>2</sub>	41.65	67.53	63.43	70.60	60.80
P <sub>3</sub>	68.53	70.08	50.34	52.78	60.43
Mean	55.36	58.06	53.95	60.00	
CD (0.05) P; 0.130		CD (0.05) B; 0.130		CD (0.05) PxB; 0.261	
SE (m) P; 0.046		SE (m)	SE (m) PxB; 0.09		B; 0.091

# 4.1.2 Yield parameters

# 4.1.2.1 Number of pods per plant

Number of pods per plant is significantly influenced by different levels of P and B. Increase in pod number per plant can be observed with increased dose of P

application (table 14). The treatment, P<sub>3</sub>B<sub>2</sub> (P, 90 kg ha<sup>-1</sup> and B, 5 kg ha<sup>-1</sup>) produced highest number of pods per plant. Number of pods increased with increased dose of P application.

#### 4.1.2.2 Yield

The data given in table 15 indicates that yield was significantly influenced by application of different doses of P and B fertilizer application. The highest yield (3.66 t ha<sup>-1</sup>) was recorded in  $P_3$   $B_1$  (P, 90 kg ha<sup>-1</sup> and B, 5 kg ha<sup>-1</sup>). The lowest yield (3.17 t ha<sup>-1</sup>) was in  $P_0$   $B_3$  (P, 0 kg ha<sup>-1</sup> and B, 15 kg ha<sup>-1</sup>). Yield was increased with increased dose of P application.

### 4.2 PHYSICO - CHEMICAL CHARACTERISTICS OF SOIL

# 4.2.1 Soil pH

The effect of application of different levels of P and B on soil pH is given in the table 16. Data showed that none of main effects and interaction effect has significant influence on the pH. There was only a slight change in pH after the harvest. pH varied from 7.80 to 7.85.

#### 4.2.2 Electrical conductivity

The effect of application of different levels of P and B on soil EC is given in the table 17. The main effect of different levels of P and B has no effect on EC, whereas the interaction effects were found to be significant. EC of soil was maximum in treatment,  $P_1 B_0$  (0.298 dS m<sup>-1</sup>).

#### 4.2.3 Organic carbon

The data on effect of application of different levels of P and B on soil OC is given in the table 18. Data showed that both main effects and interaction effect has no significant influence on OC content.

Table 14: Effect of P and B application on number of pods per plant

T<sub>1</sub>: 16.35

	$\mathbf{B}_0$	$\mathbf{B}_1$	$\mathbf{B}_2$	$\mathbf{B}_3$	Mean	
$P_0$	13.33	13.25	12.93	12.38	12.97	
$\mathbf{P_1}$	15.35	15.08	14.48	14.30	14.80	
P <sub>2</sub>	19.48	19.08	18.53	18.23	18.83	
P <sub>3</sub>	21.53	22.35	23.33	21.38	22.14	
Mean	17.42	17.44	17.31	16.57		
CD (0.05) P; 0.103		CD (0.05) B; 0.103		CD (0.05) PxB; 0.207		
SE (m) P; 0.036		SE (m) B;	0.036	SE (m) PxB:	SE (m) PxB; 0.072	

Table 15: Effect of P and B application on groundnut yield (t ha<sup>-1</sup>)

 $T_1$ : 3.40

	$\mathbf{B}_{0}$	$\mathbf{B}_{1}$	$\mathbf{B}_2$	<b>B</b> <sub>3</sub>	Mean
$\mathbf{P_0}$	3.28	3.22	3.17	3.02	3.17
$\mathbf{P}_1$	3.34	3.33	3.30	3.40	3.34
$\mathbf{P}_{2}$	3.45	3.52	3.52	3.43	3.48
P <sub>3</sub>	3.57	3.66	3.64	3.54	3.60
Mean	3.41	3.43	3.41	3.35	
CD (0.05) P; 0.056		CD (0.05) B; 0.056		CD (0.05) PxB; 0.113	
SE (m) P; 0.02			(m) B; 0.02 SE (m) PxB; 0.03		; 0.039

Table 16: Effect of P and B application on pH of soil

 $T_1:7.80$ 

	$\mathbf{B}_0$	$\mathbf{B}_1$	$\mathbf{B}_2$	<b>B</b> <sub>3</sub>	Mean
$\mathbf{P_0}$	7.85	7.85	7.84	7.84	7.84
$\mathbf{P}_{1}$	7.84	7.84	7.84	7.81	7.83
P <sub>2</sub>	7.84	7.85	7.83	7.84	7.84
P <sub>3</sub>	7.82	7.84	7.85	7.84	7.84
Mean	7.84	7.85	7.84	7.83	
CD (0.05)	P; NS	CD (0.05)	B; NS	CD (0.05) I	PxB; NS
SE (m) P;	0.005	SE (m) B	; 0.005	SE (m) PxI	3; 0.010

Table 17: Effect of P and B application on EC of soil (dS m<sup>-1</sup>)

 $T_1: 0.281$ 

	$\mathbf{B_0}$	<b>B</b> <sub>1</sub>	B <sub>2</sub>	<b>B</b> <sub>3</sub>	Mean
P <sub>0</sub>	0.285	0.285	0.278	0.286	0.283
<b>P</b> <sub>1</sub>	0.298	0.259	0.294	0.251	0.275
P <sub>2</sub>	0.290	0.290	0.289	0.290	0.290
P <sub>3</sub>	0.285	0.293	0.291	0.288	0.289
Mean	0.289	0.282	0.288	0.279	
CD (0.05) 1	P; NS	CD (0.05)	B; NS	CD (0.05)P	xB; 0.023
SE (m) P;	0.004	SE (m) B	; 0.004	SE (m) PxF	3; 0.008

Table 18: Effect of P and B application on organic carbon in soil (%)
T<sub>1</sub>: 1.041

	$\mathbf{B_0}$	$\mathbf{B}_1$	$\mathbf{B}_2$	B <sub>3</sub>	Mean
$P_0$	1.005	1.040	1.074	1.005	1.031
P <sub>1</sub>	1.043	1.020	1.055	1.009	1.032
P <sub>2</sub>	1.055	1.055	1.005	1.063	1.044
P <sub>3</sub>	1.082	1.013	1.090	1.086	1.067
Mean	1.046	1.032	1.056	1.040	
CD (0.05) I	P; NS	CD (0.05)	B; NS	CD (0.05)P	xB; NS
SE (m) P; 0.015		SE (m) B	; 0.015	SE (m) PxE	3; 0.030

### 4.3 NUTRIENT STATUS OF SOIL

### 4.3.1 Available nitrogen

The effects of application of different levels of P and B on available N are given in the table 19. Data showed that main effect, different levels of P and B have an influence on available nitrogen in the soil. N increased with increased dose of P application. However the interaction effect has no significant influence on available nitrogen.

### 4.3.2 Available phosphorus

The influence of application of different levels of P and B on available P is given in the table 20. Data showed that main effect of P and B and interaction effects were found to be significant. In comparison with initial P, available P was increased in all treatments due to fertilizer application. Treatment, P<sub>3</sub> B<sub>0</sub> (P, 90 kg ha<sup>-1</sup> and B, 0 kg ha<sup>-1</sup>) showed highest available P (51.31 kg ha<sup>-1</sup>). A decrease in available P with increased dose of B can be observed among treatments.

Table 19: Effect of P and B application on available nitrogen in soil (kg ha<sup>-1</sup>)

 $T_1$ : 254.02

	$\mathbf{B}_{0}$	B <sub>1</sub>	$\mathbf{B}_2$	$\mathbf{B}_3$	Mean
$P_0$	260.29	257.15	257.15	250.88	256.37
P <sub>1</sub>	260.29	257.15	254.02	254.02	256.37
P <sub>2</sub>	285.38	282.24	279.10	266.56	278.32
P <sub>3</sub>	291.65	288.51	285.38	282.24	286.94
Mean	274.40	271.26	268.91	263.42	
CD (0.05) P	; 5.793	CD (0.05)	B; 5.793	CD (0.05) Px	B; NS
SE (m) P; 2	.027	SE (m) B;	2.027	SE (m) PxB;	4.054

Table 20: Effect of P and B application on available phosphorus in soil (kg ha¹)
T<sub>1</sub>: 34.75

	$\mathbf{B_0}$	<b>B</b> <sub>1</sub>	$\mathbf{B_2}$	<b>B</b> <sub>3</sub>	Mean
$\mathbf{P_0}$	18.93	15.66	15.10	12.75	15.61
P <sub>1</sub>	32.89	29.68	28.40	23.90	28.72
P <sub>2</sub>	40.98	36.53	36.29	28.08	35.47
P <sub>3</sub>	51.31	50.36	38.15	35.36	43.80
Mean	36.03	33.06	29.49	25.02	
CD (0.05) P	0.794	CD (0.05)	B; 0.794	CD (0.05) P	xB; 1.588
SE (m) P; 0.	278	SE (m) B; 0.278 SE (m) P		SE (m) PxB	; 0.556

# 4.3.3 Available potassium

The influence of application of different levels of P and B on available K is given in the table 21. The main effect of P and B and interaction effects were found to

be significant. Treatment,  $P_1B_3$  (P, 60 kg ha <sup>-1</sup> and B, 15 kg ha <sup>-1</sup>) showed highest K content (454.72 kg ha <sup>-1</sup>)

### 4.3.4Available calcium

The influence of application of different levels of P and B on available Ca is given in the table 22. Available Ca significantly varied due to main effects of P and B and interaction effects. Available Ca in soil increased compared to initial sample due to fertilizer application. All the treatments had high available Ca. Highest available calcium was in treatment, P<sub>0</sub>B<sub>3</sub> (4,975.31 mg kg<sup>-1</sup>).

### 4.3.5Available magnesium

The influence of application of different levels of P and B on available Mg is given in the table 23. Data showed that available Mg content in soil significantly varied among different treatments due to main effects and interaction effects of P and B. Treatment, P<sub>3</sub>B<sub>1</sub> had highest available Mg (730.56 mg kg<sup>-1</sup>). Mg showed positive interaction with P application.

### 4.3.6 Available sulphur

The influence of application of different levels of P and B on available S is given in the table 24. There was a noticeable change in available S due to different levels of fertilizer application. Highest available S was in treatment, P<sub>3</sub>B<sub>3</sub> (20.76 mg kg<sup>-1</sup>). Available S is influenced by both main effects and interaction effect of P and B.

Table 21: Effect of P and B application on available potassium in soil (kg  $ha^{-1}$ ) T<sub>1</sub>: 374.00

P.	$\mathbf{B}_0$	$\mathbf{B}_1$	$\mathbf{B}_{2}$	$\mathbf{B}_3$	Mean
$P_0$	409.36	445.48	438.76	442.96	434.14
$P_1$	448.28	447.44	369.6	454.72	430.01
P <sub>2</sub>	428.12	451.08	306.88	321.72	376.95
P <sub>3</sub>	319.20	365.96	372.4	427.56	371.28
Mean	401.24	427.49	371.91	411.74	
CD (0.05) I	P; 3.759	CD (0.05)	B; 3.759	CD (0.05) Px	B;7.518
SE (m) P; 1	SE (m) P; 1.315		SE (m) B; 1.315		2.631

Table 22: Effect of P and B application on available calcium in soil (mg kg<sup>-1</sup>)
T<sub>1</sub>: 4,325.31

	$\mathbf{B_0}$	$\mathbf{B}_1$	$\mathbf{B_2}$	$\mathbf{B}_3$	Mean
$P_0$	4573.44	4574.69	4615.31	4675.31	4609.69
$P_1$	4314.38	4396.25	4464.06	4555.94	4432.66
P <sub>2</sub>	4399.06	4540.00	4523.44	4554.38	4504.22
P <sub>3</sub>	4447.81	4640.00	4650.00	4705.00	4610.70
Mean	4433.67	4537.73	4563.20	4622.66	
CD (0.05);	P 33.948	CD (0.05)	; B 23.948	CD (0.05) Px	кВ; 67.897
SE (m) P; 1	11.879	SE (m) B;	11.879	SE (m) PxB;	23.759

Table 23: Effect of P and B application on available magnesium in soil (mg kg<sup>-1</sup>)

T<sub>1</sub>: 687.46

	$\mathbf{B}_0$	<b>B</b> <sub>1</sub>	B <sub>2</sub>	B <sub>3</sub>	Mean
$\mathbf{P_0}$	715.84	694.28	681.56	697.78	697.37
P <sub>1</sub>	663.25	675.47	720.00	646.41	676.28
$P_2$	702.16	650.72	718.56	726.22	699.41
P <sub>3</sub>	728.06	725.59	730.56	711.75	723.99
Mean	702.33	686.52	712.67	695.54	
CD (0.05) I	P; 6.443	CD (0.05) B	; 6.443	CD (0.05) P	xB; 12.887
SE (m) P; 2	2.255	SE (m) B; 2	.225	SE (m) PxB	; 4.509

Table 24: Effect of P and B application on available sulphur in soil (mg kg<sup>-1</sup>)

T<sub>1</sub>:12.15

	${f B_0}$	$\mathbf{B}_1$	$\mathbf{B}_2$	<b>B</b> <sub>3</sub>	Mean
$P_0$	8.85	7.86	7.56	7.39	7.91
$\mathbf{P}_{1}$	8.63	11.74	12.02	13.20	11.40
P <sub>2</sub>	13.72	14.76	14.91	16.51	14.97
P <sub>3</sub>	18.92	19.41	20.49	20.76	19.89
Mean	12.53	13.44	13.74	14.46	
CD (0.05) P	; 0.538	CD (0.05) H	3; 0.538	CD (0.05) Px	кВ; 1.076
SE (m) P; 0.188		SE (m) B; (	).188	SE (m) PxB;	0.376

#### 4.3.7Available iron

The influence of application of different levels of P and B on available Fe is given in the table 25. Highest available Fe was in treatment, P<sub>0</sub>B<sub>3</sub> (8.27 mg kg<sup>-1</sup>). Available Fe was low in soil treated with high dose of P. The available Fe content in soil varied significantly due to the effect of different doses of P and B. Interaction effect has no significant effect on Fe content. Fe content decreased with increased dose of P application and increased with increased dose of B application.

### 4.3.8 Available manganese

The influence of application of different levels of P and B on available Mn is given in the table 26.Data showed that main effects of P and B and interaction effects affect the available Mn in soil. Treatment, P<sub>2</sub>B<sub>3</sub> showed highest available Mn (8.91 mg kg<sup>-1</sup>). Available Mn decreased with increased dose of P application. Mn increased with increased dose of B application.

### 4.3.9 Available zinc

The effect of application of different levels of P and B on available Zn is given in the table 27. Data showed that main effects of P and B and interaction effects affect the available Zn in the soil. Highest Zn was observed in treatment,  $P_0$  B<sub>0</sub> (1.07 mg kg<sup>-1</sup>). Zn was decreased with increased dose of P application.

### 4.3.10 Available copper

The influence of application of different levels of P and B on available Cu is given in the table 28. Both main effects and interaction effect were found to have no significant effect on available Cu in soil.



Table 25: Effect of P and B application on available iron in soil (mg kg<sup>-1</sup>)

 $T_1$ : 7.36

	$\mathbf{B_0}$	$\mathbf{B_1}$	$\mathbf{B}_2$	$\mathbf{B}_3$	Mean
$P_0$	8.01	8.02	8.15	8.27	8.11
$\mathbf{P_1}$	7.69	7.71	7.75	7.97	7.78
P <sub>2</sub>	7.33	7.40	7.49	7.82	7.51
P <sub>3</sub>	7.15	7.19	7.25	7.55	7.28
Mean	7.54	7.58	7.66	7.90	
CD (0.05) I	°; 0.143	CD (0.05)	B; 0.143	CD (0.05) Px	B; NS
SE (m) P; (	0.050	SE (m) B; 0.050 SE (m) PxB;		SE (m) PxB;	0.010

Table 26: Effect of P and B application on available Manganese in soil (mg kg<sup>-1</sup>)

 $T_1:7.79$ 

	$\mathbf{B}_0$	$\mathbf{B}_{1}$	B <sub>2</sub>	B <sub>3</sub>	Mean
. P <sub>0</sub>	7.99	8.01	8.39	8.91	8.32
P <sub>1</sub>	7.37	7.45	7.50	7.51	7.46
P <sub>2</sub>	7.63	7.65	7.67	7.78	7.68
P <sub>3</sub>	6.84	6.91	7.24	7.36	7.09
Mean	7.46	7.51	7.70	7.89	
CD (0.05) I	P; 0.099	CD (0.05)	B; 0.099	CD (0.05) P	xB; 0.199
SE (m) P; (	0.035	SE (m) B; 0.035		SE (m) PxB	; 0.069

Table 27: Effect of P and B application on available zinc in soil (mg kg<sup>-1</sup>)

 $T_1: 0.89$ 

	$\mathbf{B}_0$	· B <sub>1</sub>	$\mathbf{B}_2$	$\mathbf{B}_3$	Mean
P <sub>0</sub>	1.07	0.93	0.99	0.87	0.97
<b>P</b> <sub>1</sub>	0.89	0.83	0.90	0.88	0.88
P <sub>2</sub>	1.02	0.90	0.94	1.03	0.97
P <sub>3</sub>	0.77	0.64	0.76	0.79	0.75
Mean	0.94	0.83	0.90	0.89	
CD (0.05) P; 0.044		CD (0.05) B; 0.044		CD (0.05) PxB; 0.088	
SE (m) P; 0.015		SE (m) B;	SE (m) B; 0.015		0.031

Table 28: Effect of P and B application on available copper in soil (mg kg<sup>-1</sup>)

 $T_1: 2.15$ 

	$\mathbf{B}_0$	$\mathbf{B}_1$	$\mathbf{B}_2$	B <sub>3</sub>	Mean
P <sub>0</sub>	2.10	2.18	2.15	2.10	2.13
P <sub>1</sub>	2.13	2.16	2.10	2.14	2.13
P <sub>2</sub>	2.11	2.09	2.13	2.10	2.11
P <sub>3</sub>	2.10	2.21	2.11	2.08	2.13
Mean	2.11	2.16	2.12	2.11	
CD (0.05) F	CD (0.05) P; NS		CD (0.05) B; NS		kB; NS
SE (m) P; 0	SE (m) B; 0.0		0.018	SE (m) PxB;	0.037



### 4.3.11 Water soluble boron

Data showed (table 29) that water soluble B in soil significantly varied due to application of different levels of P and B in the soil. B in soil decreased with application of increased dose of P. The treatment,  $P_0B_3$  (P, 90 kg ha<sup>-1</sup> and B, 0 kg ha<sup>-1</sup>) showed highest B (0.80 mg kg<sup>-1</sup>) among treatments. B content decreased with increased dose of P application.

Table 29: Effect of P and B application on water soluble boron (mg kg<sup>-1</sup>)

 $T_1: 0.57$ 

	$\mathbf{B}_{0}$	$\mathbf{B}_1$	$\mathbf{B}_2$	$\mathbf{B}_3$	Mean	
$P_0$	0.64	0.66	0.74	0.80	0.71	
P <sub>1</sub>	0.61	0.62	0.62	0.62	0.62	
P <sub>2</sub>	0.55	0.56	0.57	0.62	0.58	
P <sub>3</sub>	0.49	0.51	0.56	0.59	0.54	
Mean	0.57	0.59	0.62	0.66		
CD (0.05) P; 0.025		CD (0.05) B; 0.025		CD (0.05) Px	B; 0.049	
SE (m) P; (	0.009	SE (m) B;	0.009	SE (m) P xB; 0.017		

### 4.4 ANALYSIS OF PLANT SAMPLES

### 4.4.1 Nitrogen content in groundnut plant

Nitrogen content of plant samples at flowering, pegging, pod development and harvest stages are given in the tables 30, 31, 32, 33 respectively. Nitrogen content of plant in all the stages was significantly influenced due to main effect and interaction effect of treatments. Plants showed highest nitrogen content at flowering stage, there after nitrogen content decreased with plant growth. Nitrogen content in

plant increased with application of increased dose of phosphorus. Highest nitrogen (2.62 per cent) was in the treatment, P<sub>3</sub>B<sub>0</sub> (P, 90kg ha<sup>-1</sup> and B, 0 kg ha<sup>-1</sup>).

Table 30: Effect of P and B application on nitrogen content in plant samples at flowering stage (%)

 $T_1: 2.18$ 

	$\mathbf{B}_0$	$\mathbf{B}_1$	$\mathbf{B}_2$	$\mathbf{B}_3$	Mean	
P <sub>0</sub>	2.23	2.14	1.98	1.97	2.08	
P <sub>1</sub>	2.23	2.21	2.21	2.21	2.21	
P <sub>2</sub>	2.44	2.36	2.17	2.01	2.25	
P <sub>3</sub>	2.62	2.42	2.30	2.25	2.40	
Mean	2.38	2.28	2.16	2.11		
CD (0.05) P; 0.012		CD (0.05	5) B; 0.012	CD (0.05)	PxB; 0.023	
SE (m) P; 0.004		SE (m)	SE (m) B; 0.004		SE (m) PxB; 0.008	

Table 31: Effect of P and B application on nitrogen content in plant samples at pegging stage (%)

 $T_1$ : 2.00

	$\mathbf{B}_0$	$\mathbf{B}_1$	$\mathbf{B_2}$	<b>B</b> <sub>3</sub>	Mean
P <sub>0</sub>	1.98	1.93	1.82	1.81	1.88
P <sub>1</sub>	1.97	1.93	1.98	1.93	1.95
P <sub>2</sub>	2.15	2.07	1.94	1.82	1.99
P <sub>3</sub>	2.31	2.21	2.06	2.01	2.15
Mean	2.10	2.04	1.95	1.89	
CD (0.05) P; 0.008		CD (0.05) B; 0.008		CD (0.05) PxB; 0.015	
SE (m) P; 0.003		SE (m)	SE (m) B; 0.003		xB; 0.005

Table 32: Effect of P and B application on nitrogen content in plant samples at pod setting stage (%)

T<sub>1</sub>: 1.79

	${f B}_0$	$\mathbf{B}_1$	$\mathbf{B}_2$	B <sub>3</sub>	Mean
P <sub>0</sub>	1.73	1.73	1.66	1.65	1.69
P <sub>1</sub>	1.72	1.65	1.76	1.65	1.69
P <sub>2</sub>	1.86	1.79	1.70	1.62	1.74
P <sub>3</sub>	2.00	2.00	1.82	1.78	1.90
Mean	1.83	1.79	1.73	1.68	
CD (0.05) P; 0.012		CD (0.05) B; 0.012		CD (0.05)	PxB; 0.023
SE (m) P; 0.004		SE (m)	B; 0.004	SE (m) PxB; 0.008	

Table 33: Effect of P and B application on nitrogen content in plant samples at harvest stages (%)

 $T_1$ : 1.61

	$\mathbf{B}_0$	$\mathbf{B}_1$	$\mathbf{B_2}$	$\mathbf{B}_3$	Mean
$P_0$	1.53	1.53	1.50	1.51	1.52
P <sub>1</sub>	1.53	1.49	1.53	1.49	1.51
P <sub>2</sub>	1.64	1.55	1.51	1.47	1.54
P <sub>3</sub>	1.75	1.75	1.61	1.55	1.66
Mean	1.61	1.58	1.54	1.50	
CD (0.05) P; 0.009		CD (0.05) B; 0.009		CD (0.05)	PxB; 0.018
SE (m) P; 0.003		SE (m)	B; 0.003	SE (m) P	xB; 0.006

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## 4.4.2 Phosphorus content in plant

Phosphorus content in plant sample at flowering, pegging, pod development and harvest stages are given in the tables 34, 35, 36, 37 respectively. P content in plant was increased with increased dose of fertilizer application. P content in all the stages significantly varied due to main effect and interaction effect. The treatment, P<sub>3</sub> B<sub>1</sub> (P, 90kg ha<sup>-1</sup> and B, 5 kg ha<sup>-1</sup>) showed highest P content (0.37 per cent at flowering stage, 0.34 per cent at pegging stage, 0.30 per cent at pod development stage and 0.27 per cent at harvest stage).

Table 34: Effect of P and B application on phosphorus content in plant samples at flowering stage (%)

 $T_1: 0.33$ 

	$\mathbf{B}_{0}$	<b>B</b> <sub>1</sub>	B <sub>2</sub>	B <sub>3</sub>	Mean
$P_0$	0.27	0.25	0.24	0.21	0.24
. P <sub>1</sub>	0.30	0.30	0.30	0.30	0.30
P <sub>2</sub>	0.33	0.32	0.32	0.32	0.32
P <sub>3</sub>	0.36	0.37	0.33	0.34	0.35
Mean	0.31	0.31	0.30	0.29	
CD (0.05) P; 0.011		CD (0.05) B; 0.011		CD (0.05) PxB; 0.02	
SE (m) P; 0.004		SE (m) B; 0.004		SE (m) PxB; 0.007	

Table 35: Effect of P and B application on phosphorus content in plant samples at pegging stage (%)

T<sub>1</sub>: 0.33

	$\mathbf{B}_0$	<b>B</b> <sub>1</sub>	B <sub>2</sub>	<b>B</b> <sub>3</sub>	Mean
$P_0$	0.23	0.22	0.20	0.18	0.21
$\mathbf{P}_{1}$	0.25	0.25	0.26	0.27	0.26
P <sub>2</sub>	0.31	0.29	0.28	0.27	0.29
P <sub>3</sub>	0.33	0.34	0.31	0.31	0.32
Mean	0.28	0.27	0.26	0.26	
CD (0.05	CD (0.05) P; 0.005		CD (0.05) B; 0.005		PxB; 0.011
SE (m)	SE (m) P; 0.002		SE (m) B; 0.002		xB; 0.004

# Table 36: Effect of P and B application on phosphorus content in plant samples at pod setting stage (%)

T<sub>1</sub>:0.29

	$\mathbf{B}_0$	$\mathbf{B}_1$	B <sub>2</sub>	B <sub>3</sub>	Mean
$P_0$	0.20	0.19	0.17	0.14	0.17
$\mathbf{P}_{1}$	0.21	0.21	0.22	0.24	0.22
P <sub>2</sub>	0.28	0.27	0.24	0.22	0.25
P <sub>3</sub>	0.29	0.30	0.29	0.29	0.29
Mean	0.24	0.24	0.23	0.22	
CD (0.05)	CD (0.05) P; 0.006		) B; 0.006	CD (0.05) PxB; 0.011	
SE (m)	SE (m) P; 0.002		B; 0.002	SE (m) PxB; 0.004	

Table 37: Effect of P and B application on phosphorus content in plant samples at harvest stage (%)

 $T_1$ : 0.26

	$\mathbf{B}_0$	$\mathbf{B}_1$	$\mathbf{B}_2$	B <sub>3</sub>	Mean
$P_0$	0.17	0.16	0.14	0.12	0.15
$\mathbf{P_1}$	0.18	0.18	0.19	0.19	0.18
P <sub>2</sub>	0.24	0.23	0.21	0.19	0.22
P <sub>3</sub>	0.26	0.27	0.2	0.2	0.26
Mean	0.21	0.20	0.19	0.19	
CD (0.05	CD (0.05) P; 0.007		CD (0.05) B; 0.007		PxB; 0.013
SE (m)	SE (m) P; 0.002		3; 0.002	SE (m) Px	В; 0.005

### 4.4.3 Potassium content in plant

Potassium content in plant samples was significantly influenced by application of P and B. The treatment, P<sub>2</sub>B<sub>3</sub> (P, 75kg ha <sup>-1</sup> and B, 15 kg ha <sup>-1</sup>) had highest K content at flowering (2.21 per cent). At pegging, pod development and harvesting stages treatment, P<sub>2</sub>B<sub>0</sub> (P, 75kg ha <sup>-1</sup> and B, 0 kg ha <sup>-1</sup>) showed highest K content.

Table 38: Effect of P and B application on plant potassium content at flowering stage (%)

 $T_1$ : 2.18

	$\mathbf{B_0}$	$\mathbf{B}_1$	$\mathbf{B}_2$	$\mathbf{B}_3$	Mean
$P_0$	2.06	2.08	1.95	1.65	1.94
P <sub>1</sub>	2.00	1.87	1.99	1.86	1.93
P <sub>2</sub>	2.10	1.81	2.01	2.21	2.03
P <sub>3</sub>	1.96	2.19	1.81	2.03	2.00
Mean	2.03	1.99	1.94	1.94	
CD (0.05)	CD (0.05) P; 0.039		CD (0.05) B; 0.039		PxB; 0.077
SE (m)	SE (m) P; 0.014		SE (m) B; 0.014		кВ; 0.028

# Table 39: Effect of P and B application on plant potassium content at pegging stage (%)

T<sub>1</sub>: 1.88

	$\mathbf{B_0}$	$\mathbf{B}_1$	$\mathbf{B_2}$	$\mathbf{B}_3$	Mean
$\mathbf{P_0}$	1.70	1.83	1.59	1.67	1.70
$\mathbf{P}_1$	1.85	1.77	1.85	1.82	1.82
P <sub>2</sub>	1.95	1.73	1.90	1.93	1.88
P <sub>3</sub>	1.72	1.78	1.72	1.85	1.77
Mean	1.81	1.78	1.76	1.82	
CD (0.05)	CD (0.05) P; 0.023		B; 0.023	CD (0.05) PxB; 0.046	
SE (m)	P; 0.008	SE (m) B; 0.008		SE (m) PxB; 0.016	

Table 40: Effect of P and B application on plant potassium content at pod setting stage (%)

 $T_1$ : 1.58

	$\mathbf{B}_{0}$	$\mathbf{B}_1$	$\mathbf{B}_2$	$\mathbf{B}_3$	Mean
$\mathbf{P_0}$	1.34	1.59	1.38	1.53	1.46
P <sub>1</sub>	1.71	1.66	1.71	1.78	1.71
P <sub>2</sub>	1.80	1.65	1.79	1.64	1.72
P <sub>3</sub>	1.48	1.37	1.63	1.68	1.54
Mean	1.58	1.57	1.63	1.66	×
CD (0.05)	CD (0.05) P; 0.025		CD (0.05) B; 0.025		PxB; 0.051
SE (m) P	SE (m) P; 0.009		SE (m) B; 0.009		xB; 0.018

Table 41: Effect of P and B application on plant potassium content at harvest stage (%)

T<sub>1</sub>: 1.49

	B <sub>0</sub>	B <sub>1</sub>	B <sub>2</sub>	B <sub>3</sub>	Mean
$P_0$	1.26	1.50	1.27	1.46	1.37
$P_1$	1.62	1.54	1.59	1.66	1.60
P <sub>2</sub>	1.69	1.54	1.66	1.54	1.61
P <sub>3</sub>	1.35	1.31	1.54	1.58	1.44
Mean	1.48	1.47	1.51	1.56	
CD (0.05)	CD (0.05) P; 0.029		CD (0.05) B; 0.029		xB; 0.059
SE (m) I	SE (m) P; 0.010		SE (m) B; 0.010		B; 0.021

## 4.4.4 Calcium content in plant

Calcium content in plant samples at all the four stages were found to be significantly varied due to application of different levels of fertilizers. Ca content was highest in treatment having low P. P<sub>0</sub>B<sub>3</sub> (P, 0 kg ha<sup>-1</sup> and B, 15 kg ha<sup>-1</sup>) showed highest Ca content at flowering (1.98 per cent), pegging (1.61 per cent), pod setting (1.24 per cent) and harvest(1.01 per cent) stages.

Table 42: Effect of P and B application on plant calcium content at flowering stage (%)

 $T_1$ : 1.51

	$\mathbf{B_0}$	$\mathbf{B}_1$	$\mathbf{B}_2$	$\mathbf{B}_3$	Mean
$\mathbf{P_0}$	1.84	1.87	1.98	1.98	1.91
$\mathbf{P}_1$	1.52	1.53	1.63	1.73	1.60
P <sub>2</sub>	1.55	1.70	1.70	1.72	1.67
P <sub>3</sub>	1.56	1.76	1.79	1.83	1.73
Mean	1.62	1.71	1.77	1.81	
CD (0.05	CD (0.05) P; 0.029		В; 0.029	CD (0.05) F	PxB; 0.058
SE (m)	SE (m) P; 0.010		B; 0.010	SE (m) PxB; 0.020	

Table 43: Effect of P and B application on plant calcium content at pegging stage (%)

T<sub>1</sub>: 1.14

	$\mathbf{B}_0$	$\mathbf{B}_1$	$\mathbf{B}_2$	B <sub>3</sub>	Mean
$P_0$	1.50	1.52	1.60	1.61	1.56
P <sub>1</sub>	1.09	1.16	1.24	1.34	1.21
P <sub>2</sub>	1.18	1.30	1.28	1.31	1.27
P <sub>3</sub>	1.20	1.39	1.43	1.45	1.37
Mean	1.24	1.34	1.38	1.43	
CD (0.05)	CD (0.05) P; 0.018		CD (0.05) B; 0.018		PxB; 0.037
SE (m) I	SE (m) P; 0.006		SE (m) B; 0.006		xB; 0.012

Table 44: Effect of P and B application on plant calcium content at pod setting stage (%)

 $T_1: 0.76$ 

	$\mathbf{B_0}$	<b>B</b> <sub>1</sub>	$\mathbf{B}_2$	$\mathbf{B}_3$	Mean
$\mathbf{P_0}$	1.15	1.18	1.22	1.24	1.20
$\mathbf{P_1}$	0.66	0.80	0.84	0.95	0.81
$P_2$	0.81	0.90	0.85	0.90	0.86
$P_3$	0.84	1.02	1.07	1.07	1.00
Mean	0.87	0.97	1.00	1.04	
CD (0.05)	CD (0.05) P; 0.023		CD (0.05) B;0.023		PXB; 0.045
SE (m) P	SE (m) P; 0.008		SE (m) B; 0.008		кВ; 0.016



Table 45: Effect of P and B application on plant calcium content at harvest stage (%)

 $T_1: 0.48$ 

	$\mathbf{B_0}$	$\mathbf{B}_1$	$\mathbf{B}_2$	$\mathbf{B}_3$	Mean
$P_0$	0.87	0.89	0.99	1.01	0.94
P <sub>1</sub>	0.46	0.55	0.64	0.69	0.58
P <sub>2</sub>	0.62	0.70	0.66	0.70	0.67
P <sub>3</sub>	0.65	0.81	0.81	0.83	0.77
Mean	0.65	0.74	0.78	0.81	
CD (0.05)	CD (0.05) P; 0.029		CD (0.05) B;0.029		PxB; 0.058
SE (m)	SE (m) P; 0.010		SE (m) B; 0.010		в; 0.020

### 4.4.5 Magnesium content in plant

Magnesium content in plant samples was significantly influenced by application of P and B. The treatment,  $P_0B_1$  (P, 0 kg ha <sup>-1</sup> and B, 10 kg ha <sup>-1</sup>) had highest Mg content at flowering (2.97 per cent). At pegging stage, treatment,  $P_1B_1$  (P, 60 kg ha <sup>-1</sup> and B, 10 kg ha <sup>-1</sup>) had highest Mg content (2.02 per cent). pod development and harvesting stages treatment,  $P_2B_0$  (P, 75kg ha <sup>-1</sup> and B, 0 kg ha <sup>-1</sup>) showed highest Mg content.

Table 46: Effect of P and B application on plant magnesium content at flowering stage (%)

 $T_1$ : 2.41

	$\mathbf{B}_{0}$	$\mathbf{B_1}$	B <sub>2</sub>	B <sub>3</sub>	Mean
P <sub>0</sub>	2.79	2.97	2.73	3.39	2.97
P <sub>1</sub>	2.46	2.62	2.97	2.67	2.68
P <sub>2</sub>	2.80	2.73	2.92	2.64	2.77
P <sub>3</sub>	2.53	2.85	2.38	2.63	2.59
Mean	2.65	2.79	2.75	2.83	*
CD (0.05	CD (0.05) P; 0.060		CD (0.05) B; 0.060		PxB; 0.120
SE (m)	SE (m) P; 0.020		SE (m) B; 0.020		xB; 0.039

Table 47: Effect of P and B application on plant magnesium content at pegging stage (%)

T<sub>1</sub>: 1.68

s	$\mathbf{B_0}$	<b>B</b> <sub>1</sub>	B <sub>2</sub>	B <sub>3</sub>	Mean
P <sub>0</sub>	1.95	2.02	1.86	2.18	2.00
P <sub>1</sub>	1.66	1.82	1.97	1.83	1.82
P <sub>2</sub>	1.85	1.84	1.92	1.84	1.86
P <sub>3</sub>	1.79	1.98	1.72	1.75	1.81
Mean	1.81	1.91	1.86	1.89	
CD (0.05	CD (0.05) P; 0.031		CD (0.05) B; 0.031		PxB; 0.061
SE (m)	SE (m) P; 0.010		SE (m) B; 0.010		кВ; 0.020

Table 48: Effect of P and B application on plant magnesium content at pod setting stage (%)

 $T_1$ : 0.97

	$\mathbf{B_0}$	$\mathbf{B_1}$	B <sub>2</sub>	B <sub>3</sub>	Mean	
P <sub>0</sub>	1.09	1.07	0.98	0.97	1.03	
$P_1$	0.85	1.03	0.97	0.97	0.95	
$P_2$	0.89	0.94	0.91	1.03	0.94	
P <sub>3</sub>	1.05	1.09	1.06	0.87	1.01	
Mean	0.97	1.03	0.98	0.96		
CD (0.05	CD (0.05) P; 0.024		CD (0.05) B; 0.024		CD (0.05) PxB; 0.048	
SE (m)	SE (m) P; 0.008		SE (m) B; 0.008		В; 0.016	

# Table 49: Effect of P and B application on plant magnesium content at harvest stage (%)

 $T_1$ : 0.86

	$\mathbf{B_0}$	$\mathbf{B}_1$	B <sub>2</sub>	B <sub>3</sub>	Mean
$P_0$	1.03	0.98	0.88	0.86	0.94
P <sub>1</sub>	0.75	0.91	0.86	0.86	0.84
$P_2$	0.79	0.84	0.81	0.92	0.84
P <sub>3</sub>	0.94	0.99	0.91	0.79	0.90
Mean	0.88	0.93	0.86	0.86	
CD (0.05	CD (0.05) P; 0.022		CD (0.05) B; 0.022		PxB; 0.044
SE (m)	P; 0.007	SE (m) B; 0.007		SE (m) P	xB; 0.014

### 4.4.6 Sulphur content in plant

Sulphur content of plant at different stages varied significantly due to main effect and interaction effect. S content at all the stages were influenced by treatments. The treatment having high P produced high S content. Treatment, P<sub>3</sub> B<sub>3</sub> (P, 90 kg ha<sup>-1</sup> and B, 15 kg ha<sup>-1</sup>) showed highest S content at flowering (0.83 per cent) and pegging (0.60 per cent) stages and P<sub>3</sub>B<sub>2</sub> (P, 90 kg ha<sup>-1</sup> and B 10 kg ha<sup>-1</sup>) showed highest S content at pod setting(0.44 per cent) and harvest stages (0.39 per cent).

Table 50: Effect of P and B application on plant sulphur content at flowering stage (%)

 $T_1: 0.53$ 

	$\mathbf{B_0}$	$\mathbf{B}_1$	$\mathbf{B}_2$	B <sub>3</sub>	Mean
$P_0$	0.46	0.43	0.39	0.38	0.41
<b>P</b> <sub>1</sub>	0.41	0.47	0.49	0.51	0.47
P <sub>2</sub>	0.52	0.53	0.54	0.58	0.54
P <sub>3</sub>	0.57	0.61	0.62	0.83	0.66
Mean	0.49	0.51	0.51	0.57	
CD (0.05) P	CD (0.05) P; 0.009		) B; 0.009	CD (0.05)	PxB; 0.019
SE (m) P;	SE (m) P; 0.003		B; 0.003	SE (m) PxB; 0.007	

Table 51: Effect of P and B application on plant sulphur content at pegging stage (%)

T<sub>1</sub>: 0.29

	$\mathbf{B_0}$	$\mathbf{B}_1$	$\mathbf{B_2}$	$\mathbf{B}_3$	Mean
$\mathbf{P_0}$	0.34	0.32	0.30	0.29	0.31
P <sub>1</sub>	0.32	0.35	0.37	0.40	0.36
P <sub>2</sub>	0.40	0.43	0.42	0.44	0.42
P <sub>3</sub>	0.44	0.47	0.53	0.60	0.51
Mean	0.37	0.39	0.40	0.43	
CD (0.05) P;	CD (0.05) P; 0.007		) B;0.007	CD (0.05) I	PxB; 0.014
SE (m) P; (	SE (m) P; 0.002		B; 0.002	SE (m) Px	в; 0.005

Table 52: Effect of P and B application on plant sulphur content at pod setting stage (%)

T<sub>1</sub>: 0.20

	$\mathbf{B}_0$	<b>B</b> <sub>1</sub>	B <sub>2</sub>	B <sub>3</sub>	Mean	
P <sub>0</sub>	0.22	0.21	0.21	0.20	0.21	
P <sub>1</sub>	0.22	0.24	0.25	0.28	0.25	
P <sub>2</sub>	0.27	0.28	0.30	0.31	0.29	
P <sub>3</sub>	0.32	0.34	0.44	0.36	0.36	
Mean	0.26	0.27	0.30	0.29		
CD (0.05) P;	CD (0.05) P;0.016		) B;0.016	CD (0.05) PxB; 0.031		
SE (m) P; 0	SE (m) P; 0.005		SE (m) B; 0.005		SE (m) PxB; 0.011	

Table 53: Effect of P and B application on plant sulphur content at harvest stage (%)

 $T_1: 0.27$ 

	$\mathbf{B}_0$	$\mathbf{B}_1$	$\mathbf{B}_2$	<b>B</b> <sub>3</sub>	Mean	
P <sub>0</sub> ac	0.19	0.18	0.18	0.18	0.18	
P <sub>1</sub>	0.19	0.21	0.22	0.26	0.22	
P <sub>2</sub>	0.24	0.27	0.26	0.28	0.26	
P <sub>3</sub>	0.28	0.30	0.39	0.33	0.33	
Mean	0.23	0.24	0.26	0.26		
CD (0.05) P;	CD (0.05) P; 0.011		CD (0.05) B; 0.011		PxB; 0.023	
SE (m) P; (	SE (m) P; 0.004		SE (m) B; 0.004		SE (m) PxB; 0.008	

## 4.4.7 Iron content in groundnut plant

Iron content at all stages was significantly affected by main effect and interaction effect.  $P_0B_3$  (P, 0kg ha<sup>-1</sup> and B, 15 kg ha<sup>-1</sup>) showed highest Fe content at flowering stage (497.50 mg kg<sup>-1</sup>). At pegging and harvesting stage treatment,  $P_0B_0$  (P, 0 kg ha<sup>-1</sup> and B, 0 kg ha<sup>-1</sup>) showed highest Fe content. At pod development stage treatment  $P_0B_2$  (P, 0 kg ha<sup>-1</sup> and B, 10 kg ha<sup>-1</sup>) showed highest Fe content.

Table 54: Effect of P and B application on plant iron content at flowering stage (mg kg<sup>-1</sup>)

T<sub>1</sub>: 425.52

	$\mathbf{B_0}$	$\mathbf{B}_1$	$\mathbf{B_2}$	<b>B</b> <sub>3</sub>	Mean	
$\mathbf{P_0}$	424.63	366.04	388.41	497.50	419.14	
P <sub>1</sub>	458.54	417.13	390.60	376.47	410.68	
P <sub>2</sub>	383.75	380.63	312.00	418.66	373.76	
P <sub>3</sub>	359.28	336.13	451.85	454.41	400.42	
Mean	406.55	374.98	385.71	436.76	8	
CD (0.05) I	CD (0.05) P; 14.021		CD (0.05) B;14.021		CD (0.05) PxB; 28.041	
SE (m) P	; 4.906	SE (m)	SE (m) B; 4.906 SE		m) PxB; 9.812	

# Table 55: Effect of P and B application on plant iron content at pegging stage (mg kg<sup>-1</sup>)

T<sub>1</sub>: 414.98

	$\mathbf{B}_0$	$\mathbf{B}_1$	$\mathbf{B}_2$	B <sub>3</sub>	Mean	
$P_0$	400.69	306.65	382.65	395.82	371.45	
P <sub>1</sub>	375.30	371.13	314.96	355.49	354.22	
P <sub>2</sub>	352.12	345.99	304.02	340.77	335.72	
P <sub>3</sub>	327.71	332.22	388.08	331.33	344.84	
Mean	363.95	339.00	347.43	355.85		
CD (0.05)	CD (0.05) P; 9.533		CD (0.05) B; 9.533		CD (0.05) PxB; 19.066	
SE (m) P	SE (m) P; 3.336		SE (m) B; 3.336		SE (m) PxB; 6.672	

Table 56: Effect of P and B application on plant iron content at pod setting stage (mg kg<sup>-1</sup>)

T<sub>1</sub>: 378.56

	$\mathbf{B_0}$	B <sub>1</sub>	$\mathbf{B}_2$	B <sub>3</sub>	Mean
$P_0$	376.75	247.25	376.88	294.13	323.75
P <sub>1</sub>	292.06	325.13	239.31	334.50	297.75
P <sub>2</sub>	320.47	311.35	296.04	262.88	297.68
P <sub>3</sub>	296.13	328.32	324.31	208.25	289.25
Mean	321.35	303.01	309.13	274.94	
CD (0.05)	CD (0.05) P; 13.267		CD (0.05) B;13.267		xB; 26.534
SE (m)	P; 4.643	SE (m)	SE (m) B; 4.643		xB; 9.285

# Table 57: Effect of P and B application on plant iron content at harvest stage (mg kg<sup>-1</sup>)

T<sub>1</sub>: 348.36

	$\mathbf{B_0}$	$\mathbf{B}_1$	B <sub>2</sub>	B <sub>3</sub>	Mean	
P <sub>0</sub>	359.28	233.42	357.76	275.89	306.59	
P <sub>1</sub>	272.96	308.51	229.29	325.70	284.11	
P <sub>2</sub>	315.31	305.39	292.31	247.68	290.17	
P <sub>3</sub>	276.76	326.81	304.48	199.31	276.84	
Mean	306.08	293.53	295.96	262.15		
CD (0.05) P	CD (0.05) P; 9.327		CD (0.05) B;9.327		xB; 18.654	
SE (m) P;	SE (m) P; 3.264		SE (m) B; 3.264		SE (m) PxB; 6.528	

## 4.4.8 Manganese content in groundnut plant

Manganese content in plant sample at flowering, pegging, pod development and harvest stages are given in the tables 58, 59, 60, 61 respectively. Mn content was high in treatment having low P. The treatment, P<sub>1</sub>B<sub>3</sub> (P, 0 kg ha<sup>-1</sup> and B, 15 kg ha<sup>-1</sup>) showed highest Mn content at flowering (82.25 mg kg<sup>-1</sup>), pegging (74.38 mg kg<sup>-1</sup>), pod setting (66.5 mg kg<sup>-1</sup>) and harvest (62.38 mg kg<sup>-1</sup>) stage. Mn content was found to be significantly different in all stages. But interaction effect at harvest stage was non-significant.

Table 58: Effect of P and B application on plant manganese content at flowering stage (mg kg<sup>-1</sup>)

T<sub>1</sub>: 74.25

	$\mathbf{B}_0$	$\mathbf{B}_1$	$\mathbf{B}_2$	<b>B</b> <sub>3</sub>	Mean
$\mathbf{P_0}$	66.00	71.19	72.69	74.25	71.03
P <sub>1</sub>	77.63	78.88	81.63	82.25	80.09
P <sub>2</sub>	60.94	64.69	65.06	74.81	66.38
P <sub>3</sub>	58.06	58.81	59.25	60.56	59.17
Mean	65.66	68.39	69.66	72.97	
CD (0.05)	CD (0.05) P; 2.054		CD (0.05) B; 2.054		PxB; 4.109
SE (m) P	SE (m) P; 0.719		SE (m) B; 0.719		xB; 1.438

Table 59: Effect of P and B application on plant manganese content at pegging stage (mg kg<sup>-1</sup>)

T<sub>1</sub>: 70.00

	$\mathbf{B_0}$	<b>B</b> <sub>1</sub>	B <sub>2</sub>	<b>B</b> <sub>3</sub>	Mean
$\mathbf{P_0}$	62.25	64.97	66.09	67.50	65.20
<b>P</b> <sub>1</sub>	70.69	72.13	73.81	74.38	72.75
P <sub>2</sub>	54.16	57.91	61.28	65.41	59.69
P <sub>3</sub>	50.34	50.97	52.19	52.91	51.60
Mean	59.36	61.49	63.35	65.05	
CD (0.05)	CD (0.05) P; 1.197		CD (0.05) B; 1.197		PxB; 2.394
SE (m) P	; 0.419	SE (m)	B; 0.419	SE (m) PxB; 0.8	

Table 60: Effect of P and B application on plant manganese content at pod setting stage (mg kg<sup>-1</sup>)

T<sub>1</sub>: 67.16

	$\mathbf{B}_{0}$	$\mathbf{B}_1$	$\mathbf{B_2}$	B <sub>3</sub>	Mean	
$\mathbf{P_0}$	58.50	58.75	59.50	60.75	59.38	
.P <sub>1</sub>	63.75	65.38	66.00	66.50	65.41	
P <sub>2</sub>	47.38	51.13	57.50	56.00	53.00	
P <sub>3</sub>	42.63	43.13	45.14	45.25	44.03	
Mean	53.06	54.59	57.03	57.13		
CD (0.05) P; 1.725		CD (0.05) B; 1.725		CD (0.05) PxB; 3.451		
SE (m) P	SE (m) P; 0.604		SE (m) B; 0.604		SE (m) PxB; 1.208	

Table 61: Effect of P and B application on plant manganese content at harvest stage (mg kg<sup>-1</sup>)

T<sub>1</sub>: 65.81

	$\mathbf{B}_0$	<b>B</b> <sub>1</sub>	B <sub>2</sub>	B <sub>3</sub>	Mean
$\mathbf{P_0}$	55.06	55.50	57.44	56.75	56.19
P <sub>1</sub>	60.88	61.44	62.13	62.38	61.70
P <sub>2</sub>	44.38	48.13	53.50	52.00	49.50
P <sub>3</sub>	39.25	40.56	41.56	42.00	40.84
Mean	49.89	51.41	53.66	53.28	
CD (0.05)	CD (0.05) P; 1.596		CD (0.05) B; 1.596		PxB; NS
SE (m) P	SE (m) P; 0.558		SE (m) B; 0.558		xB; 1.117

## 4.4.9 Zinc content in plant

Zinc content in plant samples at flowering, pegging, pod development and harvesting stages were significantly influenced by main effect and interaction effect. Zn content was highest in treatment having low P. At flowering stage, P<sub>0</sub>B<sub>3</sub> (P, 0 kg ha<sup>-1</sup> and B, 15 kg ha<sup>-1</sup>) showed highest Zn content (53.06 mg kg<sup>-1</sup>). Treatment, P<sub>0</sub>B<sub>2</sub> (P, 0 kg ha<sup>-1</sup> and B, 10 kg ha<sup>-1</sup>) showed highest Zn at pegging (50.00 mg kg<sup>-1</sup>), pod setting (47.63 mg kg<sup>-1</sup>) and harvest (43.48 mg kg<sup>-1</sup>) stages.

Table 62: Effect of P and B application on plant zinc content at flowering stage (mg kg<sup>-1</sup>)

T<sub>1</sub>: 50.50

$\mathbf{B_0}$	$\mathbf{B}_1$	$\mathbf{B_2}$	$\mathbf{B}_3$	Mean	
49.13	46.88	52.38	53.06	50.36	
41.44	41.38	42.25	41.88	41.73	
40.56	41.81	41.63	44.13	42.03	
39.75	38.31	37.63	37.06	38.19	
42.72	42.09	43.47	44.03		
CD (0.05) P; 0.914		CD (0.05) B; 0.914		PxB; 1.827	
SE (m) P; 0.320		SE (m) B; 0.320		SE (m) PxB; 0.639	
	49.13 41.44 40.56 39.75 42.72 P; 0.914	49.13 46.88 41.44 41.38 40.56 41.81 39.75 38.31 42.72 42.09 P; 0.914 CD (0.05	49.13 46.88 52.38  41.44 41.38 42.25  40.56 41.81 41.63  39.75 38.31 37.63  42.72 42.09 43.47  P; 0.914 CD (0.05) B; 0.914	49.13       46.88       52.38       53.06         41.44       41.38       42.25       41.88         40.56       41.81       41.63       44.13         39.75       38.31       37.63       37.06         42.72       42.09       43.47       44.03         P; 0.914       CD (0.05) B; 0.914       CD (0.05) B	

Table 63: Effect of P and B application on plant zinc content at pegging stage (mg kg<sup>-1</sup>)

 $T_1$ : 42.38

	$\mathbf{B_0}$	$\mathbf{B_{1}}$	B <sub>2</sub>	B <sub>3</sub>	Mean	
$P_0$	47.82	46.19	50.00	48.03	48.01	
$P_1$	40.13	40.57	39.76	39.75	40.05	
P <sub>2</sub>	37.32	38.41	37.38	36.88	37.50	
P <sub>3</sub> '	34.44	35.72	35.00	34.35	34.88	
Mean	39.92	40.22	40.54	39.75		
CD (0.05) I	CD (0.05) P; 0.559		CD (0.05) B; 0.559		PxB; 1.119	
SE (m) P;	SE (m) P; 0.196		SE (m) B; 0.196		SE (m) PxB; 0.391	

Table 64: Effect of P and B application on plant zinc content at pod setting stage (mg kg<sup>-1</sup>)

T<sub>1</sub>: 33.00

	$\mathbf{B_0}$	$\mathbf{B}_1$	$\mathbf{B}_2$	B <sub>3</sub>	Mean	
$P_0$	46.50	45.50	47.63	43.00	45.66	
P <sub>1</sub>	38.81	39.75	37.25	37.63	38.36	
P <sub>2</sub>	34.06	35.00	33.13	29.63	32.95	
P <sub>3</sub>	29.13	33.13	32.38	31.63	31.56	
Mean	37.13	38.34	37.59	35.47		
CD (0.05) P	CD (0.05) P; 0.712		CD (0.05) B; 0.712		CD (0.05) PxB; 1.424	
SE (m) P;	SE (m) P; 0.249		SE (m) B; 0.249		SE (m) PxB; 0.498	

# Table 65: Effect of P and B application on plant zinc content at harvest stage (mg kg<sup>-1</sup>)

T<sub>1</sub>: 30.30

	$\mathbf{B}_0$	$\mathbf{B}_1$	B <sub>2</sub>	B <sub>3</sub>	Mean	
$P_0$	41.94	41.00	43.48	39.81	41.56	
<b>P</b> <sub>1</sub>	35.56	36.00	33.75	33.56	34.72	
P <sub>2</sub>	30.81	30.94	30.06	26.69	29.63	
P <sub>3</sub>	26.19	30.13	28.35	28.13	28.20	
Mean	33.63	34.52	33.91	32.05		
CD (0.05) I	CD (0.05) P; 0.393		CD (0.05) B; 0.393		CD (0.05) PxB; 0.785	
SE (m) P;	SE (m) P; 0.137		SE (m) B; 0.137		SE (m) PxB; 0.275	

## 4.4.10 Copper content in plant

Copper content in plant samples at flowering, pegging, pod development and harvest stages are given in the tables 66, 67, 68, 69 respectively. Cu content at flowering and pegging stage was significant due to main effect of P and interaction effect and the main effect of B had no significant influence on Cu content at all these stages. At pod development stage and harvest stage Cu content was non-significant due to both main effects

Table 66: Effect of P and B application on plant copper content at flowering stage (mg kg<sup>-1</sup>)

 $T_1$ : 20.00

	$\mathbf{B}_{0}$	$\mathbf{B}_1$	$\mathbf{B}_2$	<b>B</b> <sub>3</sub>	Mean	
$\mathbf{P_0}$	18.38	18.38	19.25	17.63	18.41	
$P_1$	17.25	17.25	16.25	19.75	17.63	
$P_2$	19.63	18.38	19.50	17.00	18.63	
P <sub>3</sub>	15.63	15.75	16.38	18.13	16.47	
Mean	17.72	17.44	17.84	18.13		
CD (0.05) I	CD (0.05) P; 1.100		CD (0.05) B; NS		CD (0.05) PxB; 2.199	
SE (m) P;	SE (m) P; 0.385		SE (m) B; 0.385		SE (m) PxB; 0.770	

Table 67: Effect of P and B application on plant copper content at pegging stage (mg kg<sup>-1</sup>)

T<sub>1</sub>: 18.84

	$\mathbf{B}_{0}$	<b>B</b> <sub>1</sub>	$\mathbf{B}_{2}$	$\mathbf{B}_3$	Mean	
$P_0$	16.13	16.06	15.88	15.41	15.87	
$\mathbf{P}_{1}$	15.47	15.50	14.72	16.22	15.48	
P <sub>2</sub>	16.72	16.13	16.31	15.06	16.06	
P <sub>3</sub>	14.00	14.44	15.16	15.97	14.89	
Mean	15.58	15.53	15.52	15.66		
CD (0.05)	CD (0.05) P; 0.658		CD (0.05) B; NS		CD (0.05) PxB; 1.315	
SE (m)	SE (m) P; 0.230		) B; 0.230	SE (m) PxB; 0.460		

# Table 68: Effect of P and B application on plant copper content at pod setting stage (mg kg<sup>-1</sup>)

T<sub>1</sub>: 14.19

	$\mathbf{B}_0$	<b>B</b> <sub>1</sub>	B <sub>2</sub>	B <sub>3</sub>	Mean
$P_0$	13.88	13.75	12.50	13.19	13.33
P <sub>1</sub>	13.69	13.75	13.19	12.69	13.33
P <sub>2</sub>	13.81	13.88	13.13	13.13	13.48
P <sub>3</sub>	12.38	13.13	13.94	13.81	13.31
Mean	13.44	13.63	13.19	13.20	
CD (0.0	CD (0.05) P; NS		CD (0.05) B; NS		PxB; 0.985
SE (m)	SE (m) P; 0.172		(m) B; 0.172 SE (m) PxB; (		xB; 0.345

Table 69: Effect of P and B application on plant copper content at harvest stage (mg kg<sup>-1</sup>)

T<sub>1</sub>: 11.69

	$\mathbf{B_0}$	$\mathbf{B}_1$	$\mathbf{B}_2$	<b>B</b> <sub>3</sub>	Mean	
P <sub>0</sub>	11.63	11.81	10.50	10.94	11.22	
P <sub>1</sub>	11.69	11.75	11.19	10.69	11.33	
P <sub>2</sub>	11.81	11.89	11.01	10.85	11.39	
P <sub>3</sub>	10.43	11.12	11.90	11.91	11.34	
Mean	11.39	11.64	11.15	11.10	5	
CD (0.05)	CD (0.05) P; NS		CD (0.05) B; NS		CD (0.05) PxB; 1.006	
SE (m) P	SE (m) P; 0.176		SE (m) B; 0.176		xB; 0.352	

### 4.4.11 Boron content in plant

Boron content at flowering, pegging, pod development and harvest stages are given in the tables 70, 71, 72 73 respectively. The B content was significantly influenced by different levels of P and B application. The treatment,  $P_0B_3$  (P, 0 kg ha<sup>-1</sup> and B, 15 kg ha<sup>-1</sup>) produced highest B content at flowering (21.47 mg kg<sup>-1</sup>), pegging (17.43 mg kg<sup>-1</sup>), pod development (13.38 mg kg<sup>-1</sup>) and at harvest (11.45 mg kg<sup>-1</sup>) stages.

Table 70: Effect of P and B application on plant boron content at flowering stage (mg kg<sup>-1</sup>)

T<sub>1</sub>: 20.66

	$\mathbf{B}_0$	$\mathbf{B}_{1}$	$\mathbf{B}_2$	$\mathbf{B}_3$	Mean	
$\mathbf{P}_{0}$	19.82	20.26	20.63	21.47	20.54	
$\mathbf{P}_1$	18.60	19.07	19.41	19.47	19.13	
$\mathbf{P}_{2}$	17.41	18.19	18.35	19.22	18.29	
P <sub>3</sub>	16.01	17.10	17.78	18.38	17.31	
Mean	17.96	18.65	19.04	19.63		
CD (0.05) 1	CD (0.05) P; 0.205		CD (0.05) B; 0.205		CD (0.05) PxB; 0.410	
SE (m) P	SE (m) P; 0.072		SE (m) B; 0.072		SE (m) PxB; 0.144	

# Table 71: Effect of P and B application on plant boron content at pegging stage (mg kg<sup>-1</sup>)

T<sub>1</sub>:16.96

	$\mathbf{B}_0$	$\mathbf{B}_1$	$\mathbf{B}_2$	$\mathbf{B}_3$	Mean
$\mathbf{P_0}$	16.07	16.55	16.80	17.43	16.71
P <sub>1</sub>	15.10	15.38	15.71	15.74	15.48
P <sub>2</sub>	13.83	14.55	14.73	15.51	14.65
P <sub>3</sub>	13.24	13.27	14.22	14.77	13.88
Mean	14.56	14.94	15.36	15.86	
CD (0.05) P; 0.131		CD (0.05) B; 0.131		CD (0.05) PxB; 0.261	
SE (m) P; 0.046		SE (m) B; 0.046		SE (m) PxB; 0.091	

Table 72: Effect of P and B application on plant boron content at pod setting stage (mg kg<sup>-1</sup>)

T<sub>1</sub>:13.00

	$\mathbf{B_0}$	$\mathbf{B}_1$	$\mathbf{B}_2$	$\mathbf{B}_3$	Mean
$P_0$	12.32	12.85	12.97	13.38	12.88
P <sub>1</sub>	11.60	11.69	12.00	12.00	11.82
P <sub>2</sub>	10.25	10.91	11.10	11.78	11.01
P <sub>3</sub>	10.47	9.44	10.66	11.16	10.43
Mean	11.16	11.22	11.68	12.08	A
CD (0.05) F	CD (0.05) P; 0.156		B; 0.156	CD (0.05) PxB; 0.313	
SE (m) P;	SE (m) P; 0.055		SE (m) B; 0.055		xB; 0.109

Table 73: Effect of P and B application on plant boron content at harvest stage  $(mg\ kg^{-1})$ 

T<sub>1</sub>: 10.76

	$\mathbf{B_0}$	<b>B</b> <sub>1</sub>	B <sub>2</sub>	B <sub>3</sub>	Mean
$P_0$	10.36	10.83	10.98	11.45	10.90
$\mathbf{P}_1$	9.69	9.64	10.10	10.00	9.86
P <sub>2</sub>	8.25	8.94	9.12	10.12	9.11
P <sub>3</sub>	8.44	8.09	8.68	9.18	8.60
Mean	9.18	9.37	9.72	10.19	
CD (0.05) I	CD (0.05) P; 0.142		5) B; 0.142	CD (0.05)	PxB; 0.285
SE (m) P;	SE (m) P; 0.050		SE (m) B; 0.050		xB; 0.100

#### 4.5 NUTRIENT CONTENT IN POD

#### 4.5.1 Nitrogen content in pod

Nitrogen content in pod varied significantly due to main effect and interaction effect. N content was increased with increased dose of P application (table 74). Highest N content was observed in P<sub>3</sub>B<sub>0</sub> (P, 90 kg ha<sup>-1</sup> B, 0 kg ha<sup>-1</sup>) which had 3.89 per cent.

# 4.5.2 Phosphorus content in pod

Data (table 75) on P content in pod showed that treatments varied significantly due to main effect and interaction effect. Treatment, P<sub>3</sub>B<sub>1</sub> (P, 90 kg ha<sup>-1</sup> B, 5 kg ha<sup>-1</sup>) showed highest P content. P content in pod increased with increased dose of P application.

#### 4.5.3 Potassium content in pod

Effect of application of P and B on K content in pod is shown in the table 76. The K content varied significantly due to main effect and interaction effect. Treatment, P<sub>2</sub> B<sub>3</sub> (P, 75 kg ha<sup>-1</sup> B, 15 kg ha<sup>-1</sup>) showed highest K content.

Table 74: Effect of P and B application on nitrogen content in pod (%)

 $T_1$ : 3.47

	$\mathbf{B}_{0}$	$\mathbf{B}_1$	$\mathbf{B_2}$	В3	Mean
P <sub>0</sub>	3.60	3.41	3.38	3.35	3.43
P <sub>1</sub>	3.66	3.51	3.49	3.57	3.55
P <sub>2</sub>	3.89	3.82	3.41	3.39	3.63
P <sub>3</sub>	3.91	3.87	3.78	3.71	3.81
Mean	3.76	3.65	3.51	3.50	
CD (0.05) P; 0.009		CD (0.05) B; 0.009		CD (0.05) PxB; 0.017	
SE (m) P; 0.003		SE (m) B; 0.003		SE (m) PxB; 0.006	

Table 75: Effect of P and B application on phosphorus content in pod (%)

 $T_1: 0.40$ 

	$\mathbf{B}_0$	B <sub>1</sub>	$\mathbf{B_2}$	B <sub>3</sub>	Mean
$P_0$	0.35	0.35	0.35	0.34	0.35
$\mathbf{P_1}$	0.36	0.37	0.36	0.37	0.36
P <sub>2</sub>	0.40	0.38	0.39	0.39	0.39
P <sub>3</sub>	0.42	0.42	0.40	0.41	0.41
Mean	0.38	0.38	0.38	0.38	
CD (0.05) P; 0.002		CD (0.05) B; 0.002		CD (0.05) PxB; 0.003	
SE (m) P; 0.001		SE (m) B; 0.001		SE (m) PxB; 0.001	

Table 76: Effect of P and B application on potassium content in pod (%)

 $T_1: 0.99$ 

	$\mathbf{B}_{0}$	$\mathbf{B}_1$	$\mathbf{B_2}$	$\mathbf{B}_3$	Mean
$P_0$	0.98	0.99	0.93	0.90	0.95
$\mathbf{P_1}$	0.95	0.93	0.96	0.91	0.94
P <sub>2</sub>	0.98	0.91	0.96	1.00	0.96
P <sub>3</sub>	0.93	0.99	0.92	0.97	0.95
Mean	0.96	0.95	0.94	0.95	
CD (0.05) P; 0.001		CD (0.05) B; 0.001		CD (0.05) PxB; 0.002	
SE (m) P; 0.00			SE (m) B; 0.00		; 0.001

# 4.5.4 Calcium content in pod

Effect of application of P and B on Ca content in pod is shown in the table 77. Calcium content varied significantly due to the application of P and B fertilizers. Ca showed a synergistic interaction with P. Ca content increased with increased dose of P application. Highest Ca content was noted in P<sub>3</sub>B<sub>3</sub> (P, 90 kg ha<sup>-1</sup> B, 15 kg ha<sup>-1</sup>).

# 4.5.5 Magnesium content in pod

Effect of application of P and B on Mg content in pod is shown in the table 78. Mg content varied significantly due to the application of P and B fertilizers. Highest Mg content was observed in  $P_0B_2$  (P, 0 kg ha<sup>-1</sup> B, 10 kg ha<sup>-1</sup>).

# 4.5.6 Sulphur content in pod

Sulphur content varied significantly due to the application of P and B fertilizers. The S showed a synergistic interaction with P. S content increased with increased dose of P application (table 79). Highest S content was noted in P<sub>3</sub>B<sub>3</sub> (P, 90 kg ha<sup>-1</sup> B, 15 kg ha<sup>-1</sup>).

Table 77: Effect of P and B application on calcium content in pod (%)

 $T_1: 0.18$ 

	$\mathbf{B}_0$	$\mathbf{B}_1$	$\mathbf{B}_2$	$\mathbf{B}_3$	Mean
$P_0$	0.145	0.130	0.140	0.120	0.134
<b>P</b> <sub>1</sub>	0.150	0.120	0.150	0.150	0.143
P <sub>2</sub>	0.180	0.160	0.170	0.160	0.168
P <sub>3</sub>	0.200	0.220	0.190	0.210	0.205
Mean	0.169	0.158	0.163	0.160	
CD (0.05) P; 0.003		CD (0.05) B; 0.003		CD (0.05) PxB; 0.006	
SE (m) P; 0.001		SE (m) B; 0.001		SE (m) PxB; 0.002	

Table 78: Effect of P and B application on magnesium content in pod (%)

 $T_1: 0.25$ 

	$\mathbf{B}_{0}$	$\mathbf{B}_1$	$\mathbf{B}_2$	$\mathbf{B}_3$	Mean
$P_0$	0.22	0.25	0.28	0.25	0.25
$\mathbf{P_1}$	0.27	0.24	0.26	0.24	0.25
P <sub>2</sub>	0.26	0.25	0.27	0.30	0.27
P <sub>3</sub>	0.22	0.26	0.24	0.27	0.25
Mean	0.24	0.25	0.26	0.27	2
CD (0.05) P; 0.006		CD (0.05) B; 0.006		CD (0.05) P	xB; 0.011
SE (m) P; 0.002		SE (m) B; 0.002		SE (m) PxB; 0.004	

Table 79: Effect of P and B application on sulphur content in pod (%)

 $T_1: 0.44$ 

	$\mathbf{B}_{0}$	$\mathbf{B}_1$	$\mathbf{B}_2$	$\mathbf{B}_3$	Mean
$P_0$	0.38	0.36	0.39	0.45	0.40
$\mathbf{P}_1$	0.41	0.43	0.46	0.47	0.44
P <sub>2</sub>	0.44	0.48	0.48	0.50	0.47
P <sub>3</sub>	0.53	0.65	0.55	0.66	0.60
Mean	0.44	0.48	0.47	0.52	
CD (0.05) P; 0.002		CD (0.05) B; 0.002		CD (0.05) PxB; 0.004	
SE (m) P; 0.001		SE (m) B; 0.001		SE (m) PxB; 0.001	

# 4.5.7 Iron content in pod

Effect of application of P and B on Fe content in pod is shown in the table 80. Fe content was influenced by both main effect and interaction effect. Highest Fe content was noted in  $P_0B_2$  (P, 0 kg ha<sup>-1</sup> B, 15 kg ha<sup>-1</sup>).

# 4.5.8 Manganese content in pod

Manganese content varied significantly due to main effect and interaction effect. Mn content decreased with increased dose of P application (table 81). The highest Mn content was observed in  $P_2B_2$  (P, 75 kg ha<sup>-1</sup> B, 15 kg ha<sup>-1</sup>).

# 4.5.9 Zinc content in pod

Data (table 82) on Zn content showed that Zn was influenced by P and B application. Zn content decreased with increased dose of P application. Highest Zn was noted in P<sub>0</sub> B<sub>3</sub> (P, 0 kg ha<sup>-1</sup> B, 15 kg ha<sup>-1</sup>).

# 4.5.10 Copper content in pod

Copper content was found to be non-significant due to main effect and significant due to interaction effect (table 83).

# 4.5.11 Boron content in pod

Data (table 84) on B content showed that B had influenced by P and B application. B content decreased with increased dose of P application. Highest B was noted in P<sub>0</sub>B<sub>3</sub> (P, 0 kg ha<sup>-1</sup> B, 15 kg ha<sup>-1</sup>).

Table 80: Effect of P and B application on iron content in pod (mg kg<sup>-1</sup>)

 $T_1$ : 253.25

	$\mathbf{B_0}$	$\mathbf{B}_1$	$\mathbf{B}_2$	$\mathbf{B}_3$	Mean
$\mathbf{P_0}$	263.00	207.75	254.25	300.75	256.44
P <sub>1</sub>	295.50	198.00	212.75	228.00	233.56
P <sub>2</sub>	208.25	138.75	233.00	217.50	199.38
P <sub>3</sub>	256.75	114.00	128.25	186.75	171.44
Mean	255.88	164.63	207.06	233.25	
CD (0.05) P; 0.601		CD (0.05) B; 0.601		CD (0.05) PxB; 1.201	
SE (m) P; 0.197		SE (m) B; 0.197		SE (m) PxB; 0.395	

Table 81: Effect of P and B application on manganese content in pod (mg kg-1)

T<sub>1</sub>: 19.00

	$\mathbf{B}_0$	B <sub>1</sub>	B <sub>2</sub>	B <sub>3</sub>	Mean
$P_0$	20.25	20.75	21.50	21.75	21.06
P <sub>1</sub>	23.25	24.25	25.75	25.75	24.75
$P_2$	18.50	19.00	19.50	22.75	19.94
P <sub>3</sub>	15.25	17.50	18.00	18.00	17.19
Mean	19.31	20.38	21.19	22.06	
CD (0.05) P; 0.277		CD (0.05) B; 0.277		CD (0.05) PxB; 0.553	
SE (m) P; 0.091		SE (m) B; 0.091		SE (m) PxB; 0.182	

# Table 82: Effect of P and B application on zinc content in pod (mg kg<sup>-1</sup>)

T<sub>1</sub>: 58.50

	$\mathbf{B}_{0}$	$\mathbf{B}_1$	$\mathbf{B}_2$	<b>B</b> <sub>3</sub>	Mean
$P_0$	55.75	56.50	59.75	61.00	58.25
<b>P</b> <sub>1</sub>	50.50	50.00	54.00	53.50	52.00
P <sub>2</sub>	49.50	52.50	50.50	54.00	51.63
P <sub>3</sub>	49.00	47.50	47.75	46.50	47.69
Mean	51.19	51.63	53.00	53.75	
CD (0.05) P; 0.277		CD (0.05) B; 0.277		CD (0.05) PxB; 0.553	
SE (m) P; 0.091		SE (m) B; 0.091		SE (m) PxB; 0.182	

Table 83: Effect of P and B application on copper content in pod (mg kg<sup>-1</sup>)

T<sub>1</sub>: 14.00

	$\mathbf{B}_{0}$	$\mathbf{B}_1$	$\mathbf{B}_2$	B <sub>3</sub>	Mean
P <sub>0</sub>	13.25	11.50	9.25	11.00	11.25
P <sub>1</sub>	9.75	11.00	10.75	9.25	10.19
P <sub>2</sub>	11.75	11.75	10.50	10.25	11.06
P <sub>3</sub>	9.25	9.25	13.50	10.25	10.56
Mean	11.00	10.88	11.00	10.19	
CD (0.05) P; NS		CD (0.05) B; NS		CD (0.05) PxB; 2.842	
SE (m) P; 0.467		SE (m) B; 0.467		SE (m) PxB; 0.934	

Table 84: Effect of P and B application on boron content in pod (mg kg<sup>-1</sup>)

 $T_1$ : 17.45

	$\mathbf{B}_0$	<b>B</b> <sub>1</sub>	$\mathbf{B}_2$	<b>B</b> <sub>3</sub>	Mean
$\mathbf{P_0}$	17.13	17.54	18.38	20.27	18.33
$\mathbf{P}_1$	16.56	17.36	17.97	19.91	17.95
P <sub>2</sub>	16.68	17.15	17.65	19.14	17.65
P <sub>3</sub>	15.97	16.93	17.04	17.42	16.84
Mean	16.58	17.24	17.76	19.18	
CD (0.05) P; 0.022		CD (0.05) B; 0.022		CD (0.05) PxB; 0.045	
SE (m) P; 0.007		SE (m) B; 0.	SE (m) B; 0.007		; 0.015

#### 4.6 UPTAKE OF NUTRIENTS BY PLANT

# 4.6.1 Nitrogen uptake by plant

Uptake of N by plant varied significantly due to different levels of fertilizer application. Significantly highest N uptake by plant was noticed in treatment,  $P_3B_0$  (P, 90 kg ha<sup>-1</sup> B, 0 kg ha<sup>-1</sup>) at all stages.

Table 85: Effect of P and B application on nitrogen uptake by plant at flowering stage (kg ha<sup>-1</sup>)

1: 12.44				Я		
	$\mathbf{B}_0$	B <sub>1</sub>	B <sub>2</sub>	B <sub>3</sub>	Mean	
$P_0$	14.21	10.93	9.12	6.75	10.25	
P <sub>1</sub>	15.48	13.78	12.56	13.81	13.91	
P <sub>2</sub>	23.85	20.02	12.08	9.29	16.31	
P <sub>3</sub>	27.08	23.13	19.46	17.03	21.68	
Mean	20.16	16.96	13.30	11.72		
CD (0.05) P; 0.225		CD (0.05) B; 0.225		CD (0.05) PxB; 0.450		
SE (m) F	SE (m) P; 0.079		SE (m) B; 0.079		SE (m) PxB; 0.157	

Table 86: Effect of P and B application on nitrogen uptake by plant at pegging stage (kg ha<sup>-1</sup>)

T <sub>1</sub> : 35.49					
8.	$\mathbf{B_0}$	, B <sub>1</sub>	B <sub>2</sub>	B <sub>3</sub>	Mean
$P_0$	36.36	33.08	30.22	27.87	31.88
$\mathbf{P}_{1}$	37.35	35.16	35.06	36.22	35.94
P <sub>2</sub>	46.76	42.45	34.00	30.20	38.35
P <sub>3</sub>	51.62	47.59	42.14	39.40	45.19
Mean	43.02	39.57	35.35	33.42	
CD (0.05)	P; 0.340	CD (0.05	5) B; 0.340	CD (0.05)	PxB; 0.680
SE (m) I	SE (m) P; 0.119		SE (m) B; 0.119		xB; 0.238

Table 87: Effect of P and B application on nitrogen uptake by plant at pod setting stage (kg ha<sup>-1</sup>)

	$\mathbf{B}_0$	$\mathbf{B}_1$	$\mathbf{B_2}$	$\mathbf{B}_3$	Mean
P <sub>0</sub>	49.12	41.40	38.77	39.34	42.16
$\mathbf{P}_1$	49.38	43.76	43.45	43.40	44.99
P <sub>2</sub>	64.52	58.63	42.49	38.84	51.12
P <sub>3</sub>	78.99	68.57	54.30	51.43	63.32
Mean	60.50	53.09	44.75	43.25	
CD (0.05) P; 1.315		CD (0.05) B; 1.315		CD (0.05) PxB; 2.63	
SE (m) P; 0.460		SE (m) B; 0.460		SE (m) PxB; 0.920	

Table 88: Effect of P and B application on nitrogen uptake by plant at harvest stage (kg ha<sup>-1</sup>)

: 49.15					
	$\mathbf{B}_{0}$	$\mathbf{B}_1$	$\mathbf{B_2}$	$\mathbf{B}_3$	Mean
$\mathbf{P_0}$	52.73	47.32	44.84	44.27	47.29
P <sub>1</sub>	54.76	48.74	47.40	48.49	49.85
P <sub>2</sub>	67.48	60.79	46.24	43.95	54.61
P <sub>3</sub>	79.43	68.70	59.21	55.51	65.71
Mean	63.60	56.38	49.42	48.05	
CD (0.05) P; 1.013		CD (0.05) B; 1.013		CD (0.05)PxB; 2.025	
SE (m) P	; 0.354	SE (m)	B; 0.354	SE (m) PxB; 0.709	

# 4.6.2 Phosphorus uptake by plant

Uptake of P by plant varied significantly due to different levels of fertilizer application. Significantly highest P uptake by plant was noticed in treatment,  $P_3B_0$  (P, 90 kg ha<sup>-1</sup> B, 0 kg ha<sup>-1</sup>) at all stages.

Table 89: Effect of P and B application on phosphorus uptake by plant at flowering stage (kg ha<sup>-1</sup>)

	$\mathbf{B_0}$	$\mathbf{B}_1$	$\mathbf{B_2}$	$\mathbf{B_3}$	Mean
$P_0$	1.71	1.26	1.10	0.74	1.20
$\mathbf{P}_{1}$	2.06	1.88	1.71	1.90	1.89
P <sub>2</sub>	3.20	2.68	1.78	1.46	2.28
P <sub>3</sub>	3.74	3.58	2.80	2.59	3.17
Mean	2.68	2.35	1.85	1.67	
CD (0.05) P; 0.076		CD (0.05) B;0.076		CD (0.05);PxB 0.15	
SE (m) P; 0.027		SE (m) B; 0.027		SE (m) PxB; 0.053	

Table 90: Effect of P and B application on phosphorus uptake by plant at pegging stage (kg ha<sup>-1</sup>)

	$\mathbf{B_0}$	$\mathbf{B_1}$	$\mathbf{B_2}$	$\mathbf{B_3}$	Mean
$\mathbf{P_0}$	4.24	3.70	3.38	2.74	3.51
$P_1$	4.74	4.61	4.60	5.06	4.75
P <sub>2</sub>	6.64	5.94	4.93	4.45	5.49
P <sub>3</sub>	7.33	7.22	6.28	6.15	6.74
Mean	5.74	5.37	4.80	4.60	
CD (0.05)	P;0.111	CD (0.0	5) B;0.111	CD (0.05)	;PxB 0.22
SE (m) P; 0.039		SE (m)	B; 0.039	SE (m) P	xB; 0.078

Table 91: Effect of P and B application on phosphorus uptake by plant at pod setting stage (kg ha<sup>-1</sup>)

	$\mathbf{B}_{0}$	B <sub>1</sub>	$\mathbf{B}_{2}$	B <sub>3</sub>	Mean	
P <sub>0</sub>	5.54	4.49	3.92	3.35	4.32	
P <sub>1</sub>	5.98	5.52	5.38	6.18	5.76	
P <sub>2</sub>	9.84	8.70	6.06	5.26	7.47	
P <sub>3</sub>	11.54	10.23	8.51	8.25	9.63	
Mean	8.23	7.23	5.96	5.76		
CD (0.05) P; 0.273		CD (0.05) B; 0.273		CD (0.05) PxB; 0.54		
SE (m) P; 0.096		SE (m)	SE (m) B; 0.096		SE (m) PxB; 0.191	

Table 92: Effect of P and B application on phosphorus uptake by plant at harvest stage (kg ha<sup>-1</sup>)

	$\mathbf{B_0}$	B <sub>1</sub>	$\mathbf{B_2}$	B <sub>3</sub>	Mean
P <sub>0</sub>	5.93	4.95	4.19	3.45	4.63
P <sub>1</sub>	6.35	5.82	5.81	6.11	6.02
P <sub>2</sub>	9.91	9.05	6.53	5.62	7.78
P <sub>3</sub>	11.93	10.43	9.48	9.52	10.34
Mean	8.53	7.56	6.50	6.17	
CD (0.05) P;0.263		CD (0.05) B;0.263		CD (0.05);PxB 0.525	
SE (m) P; 0.092		CD (0.05	CD (0.05) B;0.092		xB; 0.184

# 4.6.3 Potassium uptake by plant

Potassium uptake by plant was found to be significant in all treatments due to main effect and interaction effect. The treatment,  $P_3B_1$  (P, 90 kg ha<sup>-1</sup> B, 5 kg ha<sup>-1</sup>)

showed highest K uptake at flowering stage (39.08 kg ha<sup>-1</sup>). At pegging, pod setting and harvest stages, P<sub>2</sub>B<sub>0</sub> (P, 90 kg ha<sup>-1</sup> B, 5 kg ha<sup>-1</sup>) showed highest uptake.

Table 93: Effect of P and B application on potassium uptake by plant at flowering stage (kg ha<sup>-1</sup>)

1: 12.43						
	$\mathbf{B}_0$	$\mathbf{B}_1$	$\mathbf{B}_{2}$	B <sub>3</sub>	Mean	
P <sub>0</sub>	13.17	10.65	8.99	5.66	9.62	
$\mathbf{P}_{1}$	13.92	11.68	11.31	11.59	12.12	
P <sub>2</sub>	20.49	15.35	11.20	10.23	14.32	
P <sub>3</sub>	20.28	20.85	15.35	15.39	17.97	
Mean	16.97	14.63	11.71	10.72	,	
CD (0.05) P; 0.381		CD (0.05) B; 0.381		CD (0.05) PxB; 0.762		
SE (m) P; 0.133		SE (m)	B; 0.133	SE (m) PxB; 0.267		

Table 94: Effect of P and B application on potassium uptake by plant at pegging stage (kg ha<sup>-1</sup>)

Γ <sub>1</sub> : 33.26					
	$\mathbf{B_0}$	$\mathbf{B}_1$	$\mathbf{B_2}$	$\mathbf{B}_3$	Mean
$\mathbf{P_0}$	31.26	31.37	27.73	24.50	28.71
P <sub>1</sub>	35.12	32.18	32.69	34.12	33.53
P <sub>2</sub>	42.33	35.44	33.33	32.05	35.78
P <sub>3</sub>	38.49	38.31	35.27	36.27	37.08
Mean	36.80	34.32	32.26	31.73	41
CD (0.05) P;0.546		CD (0.05)	B;0.546	CD (0.05) PxB; 1.091	
SE (m) P;			0.191	SE (m) PxB; 0.382	

Table 95: Effect of P and B application on potassium uptake by plant at pod setting stage (kg ha<sup>-1</sup>)

1: 41.06				ē .	
	$\mathbf{B_0}$	$\mathbf{B}_1$	$\mathbf{B}_2$	<b>B</b> <sub>3</sub>	Mean
$P_0$	38.00	37.95	31.61	36.58	36.04
P <sub>1</sub>	49.78	43.08	41.70	46.78	45.33
P <sub>2</sub>	62.41	54.21	44.59	39.22	50.11
P <sub>3</sub>	56.42	47.18	48.70	48.46	50.19
Mean	51.65	45.60	41.65	42.76	
CD (0.05) I	P; 1.435	CD (0.05)	B; 1.435	CD (0.05)	PxB; 2.869
SE (m) P; 0.502		SE (m) B; 0.502		SE (m) PxB; 1.004	

Table 96: Effect of P and B application on potassium uptake by plant at harvest stage (kg ha<sup>-1</sup>)

45.40					
	B <sub>0</sub>	$\mathbf{B}_1$	B <sub>2</sub>	B <sub>3</sub>	Mean
P <sub>0</sub>	43.44	46.24	38.02	42.80	42.62
P <sub>1</sub>	57.99	50.49	49.33	54.04	52.96
P <sub>2</sub>	69.87	60.48	50.99	45.99	56.83
P <sub>3</sub>	61.15	51.49	56.53	56.68	56.46
Mean	58.11	52.17	48.72	49.88	
CD (0.05) P; 1.336		CD (0.05) B; 1.336		CD (0.05) PxB; 2.672	
SE (m) P; 0.468		SE (m) B; 0.468		SE (m) PxB; 0.935	

# 4.6.4 Calcium uptake by plant

Calcium uptake by plant varied significantly due to main effect and interaction effect. Highest Ca uptake was noted in P<sub>3</sub>B<sub>1</sub> (P, 90 kg ha<sup>-1</sup> B, 5 kg ha<sup>-1</sup>)

during all stages. Ca uptake increased as plants grews and there was a slight decrease at harvest stage. Main effect of B was non -significant at harvest stage.

Table 97: Effect of P and B application on calcium uptake by plant at flowering stage (kg ha<sup>-1</sup>)

	$\mathbf{B}_{0}$	$\mathbf{B_1}$	$B_1$ $B_2$	B <sub>3</sub>	Mean
$\mathbf{P_0}$	11.75	9.55	9.09	6.79	9.29
$\mathbf{P}_{1}$	10.56	9.52	9.28	10.77	10.03
P <sub>2</sub>	15.16	14.41	9.46	7.95	11.74
P <sub>3</sub>	16.14	16.78	15.15	13.87	15.49
Mean	13.40	12.56	10.75	9.84	
CD (0.05)	P; 0.288	CD (0.05	) B; 0.288	CD (0.05)	PxB; 0.576
SE (m) I	SE (m) P; 0.101		SE (m) B; 0.101		xB; 0.202

Table 98: Effect of P and B application on calcium uptake by plant at pegging stage (kg ha<sup>-1</sup>)

T <sub>1</sub> : 17.04					
	$\mathbf{B_0}$	$\mathbf{B}_1$	$\mathbf{B_2}$	$\mathbf{B}_3$	Mean
$P_0$	27.58	26.06	26.57	24.84	26.26
P <sub>1</sub>	20.66	21.24	21.94	25.17	22.25
P <sub>2</sub>	25.74	26.63	22.44	21.78	24.15
P <sub>3</sub>	26.87	29.95	29.28	28.44	28.64
Mean	25.21	25.97	25.06	25.06	
CD (0.05)	P; 0.437	CD (0.05)	B; 0.437	CD (0.05)	PxB; 0.874
SE (m) I	P; 0.153	SE (m) B; 0.153		SE (m) PxB; 0.306	

Table 99: Effect of P and B application on calcium uptake by plant at pod setting stage (kg ha<sup>-1</sup>)

	$\mathbf{B}_0$	$\mathbf{B}_1$	B <sub>2</sub>	<b>B</b> <sub>3</sub>	Mean
P <sub>0</sub>	32.73	28.20	28.56	29.52	29.75
P <sub>1</sub>	18.98	21.15	20.83	25.03	21.50
P <sub>2</sub>	28.06	29.35	21.23	21.47	25.03
P <sub>3</sub>	33.13	35.06	31.81	31.03	32.76
Mean	28.22	28.44	25.61	26.76	
CD (0.05)	P; 0.713	CD (0.05)	B; 0.713	CD (0.05)	PxB; 1.42
SE (m) P; 0.250		SE (m) B; 0.250		SE (m) PxB; 0.499	

Table 100: Effect of P and B application on calcium uptake by plant at harvest stage (kg ha<sup>-1</sup>)

	$\mathbf{B_0}$	$\mathbf{B_1}$	$\mathbf{B_2}$	$\mathbf{B}_3$	Mean
P <sub>0</sub>	29.85	27.53	29.56	29.66	29.15
P <sub>1</sub>	16.38	17.96	19.90	22.35	19.15
P <sub>2</sub>	25.37	27.45	20.28	21.04	23.53
P <sub>3</sub>	29.31	31.78	29.91	29.93	30.23
Mean	25.23	26.18	24.91	25.74	
CD (0.05)	P; 1.147	CD (0.05	) B; NS	CD (0.05)	PxB; 2.293
SE (m) P; 0.401		SE (m) B; 0.401		SE (m) PxB; 0.802	

### 4.6.5 Magnesium uptake by groundnut plant

Application of P and B fertilizers significantly affected Mg uptake by plant. The treatment, P<sub>3</sub>B<sub>1</sub> (P, 90 kg ha<sup>-1</sup> B, 5 kg ha<sup>-1</sup>) had highest Mg uptake during flowering and pegging stages. During pod setting and harvest, highest Mg uptake was noted in P<sub>3</sub>B<sub>0</sub> (P, 90 kg ha<sup>-1</sup> B, 0 kg ha<sup>-1</sup>). The main effect and interaction effect was found to be significant at all stages except at harvest where, main effect of B was non-significant.

Table 101: Effect of P and B application on Magnesium uptake by plant at flowering stage (kg ha<sup>-1</sup>)

Γ <sub>1</sub> : 16.78					=
	$\mathbf{B_0}$	$\mathbf{B}_1$	$\mathbf{B}_2$	B <sub>3</sub>	Mean
$\mathbf{P_0}$	17.78	15.00	12.40	11.41	14.15
$\mathbf{P}_1$	17.42	16.11	17.08	16.71	16.83
P <sub>2</sub>	27.39	23.05	16.15	12.41	19.75
P <sub>3</sub>	25.78	27.51	20.20	20.06	23.39
Mean	22.09	20.41	16.46	15.15	
CD (0.05) P; 0.533		CD (0.05) B; 0.533		CD (0.05) PxB; 1.065	
SE (m) P;	0.175	SE (m)	В; 0.175	SE (m) PxB	; 0.350

Table 102: Effect of P and B application on Magnesium uptake by plant at pegging stage (kg ha<sup>-1</sup>)

T <sub>1</sub> : 33.41	Γ <sub>1</sub> : 33.41						
	$\mathbf{B_0}$	<b>B</b> <sub>1</sub>	$\mathbf{B}_2$	B <sub>3</sub>	Mean		
$P_0$	35.71	34.44	30.66	33.36	33.54		
$P_1$	31.53	33.05	34.89	33.24	33.18		
P <sub>2</sub>	40.13	37.46	33.57	30.64	35.45		
P <sub>3</sub>	39.66	42.65	35.19	34.30	37.95		
Mean	36.76	36.90	33.58	32.88			
CD (0.05) I	CD (0.05) P; 0.581		CD (0.05) B; 0.581		B; 1.163		
SE (m) P;	SE (m) P; 0.191		B; 0.191	SE (m) PxB; 0.382			

Table 103: Effect of P and B application on Magnesium uptake by plant at pod setting stage (kg ha<sup>-1</sup>)

T <sub>1</sub> : 23.22					
×	$\mathbf{B}_0$	B <sub>1</sub>	B <sub>2</sub>	В3	Mean
$P_0$	31.74	25.95	22.65	21.74	25.52
P <sub>1</sub>	24.84	27.37	24.18	26.23	25.65
P <sub>2</sub>	29.92	29.33	22.88	23.43	26.39
P <sub>3</sub>	41.55	37.45	30.16	24.97	33.53
Mean	32.01	30.02	24.96	24.09	
CD (0.05) P	P; 0.849	CD (0.05)	B; 0.849	CD (0.05) Pxl	B; 1.699
SE (m) P;	0.279	SE (m) B	3; 0.279	SE (m) PxB	; 0.559

Table 104: Effect of P and B application on Magnesium uptake by plant at harvest stage (kg ha<sup>-1</sup>)

:					
	$\mathbf{B}_{0}$	$\mathbf{B}_1$	$\mathbf{B}_2$	B <sub>3</sub>	Mean
$P_0$	36.07	29.98	25.31	25.02	29.09
$\mathbf{P}_1$	26.37	29.91	26.62	28.13	27.75
$P_2$	33.24	32.88	25.17	27.69	29.74
$P_3$	42.79	38.94	33.11	28.93	35.94
Mean	34.61	32.92	27.55	27.44	
CD (0.05) P	CD (0.05) P; 0.913		3; 0.913	CD (0.05) Pxl	B; 1.826
SE (m) P;	0.300	SE (m) B;	0.300	SE (m) PxB; 0.600	

# 4.6.6 Sulphur uptake by groundnut plant

Application of P and B fertilizers significantly affected S uptake by plant. The treatment, P<sub>3</sub>B<sub>3</sub> (P, 90 kg ha<sup>-1</sup> B, 15 kg ha<sup>-1</sup>) had highest S uptake during flowering and pegging stages. During pod setting and harvest, highest S uptake was noted in P<sub>3</sub>B<sub>2</sub> (P, 90 kg ha<sup>-1</sup> B, 10 kg ha<sup>-1</sup>). The main effect and interaction effect was found to be significant at all stages except at harvest where, main effect of B was non-significant.

Table 105: Effect of P and B application on sulphur uptake by plant at flowering stage (kg ha<sup>-1</sup>)

$\Gamma_1$ : 1.56					
	$\mathbf{B_0}$	$\mathbf{B}_1$	$\mathbf{B}_{2}$	$\mathbf{B}_3$	Mean
$\mathbf{P}_0$	2.95	2.18	1.78	1.30	2.05
P <sub>1</sub>	2.87	2.90	2.76	3.19	2.93
P <sub>2</sub>	5.08	4.52	3.02	2.68	3.82
P <sub>3</sub>	5.90	5.83	5.26	6.27	5.81
Mean	4.20	3.85	3.20	3.36	
CD (0.05) I	P; 0.089	CD (0.05	) B; 0.089	CD (0.05) P	xB; 0.178
SE (m) P;	SE (m) P; 0.031		B; 0.031	SE (m) PxB; 0.062	

Table 106: Effect of P and B application on sulphur uptake by plant at pegging stage (kg ha<sup>-1</sup>)

	$\mathbf{B_0}$	$\mathbf{B}_1$	$\mathbf{B}_2$	$\mathbf{B}_3$	Mean
$\mathbf{P_0}$	6.25	5.44	4.94	4.48	5.27
$\mathbf{P_1}$	5.97	6.43	6.46	7.43	6.57
$\mathbf{P}_2$	8.62	8.86	7.36	7.38	8.05
P <sub>3</sub>	9.89	10.18	10.80	11.65	10.63
Mean	7.68	7.73	7.39	7.73	
CD (0.05)	P; 0.169	CD (0.05	) B; 0.169	CD (0.05) P	xB; 0.338
SE (m) P; 0.059		SE (m)	SE (m) B; 0.059		B; 0.118

Table 107: Effect of P and B application on sulphur uptake by plant at pod setting stage (kg ha<sup>-1</sup>)

CD (0.05) P; 0.548 SE (m) P; 0.192		CD (0.05) B; 0.548  SE (m) B; 0.192		CD (0.05) PxB; 1.096 SE (m) PxB; 0.192	
Mean	8.58	8.06	7.83	7.53	DD- 1 004
P <sub>3</sub>	12.43	11.52	12.97	10.49	11.85
	122 24 0				
P <sub>2</sub>	9.47	9.29	7.37	7.36	8.37
$\mathbf{P}_{1}$	6.26	6.39	6.06	7.42	6.53
$P_0$	6.18	5.03	4.91	4.85	5.24
	$\mathbf{B_0}$	$\mathbf{B}_1$	$\mathbf{B_2}$	$\mathbf{B}_3$	Mean

Table 108: Effect of P and B application on sulphur uptake by plant at harvest stage (kg ha<sup>-1</sup>)

CD (0.05) P; 0.494 SE (m) P; 0.173		,	CD (0.05) B; NS SE (m) B; 0.173		B; 0.346
CD (0.05) P	· 0 494	CD (0.0	5) R. NS	CD (0.05) P2	R · 0 988
Mean	9.05	8.72	8.57	8.49	
P <sub>3</sub>	12.83	11.72	14.36	11.96	12.72
$P_2$	10.02	10.64	7.87	8.31	9.21
$\mathbf{P}_1$	6.72	6.89	6.66	8.31	7.14
$P_0$	6.63	5.64	5.37	5.37	5.75
	$\mathbf{B_0}$	$\mathbf{B}_1$	$\mathbf{B_2}$	$\mathbf{B}_3$	Mear

# 4.6.7 Iron uptake by plant

Uptake of Fe by plant at different stages is shown in the tables 109, 110, 111, 112. Uptake of Fe varied significantly due to interaction effect and main effect. P<sub>3</sub>B<sub>2</sub> (P, 90 kg ha<sup>-1</sup> B, 10 kg ha<sup>-1</sup>) showed highest Fe uptake at flowering and pegging stages. P<sub>3</sub> B<sub>0</sub> (P, 90 kg ha<sup>-1</sup> B, 0 kg ha<sup>-1</sup>) showed highest uptake at pod setting stage and P<sub>2</sub> B<sub>0</sub> (P, 75 kg ha<sup>-1</sup> B, 0 kg ha<sup>-1</sup>) showed highest uptake at harvest stage.

Table 109: Effect of P and B application on iron uptake by plant at flowering stage (g ha<sup>-1</sup>)

71:338.87			ii.			
	$\mathbf{B}_0$	B <sub>1</sub>	B <sub>2</sub>	B <sub>3</sub>	Mean	
$\mathbf{P_0}$	271.31	187.33	179.00	170.90	202.13	
$P_1$	318.88	259.86	222.49	235.33	259.14	
P <sub>2</sub>	374.79	323.07	173.68	193.82	266.34	
P <sub>3</sub>	371.67	320.84	383.17	344.54	355.05	
Mean	334.16	272.77	239.58	236.15		
CD (0.05)	CD (0.05) P; 10.217		CD (0.05) B; 10.217		CD (0.05)PxB; 20.435	
SE (m)	P; 3.575	SE (m)	B; 3.575	SE (m) PxB; 7.151		

Table 110: Effect of P and B application on iron uptake by plant at pegging stage (g ha<sup>-1</sup>)

	$\mathbf{B}_0$	$\mathbf{B}_{1}$	$\mathbf{B}_2$	B <sub>3</sub>	Mean
$P_0$	736.72	524.95	635.42	610.84	626.98
P <sub>1</sub>	711.39	676.53	557.28	665.79	652.75
P <sub>2</sub>	766.11	708.75	534.00	566.69	643.89
P <sub>3</sub>	732.27	715.86	794.72	648.77	722.90
Mean	736.62	656.52	630.35	623.02	*1
CD (0.05) P; 18.605		CD (0.05) B; 18.605		CD (0.05) PxB; 37.210	
SE (m) P; 6.510		SE (m)	B; 6.510	SE (m) PxB; 13.021	

Table 111: Effect of P and B application on iron uptake by plant at pod setting stage (g ha<sup>-1</sup>)

Mean	1048.74 P; 44.940	901.59 CD (0.05) I	794.49	703.46 CD (0.05) P	
P <sub>3</sub>	1168.24	1128.65	966.25	602.18	966.33
P <sub>2</sub>	1115.54	1020.52	739.44	629.08	876.14
P <sub>1</sub>	840.87	865.24	591.74	879.08	794.23
$P_0$	1070.32	591.94	880.53	703.51	811.58
	$\mathbf{B_0}$	$\mathbf{B}_1$	$\mathbf{B}_2$	$\mathbf{B}_3$	Mean

Table 112: Effect of P and B application on iron uptake by plant at harvest stage (g ha<sup>-1</sup>)

T <sub>1</sub> :1242.22			T			
	$\mathbf{B_0}$	$\mathbf{B}_1$	$\mathbf{B_2}$	$\mathbf{B_3}$	Mean	
$P_0$	1236.32	721.86	1070.52	810.20	959.72	
P <sub>1</sub>	976.86	1012.02	710.15	1061.90	940.23	
P <sub>2</sub>	1298.36	1200.84	897.97	741.67	1034.71	
P <sub>3</sub>	1257.90	1286.78	1121.69	716.51	1095.72	
Mean	1192.36	1055.37	950.08	832.57		
CD (0.05)	CD (0.05) P; 36.848		CD (0.05) B; 36.848		CD (0.05) PxB; 73.695	
SE (m) P	SE (m) P; 12.894		SE (m) B; 12.894		SE (m) PxB; 25.788	

# 4.6.8 Manganese uptake by groundnut plants

Manganese uptake was significantly influenced by main effect and interaction effect. The treatment,  $P_3B_0$  (P, 90 kg ha<sup>-1</sup> B, 0 kg ha<sup>-1</sup>) showed highest Mn uptake at flowering stage. At pegging highest uptake was observed in  $P_1B_3$  (P, 60 kg ha<sup>-1</sup> B, 15 kg ha<sup>-1</sup>) and at pod setting and harvest, the highest Mn uptake was in treatment  $P_1B_0$  (P, 60 kg ha<sup>-1</sup> B, 0 kg ha<sup>-1</sup>)

Table 113: Effect of P and B application on manganese uptake by plant at flowering stage (g ha<sup>-1</sup>)

	$\mathbf{B_0}$	$\mathbf{B_1}$	$\mathbf{B}_2$	$\mathbf{B}_3$	Mean
$\mathbf{P_0}$	42.14	36.45	33.49	25.48	34.39
P <sub>1</sub>	53.98	49.13	46.49	51.38	50.24
$P_2$	59.47	54.89	36.22	34.59	46.29
P <sub>3</sub>	60.10	56.18	50.29	45.88	53.11
Mean	53.92	49.16	41.62	39.33	
CD (0.05) P; 1.775		CD (0.05) B; 1.775		CD (0.05) PxB; 3.551	
SE (m) P:	SE (m) P; 0.621		3; 0.621	SE (m) PxB; 1.242	

Table 114: Effect of P and B application on manganese uptake by plant at pegging stage (g ha<sup>-1</sup>)

T <sub>1</sub> :121.51					
	$\mathbf{B}_0$	$\mathbf{B}_1$	$\mathbf{B}_2$	B <sub>3</sub>	Mean
$P_0$	114.44	111.24	109.76	104.16	109.90
$\mathbf{P}_1$	134.00	131.49	130.63	139.40	133.88
$P_2$	117.85	118.64	107.66	108.74	113.22
P <sub>3</sub>	112.51	109.85	106.90	103.58	108.21
Mean	119.70	117.80	113.73	113.97	
CD (0.05)	CD (0.05) P; 2.735		B; 2.735	CD (0.05) PxB; 5.471	
SE (m) P	; 0.957	SE (m) I	3; 0.957	SE (m) PxB; 1.914	

Table 115: Effect of P and B application on manganese uptake by plant at pod setting stage (g ha<sup>-1</sup>)

T <sub>1</sub> :193.22					
	$\mathbf{B}_0$	$\mathbf{B}_1$	B <sub>2</sub>	B <sub>3</sub>	Mean
$P_0$	166.01	140.50	138.92	145.21	147.66
P <sub>1</sub>	183.59	173.87	163.16	174.70	173.83
P <sub>2</sub>	164.84	167.84	143.66	133.91	152.56
P <sub>3</sub>	168.16	148.28	134.71	130.94	145.52
Mean	170.65	157.62	145.11	146.19	
CD (0.05)	CD (0.05) P; 6.009		B; 6.009	CD (0.05) PxB; 12.017	
SE (m) P	; 2.103	SE (m) I	3; 2.103	SE (m) PxB; 4.205	

Table 116: Effect of P and B application on manganese uptake by plant at harvest stage (g ha<sup>-1</sup>)

	SE (m) P; 2.194		SE (m) B; 2.194		SE (m) PxB; 4.387	
CD (0.05) P;	6.269	CD (0.05)	B; 6.269	CD (0.05)	PxB; 12.538	
Mean	192.23	180.43	170.41	169.16		
$P_3$	178.41	159.41	153.04	150.97	160.46	
$P_2$	183.30	189.29	164.32	155.71	173.15	
$\mathbf{P}_{1}$	217.92	201.35	192.55	203.32	203.78	
$\mathbf{P_0}$	189.31	171.66	171.75	166.64	174.84	
	$\mathbf{B_0}$	$\mathbf{B}_1$	$\mathbf{B}_2$	$\mathbf{B}_3$	Mean	

# 4.6.9 Zinc uptake by groundnut plants

Zinc uptake varied significantly due to application of P and B fertilizers. The treatment  $P_3B_0$  (P, 90 kg ha<sup>-1</sup> B, 0 kg ha<sup>-1</sup>) showed highest zinc uptake at flowering stage. Whereas at pegging, pod setting and harvest stage,  $P_0B_0$  (P, 0 kg ha<sup>-1</sup> B, 0 kg ha<sup>-1</sup>) showed highest Zn uptake.

Table 117: Effect of P and B application on zinc uptake by plant at flowering stage (g ha<sup>-1</sup>)

: 28.05	n	D	D	- D	Maan	
	$\mathbf{B_0}$	$\mathbf{B}_1$	$\mathbf{B_2}$	$\mathbf{B_3}$	Mean	
$\mathbf{P_0}$	29.70	23.99	24.13	18.20	24.01	
$\mathbf{P}_1$	28.83	25.29	24.07	26.14	26.08	
P <sub>2</sub>	39.10	35.48	23.15	20.41	29.53	
P <sub>3</sub>	41.12	36.58	31.88	28.10	34.42	
Mean	34.69	30.33	25.81	23.21		
CD (0.05)	CD (0.05) P; 0.786		CD (0.05) B; 0.786		CD (0.05) PxB; 1.573	
SE (m) F	2: 0.275	SE (m)	B; 0.275	SE (m) PxB; 0.550		

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Table 118: Effect of P and B application on zinc uptake by plant at pegging stage (g ha<sup>-1</sup>)

72.78				8	
	$\mathbf{B_0}$	B <sub>1</sub>	B <sub>2</sub>	B <sub>3</sub>	Mean
$\mathbf{P_0}$	87.91	79.07	83.04	74.12	81.03
$\mathbf{P}_1$	76.07	73.95	70.35	74.50	73.72
$\mathbf{P}_2$	81.19	78.69	65.65	61.31	71.71
P <sub>3</sub>	76.96	76.98	71.67	67.25	73.22
Mean	80.53	77.17	72.68	69.29	
CD (0.05) P; 1.227		CD (0.05) B; 1.227		CD (0.05) PxB; 2.454	
SE (m) I	P; 0.429	SE (m) B; 0.429		SE (m) PxB; 0.859	

Table 119: Effect of P and B application on zinc uptake by plant at pod setting stage (g ha<sup>-1</sup>)

1: 85.83					
	$\mathbf{B}_{0}$	<b>B</b> <sub>1</sub>	$\mathbf{B}_2$	<b>B</b> <sub>3</sub>	Mean
$\mathbf{P_0}$	132.18	108.98	111.27	102.96	113.85
$\mathbf{P}_1$	111.77	105.75	92.05	98.93	102.12
P <sub>2</sub>	118.51	114.94	82.76	70.96	96.79
P <sub>3</sub>	114.89	113.86	96.66	91.44	104.21
Mean	119.34	110.88	95.68	91.07	e .
CD (0.05) P; 3.804		CD (0.05) B; 3.804		CD (0.05) PxB; 7.609	
SE (m) I	P; 1.331	SE (m)	B; 1.331	SE (m) PxB; 2.662	

Table 120: Effect of P and B application on zinc uptake by plant at harvest stage (g ha<sup>-1</sup>)

	D	$\mathbf{B}_1$	$\mathbf{B}_2$	<b>B</b> <sub>3</sub>	Mean
	$\mathbf{B_0}$	$\mathbf{D}_1$	<b>D</b> 2	В3	Mean
$\mathbf{P_0}$	144.32	126.82	130.08	116.92	129.53
$\mathbf{P}_{1}$	127.26	117.97	104.56	109.42	114.80
P <sub>2</sub>	127.24	121.71	92.36	79.91	105.31
P <sub>3</sub>	119.01	118.62	104.46	101.02	110.78
Mean	129.46	121.28	107.86	101.82	
CD (0.05)	P; 2.499	CD (0.05)	B; 2.499	CD (0.05)	PxB; 4.998
SE (m) I	P; 0.874	SE (m) B; 0.874		SE (m) PxB; 1.749	

# 4.6.10 Copper uptake by groundnut plant

Uptake of Cu was significant at all the stages. The treatment,  $P_2B_0$  (P, 75 kg ha<sup>-1</sup> B, 0 kg ha<sup>-1</sup>) showed highest uptake at flowering and pegging and pod setting stages. Highest uptake during harvest stage was noted in  $P_3B_0$  (P, 90 kg ha<sup>-1</sup> B, 0 kg ha<sup>-1</sup>). Uptake was decreased from pod setting to harvest stage.

Table 121: Effect of P and B application on copper uptake by plant at flowering stage (g ha<sup>-1</sup>)

	$\mathbf{B_0}$	$\mathbf{B_1}$	B <sub>2</sub>	$\mathbf{B}_3$	Mean
$\mathbf{P_0}$	8.86	7.04	5.76	4.53	6.55
P <sub>1</sub>	9.52	8.57	7.50	7.91	8.37
P <sub>2</sub>	13.49	11.78	7.30	6.08	9.66
P <sub>3</sub>	12.82	12.53	11.81	10.47	11.91
Mean	11.17	9.98	8.09	7.25	
CD (0.05) P; 0.412		CD (0.05) B; 0.412		CD (0.05)	PxB; 0.823
SE (m) P; 0.144		SE (m) I	SE (m) B; 0.144		xB; 0.288

Table 122: Effect of P and B application on copper uptake by plant at pegging stage (g ha<sup>-1</sup>)

	${f B_0}$	$\mathbf{B}_1$	$\mathbf{B}_2$	$\mathbf{B}_3$	Mean
P <sub>0</sub>	29.64	27.49	26.37	23.78	26.82
$\mathbf{P}_1$	29.33	28.26	26.05	30.44	28.52
P <sub>2</sub>	36.39	33.04	28.66	25.05	30.78
P <sub>3</sub>	31.29	31.12	31.04	31.27	31.18
Mean	31.66	29.98	28.03	27.63	
CD (0.05) P; 1.272		CD (0.05) B;1.272		CD (0.05) F	PxB; 2.544
SE (m) P; 0.445		SE (m) B; 0.445		SE (m) PxB; 0.890	

Table 123: Effect of P and B application on copper uptake by plant at pod setting stage (g ha<sup>-1</sup>)

	D	D	D	D	Maan
	$\mathbf{B_0}$	$\mathbf{B}_1$	$\mathbf{B_2}$	$\mathbf{B}_3$	Mean
$\mathbf{P_0}$	52.36	44.07	44.97	42.14	45.88
$\mathbf{P}_1$	49.71	45.90	40.21	51.91	46.93
P <sub>2</sub>	68.26	60.25	48.70	40.71	54.48
P <sub>3</sub>	61.63	54.12	48.78	52.41	54.23
Mean	57.99	51.08	45.66	46.79	
CD (0.05) P; 3.425		CD (0.05) B; 3.425		CD (0.05) P	xB; 6.850
SE (m) P; 1.198		SE (m) B; 1.198		SE (m) PxB; 2.397	

Table 124: Effect of P and B application on copper uptake by plant at harvest stage (g ha<sup>-1</sup>)

:44.82					
	$\mathbf{B}_0$	<b>B</b> <sub>1</sub>	$\mathbf{B}_2$	<b>B</b> <sub>3</sub>	Mean
P <sub>0</sub>	38.37	36.34	32.02	31.95	34.67
$\mathbf{P}_{1}$	41.60	38.49	36.00	36.67	38.19
P <sub>2</sub>	48.25	47.01	34.85	33.23	40.83
P <sub>3</sub>	49.93	45.28	43.95	42.88	45.51
Mean	44.53	41.78	36.70	36.18	
CD (0.05) P; 2.304		CD (0.05) B; 2.304		CD (0.05) P	xB; 4.609
SE (m) P; 0.806		SE (m)	B; 0.806	SE (m) PxB; 1.613	

### 4.6.11 Boron uptake by groundnut plant

Uptake of B was significantly influenced by main effect and interaction effect. Treatment,  $P_2B_0$  (P, 75 kg ha<sup>-1</sup> B, 0 kg ha<sup>-1</sup>) showed highest uptake at flowering and pegging stages. Treatment,  $P_3B_0$  (P, 90 kg ha<sup>-1</sup> B, 0 kg ha<sup>-1</sup>) showed highest uptake at pod setting and harvest stages.

Table 125: Effect of P and B application on boron uptake by plant at flowering stage (g ha<sup>-1</sup>)

Γ <sub>1</sub> : 11.91						
	$\mathbf{B_0}$	$\mathbf{B_1}$	$\mathbf{B_2}$	$\mathbf{B}_3$	Mean	
$\mathbf{P_0}$	12.65	10.37	9.50	7.37	9.97	
$\mathbf{P}_1$	12.93	11.88	11.06	12.16	12.01	
P <sub>2</sub>	16.99	15.44	10.21	8.89	12.88	
P <sub>3</sub>	16.56	16.33	15.07	13.93	15.47	
Mean	14.78	13.50	11.46	10.59		
CD (0.05)	CD (0.05) P; 0.276		CD (0.05) B; 0.276		PxB; 0.552	
SE (m) P	SE (m) P; 0.096		SE (m) B; 0.096		SE (m) PxB; 0.193	

Table 126: Effect of P and B application on boron uptake by plant at pegging stage (g ha<sup>-1</sup>)

T <sub>1</sub> : 30.01						
	$\mathbf{B_0}$	<b>B</b> <sub>1</sub>	$\mathbf{B}_2$	B <sub>3</sub>	Mean	
$\mathbf{P_0}$	29.54	28.34	27.91	26.89	28.17	
$\mathbf{P_1}$	28.62	28.04	27.80	29.50	28.49	
P <sub>2</sub>	30.10	29.81	25.87	25.78	27.89	
P <sub>3</sub>	29.59	28.61	29.12	28.92	29.06	
Mean	29.46	28.70	27.67	27.77		
CD (0.05) P; 0.360		CD (0.05) B; 0.360		CD (0.05)	PxB; 0.721	
SE (m) P	SE (m) P; 0.237		SE (m) B; 0.237		SE (m) PxB; 0.474	

Table 127: Effect of P and B application on boron uptake by plant at pod setting stage (g ha<sup>-1</sup>)

CD (0.05) P; 0.938 SE (m) P; 0.328		SE (m) B; 0.32	CD (0.05) PxB; 1.875 SE (m) PxB; 0.656		
Mean	36.34	32.53	29.88	31.00	D-D. 1 975
P <sub>3</sub>	41.31	32.45	31.80	32.28	34.46
P <sub>2</sub>	35.65	35.82	27.73	28.19	31.85
$\mathbf{P_1}$	33.39	31.10	29.68	31.54	31.43
$\mathbf{P_0}$	35.00	30.77	30.31	31.99	32.01
	$\mathbf{B}_{0}$	$\mathbf{B}_1$	$\mathbf{B}_2$	$\mathbf{B}_3$	Mean

Table 128: Effect of P and B application on boron uptake by plant at harvest stage (g ha<sup>-1</sup>)

T <sub>1</sub> : 33.59									
	$\mathbf{B_0}$	<b>B</b> <sub>1</sub>	<b>B</b> <sub>2</sub>	B <sub>3</sub>	Mean				
$\mathbf{P_0}$	35.65	33.48	32.86	33.61	33.90				
$P_1$	34.66	31.62	31.29	32.60	32.54				
P <sub>2</sub>	34.07	35.16	28.02	30.30	31.89				
P <sub>3</sub>	38.35	31.84	31.96	32.98	33.78				
Mean	35.68	33.02	31.03	32.37					
CD (0.05) P; 0.764		CD (0.05) B; 0.764		CD (0.05) PxB; 1.527					
SE (m) P; 0.267		SE (m) B; 0.267		SE (m) P	SE (m) PxB; 0.534				

#### 4.7 NUTRIENT UPTAKE BY POD

#### 4.7.1 Nitrogen uptake by pod

Data (table 129) on N uptake by pod showed that, N uptake by pod varied significantly due to application of different levels of P and B. The treatment,  $P_3B_1$  (P, 90 kg ha<sup>-1</sup> B, 5 kg ha<sup>-1</sup>) showed highest N uptake.

### 4.7.2 Phosphorus uptake by pod

Uptake of P by plant at different stages was shown in the table 130. P uptake by pod varied significantly due to application of different levels of P and B. The treatment, P<sub>3</sub>B<sub>1</sub> (P, 90 kg ha<sup>-1</sup> B, 5 kg ha<sup>-1</sup>) showed highest P uptake.

Table 129: Effect of P and B application on nitrogen uptake by pod (kg ha<sup>-1</sup>)

T<sub>1</sub>: 67.26

	$\mathbf{B_0}$	$\mathbf{B}_1$	$\mathbf{B}_2$	B <sub>3</sub>	Mean
$\mathbf{P_0}$	73.58	67.73	64.29	61.05	66.66
$\mathbf{P}_{1}$	78.54	72.31	71.86	77.79	75.13
P <sub>2</sub>	99.57	104.72	99.88	78.07	95.56
P <sub>3</sub>	123.48	126.93	123.14	111.67	121.30
Mean	93.79	92.92	89.79	82.15	-
CD (0.05) P; 0.627		CD (0.05) B; 0.627		CD (0.05) Px	B; 1.255
SE (m) P;	SE (m) P; 0.206 SE (m) B; 0.206		06	SE (m) PxB;	0.412

# Table 130: Effect of P and B application on phosphorus uptake by pod (kg ha<sup>-1</sup>)

`T<sub>1</sub>: 7.83

	$\mathbf{B}_0$	$\mathbf{B}_{1}$	$\mathbf{B}_2$	B <sub>3</sub>	Mean
$\mathbf{P_0}$	7.19	6.95	6.61	6.22	6.74
P <sub>1</sub>	7.65	7.69	7.37	8.05	7.69
P <sub>2</sub>	10.14	10.38	11.53	8.96	10.25
P <sub>3</sub>	13.14	13.91	13.18	12.28	13.13
Mean	9.53	9.73	9.67	8.88	
CD (0.05) P; 0.066		CD (0.05) B; 0.066		CD (0.05) Px	B; 0.131
SE (m) P;	0.022	SE (m) B; 0.0	22	SE (m) PxB;	0.044

# 4.7.3 Potassium uptake by pod

Potassium uptake varied significantly due to main effect and interaction effect (table 131). The treatment, P<sub>3</sub>B<sub>1</sub> (P, 90 kg ha<sup>-1</sup> B, 5 kg ha<sup>-1</sup>) showed highest K uptake.

# 4.7.4 Calcium uptake by pod

Calcium uptake varied significantly due to main effect and interaction effect (table 132). The treatment,  $P_3B_1$  (P, 90 kg ha<sup>-1</sup> B, 5 kg ha<sup>-1</sup>) showed highest Ca uptake.

# 4.7.5 Magnesium uptake by pod

Uptake of Mg by plant varied significantly due to different levels of fertilizer application (table 133). Significantly highest Mg uptake by plant was noticed in treatment,  $P_3B_0$  (P, 90 kg ha<sup>-1</sup> B, 0 kg ha<sup>-1</sup>).

### 4.7.6 Sulphur uptake by pod

Uptake of S by pod varied significantly due to different levels of fertilizer application (table 134). Significantly highest S uptake by pod was noticed in treatment, P<sub>3</sub>B<sub>1</sub> (P, 90 kg ha<sup>-1</sup> B, 5 kg ha<sup>-1</sup>).

Table 131: Effect of P and B application on potassium uptake by pod (kg ha<sup>-1</sup>)
T<sub>1</sub>:19.24

	B <sub>0</sub>	$\mathbf{B}_1$	B <sub>2</sub>	B <sub>3</sub>	Mean
$P_0$	20.01	19.67	17.72	16.46	18.46
P <sub>1</sub>	20.32	19.12	19.81	19.91	19.79
P <sub>2</sub>	25.13	24.90	28.24	22.94	25.30
P <sub>3</sub>	29.27	32.49	29.93	29.16	30.21
Mean	23.68	24.04	23.93	22.12	
CD (0.05)	P; 0.129	CD (0.05) B; 0.129		CD (0.05) Px	B; 0.257
SE (m) P	m) P; 0.042 SE (m) B; 0.0		042	SE (m) PxB;	0.085

Table 132: Effect of P and B application on Ca uptake by pod (kg ha<sup>-1</sup>)

 $T_1$ : 3.25

	$\mathbf{B_0}$	$\mathbf{B_1}$	$\mathbf{B}_2$	B <sub>3</sub>	Mean
$P_0$	2.96	2.59	2.67	2.19	2.60
$\mathbf{P_1}$	3.22	2.48	3.09	3.27	3.02
P <sub>2</sub>	4.61	4.39	4.98	3.69	4.42
P <sub>3</sub>	6.32	7.23	6.20	6.32	6.52
Mean	4.28	4.17	4.23	3.87	
CD (0.05) P; 0.048		CD (0.05) B; 0.048		CD (0.05) PxB; 0.097	
SE (m) P; 0.016		SE (m) B; 0.016		SE (m) PxB; 0.032	

Table 133: Effect of P and B application on Mg uptake by pod (kg ha<sup>-1</sup>)

 $T_1$ : 5.51

	$\mathbf{B_0}$	B <sub>1</sub>	$\mathbf{B}_2$	<b>B</b> <sub>3</sub>	Mean
P <sub>0</sub>	4.50	4.97	5.33	4.56	4.84
P <sub>1</sub>	5.79	4.95	5.36	5.24	5.34
P <sub>2</sub>	6.66	6.86	7.91	6.91	7.09
P <sub>3</sub>	6.80	8.54	7.83	8.13	7.82
Mean	5.94	6.33	6.61	6.21	и
CD (0.05) P; 0.184		CD (0.05) B; 0.184		CD (0.05) PxB; 0.367	
SE (m) P; 0.060		SE (m) B; 0.060		SE (m) PxB; 0.121	

Table 134: Effect of P and B application on S uptake by pod (kg ha<sup>-1</sup>)

T<sub>1</sub>: 7.94

	$\mathbf{B_0}$	$B_1$	$\mathbf{B}_2$	<b>B</b> <sub>3</sub>	Mean
P <sub>0</sub>	7.81	7.09	7.47	8.22	7.65
P <sub>1</sub>	8.87	8.83	9.55	10.34	9.40
P <sub>2</sub>	11.20	13.09	14.17	11.43	12.47
P <sub>3</sub>	16.89	21.29	18.04	19.94	19.04
Mean	11.19	12.58	12.31	12.48	
CD (0.05) P; 0.086		CD (0.05) B; 0.086		CD (0.05) PxB; 0.171	
SE (m) P; 0.030		SE (m) B; 0.030		SE (m) PxB; 0.059	

### 4.7.7 Iron uptake by pod

Iron uptake by pod was found to be significant. Compared to other micro nutrients Fe was present in highest amount. The treatment, P<sub>2</sub>B<sub>2</sub> (P, 75 kg ha<sup>-1</sup> and B, 10 kg ha<sup>-1</sup>) showed highest Fe uptake.

# 4.7.8 Manganese uptake by pod

Uptake of Mn by pod varied significantly due to different levels of fertilizer application. Significantly highest Mn uptake by pod was noticed in treatment,  $P_3B_1$  (P, 90 kg ha<sup>-1</sup> B, 5 kg ha<sup>-1</sup>).

### 4.7.9 Zinc uptake by pod

Zinc uptake varied significantly due to main effect and interaction effect. The treatment,  $P_3B_1$  (P, 90 kg ha<sup>-1</sup> B, 5 kg ha<sup>-1</sup>) showed highest Zn uptake.

Table 135: Effect of P and B application on Fe uptake by pod (g ha<sup>-1</sup>)

T<sub>1</sub>: 358.15

	$\mathbf{B_0}$	B <sub>1</sub>	B <sub>2</sub>	$\mathbf{B}_3$	Mean
$P_0$	537.57	413.22	484.35	548.87	496.00
P <sub>1</sub>	634.14	408.47	438.69	497.50	494.70
P <sub>2</sub>	533.75	380.87	682.45	500.90	524.49
P <sub>3</sub>	811.85	374.38	418.35	562.11	541.67
Mean	629.33	394.23	505.96	527.35	
CD (0.05) P; 2.779		CD (0.05) B; 2.779		CD (0.05) PxB; 5.558	
SE (m) P; 0.914		SE (m) B; 0.	914	SE (m) PxB; 1.827	

# Table 136: Effect of P and B application on Mn uptake by pod (g ha<sup>-1</sup>)

T<sub>1</sub>: 45.13

	$\mathbf{B_0}$	<b>B</b> <sub>1</sub>	$\mathbf{B}_2$	B <sub>3</sub>	Mean
$\mathbf{P_0}$	41.39	41.27	40.96	39.69	40.83
$\mathbf{P}_1$	49.89	50.03	53.10	56.19	52.30
P <sub>2</sub>	47.41	52.16	57.12	52.39	52.27
P <sub>3</sub>	48.22	57.47	58.72	54.18	54.65
Mean	46.73	50.23	52.47	50.61	
CD (0.05) P; 0.662		CD (0.05) B; 0.662		CD (0.05) PxB; 1.323	
SE (m) P; 0.217		SE (m) B; 0.217		SE (m) PxB; 0.435	

Table 137: Effect of P and B application on Zn uptake by pod (g ha<sup>-1</sup>)

T<sub>1</sub>: 123.76

	$\mathbf{B}_{0}$	B <sub>1</sub>	$\mathbf{B}_2$	B <sub>3</sub>	Mean
P <sub>0</sub>	113.95	112.38	113.83	111.33	112.87
P <sub>1</sub>	108.37	103.15	111.35	116.74	109.90
P <sub>2</sub>	126.87	144.11	147.92	124.36	135.81
P <sub>3</sub>	154.94	155.99	155.76	139.97	151.66
Mean	126.03	128.91	132.21	123.10	
CD (0.05) P; 0.973		CD (0.05) B; 0.973		CD (0.05) PxB; 1.946	
SE (m) P; 0.320		SE (m) B; 0.320	SE (m) PxB; 0.640		.640

# 4.7.10 Copper uptake by pod

Copper uptake varied significantly due to main effect of P and interaction effect and it was non- significant due to factor B. The treatment, P<sub>3</sub>B<sub>2</sub> (P, 90 kg ha<sup>-1</sup> B, 10 kg ha<sup>-1</sup>) showed highest Cu uptake.

# 4.7.11 Boron uptake by pod

Uptake of B by pod at different stages is shown in the table 139. B uptake by groundnut pod varied significantly due to different treatments applied. Significantly highest uptake was in treatment having high P.

Table 138: Effect of application of P and B on Cu uptake by pod (g ha<sup>-1</sup>)

T<sub>1</sub>: 28.76

	$\mathbf{B}_0$	B <sub>1</sub>	$\mathbf{B_2}$	B <sub>3</sub>	Mean	
P <sub>0</sub>	27.08	22.87	17.62	20.08	21.91	
P <sub>1</sub>	20.91	22.69	22.17	20.18	21.49	
P <sub>2</sub>	30.12	32.25	30.76	23.61	29.18	
P <sub>3</sub>	29.25	30.38	44.04	30.94	33.65	
Mean	26.84	27.05	28.64	23.70		
CD (0.05) P; 3.827		CD (0.05) B; NS		CD (0.05) PxB; 7.655		
SE (m) P;	1.258	SE (m) B; 1.	258	SE (m) PxB; 2.516		

# Table 139: Effect of P and B application on B uptake by pod (g ha<sup>-1</sup>)

T<sub>1</sub>: 42.24

	$\mathbf{B}_0$	<b>B</b> <sub>1</sub>	$\mathbf{B}_2$	B <sub>3</sub>	Mean
$\mathbf{P_0}$	35.01	34.89	35.01	36.99	35.48
P <sub>1</sub>	35.54	35.80	37.05	43.44	37.96
P <sub>2</sub>	42.74	47.06	51.68	44.08	46.39
P <sub>3</sub>	50.50	55.58	55.58	52.42	53.52
Mean	40.95	43.33	44.83	44.23	
CD (0.05) P; 0.219		CD (0.05) B; 0.219		CD (0.05) PxB; 0.438	
SE (m) P; 0.072		SE (m) B; 0.072	72 SE (m) PxB; 0.144		44

# 4.8 PROTEIN CONTENT IN GROUNDNUT POD

Data on protein content in groundnut kernel is shown in the table 140. Protein content was significantly influenced by main effect and interaction effect of P and B. Highest protein content was noted in  $P_3B_0$  (P, 90 kg ha<sup>-1</sup> B, 0 kg ha<sup>-1</sup>).

Table 140: Effect of P and B application on protein content (g)

T<sub>1</sub>:22.56

	$\mathbf{B}_0$	B <sub>1</sub>	$\mathbf{B}_2$	B <sub>3</sub>	Mean
$\mathbf{P_0}$	22.50	21.28	21.09	20.91	21.45
P <sub>1</sub>	22.88	21.91	21.78	22.28	22.21
P <sub>2</sub>	24.28	23.84	21.31	21.19	22.66
P <sub>3</sub>	24.41	24.16	23.59	23.19	23.84
Mean	23.52	22.80	21.95	21.89	
CD (0.05) P; 0.054		CD (0.05) B; 0.054		CD (0.05) PxB; 0.108	
SE (m) P; 0.018		SE (m) B; 0.0	8 SE (m) PxB; 0.036		0.036

# **Discussion**

#### 5. DISCUSSION

The results of study entitled "P and B interactions in black cotton soils of Kerala with respect to groundnut (*Arachis hypogaea* L.)" presented in chapter 4 are discussed here with supporting studies conducted elsewhere and based on available literature.

#### 5.1 GROWTH AND YIELD PARAMETERS OF GROUNDNUT

### 5.1.1 Growth parameters

# 5.1.1.1 Plant height and number of leaves

Plant height and number of leaves at flowering, pegging, pod development and harvest stages were significantly affected by application of different levels of P and B fertilizers. At flowering, pegging and pod development stages, treatment T<sub>14</sub> (P<sub>3</sub>B<sub>0</sub>: P, 90 kg ha<sup>-1</sup> and B, 0 kg ha<sup>-1</sup>) gave highest plant height. At harvest stage, highest plant height was noticed in T<sub>8</sub> (P<sub>1</sub>B<sub>2</sub>: P, 60 kg ha<sup>-1</sup> and B, 0 kg ha<sup>-1</sup>). Application of P increased plant height and number of leaves. Application of soluble P increased the availability of soluble phosphate and enhanced root development and there by enhanced nutrient uptake and resulted in improved plant growth. A higher level of P is important for root growth, root formation and N fixation (Lakshmamma and Raj, 1997). Similar results were reported by Punnoose (1968), Sabale and Khuspe (1986), Patel *et al.* (1981) and Juan *et al.* (1986). Higher content of P in soil increases the N fixation which in turn enhances the plant growth (Balasubramanian and Palaniappan, 1991). This was in conformity with the findings of Punnoose (1968), Singh and Ahuja (1985) and Rayar (1986). Basha and Rao (1980) observed reduction in plant height and shoot length of groundnut plant under P deficiency.

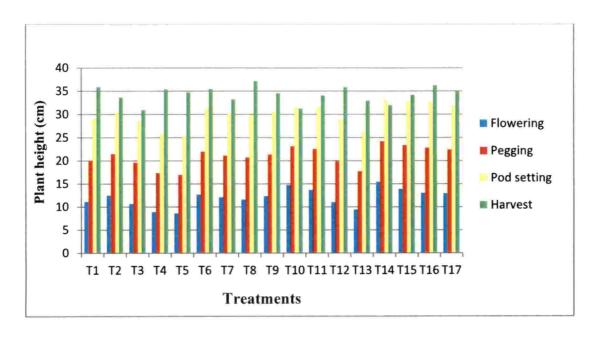


Fig 1: Effect of application of P and B on plant height

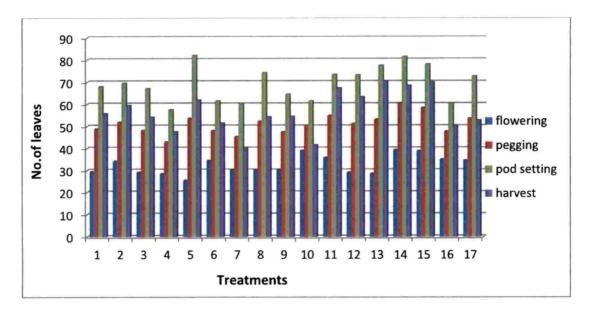


Fig 2: Effect of application of P and B on number of leaves

### 5.1.2 Yield parameters

# 5.1.2.1 Number of pods per plant and yield

There was significant variation in number of pods due to application of various levels of fertilizers. Treatment, T<sub>16</sub> (P<sub>3</sub> B<sub>2</sub>: P, 90 kg ha<sup>-1</sup> and B, 10 kg ha<sup>-1</sup>) produced maximum number of pods. Number of pods was highest in treatment receiving high dose of P (90 kg ha<sup>-1</sup>). Application of P at 90 kg ha<sup>-1</sup> and B at 5 kg ha<sup>-1</sup> resulted in highest yield (3.66 t ha<sup>-1</sup>) whereas yield from present university recommendation was 3.45 t ha<sup>-1</sup>. There was an increase of 0.21 t ha<sup>-1</sup> in yield on application of P at 90 kg ha<sup>-1</sup> and B at 5 kg ha<sup>-1</sup> than present recommendation. Banerjee *et al.* (1967) and Puri (1969) also reported the same. They reported that application of P increased number of pods and yield. Application of 90 kg P<sub>2</sub>O<sub>5</sub> ha<sup>-1</sup> recorded highest yield of dry pod. Samtana *et al.* (1994) also reported improvement in yield attributes by addition of P. This was due to formation and proliferation of new roots and improvement in their functional activity. P stimulates setting of pods, decreases the number of unfilled pods (pops) and hastens the maturity of the crop.

Application of P increases rate of N fixation and improves the N availability. Increase in N also has a role in improved yield. Plants having high N availability produce more number of flowers and pegs (Saradhi *et al.*, 1990) and results in an improved yield. N has significant role in the number of pods per plant and number of filled pods per plant. Jadhar and Narkhende (1980), Reddy *et al.* (1984) and Patel *et al.* (1994) also observed the same.

#### 5.2 EFFECT OF DIFFERENT LEVELS OF P AND B ON SOIL PROPERTIES

#### 5.2.1 Soil pH

Application of P and B had no significant influence on soil pH. However soil pH decreased slightly compared to initial status. It might be due to the application of acid forming fertilizers, urea and SSP.

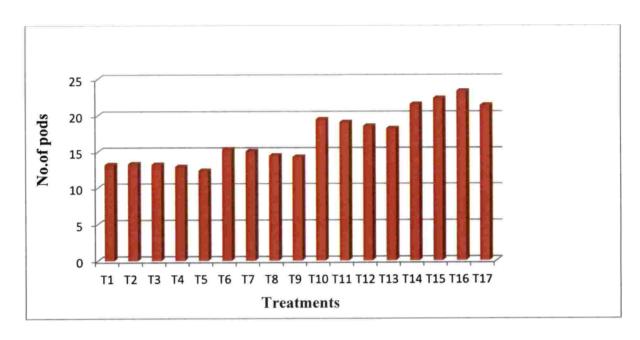


Fig 3: Effect of application of P and B on number of pods per plant

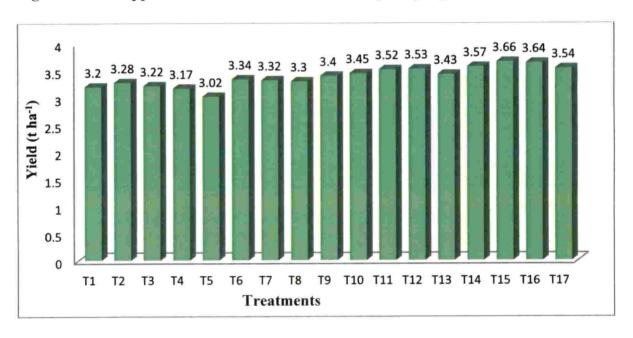


Fig 4: Effect of application of P and B on yield of groundnut

#### 5.2.2 EC

There was a slight increase in EC due to application of soluble fertilizers.

## 5.2.3 Organic carbon

There was no significant influence in the application of P and B fertilizers on organic carbon.

#### 5.2.4 Available nitrogen

Application of P and B significantly affected available N status of soil. The requirement of P in nodulating legumes is higher compared to non-nodulating crops as it plays a significant role in nodule formation and fixation of atmospheric N (Brady and Weil, 2002). Available N status of soil increased with increased dose of application of P. P has a positive interaction with N and plant development (Sumner and Farina, 1986). This might be due to the increased N fixation. Balasubramanian and Palaniappan (1991) reported that higher the concentration of P, the higher will be the amount of N fixed. N has decreased with increased dose of B.

#### 5.2.5 Available phosphorus

Application of P and B significantly affected the available P status of soil. Compared to initial available P, available P was increased in all treatments due to application of phosphatic fertilizers. Available P status of control plot also increased than initial P due to application of manures. Lowest P was recorded in treatment without P application and highest P was noted in treatment with high dose of P application. Available P status of soil was affected by B application also.

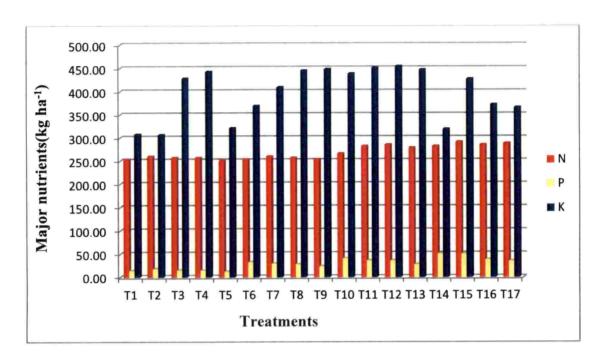


Fig 5: Effect of application of P and B on major nutrients in soil

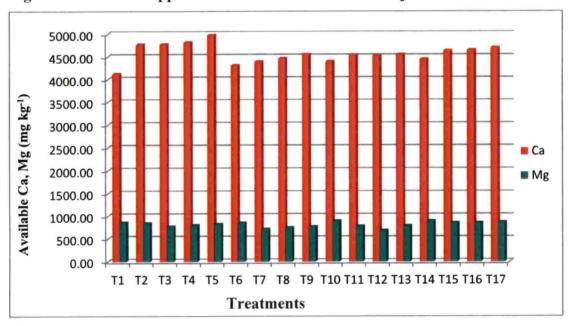


Fig 6: Effect of application of P and B on available Ca and Mg in soil

Available P decreased with increased dose of B application. An antagonism was observed between P and B. Comparable results were reported by Bingham *et al.* (1958) in citrus and May and Pritts (1993) in strawberry.

#### 5.2.6 Available potassium

Compared to initial status, available K in all treatments was increased due to application of MOP. The increase in K content after harvest was due to immediate release of K to available pool of K from FYM and high dose of K. Available K status of soil was affected by Mg content rather than P. Treatment which has highest Mg showed lowest K. Generally high pH soil has high Mg which decreases the K status of soil. This result was in conformity with Brady and Weil (2002) and Daliparthy *et al.*, (2008). In calcareous soil, K availability is limited by high pH and high concentrations of Ca and Mg (Brady and Weil, 2002). This might be due to increased competition between Ca, Mg and K for exchangeable sites. Application of P and B reduced available K content in soil mainly because of enhanced plant uptake.

#### 5.2.7 Available calcium

Experimental field had high Ca content since it is a calcareous soil with high soil pH. Available Ca status in all treatments increased due to application of SSP. There was an increase in available Ca with increased dose of P application. This might be due to the presence of Ca in SSP. There was an increase in Ca with increased dose of B application due to positive interaction between Ca and B.

#### 5.2.8 Available magnesium

The initial Mg content of soil was high. Mg had a positive interaction with P. Mg plays a positive role in P availability and recovery of P by crops in calcareous soil. Mg can alter the formation of Ca phosphate precipitates and increase P availability (Marion and Babbcock, 1977). Kuo and Milkkelsen (1979) showed that Mg may interfere with P adsorption on CaCO<sub>3</sub> surface by altering some of adsorption

sites on CaCO<sub>3</sub> surface, due to lower affinity of phosphate to Mg<sup>2+</sup> in comparison with Ca<sup>2+</sup> which causes decreased P adsorption by CaCO<sub>3</sub>. Comparable results were reported by Al-Lami (1999) and Dowood (1982) also.

#### 5.2.9 Available sulphur

There was a noticeable increase in available S from  $T_1$  to  $T_{17}$  due to increased dose of SSP application. Synergistic interaction between S and P was noted.

#### 5.2.10 Available iron

Application of P and B significantly influenced Fe status of soil. Treatment having high B showed highest available Fe. Application of B enhanced available Fe content in soil whereas heavy application of P reduced availability of Fe. This might be due to the formation of Fe-phosphate which reduced the Fe availability. This was in conformity with result of Ayed, (1970). Loneragan *et al.* (1979) reported that application of large amount of P raises P: Fe ratio and leads to deficiency of these nutrients.

#### 5.2.11 Available manganese

Available Mn was high in treatment with low P. Application of P reduced available Mn status of soil. It may be due to the formation of insoluble Mn-phosphate. High available P induces deficiency of Mn in maize (Adriano and Murphy, 1970) and in potato (Adriano *et al.*, 1971).

#### 5.2.12 Available zinc

Application of P and B significantly affected Zn content. Highest Zn was noticed in treatment,  $T_2$  ( $P_0B_0 - P$ , 0 kg ha<sup>-1</sup> and B, 0 kg ha<sup>-1</sup>) and lowest was in treatment,  $T_{15}$  ( $P_3B_1$ - P, 90 kg ha<sup>-1</sup> and B, 5 kg ha<sup>-1</sup>). Available Zn decreased with increase in P. P forms chemical bond with Zn, at high levels of P and P binds large quantity of Zn resulting in P induced Zn deficiency that leads to reduced shoot

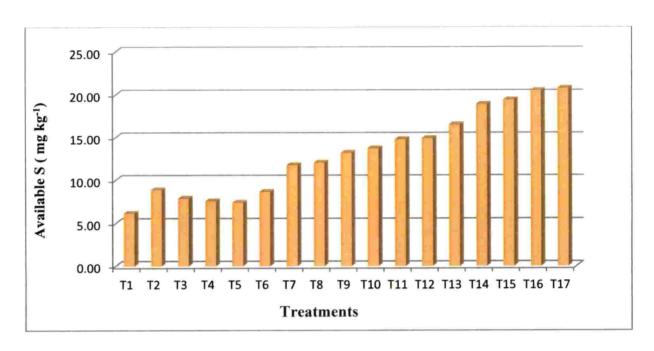


Fig 7: Effect of application of P and B on available S in soil

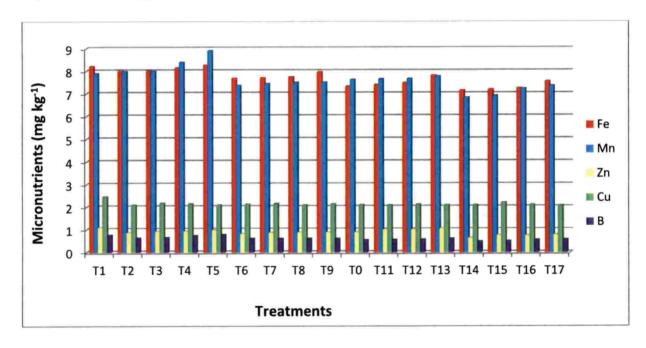


Fig 8: Effect of application of P and B on micronutrients in soil

growth. The findings of Adriano and Murphy (1970) in maize and Adriano *et al.* (1971) in potato were in conformity with the results obtained. They found that high P induced Zn deficiency.

### 5.2.13 Available copper

Effect of available Cu was non-significant for main effect and interaction effect.

#### 5.2.14 Available boron

A significant positive decrease in available B was noticed in treatment having high P. Highest B was observed in treatment, T<sub>5</sub> (P<sub>0</sub>B<sub>3</sub>: P, 0 kg ha<sup>-1</sup> and B, 15 kg ha<sup>-1</sup>) and lowest was in treatment, T<sub>14</sub> (P<sub>3</sub>B<sub>0</sub>: P, 90 kg ha<sup>-1</sup> and B, 0 kg ha<sup>-1</sup>). There was antagonistic interaction between B and P. This was in conformity with results of Bingham *et al.* (1958) in citrus and May and Pritts (1993) in strawberry.

#### 5.3 EFFECT OF APPLICATION P AND B ON PLANT NUTRIENT CONTENT

Nutrient content in groundnut was significantly different for all nutrients. Since the fertilizer used for the supply of P is SSP, besides P, S and Ca were also added which improved the nutrient status of soil as well as nutrient content in plants. Significantly highest N was noted in  $T_{14}$  ( $P_3$   $B_0$ ). Significantly highest P was noted in  $T_{15}$  ( $P_3$   $B_1$ ). Highest K was noted in  $T_{13}$  ( $P_2$   $B_3$ ).  $T_{17}$  ( $P_3$   $B_3$ ) showed highest S content.  $T_5$  ( $P_0$   $B_3$ ) showed highest Ca, Fe, Zn and B content. Significantly highest Mn was noticed in  $T_9$  ( $P_1$   $B_3$ ). Significantly highest N, P, and S content were observed in treatments with high dose of P (90 kg ha<sup>-1</sup>). Highest K content was noted in treatment having medium P (75 kg ha<sup>-1</sup>).

Low P status of soil decreased N, P, K and Ca contents in 30 day old groundnut plants (Basha and Rao, 1980). Higher content of N, P, S and K in these treatments helped root growth and enhanced uptake of nutrients and increased dry

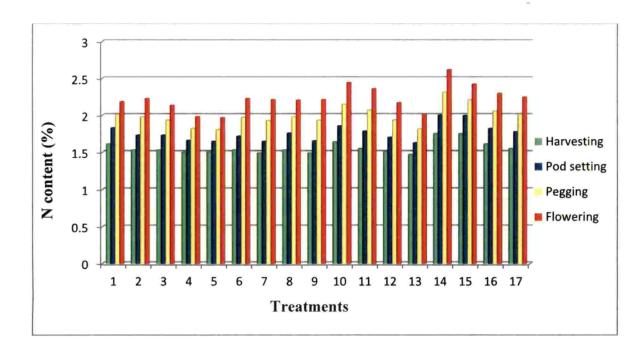


Fig 9: Effect of application of P and B on nitrogen content in plant at different stages

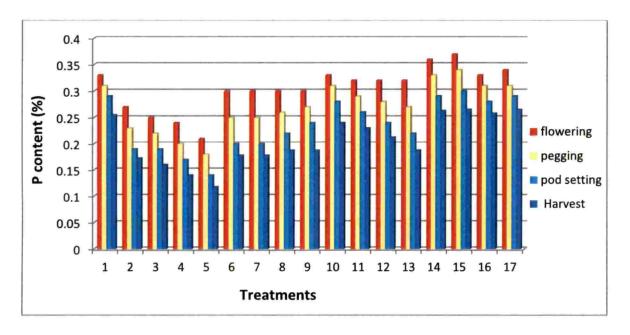


Fig 10: Effect of application of P and B on phosphorus content in plant at different stages

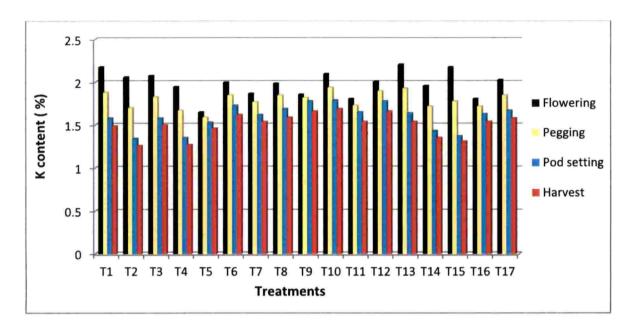


Fig 11: Effect of application of P and B on potassium content in plant at different stages

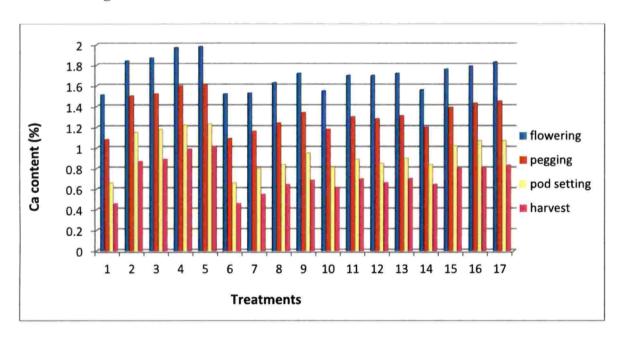


Fig 12: Effect of application of P and B on calcium content in plant at different stages

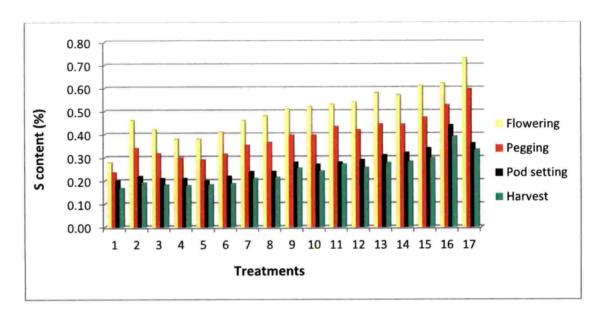


Fig 13: Effect of application of P and B on sulphur content in plant at different stages

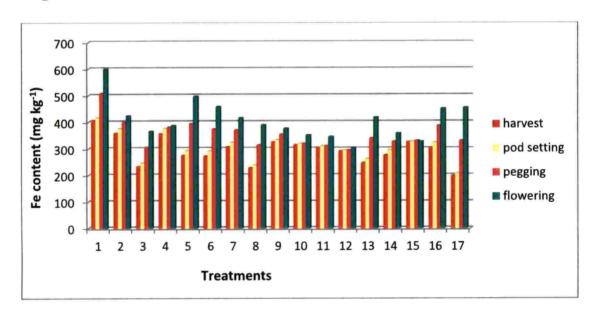


Fig 14: Effect of application of P and B on iron content in plant at different stage

matter production and yield. The findings of Nakagawa *et al.* (1981) and Patel *et al.* (1981) and also supported the result.

Nutrient content was decreased from flowering to harvest stage. Highest nutrient content was noticed in flowering stage and gradually decreased towards harvest stage. The findings of Chahal *et al.* (1983) also were in conformity with this result. Reddy and Murthy (1985) found that plant N content decreased as crop grew older.

The uptake of nutrients is associated with the metabolic activities of the plant and depends on the concentration and distribution of nutrient ions in the plant system (Manasa *et al.*, 2015). Uptake of nutrients at various stages was significantly affected by application of P and B. The uptake of N was increased from flowering to harvest with increased dry matter production. Yakadri and Satyanarayana (1992) reported that there is a close relationship between nutrient uptake and dry matter production in groundnut. Higher uptake of N was noticed in treatment having high P and this might be due to the fixation of N in presence of P which leads to better root formation. Increased dry matter production was due to better root formation which in turn enhanced higher absorption of N from soil and improved metabolic activity inside the plant (Laxminarayana, 2004). Uptake of N was more in pod compared to plant.

Uptake of P by groundnut was highest in P<sub>3</sub>B<sub>0</sub> (P 90 kg ha<sup>-1</sup> B 0 kg ha<sup>-1</sup>) at all stages. Highest uptake was noted in treatment having highest P content. The availability of nutrients increased with increase in the dose of P fertilizer. P uptake by the plant was less than the N uptake. Uptake of P increased with advancement in growth due to high dry matter production. Uptake of P was maximum at pod setting stage. Loganathan *et al.* (1996) reported that groundnut plant absorbed 10 per cent of P at vegetative stage, 40-50 per cent at reproductive stage and remaining P at reproductive to harvest of the crop. Uptake of P by pod was higher than uptake by plant.

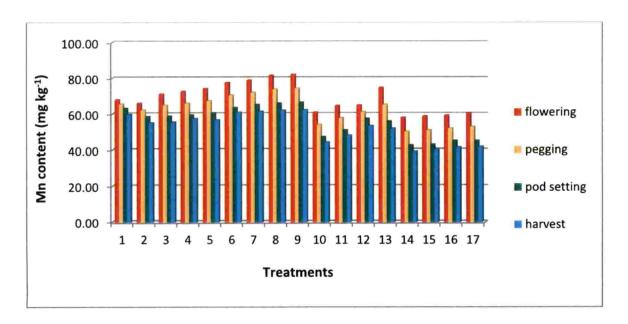


Fig 15: Effect of application of P and B on Manganese content in plant at different stages

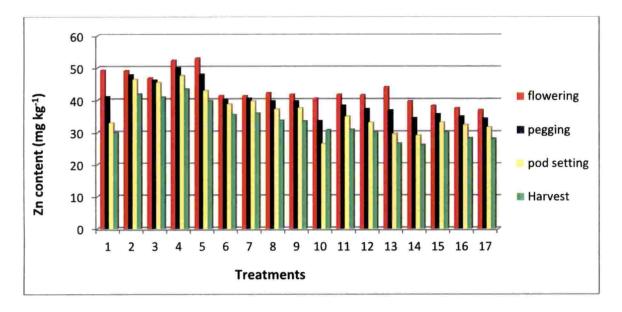


Fig 16: Effect of application of P and B on zinc content in plant at different stages

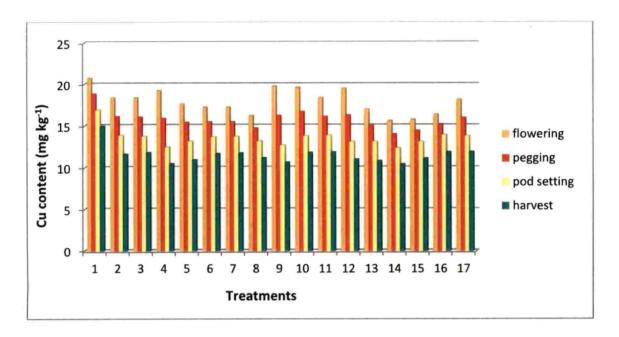


Fig 17: Effect of application of P and B on copper content in plant at different stages

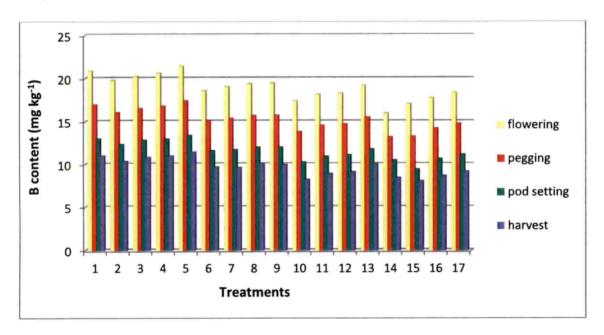


Fig 18: Effect of application of P and B on boron content in plant at different stages

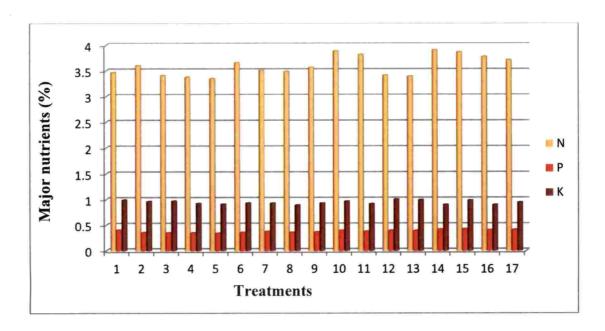


Fig 19: Effect of application of P and B on major nutrients content in pod

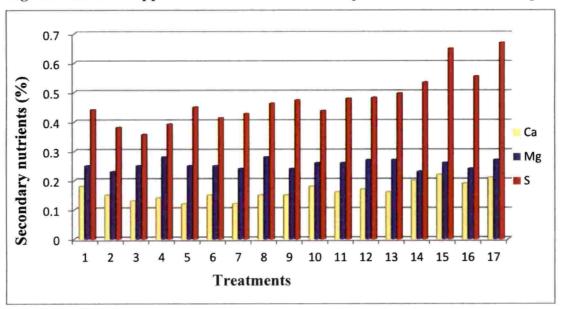


Fig 20: Effect of application of P and B on secondary nutrients content in pod

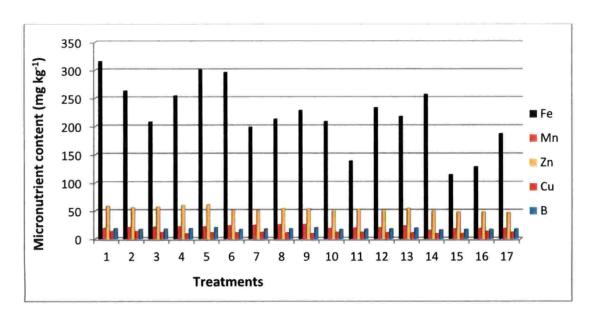


Fig 21: Effect of application of P and B on micro nutrients content in pod

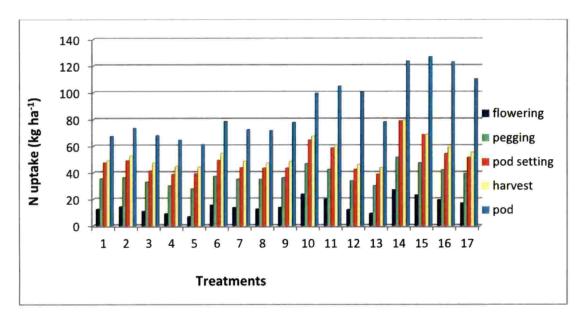


Fig 22: Effect of application of P and B on N uptake by plant at different stages and by pod

Highest K was noted in treatment T<sub>13</sub> (P<sub>2</sub> B<sub>3</sub>) which was supplied with medium doses of P (75 kg ha<sup>-1</sup>). Uptake of K was less in treatment having high Mg and Ca. K, Ca and Mg compete with each other and addition of any one of them will reduce the uptake rate of the other two (Ranade and Malvi, 2011). Uptake of K by plant was higher than uptake by pods. The haulm retains major part of K accumulated during vegetative growth indicating its utilization for structural and developmental processes and allowed little translocation of potassium towards reproductive parts and hence kernel contains less amount of K (Yakadri and Satyanarayana, 1992).

Uptake of Ca and Mg increased with advance in the age of the crop up to harvest due to the increased dry matter production. Groundnut crop was a heavy feeder of Ca. 90 per cent of Ca uptake was during flowering and pod formation stages (Sarkar *et al.*, 1999).

At harvest, Ca and Mg uptake was more in plant than in pod. Ca taken up by the plant will remain in the leaf tissues, and will not move from the leaves to the developing pods where its requirement was high (Ca is immobile in plant system). So the uptake by pods was lesser compared to plant (Meena *et al.*, 2007).

Mg shows positive interaction with N and P (Ranade and Malvi, 2011). Mg uptake was less compared to Ca except pod uptake. Generally, the binding strengths of potassium and Ca are much stronger than magnesium and they easily out-compete magnesium for exchange sites, so the uptake of magnesium was low compared to Ca uptake, but pod uptake of magnesium at harvest was more compared to Ca, because of Ca immobility towards the reproductive parts in plant (Meena *et al.*, 2007). The haulm retains magnesium accumulated during vegetative growth indicating its utilization for structural and developmental processes, with less translocation of magnesium towards reproductive parts; hence kernels contain least amount of magnesium compared to haulm (Babu *et al.*, 2007).

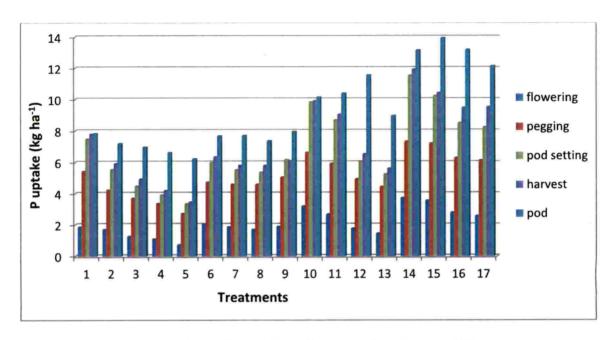


Fig 23 Effect of application of P and B on P uptake by plant at different stages and by pod

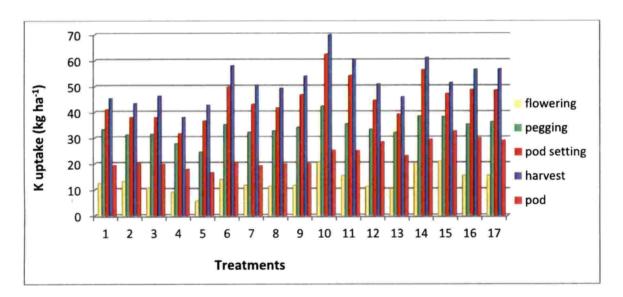


Fig 24 Effect of application of P and B on K uptake by plant at different stages and by pod

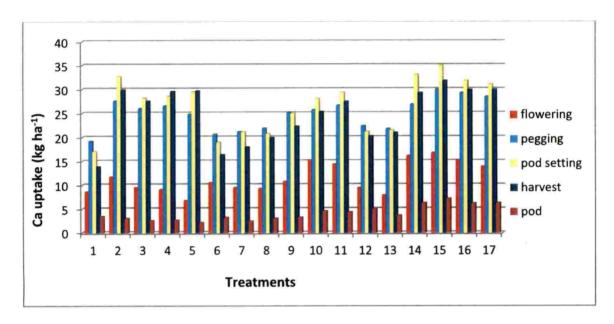


Fig 25 Effect of application of P and B on Ca uptake by plant at different stages and by pod

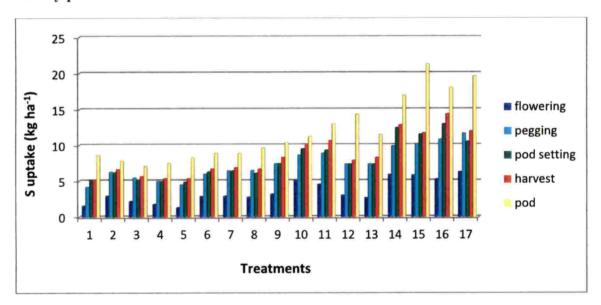


Fig 26 Effect of application of P and B on S uptake by plant at different stages and by pod

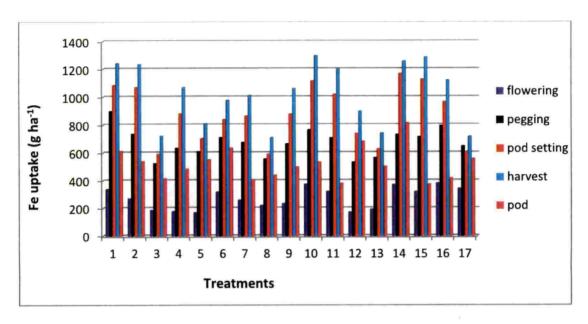


Fig 27 Effect of application of P and B on Fe uptake by plant at different stages and pod

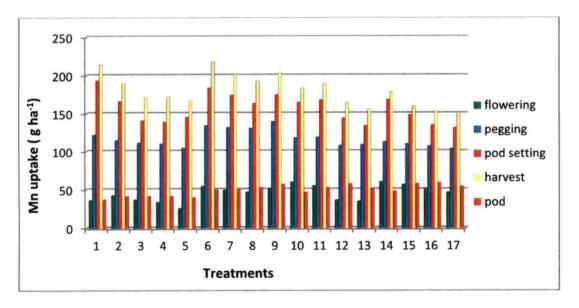


Fig 28: Effect of application of P and B on Mn uptake by plant at different stages and pod

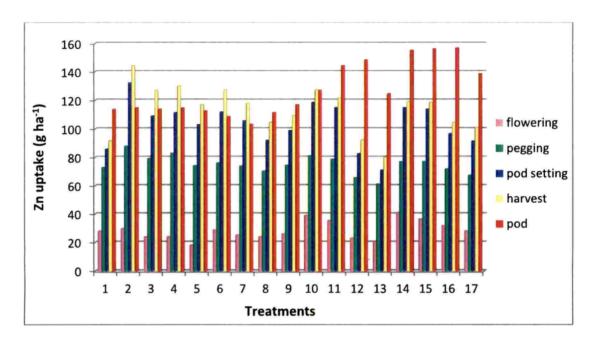


Fig 29 Effect of application of P and B on Zn uptake by plant at different stages and pod

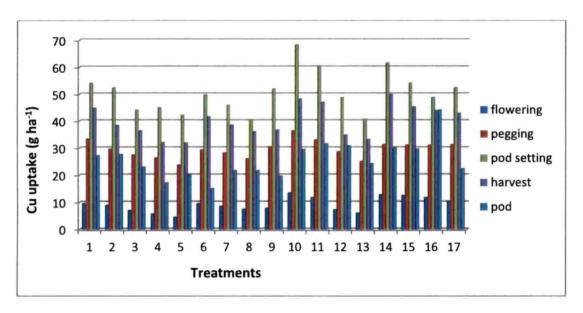


Fig 30 Effect of application of P and B on Cu uptake by plant at different stages and pod

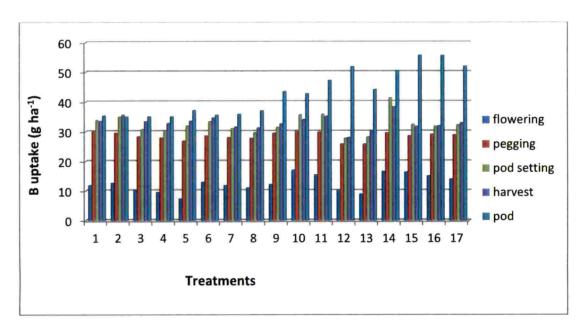


Fig 31 Effect of application of P and B on B uptake by plant at different stages and pod

The results revealed that the uptake of S by groundnut increased with the advancement in the age of the crop. This was mainly due to increased dry matter production. S uptake was more in pod than in plant and this might be due to S involvement in amino acids, proteins and oil synthesis in pod.

Rate of micro nutrient uptake was higher from 30 to 90 DAS and lower at harvest. At harvest, iron, zinc and copper uptake was more in pod than in haulm, whereas uptake was more in haulm than in pod in case of manganese and boron. Uptake of zinc by pod was higher than that of plant and this might be due to zinc involvement in amino acids and protein metabolism in pod. The data on uptake by haulm and pod at harvest stage revealed that uptake of B by plant was higher than that of pods. The B taken up by the plant will remain in the leaf tissues, and will not move from the leaves to the developing pods where its requirement is high (B was immobile in plant system), so the uptake by pods was lesser compared to haulm (Meena *et al.*, 2007). Since B is immobile in plant, B uptake was more by plant than pod.

# <u>Summary</u>

#### 6. SUMMARY

In Kerala, black soils are seen in Chittur taluk of Palakkad district occupying an area of 2000 ha. These are sufficient in all nutrients except phosphorus (P) and boron (B). So availability of P and B is one of the yield limiting factors in this soil. Finding the interaction between B and P in black soils would help to understand the factors determining the availability of these nutrients to crops.

Soil samples were collected from different locations of Chittur and analyzed for available P and B. Soil that showed deficiencies of both P and B was selected for field experiment. The study consisted of a field experiment with groundnut variety, K-6 in black cotton soils of Chittur, Palakkad followed by analysis of soil, plant and pod samples taken from the experimental field. Experiment was laid out in factorial RBD with 17 treatments and 4 replications. Treatment combinations were made with four levels of P and four levels of B with soil test based recommendations as control. N and K levels are kept same (based on POP recommendations of KAU) for all treatments except for the first treatment where soil test based recommendations was given. Single super phosphate (SSP) was used as source of P and borax was used as source of B.

- Soil was sufficient in all nutrients except P and B.
- Application of P at 90 kg ha<sup>-1</sup> increased plant height and number of leaves per plant.
- Application of P at 90 kg ha<sup>-1</sup> and B at 10 kg ha<sup>-1</sup> resulted in highest number of pods
- Application of P at 90 kg ha<sup>-1</sup> and B at 5 kg ha<sup>-1</sup> resulted in highest yield (3.66 t ha<sup>-1</sup>).

- Effect of application of different doses of P and B was non-significant for pH,
   EC and OC.
- N content in soil increased by increasing levels of P and decreased by increased dose of B.
- Application of P decreased B content in soil and vice versa. P and B have negative interaction due to anionic competition.
- K content decreased with increased dose of P and B application.
- Ca content in soil was increased with increased levels of P due to the addition of Ca through SSP. Ca increased with increased dose of B.
- Mg has synergistic interaction with P.
- S has a positive interaction with P and B. The available S was increased with increased dose of P and B application. Noticeable increase in S content was occurred through addition of SSP.
- Application of P reduced the availability of Fe, Mn and Zn in soil due to the formation of insoluble compounds like Fe-P, Mn-P and Zn-P.
- Cu content in soil was non-significant to the addition of different levels of P and B.
- Plant nutrient content showed a decreasing trend from flowering to harvest stage.
- Content of N, P, K, Ca and S was increased with increased levels of P due to enhanced availability from soil.
- Content of Fe, Mn, Zn and B decreased with increased dose of P due to reduced availability and competition.
- Uptake of nutrients followed increasing trend from flowering to harvest due to increased dry matter production.
- Uptake of N, P, S, Zn and B were higher in pod than plant.
- Application of P at 90 kg ha<sup>-1</sup> and B at 0 kg ha<sup>-1</sup> resulted in highest protein content.

# **Future line of work**

- Conduct OFT in different location before recommending for the farmers for adoption.
- Fractionation of phosphorus and boron in Vertisol.
- Influence of phosphorus and boron application on biological properties of black soils.
- Interaction study of phosphorus and boron using different sources of nutrients.

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# References

#### REFERENCES

- Adams, P. and Winsor, G. W. 1974. Some response of glasshouse tomatoes to boron. *J. Hortic. Sci.* 49(4): 355-363.
- Adriano, D. C. and Murphy, L. S. 1970. Effects of ammonium polyphosphates on yield and Chemical Composition of Irrigated Corn. J. Agron. 62(5): 561-567.
- Adriano, D. C., Paulsen, G. M. and Murphy, L. S. 1971. Phosphorus-Iron and Phosphorus-Zinc Relationships in Corn (*Zea mays* L.) Seedlings as Affected by Mineral Nutrition. *J. Agron.* 63(1): 36-39.
- Ae, N., Otani, T., Makino, T. and Tazawa, J. 1996. Role of cell wall of groundnut roots in solubilizing sparingly soluble phosphorus in soil. *Plant Soil*. 186(2): 197-204.
- Agasimani, C. A. and Hosmani, M. M. 1989. Response of groundnut crop to stand geometry in rice fallows in coastal sandy soils of Uttara Kannoda district in Karnataka. *J. Agric. Sci.* 2(1-2): 7-11.
- Al-lami, A.S.J. 1999. Evaluation of magnesium supplying power in plastic houses soils. PhD Dissertation, Baghdad University, 136p.
- Ayed, I. A. 1970. A study of the mobilization of iron in tomato roots by chelate treatments. *Plant Soil*. 32(1-3): 18-26.

- Babu, K. G., Munaswamy, V., John, K. and Raju, P. A. 2007. Effect of applied nutrients on major (N, P, K) and secondary (Ca, Mg, S) nutrients concentration at different growth stages of rainfed groundnut (*Arachis hypogaea* L.) in Alfisols. *J. Oilseeds Res.* 24(1): 84-87.
- Bala, H.M.B., Ogunlela, V.B., Kuchinda, N.C. and Tanimu, B. 2011. Response of two groundnut (*Arachis hypogaea* L.) Varieties to sowing date and NPK fertilizer rate in a semi-arid environment: Yield and yield attributes. *Asian J. Crop Sci.* 3(3): 130-134.
- Balasubramanian, P. and Palaniappan, S. 1991. Effect of population density, fertilizer levels and time of application on rice (*oryza sativa*) groundnut (*Arachis hypogaea* L.). *Indian J. Agron.* 36: 218-221.
- Banerjee, H. T., Das, M. and Bhattacharjee, T. K. 1967. Nutrition of laterite zone groundnut in West Bengal. *Fert. News* 12 (9): 41-42.
- Basha, S. M. and Rao, G. R. 1980. Effect of potassium deficiency on growth and metabolism of peanut (*Arachis hypogaea* L.) plants. *Proceedings: Plant Sci.* 89(5): 415-420.
- Bell, M.J. 1985. Phosphorus nutrition of peanut (*Arachis hypogea* L.) on cockatoo sandy soils of the ord river irrigation area. *Aust. J. Exp. Agric.* 25: 649-653.
- Bharti, B., Sonawane, P., Nawalkar, S. and Patil, V.D. 2010. Effect of micronutrients on growth and yield of groundnut. *J. Soils Crops* 20(2): 269-273.

- Bingham, F. T., Martin, J. P. and Chastain. J. A. 1958. Effect of phosphorus fertilization of California soils on minor element nutrition of cirtus. *Soil Sci*. 86: 24-31.
- Black, C. A. 1965. *Method of Soil Analysis part 11 Agronomy Monograph No.9*, American Society of Agronomy, Madison, Wisconsin, 148p.
- Boote, K. J., Graw, R. L. and Mc. Cloud, D. E. 1985. Seasonal trends in carbohydrate and N concentrations of plant parts of Florunner peanut and Bragg soyabean. *Proceedings, Am. peanut res. and educ. soc.* 17: 70.
- Brady, N.C. and Weil, R.R., 2002. The nature and properties of soils. Prentice Hall, Upper Saddle River, NJ. *The nature and properties of soils. 13th ed. Prentice Hall, Upper Saddle River, NJ.*
- Chahal, R.S., Singh, S. and Singh, M. 1983. Nitrogen, phosphorus and potassium uptake in groundnut at various stages of plant growth and levels of phosphorus. *Indian J. Plant Physiol.* 26(2): 220-225.
- Chatterjee, C., Sinhe, P. and Agarwala, S. C. 1990. Interactive effect of boron and phosphorus on growth and metabolism of maize grown in refined sand. *Can. J. Plant Sci.* 70: 455-460.
- Chauhan, Y.S., Jain, V.K., Khandedar, M.P. and Jain, P.C. 1987. Response of groundnut varieties to phosphorus fertilization. *Madras Agric. J.* 74: 261-264.

- Chavan, L.S. and Kalra, G.S. 1983. Effect of phosphorus and potassium levels under varying row spacing on yield, quality and nutrient uptake by groundnut (*Arachis hypogaea* L) variety TG-1 under high rainfall conditions of Konkan region of Maharashtra. *Indian J. Agric. Res.* 17 (1 & 2): 62-68.
- Choudhary, C. H. V. S. K. 1979. Studies on the effect of different levels of nitrogen, phosphorus and potash on growth, yield and quality of irrigated groundnut. Thesis abstract 3: 30-31.
- Cox, F.R., Adams, J.F. and Tucker, B.B. 1982. Liming, fertilization and mineral nutrition. In: H.E. Pette and C.T. young (Eds.) *Peanut Science and Technology*, American Peanut Research and Education Society Inc. Yoakum, Texas, U.S.A. pp. 139-163.
- Csinos, A. S. and Gaines, T. P. 1986. Peanut pod rot complex: A geocarposphere nutrient imbalance. *Plant Dis.* 70(6): 525-529.
- Dahatonde, B. N. 1982. Effect of nitrogen and phosphate fertilization on yield and yield contributory characters of groundnut (*Arachis hypogaea L.*). *Punjab Krishi Vidyapeeth Res. J.* 6(1): 31-33.
- Daliparthy, J., Barker, A.V. and Mondal, S.S. 2008. Potassium fractions with other nutrients in crops: a review focusing on the tropics. *J. Plant Nutr.* 17 (11): 1859-1886.
- Dart, P. J., Mc Donald, D. and Gibbons, R. W.1983. Groundnut production systems with some implication from recent research, Paper presented at Indonesia National Planning Seminar on Palawija, Yogyakarta, March.

- Das, H. N. 1982. Effect of nitrogen, phosphorus, potassium and calcium on growth and yield of groundnut. *J. Res. Assam Agric. Univ.* 3(1): 15-18.
- Dholaria, S.J., Joshi, S.N. and Kabaria, M.M., 1972. Correlation of yield and yield-contributory characters in groundnut grown under high and low fertility levels. *Indian J. Agric. Sci.* 42(6): 467-470.
- Dowood, M. A. 1982. Magnesium and Phosphorus Studies In Dohuk and Alhawler Soil and Its Relation to Grass Tetany. M Sc. Dissertation, University of Mosul, 129p.
- Dubey, S. K. 1997. Co-inoculation of phosphorus bacteria with Bradyrhizobiumjaponicum to increase phosphate availability to rainfed soybean in Vertisol. *J. Indian Soc. Soil Sci.* 45: 506-509.
- Elliott, G. C. and Laeuchli, A. 1985. Phosphorus Efficiency and Phosphate-Iron Interaction in Maize. *J. Agron.* 77(3): 399-403.
- Fageria, N. K. 1983. Ionic interactions in rice plants from dilute solutions. *Plant Soil* 70(3): 309-316.
- Friesen, D. K., Miller, M. H and Juo, A. S. R. 1980. Liming and lime-phosphorus zinc interactions in two Nigerian Ultisols- effects on maize root and shoot growth. *Soil Sci. Soc. Am. J.* 44: 1127-1232.
- Geethalakshmi, V., Lourduraj, A. C., Joel, A. J. and Rajamanickam, K. 1993. Nutrient management in groundnut. *Madras Agric. J.* 80(7): 412-414.

- Ghadekar, S. R., Wankhede, R. G., Miskin, R. B. and Das, S. N. 1993. Performance of summer groundnut under different levels of irrigation and fertilizers. *J. Soils Crops* 3(2): 121-124.
- Giri, U., Mohammad H. S. A., Nanda, M. K. and Bandyopadhyay, P. 2014. Productivity and nutrient uptake of summer groundnut (*Arachis hypogaea* L.) towards different levels of irrigation and sulphur. *J. Crop Weed*. 10(2): 248-251.
- Gomez, K.A. and Gomez, A. A. 1984. Statistical Procedures for Agricultural Research (2<sup>nd</sup> Ed.). John Wiley and Sons Inc., New York, 56p.
- Gunes, A. and Alpaslan, M. 2000. Boron uptake and toxicity in maize genotype in relation to boron and phosphorus supply. *J. Plant Nutr.* 23(4): 541-550.
- Gupta, V.K. and Raj, H. 1983. Response of groundnut genotypes to different levels of zinc. *j. Crop Physiol.* 1(1): 55-67.
- Hadwani, G.J. and Gundalia, J.D. 2005. Effect of N, P and K levels on yield, nutrient content, uptake and quality of summer groundnut grown on Typic Haplustepts. J. Indian Soc. Soil Sci. 53(1): 125-128.
- Hallock, D. L. and Garren, K. H. 1968. Pod Breakdown, Yield, and Grade of Virginia Type Peanuts as Affected by Ca, Mg, and K Sulfates . J. Agron. 60(3): 253-257.

- Hosseini, S.M., Maftoun, M., Karimian, N., Ronaghi, A. and Emam, Y. 2007. Effect of zinc× boron interaction on plant growth and tissue nutrient concentration of corn. *J. plant nutr.* 30(5): 773-781.
- Islam, M.S. and Noor, S. 1982. Performance of groundnut under different levels of phosphate fertilization in grey floodplain of Jamalpur. *Bangladesh J. Agric. Res.* 1: 35-37.
- Jackson, M. L. 1973. Soil Chemical Analysis, Prentice Hall of India, Pvt. Ltd., New Delhi, 498p.
- Jadhar, A. S. and Narkhede, B. N. 1980. Influence of nitrogen and phosphorus fertilization and growth, yield and quality of groundnut. *Madras agric. J.* 67: 131-135.
- Juan, A. R., Curayag, L. J. and Pava, H. M. 1986. Influence of phosphorus fertilization on pod yield and seed quality of three peanut varieties (*Arachis hypogea L.*). J. Agric. Food Nutri. 8(1): 33-62.
- Kabir, R., Yeasmin, S., Islam, A.K.M.M. and Sarkar, M.R. 2013. Effect of phosphorus, calcium and boron on the growth and yield of groundnut (*Arachis hypogea* L.). *Int. J. Bio-Sci. Bio-Technol.* 5(3): 51-60.
- Kachot, M. A., Patel, J. C. and Malavia, D. D. 1984. Response of summer groundnut to irrigation scheduling based on IW/CPE ratio under varying levels of nitrogen and phosphorus. *Indian J. Agron.* 67 (5): 234-236

- Kamalakannan, P. and Ravichandran, M. 2013. Effect of organic and inorganic sources of nutrients on growth and yield of groundnut in two different textured soils. *Asian J. Soil Sci.* 8(1): 88-93.
- Kamara, E.G., Olympio, N.S. and Asibuo, J.Y. 2011. Effect of calcium and phosphorus fertilizer on the growth and yield of groundnut (*Arachis hypogaea* L.). *Int. Res. J. Agric. Sci. Soil Sci.* 1(8): 326-331.
- Kaya, C., Tuna, A. L., Dikilitas, M., Ashraf, M., Koskeroglu, S. and Guneri, M. 2009. Supplementary phosphorus can alleviate boron toxicity in tomato. Sci. Hortic. 121(3): 284-288.
- Keisling, T. C., Lauer, D. A., Walker, M. E. and Henning, R. J. 1983. Visual, tissue, and soil factors associated with Zn toxicity of peanuts. J. *Agron.* 69: 765–769.
- Koch, K. and Mengal, K. 1977. Effect of K and N on utilization by spring wheat during grain protein formation. *J. Agron.* 69: 477-480.
- Krishnakumar, A. K. 1978. Study of physic-chemical characteristics of the poonthalpadam soils of Kerala. Msc (Ag) thesis, Kerala Agricultural University. P.120.
- Kuchanwar, O. D., Matte, D. B. and Kene, D. R. 1997. Evaluation of graded doses of fly-ash and fertilizers on nutrient content and uptake of groundnut grown on Vertisol. J. Soils Crops. 7: 1-3.

- Kuo, S. and Mikkelsen, D. S. 1979. Effect of magnesium phosphate adsorption by calcium carbonate. *Soil Sci.* 127: 65-69.
- Kvien, C.S., Branch, W.D., Sumner, M.E. and Csinos, A.S. 1988. Pod characteristics influencing calcium concentrations in the seed and hull of peanut. *Crop* sci. 28(4): 666-671.
- Lakshmamma, P. and Raj, A. S. 1997. Effect of cobalt, phosphorus and potassium on groundnut (*Arachis hypogaea* L.). *Legume Res.*: 20-160.
- Laxminarayana, K. 2004. Effect of organic and inorganic manures on yield and nutrient uptake of groundnut (*Arachis hypogaea* L.) in Ultisols of Mizoram. *J. Oilseeds Res.* 21(2): 280-283.
- Lindsay, W.L. and Norvell, W.A. 1978. Development of a DTPA Soil Test for Zinc, Iron, Manganese, and Copper. *Soil Sci. Soc. Am. J.* 42(3): 421-428.
- Loganathan, S. and Krishnamoorthy, V. S. 1980. Potash application to groundnut. *Madras Agric J.* 67(9): 610-612.
- Loganathan, S., Helkiah, L. J. and Thangavelu, S. 1996. Effect of FYM, nitrogen and potash on yield and uptake of nutrients in groundnut. *Madras Agric. J.* 83(8): 484-487.
- Loneragan, J. F., Grove, T. S., Robson, A. D. and Snowball, K. 1979. Phosphorus Toxicity as a Factor in Zinc-Phosphorus Interactions in Plants. *Soil Sci. Soc. Am. J.* 43(5): 966-972.

- Loneragan, J. F., Grunes, D. L., Welch, R. M., Aduayi, E. A., Tengah, A., Lazar, V. A. and Cary, E. E. 1982. Phosphorus accumulation and toxicity in leaves in relation to zinc supply. Soil Sci. Soc. Am. J. 46(2): 345-352.
- Lundergardh, H. 1934. Mineral Nutrition of Plants. Annu. Rev. Biochem. 3: 485-498.
- Mahajan, T. S., Chavan, A. S. and Dongale, J. H. 1994. Effect of boron on yield and quality of groundnut (*Arachis hypogaea* L.) on lateritic soil. *Indian J. Agric*. *Sci.* 64(8): 532-535.
- Mahapatra, I. C., Singh, K. N., Pillai, K. G. and Bapat, S. R. 1985. Rice soils and their management. *Indian J. Agron.* 30(1): 1-41.
- Mahmoud, M., Shaaban, A.F., El-Sayed, A., El-Nour, E.A.M., Aly, E.S. and Mohamed, A.K. 2006. Boron/Nitrogen interaction effect on growth and yield of faba bean plants grown under sandy soil conditions. *Int. J. Agric. Res.* 1(4): 322-330.
- Mallikarjuna, G., Sudhir, K., Srikanth, K. and Srinivasamurthy, C. A. 2003.
  Phosphorus fixation capacity and its relationship with the soil characteristics in laterite soils of Karnataka. J. Indian. Soc. Soil Sci. 51(1): 23-25.
- Manasa, V., Hebsur, N. S., Malligawad, L. H., Shiva Kumar, L. and Ramakrishna, B. 2015. Effect of water soluble fertilizers on uptake of major and micro nutrients by groundnut and post-harvest nutrient status in a Vertisol of northern transition zone of Karnataka. *The Ecoscan.* 9(1&2): 01-05.

- Mandal, S., Samui, R.C. and Mondal, A. 2005. Growth, yield and yield attributes of groundnut (*Arachis hypogaea* L.) cultivars as influenced by gypsum application. *Legume Res.* 28 (2): 119-121.
- Marion, G. M. and Babbcock, K. L. 1977. The solubility of carbonates and phosphates in calcareous soil suspension. *Soil Sci. Soc. Am. J.*41: 724–728.
- Mathew, J., Nair, K. P. M. and Kuriakose, T. C. 1983. The response of groundnut to phosphorus and potassium under different water management practices. *Agric. Res. J. Kerala* 21(2): 27-31.
- May, G. M. and Pritts, M. P. 1993. Phosphorus, zinc, and boron influence yield components in Earliglow strawberry. *J. Am. Soc. Hortic. Sci.*118(1): 43-49.
- Meena, S., Malarkodi, M. and Senthilvalavan, P. 2007. Secondary and micronutrients for groundnut A Review. *Agric. Reviews*. 28(4): 295-300.
- Moraghan, J. T. and Mascagni, H. J. 1991. Environmental and soil factors affecting micronutrient deficiencies and toxicities. *Micronutr. Agric.* 2: 371-425.
- Nadaf, S.A. and Chidanandappa, H.M. 2015. Effect of zinc and boron application on distribution and contribution of zinc fractions to the total uptake of zinc by groundnut (*Arachis hypogaea* L.) in sandy loam soils of Karnataka, India. *Legume Res. Int. J.* 38(5): 598-602.
- Nagarajan, S. S. and Gopalakrishnan, S. 1958. Root inducing substance in groundnut seeds. *Current Sci.* 27(1): 29-30.

- Nair, N. P. and Sadanandan, N. 1981. Quality of kernal in groundnut varieties TMV-2 and TMV-9 as influenced by phosphorus and potassium fertilization. *Agric. Res. J. Kerala.* 19(3): 25-28.
- Nair, N.P., Sadanandan, N., Mohamedkunju, U. and Nair, K.P.M. 1981. Potash fertilization and higher yields of bunch groundnut in Kerala. *Indian Pot. J.* 6: 14-17.
- Nakagawa, J., Machedo, J. R and Toledo, F. F. DE. 1981. The effect of increasing rates of phosphorus fertilizer on groundnut (*Arachis hypogaea* L.) crops. *Cientifica* 9 (2): 227-234.
- Nijhawan, H.L. and Maini, N.S. 1966. The effect of Calcium on groundnut yields. *Indian Farming* 16: 27-29.
- Padmaja, P., Geethakumari, V. L., Nair, H. K., Chinnamma, N. P., Sashidharan, N. K., and Rajan, K. C. 1994. A glimpse to problem soils of Kerala. Kerala Agricultural University, Vellanikkara, Trissur, Kerala. Pp. 104-111.
- Page, A. L., Miller, R. H. and Keeney, D. R. (Ed.). 1982. *Methods of soil analysis*; Chemical and microbiological properties. Madison, Wisconsin, USA.
- Pant, L.M. and Katiyar, A.K. 1996. Response of groundnut of fertilizer N, FYM and Bradyrhizobium inoculation. *Agric.Sci.Digest* 16: 52-54.
- Patel, G. N., Patel, P. T., Patel, P. H., Patel, D. M., Patel, D. K. and Patel, R. M. 2009. Yield attributes, yield, quality and uptake of nutrients by summer

- groundnut (*Arachis hypogaea* L.) as influenced by sources and levels of sulphur under varying irrigation schedules. *J. Oilseeds Res.* 26(2): 119-122.
- Patel, M. S., Golaka, P. R. and Shobhana, H. K. 1981. Growth and nutrient uptake by groundnut (*Arachis hypogaea* L.) as influenced by levels of phosphorus and soil moisture. *Indian J. Agric. Chem.* 14 (1): 33-37.
- Patel, Z.G., Solanki, N.P. and Patel, R.S.1994. Effect of varying levels of seeds, N and phosphatic fertilizer on the growth and yield of summer groundnut (*Arachis hypogaea* L.). *Gujarat agric. Univ. Res. J.* 20: 148-150.
- Piper, C.S. 1966. Soil and Plant Analysis. Hans Publishers, Bombay, 368p.
- Pollard, A. S., Parr, A. J. and Loughman, B. C. 1976. Boron in relation to membrane function in higher plants. *J. Exp. Bot.* 28(4): 831-841.
- Pradhan, A.C. and Das, A.D. 1989. Study on fertilizer response of groundnut at Singhthum district of Bihar under rainfed condition. *Indian Agric. st.* 33: 159-162.
- Punnoose, K. I. 1968. Studies on the effect of nitrogen and phosphorus on the growth, yield and quality of groundnut in Red loam soils of Kerala. Msc. (Ag) thesis, Kerala Agricultural University, Thrissur, 127p.
- Puntamkar, S.S. and Bathkal, B.G. 1967. Influence of N, P and K fertilizers on composition, growth and yield of groundnut. *Indian J. Agron.* 12(4): 344-350.

- Puri, D. N. 1969. Groundnut responds well to super phosphate. Fert. News 14(7): 46-47.
- Ramanathan, K.M., Bruchholz, H., Hanora, J.F. and Subbiah, S. 1982. Influence of nitrogen and potassium on yield and quality of groundnut var. POL-2. *Indian Pot. J.* 7: 6-10.
- Rana, D. S., Kalsi, H. S., Kapur, M. L. and Bhandari, A. L. 1984. Response of groundnut (*Arachis hypogaea* L.) and raya (*Brassica juncea L.*) to fertilizer application. *Indian J. Agron.* 29(1): 122-124.
- Ranade, U. and Malvi. 2011. Interaction of micronutrients with major nutrients with special reference to potassium. *Karnataka J. Agric. Sci.* 24(1): 106-109.
- Rao, I.V.S., Babu, M.D. and Reddy, P.R. 1984. Effect of phosphorus and Rhizobium on groundnut. *Andhra Agric. J.* 31: 253-254.
- Rao, K.T., Rao, A.U. and Sekhar, D. 2013. Effect of sources and levels of sulphur on groundnut. *J. Academia and Industrial Res.* 2(5): 268-270.
- Rao, S. R. 1979. Studies on the effect of potassium, calcium, magnesium on growth and yield of irrigated groundnut (TMV-2). Msc. (Ag) thesis, S. V Agricultural College, Tirupathy, 117p.
- Rao, S. S. and Shaktawat, M. S. 2002. Effect of organic manure, phosphorus and gypsum on groundnut (*Arachis hypogaea* L.) production under rainfed condition. *Indian J. Agron.* 47(2): 234-241.

- Ravichandra, K., Jyothi, C.N., Sigh, B.J., Dawson, J. and Krupakar, A. 2015. Growth of Groundnut (Arachis hypogaea L.) and its Yield as Influenced by Foliar Spray of Boron along with Rhizobium Inoculation. *Indian J. Dryland Agric. Res. Dev.* 30(1): 60-63.
- Rayar, A. J. 1986. Response of groundnut (Arachis hypogaea L.) to application of FYM, N and P on light sandy loam Savanna soils of northern Nigeria. *Int. J. Trop. Agri.* 4(1): 46-54.
- Reddy, B.B., Ramesh, S., Raju, M.S. and Rao, C.M. 1984. Response of groundnut to nitrogen fertilizer and plant densities in relation to rhizobial inoculation. *J. Oilseed Res.* 1: 233-235.
- Reddy, G. P. and Rao, S. C. 1965. Fertilizers response in groundnut. *Indian oil Seeds*. *J.* 9:274-279.
- Reddy, K.C. and Murthy, P.K. 1985. Distribution of N, P, K, Ca, Mg, S, Zn, Fe, Mn and Cu in groundnut. *Plant soil* 84(2): 269-273.
- Reddy, R., Krishnamoorthy, K. and Malewar, G. U. 1983. Response of groundnut to the application of nitrogen, phosphorus and potassium both in the presence and absence of FYM through soil and foliar spray. *Indian J. Agron.* 21(4): 321-326.
- Reddy, T. S., Reddy, S. D. and Reddy, P. G. 2011. Fertilizer management for maximizing productivity and profitability of export oriented groundnut (*Arachis hypogaea* L.). J. Res. ANGRAU. 39(4): 83-85.

- Ruksar, B., Shroff, J.C. and Shah, S.N. 2017. Effect of sources and levels of sulphur and bio-fertilizer on growth, yield and quality of summer groundnut. *Int. J. Agric. Sci.* 13(1): 67-70.
- Saeed, M. and Fox, R. L. 1979. Influence of Phosphate Fertilization on Zinc Adsorption by Tropical Soils . *Soil Sci. Soc. Am. J.* 43(4): 683-686.
- Sagare, B.N. and Bhalkar, D.V. 1986. Effects of applied sulphur and phosphorus in groundnut nutrition and their residual effects on wheat. *J. Maharashtra Agric. Univ.* 11(3): 261-263.
- Saini, J. S. and Tripathi, H. P. 1973. Effect of nitrogen and phosphorus application on the yield of spreading and erect varieties of groundnut (*Arachis hypogaea* L.). *Indian J. Agron.* 48(2): 116-118.
- Samtana, M., Bhadoria, P.B.S. and Ghosh, B. 1994. Effect of lime and phosphorus on phosphate availability and yield of groundnut (*Arachis hypogaea* L.) in Oxisol. *Indian J. Agron*. 39: 692-693.
- Sanchez, L. F. S. and Owen, E. J. B. 1978. Influence of liming and fertilization with N, P and K on yields of peanut (*Arachis hypogaea* L.) grown on soils of the high terraces of the Eastern plains of Colombia. Fert. Abstr. 13: 190-193.
- Saradhi, K., Subbiah, G., Raghavulu, P. and Rao, G.V.H. 1990. Effect of graded levels of nitrogen and phosphorus on growth and yield of groundnut on sandy soils. *Andhra Agric. J.* 37: 407-409.

- Sarkar, R.K., Chakraborty, A. and Saha, A. 1999. Effect of foliar application of potassium nitrate and calcium nitrate on groundnut (*Arachis hypogaea* L.). *Indian J. Agron.* 44(4): 809-812.
- Sebale, R.N. and Khuspe, V.S. 1986. Effects of moisture, phosphate and antitranspirants on growth, dry matter and yield of summer groundnut. *J. Maharashtra Agric. Universities* 11:13-16.
- Shelke, D. K. and Khuspe, V. S. 1980. Response of groundnut (*Arachis hypogaea* Linn.) to varying levels of irrigation, phosphorus and antitranspirant in summer. *J. Maharashtra Agric. Univ.* 5(2): 149-153.
- Shipkule, M.R., 2008. Nutrient requirement of summer groundnut by conjoint use of organic manure and inorganic fertilizers based on STCRC approach. Doctoral dissertation, Mahatma Phule Krishi Vidyapeeth, Rahuri, 138p.
- Singh, A.L. and Chaudhari, V. 2008. Macronutrient requirement of groundnut: effects on growth and yield components. *Indian J. Plant Physiol.* 11(4): 401-402.
- Singh, C. and Pathek, S. S. 1969. Groundnut responds to NPK. Fert. News 14(2): 26-27.
- Singh, F. and Oswalt, D. L. 1995. Groundnut production practices. *Skill development series* (3).
- Singh, K.P. and Ahuja, K.N. 1985. Dry matter accumulation, oil content and nutrient uptake in groundnut (*Arachis hypogea* L.) cv. T 64 as affected by fertilizers and plant density. *Indian j. agron*.

- Singh, Lal, A., Jat, R. S. and Misra, J.B. 2009. Boron fertilization is a must to enhance peanut production in India. *The proceedings of the international plant nutrition colloquium*.
- Sivasankar, A., Rao, I.V.S. and Reddy, P.R. 1981. Influence of seed size and phosphorus levels on root production and phosphorus uptake in groundnut (*Arachis hypogaea* L.). *Legume Res.* 4(2): 78-82.
- Skelton, B. J. and Shear, G. M. 1971. Calcium translocation in the peanut (*Arachis hypogaea* L.). *Indian J. Agron.* 63(3): 409-412.
- Soliman, M.F., Kostandi, S.F. and Beusichem, M.L. 1992. Infuence of Sulfurand Nitrogen Fertilizer on the Uptake of Iron, Manganese, and Zinc by Corn Plants Grown in Calcareous Soil. *Commn. Soil Sci. Plant Anal.* 23:1289-1300.
- Subbiah, B. V. and Asija, G. L. 1956. A rapid procedure for the estimation of available nitrogen in soils. *Curr. Sci.* 25: 259- 260.
- Subrahmaniyan, K., Kalaiselvan, P. and Arulmozhi, N. 2001. Response of confectionery groundnut to micronutrients. *Legume Res.* 24(2): 139-140.
- Sumner, M. E. and Farina, M. P. 1986. Phosphorus interactions with other nutrients and lime in field cropping systems. In *Advances in soil science*. Springer, New York, pp.201-236.

- Survase, D.N., Dongale, J.H. and Kadrekar, S.B. 1986. Growth, yield, quality and composition of groundnut as influenced by FYM, calcium, sulphur and boron in lateritic soil. *J. Maharashtra Agric. Univ.* 11(1): 49-51.
- Swietlik, D. 1995. Interaction between zinc deficiency and boron toxicity on growth and mineral nutrition of sour orange seedlings. *J. Plant Nutr.* 18(6): 1191-1207.
- Tavajjoh, M., Karimian, N. A., Ronaghi, A., Yaserebi, J., Hamidi, R., and Olama, V. 2016. Yield, yield components and seed quality of two rapeseed cultivars as affected by different levels of phosphorus and boron under greenhouse conditions. J. Sci. Technol. Greenh. Cult. 6(24): 99-112.
- Thimmegowda, S. 1993. Effect of fertilizer levels on growth and yield of groundnut after kharif rice. *J. Res.* 21(42): 36-38.
- Tisdale,S.L., Nelson,W.L., Beaton, J.N., and Havlin, J.L. 1993. Soil Fertility and Fertilizers (Indian Reprint, 1997). Prentice Hall of India Private Ltd., New Delhi, 634p.
- Umadevi, M., Munaswamy, V., Santaiah, V., and Prasad Rao, A. 1999. Uptake of major and secondary nutrients by groundnut as affected by sulphur levels under farmer's field conditions. *Leg. Res.* 22(1): 31-35.
- Umadevi, M., Munaswamy, V., Santaiah, V., and Prasad Rao, A. 2000. Field verification of critical available sulphur or sulphur availability index for groundnut in red soils. *J. Oilseeds Res.* 17(1): 168-170.

- Veerabhadrappa, B. H. and Yeledhalli, N. A. 2005. Effect of soil and foliar application of nutrients on growth and yield of groundnut. *Karnataka J. Agric. Sci.* 18(4): 814.
- Venkateswarlu, M. S. and Nath, V. S. 1989. NP and K requirement of groundnut during kharif and rabi seasons in Chittoor district (AP). *J. Res. APAU*, 17(1): 45-46.
- Walker, M.E. 1975. Calcium requirements for peanuts. *Communications in Soil Sci.* and Plant Analysis 6(3): 299-313.
- Walkley, A. J. and Black, C. A. 1934. An examination of the degtjareff method for determining soil organic matter and a proposed modification of the chromic acid titration method. Soil Sci. 37: 29-38.
- Watanabe, F.S. and Olsen, S.R. 1965. Test of an ascorbic acid method for determining phosphorus in water and NaHCO<sub>3</sub> extracts from soil. *Soil Sci. Soc. Am. J.* 29(6): 677-678.
- Williams, J. H. 1979. The Physiology of Groundnuts (*Arachis hypogaea* L.) cv. Egret, Nitrogen Accumulation and Distribution. *Rhodesian J. Agric. Res.* 17(1): 49-55.
- Yadav, G.L. 1990. Fertilizer requirement of groundnut (*Arachis hypogaea* L.) under clay land conditions on cultivators fields. *J. Oilseed Res.* 7: 133-135.



- Yakadri, M. and Satyanarayana, V. 1992. Response of rainfed groundnut (*Arachis hypogaea*) to potassium with varying levels of nitrogen and phosphorus. *Indian J. Agron.* 37: 202-202.
- York, E.T Jr. and Colwell, W.E. 1951. Soil properties, fertilization and maintenance of soil fertility. In *Symposium the peanut, the unpredictable legume*. Washington, pp. 122-171.

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## PHOSPHORUS AND BORON INTERACTIONS IN BLACK COTTON SOILS OF KERALA WITH RESPECT TO GROUNDNUT (Arachis hypogaea L.)

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## ABSTRACT OF THESIS

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## **ABSTRACT**

Balanced supply of nutrients is one of the most important factors determining crop yield. Sometimes the applied nutrients may not be available for plant use, as their availability depends on interaction between nutrients. When the supply of one nutrient element affects the absorption and utilization of other nutrient element, the elements are said to be in interaction and interactions may be negative or positive. In Kerala, black cotton soils are seen in Chittur taluk of Palakkad district occupying an area of approximately 2000 ha. These soils are sandy clay loam, dark, calcareous, neutral to alkaline (pH 7.0 to 8.5), high in clay content and CEC. These soils are sufficient in all nutrients except phosphorus (P) and boron (B). Finding the interaction between B and P in black soils will help to understand the factors determining the availability of these nutrients to crops.

The present study was carried out in College of Horticulture, Vellanikkara during 2016-18. The objectives of the study were to find out the interactions of boron and phosphorus in black cotton soils of Palakkad and to assess the treatment level of boron (B) and phosphorus (P) for maximizing the yield. The study consisted of a field experiment with groundnut variety, K-6 in black cotton soils of Chittur, Palakkad followed by analysis of soil, plant and pod samples taken from the experimental field. Soil samples were collected from different locations of Chittur and analyzed for available P and B. Field experiment was carried out where deficiency of both P and B was noticed.

Experiment was laid out in factorial RBD with 17 treatments and 4 replications. Treatment combinations were made with four levels of P and four levels of B with soil test based recommendations as control. N and K levels are kept same (based on POP recommendations of KAU) for all treatments except for the first treatment where soil test based recommendations was given. P<sub>0</sub> - 0 kg ha<sup>-1</sup>, P<sub>1</sub> - 60 kg ha<sup>-1</sup>, P<sub>2</sub> -75 kg ha<sup>-1</sup> and P<sub>3</sub> -90 kg ha<sup>-1</sup> were the four levels of P and B<sub>0</sub> - 0 kg ha<sup>-1</sup>,

 $B_1$  - 5 kg ha<sup>-1</sup>,  $B_2$  - 10 kg ha<sup>-1</sup> and  $B_3$  - 15 kg ha<sup>-1</sup> were the four levels of borax. Single super phosphate (SSP) was used as source of P and borax was used as source of B.

Physical characteristics of soil *viz.*, texture and bulk density were analyzed before experiment and chemical characteristics *viz.*, pH, EC, organic carbon, N, P, K, Ca, Mg, S, Fe, Mn, Zn, Cu and B were analyzed before and after the experiment. Growth parameters like plant height and number of leaves per plant were recorded at flowering, pegging, pod setting and harvest stages and yield parameters like number of pods per plant and yield were recorded at harvest stage. Plant nutrient content was analyzed and uptake was computed at different stages *viz.*, flowering, pegging, pod setting and harvest stages whereas pod nutrient content and uptake was computed only at harvest stage.

Soil nutrient status, plant nutrient content and uptake of nutrients were affected by main effect and interaction effect of P and B. Application of P at 90 kg ha<sup>-1</sup> and B at 0 kg ha<sup>-1</sup> resulted in highest plant height and number of leaves per plant. Application of P at 90 kg ha<sup>-1</sup> and B at 5 kg ha<sup>-1</sup> resulted in highest number of pods per plant and yield. Application of P at 90 kg ha<sup>-1</sup> and B 0 kg ha<sup>-1</sup> resulted in highest protein content in groundnut.

Application of increased dose of P reduced availability of B due to anionic competition. Application of P reduced the availability of Fe, Mn and Zn in soil due to the formation of insoluble compounds like Fe-P, Mn-P and Zn-P. Application of P enhanced the availability of N. K content in soil was reduced with increased dose of P application. Ca and S were increased due to its supply through fertilizer in soil.

Plant nutrient content showed a decreasing trend from flowering to harvest stage. Application of P enhanced plant and pod P content and application of B reduced plant and pod P content. Content of N, P, K, Ca and S was increased with increased levels of P and content of Fe, Mn, Zn and B was reduced with increased

dose of P. Content of N, P, S, Zn and B were higher in pod than plant. Uptake of nutrient followed an increasing trend from flowering to harvest due to increased dry matter production. The uptake of N, P, S, Zn and B by pod was higher compared to plant uptake.

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