

**“EVALUATION OF MICRONUTRIENT FORMULATION IN COWPEA
(*Vigna unguiculata* L. Walp)”**

by

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THESIS

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KERALA, INDIA

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DECLARATION

I, hereby declare that this thesis entitled “**Evaluation of micronutrient formulation in cowpea (*Vigna unguiculata* L. Walp)**” is a bonafide record of research work done by me during the course of research and the thesis has not previously formed the basis for the award to me of any degree, diploma, associate, fellowship or other similar title, of any other University or society.

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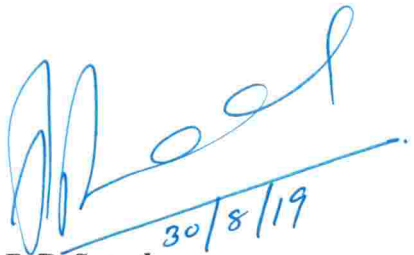
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LIST OF ABBREVIATIONS

%	-	Per cent
@	-	at the rate of
B	-	Boron
°C	-	Degree celcius
Ca	-	Calcium
CD	-	Critical difference
CEC	-	Cation Exchange Capacity
cm	-	Centimeter
cmol kg ⁻¹	-	Centimole per kilogram
COA	-	College of Agriculture
Cu	-	Copper
dS m ⁻¹	-	deci Seimens per meter
DAS	-	Days after sowing
°E	-	East
EC	-	Electrical conductivity
<i>et al</i>	-	And others
Fe	-	Iron
Fig.	-	Figure
FYM	-	Farm Yard Manure
g	-	Gram
g cm ⁻³	-	Gram per centimeter cube
ha ⁻¹	-	per hectare
hr ⁻¹	-	per hour
K	-	Potassium
KAU	-	Kerala Agricultural University
kg	-	Kilogram

kg plant ⁻¹	-	Kilogram per plant
kg ha ⁻¹	-	Kilogram per hectare
l	-	Litre
m	-	Meter
meq 100g ⁻¹	-	Milliequivalent per hundred gram
Mg	-	Magnesium
mg kg ⁻¹	-	milligram per kilogram
ml	-	Milliliter
mm	-	Millimeter
Mn	-	Manganese
Mo	-	Molybdenum
MOP	-	Muriate of Potash
N	-	Nitrogen
°N	-	North
NS	-	Not significant
O	-	Oxygen
OC	-	Organic carbon
P	-	Phosphorus
pH	-	Soil reaction
plant ⁻¹	-	Per plant
POP	-	Package of practices
ppm	-	Parts per million
S	-	Sulphur
SE(m)	-	Standard error mean
viz.	-	Namely
Zn	-	Zinc

Introduction

1. INTRODUCTION

Pulses are an important group of crops which has the capacity to provide high quality protein complementing cereal proteins in vegetarian diet. The Recommended Dietary Allowances (RDA) for adult male and female is 60 g and 55g per day respectively. India being the largest pulse crop cultivating country in the world, the country's pulse production is relatively in comparison to total cereal production. Pulses can be cultivated with minimum resources and hence it is cheaper than animal protein sources. Though pulses are grown in both kharif and rabi seasons, rabi pulses contribute more than 60 per cent of the total production. In India, pulses are cultivated in an area of 25.26 million ha with production potential of 16.47 million tonnes and yield of 652 kg ha⁻¹ (GOI, 2016). In Kerala it is cultivated in an area of 1738 ha with a production of 1433 tonnes (GOK, 2016-2017). Bengal gram, pigeon pea, green beans, black matpe, red kidney beans (rajma), black eyed peas (lobia), lentils (masoor), white peas (matar) are the major pulses grown and consumed in India.

Cowpea (*Vigna unguiculata* L. Walp) is an important pulse crop grown annually in warm climate with adequate rainfall. It can thrive well in temperate zones as well as humid tropics. Cowpea is rich in fibre, protein, iron and potassium but low in fat and calorie. It is also referred as vegetable meat because of its high protein content. Inclusion of cowpea in diet prevents cancer, anaemia and repair of muscle tissues. Cowpea is mostly grown under unirrigated condition and on marginal land, hence the yield is very low. Sufficient supply of plant nutrients according to crop needs increases the yield of crops. Yield potential of crops can be achieved by adequate supply of micronutrients along with major nutrients.

Micronutrients are essential elements in growth required by plants only in small quantities. In spite of its lower requirement in plants these are equally formidable as primary and secondary nutrients. Deficiency of micronutrients results in disrupted plant functioning and abnormal plant growth. It may further lead to yield reduction. Micronutrients are essential for the proper metabolism in animals also. Hence periodic monitoring of micronutrient status in soil must be carried out

in order to supplement them in optimum levels. The deficiency of micronutrients become prevalent quite recently affecting the productivity and stability of soils (Bell and Dell, 2008). The factors which triggers micronutrient deficiency includes intensive cropping, leaching and loss of top soil, preference of macronutrient fertilizers over organic manure and inadequate management of soil (Singh, 2003; Rattan and Sharma, 2004; De and Rai, 2005).

Micronutrients can be supplied to plants as soil application, seed treatment and foliar application. Soil application of fertilizers leads to enormous wastage of the applied nutrients through leaching, run off and gaseous loss depending on the prevailing soil conditions for crop growth. This may lead to lowering the fertilizer use efficiency. Foliar nutrition is the most advanced method in supplementing nutrition to plants. It gains attention due to its efficiency as well as cost effectiveness. Application of micronutrients in the appropriate mineral form enhances its availability in plants (Alloway, 1986; House and Welch, 1989). Formulations containing micronutrients when supplied to plants as foliar spray are found effective in correcting deficiencies and thereby increases the yield (Mona *et al.*, 2012). Foliar application of nutrients helps the plants to absorb nutrients through the stomatal openings in leaves and results in most effective use of fertilizers in a most economical way (Manasa *et al.*, 2015). Efficiency of fertiliser use and quick response of the applied nutrients results in growth enhancement as well as yield. It also reduces the chance of toxicity in plants compared to the supplementation of fertilizers through conventional methods.

Seed treatment is the method of application of fungicides, insecticide or combination of both to prevent seed borne pathogens and insects. It also aimed at ensuring the quality of crops which in turn affects the germination, growth, development and yield of crops. Micronutrient fertilizers can also be supplied as seed treatment so that the wastage of fertilizers during soil application can be significantly reduced. Application of micronutrient through seed treatment improves growth, yield and enhances the phenological events (Farooq *et al.*, 2012). In economical perspective seed treatment is the better option for which less

micronutrient is needed, it is ease in application and improves seedling growth (Singh *et al.*, 2003).

Biological nitrogen fixation accounts as a major source of nitrogen input in soils. Legume plants can fix atmospheric nitrogen by formation of nodules. Legume rhizobium symbiosis is an ideal solution to restore soil fertility (Zahran, 1999). Symbiotic association of legumes and nitrogen fixing rhizobium bacteria acts as a natural nitrogen factory of native soils. Biological nitrogen fixation is being widely exploited as it enhances the nutrient status of soil as well as increasing the growth and development of plants. Cowpea being a legume crop has the capacity to fix atmospheric nitrogen in symbiotic association with rhizobium species. Root nodules are formed in cowpea where these symbiotic bacteria are sheltered and thus nitrogen fixation occurs in areas where cowpea is cultivated. Since it restores soil fertility it is an important component of crop rotation. Cowpea is grown before cereals as it enhances the nitrogen status of the soil (Carsky *et al.*, 2002; Tarawali *et al.*, 2002; Sanginga *et al.*, 2003). All the nodules may not fix nitrogen efficiently. To enhance the nitrogen fixation capacity of nodules seeds are treated with rhizobium cultures of suitable strains according to crop species.

On this background the present investigation is proposed with the following objectives

- To evaluate the effect of micronutrient formulation through seed treatment and foliar nutrition on growth, nodulation and yield of cowpea
- To study its effect on plant nutrient uptake and residual soil nutrient status

Review of Literature

2. REVIEW OF LITERATURE

Nutrients are essential components which sustains plant as well as animal population. Plants need all 17 essential elements for its growth and development. It includes macronutrients and micronutrients. Micronutrients are referred as trace elements because plants need these nutrients in small quantities. Even though its requirement is less these nutrients are as important as macronutrients. Lack of any micronutrient element results in growth inhibition and results in complete death of plants (Mengel *et al.*, 2001). As it is involved in metabolic and other enzymatic activities their deficiency affects the growth of plants. Micronutrient status in the soil is decreasing nowadays due to the nonjudicious use of primary nutrient fertilizers. Inorder to maintain the soil nutrient status as well as nutrient requirement of the crops it is essential to supply micronutrients. The present investigation was carried to evaluate the effect of micronutrient application both as seed treatment and foliar application in growth, yield and nutrient uptake of cowpea.

2.1 IMPORTANCE OF COWPEA AS A LEGUME CROP

Cowpea (*Vigna unguiculata* L Walp) is an important vegetable crop belongs to the family Leguminosae. It is recognized as one of the major pulse crop in the world. It was domesticated primarily in Africa and later spread to different regions of the world including tropics and temperate climatic conditions (Pasquet, 1998). It grows well in tropical conditions as it can tolerate high temperature and withstand drought (Hall *et al.*, 2002; Hall, 2004). It can be grown as vegetable crop as well as for grain purpose.

In diet the nutritional quality of cowpea grain is almost same as common bean (Bressani, 1985). But folic acid and other antinutritional factors are very less in cowpea. Grain cowpea has gained importance in developing countries, facing acute shortage of fuels as grain cowpea requires less time for cooking (Hall and Ehlers, 1997).

Cowpea, a legume crop has the capacity to fix atmospheric nitrogen in symbiotic association with rhizobium species. Root nodules are formed in cowpea where the symbiotic bacteria are sheltered and fixes nitrogen in the legume cropped area. Since it restores soil fertility it is an important component of crop rotation. Cowpea is grown before cereals as it enhances the nitrogen status of the soil (Carsky *et al.*, 2002; Tarawali *et al.*, 2002; Sanginga *et al.*, 2003). All the nodules may not fix nitrogen efficiently. To enhance the nitrogen fixation capacity of nodules seeds are treated with rhizobium cultures of suitable strains according to crop species

2.2 IMPORTANCE OF MICRONUTRIENTS IN AGRICULTURE

Essential elements which are required by plants in very low concentration are referred as micronutrients. Even though these nutrients are required in very low quantities they play a prominent role in the metabolism of plants (Benepal, 1967; Katyal, 2004). Micronutrients are essential for the functioning of various enzymes involved in plant system thus helping in nutrition and yield of crops (Kazi *et al.*, 2012). The deficiency of these nutrients reduces the yield of crops tremendously (Udode- Haes *et al.*, 2012). High nutrient requirement of modern crop cultivars aggravates deficiency of micronutrients (Fageria *et al.*, 2002).

All micronutrients except boron and chlorine are metals and its uptake is influenced by several factors including environmental and microbial (Romheld and Marschner, 1986; Clark and Zeto, 2000), plant (Barber, 1995; Marschner, 1995) and soil (Lindsay, 1991; Lake *et al.*, 1984).

2.2.1 Iron

Iron is an essential micronutrient element needed for the growth of plants (Gris, 1843). It is a constituent of various enzymes including cytochrome oxidase, catalase and nitrogenase. It also helps in respiration, photosynthesis and reduction of nitrates and sulphates (Wallihan *et al.*, 1958; Reddy and Reddi, 2002).

Iron is an important constituent of leghaemoglobin in legumes (Gupta and Gupta, 2005). Leaf chlorosis due to deficiency of iron can be reduced by the application of iron fertilizers (Srivastava and Singh, 2003).

2.2.3 Manganese

Essentiality of manganese for plants was proposed by Mchargue (1922). It is essential for photosynthesis, respiration and activity of enzymes. Mn is an important component in Krebs cycle (Gupta and Gupta, 2005).

Manganese deficiency is observed in younger leaves resulting in light green mottles between the veins (Gupta and Gupta, 2005). Foliar spray of manganese sulphate at 0.1 per cent is recommended (Gupta and Gupta, 2005) for manganese deficiency.

2.2.4 Zinc

Sommer and Lipman in 1926 discovered the essential micronutrient zinc. It influences the growth and development of plants. Zinc is a limiting micronutrient in crop production. It is a constituent of enzymes which produces proteins and helps to retain the bonds of biomembranes. It is involved in the synthesis of indole acetic acid. It is essential for metabolism of carbohydrates, synthesis of proteins, internode elongation and in pollen formation (Shukla *et al.*, 2009).

Zinc deficiency results in growth retardation and little leaf (Marschner, 1995). Thus reducing the yield and quality of crops (Gupta, 1995). Zinc spray on necrotic leaves found to increase the green pigments (Srivastava and Singh, 2003).

2.2.5 Copper

Essentiality of copper as micronutrient was put forward by Lipman and MacKinney (1931). It is essential for photosynthesis and mitochondrial respiration. In chloroplasts the electron carrier proteins such as plastoquinone and plastocyanin contain Cu (Gupta and Gupta, 2005).

2.2.6 Boron

Essentiality of boron was first discovered by Warrington (1923). The major roles of boron in plants include promoting plant growth, meristem cell division, translocation of sugars, starch, nitrogen, phosphorus, development of phloem and help in calcium metabolism. Lack of boron in cells results in breakdown of phloem tubes (Edmond *et al.*, 1997).

2.2.7 Molybdenum

Essentiality of molybdenum for plants was identified by Arnon and Stout (1939). It is a component of nitrate reductase and nitrogenase. Also of enzyme oxidase which results in the conversion of abscisic acid aldehyde to ABA (Gupta and Gupta, 2005).

2.3 MICRONUTRIENTS STATUS IN INDIAN SOIL

According to Katyal and Sharma (1991) higher concentrations of micronutrients such as Fe, Mn, Zn, Cu were found in soils developed from green schist and these were too low in sandstone. Limestone, a sedimentary rock contains highest concentration of Zinc.

Soils formed on flood plain alluvium found to be lower in Zn, Cu, Mn and Fe content. Sedimentary rocks contain more Boron compared to igneous rocks (Gupta and Gupta, 1985).

Micronutrients present in the lithosphere are essential for survival of plants. The least abundant micronutrient element in lithosphere is Molybdenum (Mortvedt, 2000). Molybdenum is available to plants only at alkaline pH and it is deficient in acid soils. Poorly drained soil and peaty soils have high molybdenum content.

Unavailability of micronutrients limits growth of plants. Fe is an important micronutrient whose deficiency limits plant growth. Availability of iron to plants is

mostly affected with different cropping sequences because of frequent changes in the form of iron ie, from Fe^{2+} to Fe^{3+} .

Boron is an important micronutrient element which ranges at the rate of 7 to 630 $mg\ kg^{-1}$ in Indian soils (Prasad *et al.*, 2014). Total Zn content in Indian soils ranges from 7 to 284 ppm in entisols and vertisols respectively (Ganjir *et al.*, 1973).

2.4 MICRONUTRIENT DEFICIENCIES IN INDIAN SOILS

The advent of green revolution in India during 1960s increased the crop production in our country (Singh, 2009). Application of chemical fertilizers in fields promote higher production. The injudicious use of macronutrient fertilizers resulted in micronutrient deficiency in Indian soils. Deficiencies of nutrients including both macro and micronutrients are prevalent in Indian soils (Takkar *et al.*, 1989).

The antagonistic interaction of zinc and phosphorus is quite evident that is, soils with higher amount of phosphorus reduces the uptake of zinc from soil (Dadhich and Somani, 2007; Kizilgoz and Sakin, 2010).

Zinc is deficient in soils containing low organic matter and high pH (Rattan and Sharma, 2004). Zinc associated with solid particles cannot be taken by plants (Lake *et al.*, 1984).

Boron is an important micronutrient which is deficient in Indian soils (Sathya *et al.*, 2009). According to Katyal and Vlek (1985), B deficiency in Indian soils was initially reported as 2 per cent during 1980 and it reached to 53 per cent in 2012 (Singh, 2012). B is deficient in Indian soils with a mean of 33 per cent in the whole country (Singh, 1999; Singh, 2006).

Shukla *et al.* (2012) reported that wheat grown after rice cultivation was found to be deficient in manganese. It was observed mainly in wheat grown tracts of Punjab and Haryana. The continuous leaching of Mn was found to be the reason for Mn deficiency. Deficiency of micronutrients affects the productivity of crops which in turn affect the health of man and animals.

Plant samples were collected to study the micronutrient concentration (Shukla *et al.*, 2012). About 20,000 plants were taken for the analysis of Fe, Mn, Zn and Cu concentrations. Study showed that plants were deficient in Fe by 6 per cent, Mn by 4 per cent, Zn by 44 per cent and Cu by 10 per cent

Iron is deficient in most soils. Deficiency in soils are reflected in plants and animals as deficiency symptoms and disorders. This condition can be eliminated by increasing the levels of available iron in staple food crops (Mori, 1999).

2.5 MICRONUTRIENT STATUS IN KERALA SOILS

A major portion of Kerala soils lack boron since it is formed from acid igneous rocks (SSO, 2007). Highly soluble nature of boron in soil (Tisdale *et al.*, 1985) is one of the reason for its deficiency in Kerala, which receives a major share of rainfall during monsoon. A study conducted by Kavitha and Sujatha, 2015 in various agroecosystems of banana, coconut, nutmeg, paddy, rubber, vegetables and pepper in Thrissur district showed that all the micronutrient elements except boron are in sufficient range.

A study conducted by Suresh *et al.*, 2014 in the Northern district of Kerala, Kasaragod revealed that the soils are deficient in boron by 78 per cent, zinc by 8 and copper by 3 percentage.

Acid sulphate soils in the Kuttanad region of Kerala is found to be toxic with Fe and Al which reduces the crop yield (Thampatti *et al.*, 2005).

2.6 ROLE OF RHIZOBIUM IN GROWTH, YIELD, NODULATION AND NUTRIENT UPTAKE OF CROPS

Co-inoculation of plant growth promoting rhizobacteria and rhizobium in the soil contaminated with Cu was studied in alfalfa. The results suggested that the combination increased the growth of plants as well as Cu uptake by plants. Thus it eliminates Cu stress and improves biochemical properties of soil (Ju *et al.*, 2019).

Efficient fixation of nitrogen occurred when the legume host and rhizobium were compatible to each other. This compatibility is not only essential for infection but also for proper development and nutrient exchange mechanisms (Terpolilli *et al.*, 2012).

Seeds of chickpea were inoculated with rhizobium culture alone and in combination with N at the rate of 25 kg N per ha. The study revealed that there was significant increase in nodulation of chickpea compared to control. Increase in yield was also prominent (Tilak *et al.*, 1981).

According to Nyoki and Ndakidemi (2018), when soybean seeds were inoculated with rhizobium the uptake of macronutrients such as N, P, K and Mg was significantly increased in soybean shoots. This study also revealed that the uptake of micronutrients such as Fe, Mn, Zn, Cu increased prominently.

Wolde-meskel *et al.* (2018) studied the effect of rhizobium inoculation and phosphorus application in chickpea. Inoculation and application of P increased the grain yield of chickpea by 21 per cent and 25 per cent respectively, while the combination of both treatments increased the yield by 38 per cent.

Effect of rhizobium inoculation along with the application of N and P fertilizers was studied in summer forage cowpea. The results showed that the application of nitrogen and phosphorus fertilizers at the rate of 20 kg ha⁻¹ and 40 kg ha⁻¹ respectively along with rhizobium inoculation increased the yield and profit of summer cowpea (Amrutrao, 2018).

Inoculation of rhizobium culture in seeds of cowpea do not show significant effect on seed and fodder yield but it showed significant effect on the nitrogen content of seeds as well as P, K and Mg content of fodder (Reddy, 1981).

Plant height, number of branches and yield of garden pea were increased in seeds treated with rhizobium culture (Patel, 1994).

Number of nodules and dry weight of nodules were found to be more in cowpea plants treated with vermicompost along with rhizobial inoculation compared to FYM + rhizobial inoculation (Yadav, 2000).

Mathur (2000) reported that in mungbean inoculation of rhizobium significantly increased growth parameters. Growth parameters include nodule formation, plant height and nodule number.

Plant height, LAI, number of nodules and branches per plant were recorded more in plants inoculated with rhizobium (Thakur *et al.*, 1999). Cowpea seeds inoculated with rhizobium showed higher dry matter production compared to uninoculated seeds (Yadav and Malik, 2005).

The rate of dry matter production in cowpea was observed to increase from 39.1 to 44.5 kg ha⁻¹ per day compared to uninoculated plots with dry matter production ranges from 35.4 to 37.4 kg ha⁻¹ per day (Deshmukh and Joshi, 1973).

Yield response of cowpea treated with rhizobium culture was studied by Lehri *et al.* (1974). It was observed that the yield of cowpea increased from 17.2 to 28.7 q ha⁻¹.

When seeds of *Phaseolus aureus* were treated with rhizobium bacteria, there occurred a significant increase in shoot and root length, fresh and dry weight of plants (Madani and Singh, 1977).

In temperate conditions cowpea seeds inoculated with rhizobium showed increased number of leaves, weight and nodulation compared to the control (Pant *et al.*, 2000).

2.7 SIGNIFICANCE OF NITROGEN FIXATION IN SOIL FERTILITY

Fixation of nitrogen in soil with the help of nitrogen fixing microorganisms is referred as biological nitrogen fixation. It plays a prominent role in the establishment of crop and increase in yield. It provides nitrogen needed for the

growth and development of plants even when the crop is deprived of N fertilizers (Vargas and Hungria, 1997; Chen *et al.*, 2002).

Methods to replenish the nitrogen status of the soil are application of nitrogenous fertilizers, organic manures and growing legume crops which fix atmospheric nitrogen in symbiotic relation with rhizobium bacteria referred as Biological Nitrogen Fixation (BNF). But the rate of BNF is declining nowadays due to unjudicious use of nitrogenous fertilizers (Peoples *et al.*, 1995).

Soil erosion is one of the major reason for soil degradation which results in removal of top soil and leave the soil as infertile. Biological nitrogen fixation plays an important role in replenishing the nitrogen status of such soils (Giller and Cadisch, 1995).

In Brazil cereals were grown with N supply much less than the required quantity along with the association of diazotrophic bacteria. P, K and other micronutrients were supplied in ample quantities. It was observed that the crop was able to obtain up to 30 per cent of their required nitrogen from biological nitrogen fixation (Dobereiner, 1997).

The amount of nitrogen produced in the soil by decomposition of roots and nodules of legumes has been estimated as 3 to 120 kg N ha⁻¹yr⁻¹ or 2 to 26 per cent of biological nitrogen fixation (Ledgard and Steele, 1992). Thus improves the nitrogen status of soil.

2.8 SIGNIFICANCE OF MICRONUTRIENT SEED TREATMENT IN CROPS

Micronutrient seed treatment is equally beneficial compared to other methods of micronutrient application in crops. (Farooq *et al.*, 2012).

Pulse crops treated with Mo in the form of sodium molybdate at the rate of 10g per 25 kg of seed increased the seed yield by 45 per cent compared to non molybdenum treated seeds (Pranakrishna, 1976).

Cowpea seeds were treated with micronutrients such as zinc and boron to study its effect on growth and yield. Seeds treated with borax showed greatest plant height at 30 and 60 DAS. Seeds treated with zinc sulphate showed highest root length (18.51 cm), vigour index (4277) and seedling dry weight (0.595 g) (Masuthi *et al.*, 2009).

Common bean treated with zinc increased the yield and other traits (Kaya *et al.*, 2007). Seeds treated with micronutrients showed improved uptake of zinc and phosphorus in barley. Accumulation of dry matter was also found to be increased (Ajourri *et al.*, 2004).

Wheat seeds treated with 0.3 % zinc increased the yield by 14 per cent and also chickpea seeds treated with 0.05 % zinc increased the yield by 19 per cent (Harris *et al.*, 2008).

In kidney beans seed treatment with zinc alone did not meet the requirement of the crop (Rasmussen and Boawn, 1969). This condition depends on the amount of zinc present in the soil. In case of moderate deficiency seed treatment with zinc is economic (Harris *et al.*, 2007, 2008; Slaton *et al.*, 2001) but not suitable under severe deficiencies (Rasmussen and Boawn, 1969; Rehman *et al.*, 2012).

Rice seeds treated with 0.001 per cent and 0.1 per cent boron solutions improved the crop establishment while 0.5 per cent boron solution failed to germinate (Rehman *et al.*, 2012).

Papaya seeds treated with 2 ppm boron solution for 6 hours showed increased germination and better growth of seedlings (Deb *et al.*, 2010). Plant height, fruiting and pod yield were higher in pea seeds treated with 0.5 per cent boron solution (Kumar *et al.*, 2008).

Seed treatment with boron did not influence the yield of chickpea, lentil, rice or wheat but it increased the boron content of grains in all these crops (Johnson *et al.*, 2005).

Seed treatment is the most efficient method for the supply of molybdenum for plants compared to soil application (Donald and Spencer, 1951; Johansen *et al.*, 2006).

According to Kumar *et al.* (2004) chickpea seeds treated with molybdenum 0.5 gram per liter solution of sodium molybdate for 8 hours resulted in increase in yield by 27 per cent compared to soil application.

Seed treatment of green gram seeds with sodium molybdate along with rhizobium culture increased nodulation, nitrogen fixation, nutrient uptake, plant growth and yield (Pattanayak *et al.*, 2000).

Wheat seeds treated with manganese sulphate solutions showed improved growth, grain yield and grain manganese contents (Khalid and Malik, 1982). Seeds treated with Mn solution showed better growth and yield in *Echinacea purpurea* (L.) compared to soil application (Babaeva *et al.*, 1999).

Wheat seeds treated with CuEDTA solution increased the grain yield but seedling emergence was suppressed (Malhi, 2009). Oat seeds treated with 0.001 per cent copper sulphate solution did not show significant effect on germination but number of grains per panicle and grain weight increased resulting in 16.53 per cent increase in yield compared to untreated samples (Saric and Saciragic, 1969).

2.9 FOLIAR APPLICATION OF MICRONUTRIENTS

Application of micronutrients as foliar spray is effective in controlling deficiencies of crops because it gives immediate results compared to soil application (Torun *et al.*, 2001; Jamal *et al.*, 2006). The immediate action of foliar sprays is due to faster absorption of nutrients through leaf surface than through roots (Hashemy *et al.*, 1998).

Yield and quality of crops can be enhanced through foliar nutrition (Roemheld and El-Fouly, 1999; Sarkar *et al.*, 2007). Foliar nutrition is a better

alternative in arid and semiarid regions compared to soil application (Kaya *et al.*, 2005; Kinaci and Gulmezoglu, 2007; Babaeian *et al.*, 2011).

Fertilizer use efficiency can be increased by foliar nutrition as it requires nutrients in smaller concentrations and thus prevents toxicity and fixation (Malakouti and Tehrani, 1999; Silberbush, 2002).

Foliar feeding is the most accepted method of plant nutrition in modern world (Bernal *et al.*, 2007 and Baloch *et al.*, 2008). It is cost efficient because of its less requirement compared to soil application (Kannan, 1986 and Singh, 2007). It started during 1950 s in America and later adopted by many countries (Wiley-VCH and Wiley-VCH, 2007).

The mechanism of entry of nutrients through leaves is a step by process. It has to penetrate the leaf surface through cuticle, stomata, leaf hairs and other epidermal cells (Fernandez and Eichert, 2009).

Leaf can quickly absorb the nutrients supplied through foliar spray and transport it to phloem and xylem vessels (Hasslett *et al.*, 2001; Nasiri *et al.*, 2010). It also increases the uptake of nutrients through soil by promoting root growth (El-Fouly and El-Sayed, 1997). Thus foliar nutrition is important nowadays compared to soil application (Alam *et al.*, 2010).

Foliar feeding is important in plant metabolism as it increases photosynthetic pigments and organic components in *Cassia angustifolia* (Shitole and Dhumal, 2012).

Foliar feeding is more advisable than soil application as it is 6 to 20 times more efficient (Liew, 1988). Zn, B and Cu foliar spray is equally benefited as soil application in reducing sub soil micronutrient deficiency (Ali *et al.*, 2009; Hussanin *et al.*, 2012).

Grain yield and protein content in wheat, maize, rice, barley and sorghum was enhanced by foliar nutrition of trace elements (Boorboori *et al.*, 2012). Shoot

concentration and uptake of nutrients was found to be more in soybean plants treated with Fe and Mn foliar spray (Moosavi and Ronaghi, 2011).

Thus foliar nutrition is an effective method of supplying essential nutrients for the growth of plants.

2.10 MICRONUTRIENT APPLICATION IN GROWTH, YIELD AND NUTRIENT UPTAKE OF CROPS

Micronutrients enhances the growth and yield of crops (Simmonds, 1966). A micronutrient mixture containing Zn, B, Cu, Mg and N at 9.5, 2.6, 1.2, 2.4 and 0.46 per cent respectively was developed. Application of this mixture at 0.5 per cent in bhindi variety Varsha Uphar increased the growth and yield of crops (Mini, 2015).

El-Magid *et al.* (2000) reported that application of nitrogen, phosphorus and potassium along with foliar spray of micronutrient mixture (Fe, Mn, Cu, Zn, B, Mg, S and Mo) enhanced yield of rice.

Application of 0.1 per cent micronutrient mixture containing B, Fe, Cu, Co, Zn and Mn increased the grain yield of rice and reduced spikelet sterility (Shueadshen, 1991).

Foliar application of humic acid and zinc sulphate at 20 ppm and 25 ppm respectively produced higher yield in paddy (Durairaj, 1993). In cotton highest yield was recorded by the application of magnesium sulphate at 1 % and zinc sulphate at 0.5 % during squaring, flowering and boll formation (Katkar, 2005).

Yield of okra is increased by the foliar application of multi-micronutrient mixture (Patel *et al.*, 2009). Micronutrient mixture containing Zn, Cu, B and Fe chelated with organic components sprayed at 2 per cent increased height, leaf area and yield of okra (Datire *et al.*, 2010).

Foliar application of Agromin (Fe chelated with Zn, Cu, Mn, Mg, B and Mo) increased the growth of okra (Suryanarayana and Rao, 1981). In pumpkin fruit and seed yield was found to be increased by the foliar spray of zinc and manganese (Yousefi and Zandi, 2012).

Mishra *et al.* (2012) studied the effect of foliar spray of micronutrient mixture containing Zn, B, Fe, Mn, Cu at 100 ppm each and Mo at 50 ppm in tomato. He reported increase in number of fruits per plant and thus enhancing the yield.

Sivaiah *et al.* (2013) reported that combination spray of micronutrients or single micronutrient spray increases the plant height, fruit weight, number of branches and seed yield in tomato.

Selvi and Thiageshwari (2002) reported that micronutrient leaf nutrition increased the fruit yield of bhindi. Foliar nutrition of zinc sulphate at 0.5 % and borax at 0.2 % produced highest shoot length, number of internodes and more number of leaves per shoot of grapes (Singaram and Prabu, 2001).

Micronutrient mixture containing zinc sulphate + iron sulphate + Borax sprayed over pomegranate and found out an increase in fruits per plant and fruit weight (Afria *et al.*, 1999). Leaf length, number of leaves and floret of gladiolus was increased by the foliar spray of zinc at 0.5 per cent (Katiyar *et al.*, 2012).

Application of boron as foliar spray increased the plant height, leaf number and root weight in tomato (Verma *et al.*, 1973). Tomato cv. Dhanashree recorded highest number of fruits when 50 ppm boron was sprayed (Mishra *et al.*, 1990).

Foliar spray of boron and zinc each at 2 % and 6 % respectively increased plant height by 30 per cent compared to the control in tomato (Ejaz *et al.*, 2011). Foliar application of 1 % iron sulphate in onion showed increase in yield and plant height (Mishra *et al.*, 1990).

Plant height and root length of green gram plants treated with Fe was found to be increased (Prakash, 1998). Application of sulphur along with zinc and iron increased the growth, yield and nutrient uptake of safflower (Ravi *et al.*, 2008).

Foliar spray of 1 per cent micronutrient mixture at 30, 40 and 50 DAS increased plant height, tiller number and nutrient uptake in rice. The micronutrient mixture was comprised of 4 % Fe, 1 % Mn, 6 % Zn, 0.5 % Cu and 0.5 % B (Patel *et al.*, 2008).

Foliar spray of borax at 0.5 % in potato at 35 days after planting increased the nutrient concentration in tuber and also increased the nutrient uptake of crops (Das, 1977). Application of micronutrients such as Fe, Mn, Zn, B, Mo and Cu increased the uptake of N,P,K,Ca and Mg in soybean (Thiyageshwari and Ramanathan, 2001).

Groundnut plants treated with molybdenum at the rate of 4 g kg⁻¹ seed increased the yield of pod by 131.9 per cent compared to the control (Wankhade *et al.*, 1992). Battacharya *et al.* (1997) showed that the combined application of Mo at 3.0 kg/ha with S and Zn increased the dry weight of groundnut plants by 117 %.

Khanal *et al.* (2005) and Bhagiya *et al.* (2005) studied the effect of molybdenum in chickpeas, mungbean and groundnuts. They recorded a significant increase in pod yield and seed yield.

Gad (2012) recorded that application of molybdenum significantly increased the minerals composition such as N, P, K, Fe, Mn, Zn, Cu and Mo in groundnut seeds under different levels of nitrogen (25, 50, 75 and 100 %) compared with untreated seeds.

Groundnut treated with Mo at the rate of 2 to 4 g per kg seeds showed increased number of nodules and dry weight per plant (Wankhade *et al.*, 1992). Combined spray of micronutrients such as cobalt, boron and molybdenum enhanced

the uptake of N, P and S in pea (Singh *et al.*, 2012). It also increased the number of nodules and weight of nodules.

2.11 MICRONUTRIENT APPLICATION IN COWPEA YIELD, NODULATION, NUTRIENT UPTAKE

Foliar application of boron in cowpea showed significant increase in number of pods per plant and pod yield per plant which was recorded as 26 per cent and 13 per cent respectively and was found to be higher over the control (Chatterjee and Bandyopadhyay, 2017).

Foliar spray of 0.5 per cent FeSO_4 and 0.5 per cent ZnSO_4 at 45 days after sowing show increased seed yield in cowpea plants by 43.09 per cent compared to the control. It is then followed by combination spray of 0.5 per cent FeSO_4 and 0.5 per cent ZnSO_4 at 25 days after sowing (40.14 %). Benefit cost ratio also followed the same (Anitha *et al.*, 2005).

Foliar spray of three different concentrations (0, 1 and 2 ppm) of micronutrients such as Fe, B and Zn were sprayed at an interval of every 15 days in cowpea. Nutrient concentration and seed protein content was found to be highest in 1 ppm spray. Foliar spray with iron showed greater nutrient uptake and protein percentage in seeds compared to others (Salih, 2013).

In cowpea maximum nodulation was obtained in soils applied with 5 kg ha^{-1} and 20 kg ha^{-1} boron and manganese respectively (Rhoden and Allen, 1982). Cowpea plants supplied with Mo as spray recorded increased pod dry weight by 271.7 per cent compared to the control. Same trend was observed in soybean which showed an increase in pod weight by 258.3 per cent (Vargas and Ramirez, 1989).

Niranjana *et al.* (2005) stated that molybdenum at 2.0 g kg^{-1} seed recorded the highest growth parameters and pods yield in cowpea compared the control. Molybdenum application in cowpea plants resulted in increased number of nodules and dry weight. It also enhanced the rate of nitrogen fixation (Krishnappa *et al.*,

1992 and Noor *et al.*, 1997). Application of molybdenum on cowpea plants results in enhanced accumulation of nutrients in plant tissues (Hristozkova *et al.*, 2006).

Francois (1989) studied the effect of boron on growth of crops. He observed that the relative yield of *P. vulgaris* decreased by 12.1 per cent and seed yield of cowpea was decreased by 11.5 % with each unit increase of boron in soil solution above 1 and 2.5 mg B per litre, respectively.

Application of ZnSO₄ at the rate of 25 kg per ha through soil increased the seed yield of cowpea by 43 per cent followed by foliar spray of 0.5 per cent Zn at 25 and 45 days after sowing. Number of pods per plant and number of branches were increased resulting in yield enhancement (Patel *et al.*, 2011).

Cowpea seeds treated with *Bradyrhizobium japonicum* showed significant increase in the uptake of micronutrients by shoot (Zn = 47.6 %, Cu = 46.8 %, Fe = 24.3 % and Mn = 56.3 %), root (uptake of Zn, Cu, Fe and Mn by 54.9 %, 26.3 %, 63.7 %, and 34.8 % respectively) and pod (Zn = 36.5 %, Cu = 16.1 %, Fe = 72.5 % and Mn = 26.3 %) (Nyoki and Ndakidem, 2014).

According to Vieira *et al* (1998) application of molybdenum restricts the nodules senescence and thus maintaining a longer period of effective N₂ fixation. Foliar spray of molybdenum (0 and 40 kg ha⁻¹) and nitrogen as side dressing increased the total shoot nitrogen in common bean.

2.12 MICRONUTRIENTS IN PEST AND DISEASE CONTROL

Seeds treated with micronutrient showed resistance to soil borne pathogens in soybean. Higher the zinc content in seeds resist diseases during germination and development of seedlings (Marschner, 1995).

Essential nutrients influence the severity of diseases in plants (Huber and Graham, 1999). Optimum level of nutrients availability to plants is a better way to reduce the incidence of pest and disease in an integrated system of pest management (Graham and Webb, 1991).

Fe, Mn, Zn, Cu and B are deficient in most places of India. This results in deficiency disorders in plants. Application of nutrients to correct these disorders helps in developing resistance to diseases (Agrios, 2005).

Providing sufficient nutrients for plants reduces incidence of pest and diseases (Huber and Wilhelm, 1988). Sufficient amount of boron in plants reduces the chance of infestation while deficiency make plants susceptible (Gupta, 1993; Graham and Webb, 1991).

Manganese content reduces disease incidence. Micronutrients affects the severity of pathogens in host (Engelhard, 1990; Graham and Webb, 1991; Huber, 1980). Deficiency of micronutrients enhances disease severity (Engelhard, 1990; Graham and Webb, 1991; Huber, 1980; Fageria *et al.*, 1997; Baligar *et al.*, 1998).

Foliar spray of micronutrient solution on cucumber is a prophylactic method to avoid powdery mildew development (Reuveni *et al.*, 1998). Manganese fertilization in wheat reduced the incidence of powdery mildew and take-all (Simoglou and Dordas, 2006). Soil application of zinc reduced root rot in wheat (Grewal *et al.*, 1996).

Iron nutrition increases the vigour of plants and also inhibits the growth of pathogens in the rhizosphere. Iron nutrition reduced the severity of rust and smut in wheat (Graham and Webb, 1991; Graham, 1983).

Downey mildew of grapes can be controlled by soil application of Cu fungicides (Evans *et al.*, 2007). Ascochyta blight in beans and peas can be controlled by the nutrition of Mo in plants (Patil, 1981). In cucumber powdery mildew can be controlled by foliar application of borax, copper sulphate and manganese chloride (Reuveni *et al.*, 1998).

Materials and Methods

3. MATERIALS AND METHODS

An investigation entitled “Evaluation of micronutrient formulation in cowpea (*Vigna unguiculata* L. Walp)” was carried out at College of Agriculture, Padannakkad and Regional Agricultural Research Station (RARS) farm, Pilicode during 2017 to 2019. The objectives of the study were to evaluate the effect of micronutrient formulation through seed treatment and foliar nutrition on growth, nodulation and yield of cowpea (*Vigna unguiculata* L. Walp) and to study its effect on plant nutrient uptake and residual soil nutrient status. The study was conducted in three parts.

Part I: Standardization of micronutrient formulation for cowpea

Part II: Studies on time and dose of micronutrient formulation for seed treatment in cowpea

Part III: Field experiment to evaluate the micronutrient formulation in cowpea

3.1 STANDARDIZATION OF MICRONUTRIENT FORMULATION FOR COWPEA

Micronutrient formulation was standardized by Smt. Premalatha (2016), COA Padannakkad in tissue culture banana as part of her MSc. thesis work. It was then modified and standardized for bhindi by Ashwini (2018), MSc., COA, Padannakkad. Micronutrient formulation is a mixture of two solutions viz. solution A and solution B. Solution A is a mixture of zinc sulphate, boric acid, copper sulphate, manganous sulphate, ferrous sulphate and ammonium molybdate whereas solution B is an organic acid which act as a chelate. These two solutions have to be mixed in a specific ratio at the time of application because of the poor shelf life of micronutrient mixture. Attempt was being made to increase the shelf life of this micronutrient formulation in the combined form of both solutions A and B. Several trials were done and a combination of Solution A and Solution B were prepared so that it can be supplied to farmers as micronutrient mixture which reduces the

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difficulty of mixing solutions during field application. The micronutrient formulation can be diluted to different concentrations for seed treatment and foliar spray.

3.2 SEED TREATMENT STUDY

A laboratory study was carried out at COA, Padannakkad with different concentrations of micronutrient formulation to standardize the time and dose for seed treatment. The experiment was carried out in completely randomized design with seven treatments and three replications. Seeds treated with different concentrations of micronutrient formulation were kept in petri plates for germination studies which was then planted in pots. Each replication contained four plants and the total number of plants were 84. Soil, sand and cowdung were mixed uniformly in the ratio 1:1:1 to prepare the potting mixture. Plants were grown in the pots and observations were recorded upto three leaf stage.

3.2.1 Experimental details

Crop : Cowpea
Variety : PGCP 6 (Pant Lobia 3)
Spacing : 30 cm x 15 cm
Design : CRD
Treatments : 7
Replications : 3

3.2.2 Treatment concentrations

T₁ : Micronutrient formulation at the rate of 0.25 %
T₂ : Micronutrient formulation at the rate of 0.50 %
T₃ : Micronutrient formulation at the rate of 0.75 %

- T₄ : Micronutrient formulation at the rate of 1 %
T₅ : Micronutrient formulation at the rate of 1.5 %
T₆ : Micronutrient formulation at the rate of 2 %
T₇ : Control (Seed treatment with water)

3.2.4 Biometric observations

The biometric observations included germination percentage, days taken for germination, seedling length at three leaf stage and seedling vigour index were recorded at 15 days after planting.

3.2.4.1 Germination percentage (%)

Germination percentage of cowpea seeds was noted by keeping it in moist filter paper.

3.2.4.2 Days taken for germination

Days taken for germination were recorded by counting the number of days taken for whole seeds to germinate.

3.2.4.3 Seedling length at three leaf stage (cm)

Plant height was measured from base of the plant to tip at two weeks after planting.

3.2.4.4 Seedling vigour index

Seedling vigour index was calculated by multiplying germination percentage with seedling length.

3.3 FIELD EXPERIMENT

The field experiment was carried out at Regional Agricultural Research Station (RARS), Piliocode to evaluate the effect of micronutrient formulation in cowpea. The experiment was conducted in randomized block design with 12

treatments replicated three times. The treatment consisting of combination of four levels of seed treatment (no seed treatment, seed treatment with rhizobium, seed treatment with best concentration from experiment 1 and second best seed treatment from experiment 1) and three levels of foliar application of micronutrient (no foliar application, one foliar application at 15 DAS and two foliar application at 15 and 30 DAS). Manures and fertilizers application and other cultural practices were followed as per POP, KAU (2016) for all the treatments uniformly.

3.3.1 Location and climate of area

Geographically, the experimental site was located at 12° 12' N latitude and 75° 10' E longitude and at an altitude of 15 m above mean sea level. The region mostly experiences a warm humid tropical climate.

3.3.2 Experimental site

The soil type of experimental field was red loam. Land is mainly used for cultivation of vegetables and tuber crops. Cassava was cultivated in the plot before the commencement of experiment.

3.3.3 Design and layout

The layout of the field is given in Plate 1. The details of experiment are given below.

Crop	: Cowpea
Variety	: PGCP 6 (Pant Lobia 3)
Spacing	: 30 cm x 15 cm
Plot size	: 3 m ²
Design	: Factorial RBD
Treatments	: 12
Replications	: 3
Date of sowing	: 20 th October 2018
Date of first harvest	: 24 th December 2018

EXPERIMENTAL LAYOUT

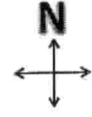
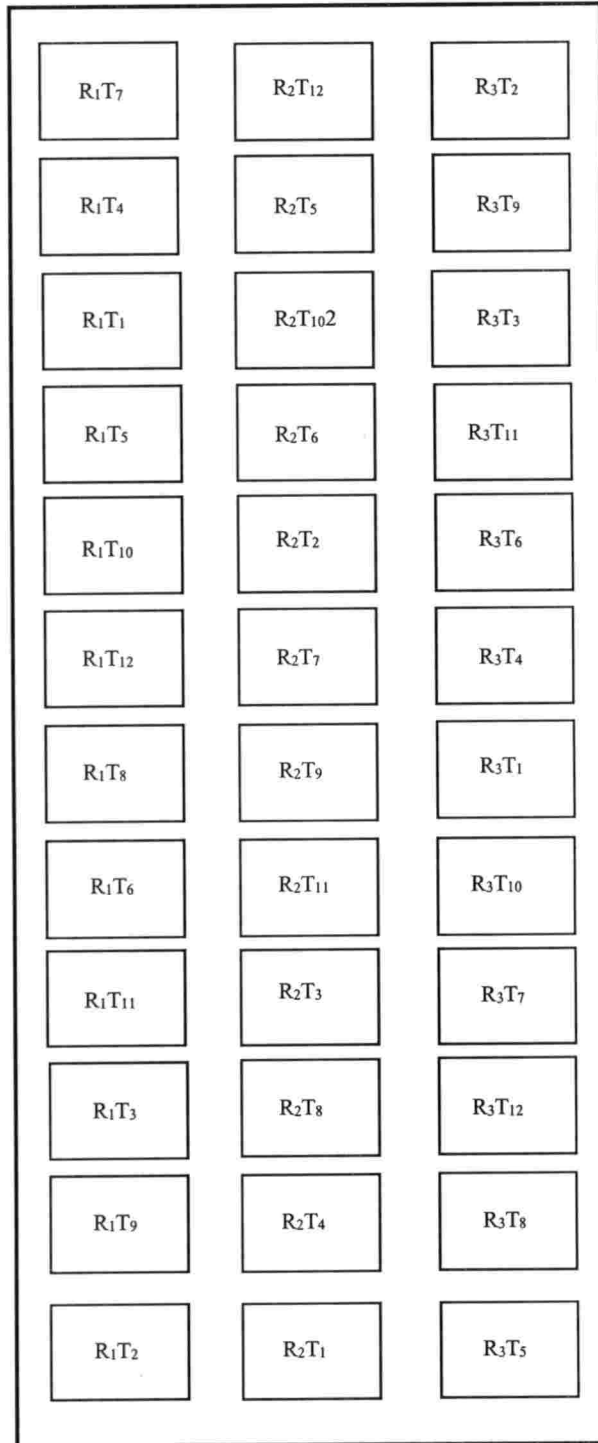


Plate: 1 – Layout of the field experiment

3.3.4 Treatments

Factor A : Seed treatment

S₁ : No seed treatment

S₂ : Seed treatment with rhizobium

S₃ : Seed treatment with micronutrient formulation (best seed treatment from experiment 1)

S₄ : Seed treatment with micronutrient formulation (second best seed treatment from experiment 1)

Factor B : Foliar application

F₁ : No foliar application

F₂ : One foliar application at 15 DAS

F₃ : Two foliar application at 15 and 30 DAS

Treatment details

T₁ : No seed treatment and no foliar application (S₁F₁)

T₂ : No seed treatment + one foliar application (S₁F₂)

T₃ : No seed treatment + two foliar application (S₁F₃)

T₄ : Seed treatment with rhizobium + no foliar application (S₂F₁)

T₅ : Seed treatment with rhizobium + one foliar application (S₂F₂)

T₆ : Seed treatment with rhizobium + two foliar application (S₂F₃)

T₇ : Seed treatment with best treatment from experiment 1 + no foliar application (S₃F₁)

T₈ : Seed treatment with best treatment from experiment 1 + one foliar application (S₃F₂)

T₉ : Seed treatment with best treatment from experiment 1 + two foliar application (S₃F₃)

T₁₀ : Seed treatment with second best treatment from experiment 1+ no foliar application (S₄F₁)

T₁₁ : Seed treatment with second best treatment from experiment 1+ one foliar application (S₄F₂)

T₁₂ : Seed treatment with second best treatment from experiment 1+ two foliar application (S₄F₃)

3.3.5 Crop cultivation details

3.3.5.1 Land preparation

A fine tilth was made by thorough ploughing of the experimental field. The stubbles were removed and field was laid out as shown in plate 1. Lime was applied during first ploughing at the rate of 250 kg ha⁻¹ based on the soil pH. Raised beds were prepared at a height of 30 cm with each plots having 2 m length and 1.5 m width. Each plots were separated by 40 cm space.

3.3.5.2 Manuring and fertilizer application

FYM was applied at the rate of 20 t ha⁻¹ and thoroughly mixed before sowing. Macro nutrient fertilizers were applied as per POP recommendation for cowpea. Nitrogen was applied at the rate of 20 t ha⁻¹ in two split doses first half at final ploughing and the next half at 15 DAS. Phosphorus and potassium were applied basally at the rate of 30 and 10 t ha⁻¹ respectively. Micronutrient formulation was sprayed according to the treatments.

3.3.5.3 Seed rate and sowing

Seeds of cowpea variety PGCP – 6 (Pant lobia 3) released by Pantnagar Agricultural University were used in the experiment. It is a short duration cowpea variety with high yield potential. Seeds were sown in furrows taken in the bed at the rate of 40 kg ha⁻¹ with a spacing of 30 cm between rows and 15 cm between plants.

3.3.5.4 Irrigation and after cultivation

Crop was irrigated throughout the growth and hand weeding was done at regular intervals.

3.3.5.5 Plant protection

Nimbecidine was sprayed to control thrips during initial growth period. Copper oxychloride 50 WP was drenched against collar rot.

3.3.5.6 Harvesting

Pods were harvested at fully matured stage. First harvesting was done at 65 DAS.

3.4 BIOMETRIC OBSERVATIONS

Biometric observations like plant height, number of branches, stem diameter were taken at flower bud initiation and at harvest. Number of nodules per plant, fresh weight and dry weight of nodules and number of days taken for first flowering were recorded. Number of pods per plant, number of seeds per pod, pod weight per plant, length of pod, dry matter per plant and root CEC were taken at harvest.

3.4.1 Plant height (cm)

Plant height was measured from base of the plant to tip of main shoot at flower bud initiation stage and at harvest using a meter scale.

3.4.2 Number of branches

Number of branches were recorded at flower bud initiation stage and at harvest.

3.4.3 Stem diameter (cm)

Stem diameter was taken at flower bud initiation stage and at harvest using a measuring tape from five index plants and the average value was noted.

3.4.4 Number of nodules per plant

Number of nodules per plant was counted at 30 DAS. Three plants were randomly uprooted from each plot and it was then washed to remove the soil and the count was taken.

3.4.5 Fresh weight of nodules (g)

Fresh weight of nodules was taken soon after separation of the nodules from roots.

3.4.6 Dry weight of nodules (g)

The nodules were kept in petri plates according to treatments and placed in an oven for 5 days at 50 degree celcius. Dry weight was recorded when the weight attains a constant value.

3.4.7 Number of days taken for first flowering

It is the day at which first flower emergence take place from date of sowing.

3.4.8 Number of pods per plant

The number of pods obtained from five index plant from each plot were recorded and the average was taken to obtain the number of pods per plant.

3.4.9 Number of seeds per pod

Ten pods from index plants were selected randomly and the number of seeds from each pod were counted and the average was taken as the number of seeds per pod.

3.4.10 Pod weight per plant (g)

Total pod weight of ten plants selected as index plants were recorded and the average value was taken as the pod weight per plant.

3.4.11 Length of pod (cm)

Ten pods were randomly selected from the index plants and measured the length using a meter scale. The mean value was taken as the length of pod.

3.4.12 Grain yield (kg ha⁻¹)

Total grain yield from each plot was recorded and the yield was calculated in kg per plot and yield in kg per hectare were calculated

3.4.12 Dry matter production (kg ha⁻¹)

Five plants were uprooted from each plot and fresh weight was recorded. The plants were kept for shade drying and then oven dried at 65° C. Dry weight was recorded to find out the dry matter production per plant which was then used for calculating dry matter production in one hectare.

3.4.13 Root CEC

The CEC of root was determined as per the standard procedure given by Mitsui and Ueda in 1963. Roots were washed in distilled water and shaken with potassium chloride and titrated against NaOH.

3.5 SOIL ANALYSIS

3.5.1 Initial soil analysis

Initial soil sample was collected from the prepared field for analysis at different places randomly and it is then pooled and reduced to 500 g and air dried. The air dried sample was sieved with 2 mm sieve and stored in an air tight container.

The soil samples were analyzed for pH, EC, organic carbon, CEC, bulk density, particle density, available macronutrients such as N, P, K and micronutrients such as Fe, Mn, Zn, Cu, B and Mo following the standard procedures in Table 1. Initial soil properties are given in Table 2.

3.5.2 Experimental soil analysis

Soil samples for laboratory analysis were collected from each plot at 45 DAS and at harvest. The samples were air dried, ground, sieved with 2 mm sieve and stored in air tight container. They were analyzed for pH, EC, organic carbon, available nutrients such as N, P, K, Fe, Mn, Zn, Cu, B and Mo as per the standard procedures as given in Table 1.

3.6 LEAF ANALYSIS

Leaf samples were collected at harvest and analyzed for nutrients such as N, P, K, Ca, Mg, S, Fe, Mn, Zn, Cu, B and Mo using standard procedures given in Table 3.

3.7 GRAIN ANALYSIS

Fully matured grains from each plots were collected and dried in an oven at 60 ° C and powdered for the analysis of N, P, K, Ca, Mg, S, Fe, Mn, Zn, Cu, B and Mo using standard procedures given in Table 3.

3.8 MICROBIOLOGICAL OBSERVATION

3.8.1 Soil biomass carbon

Soil biomass carbon was determined by fumigation extraction method as per the procedure put forward by Jenkinson and Poulson (1976). Fumigation of the collected soil samples were done with ethylene free chloroform in vacuum desiccator for 24 hours.

3.9 INCIDENCE OF PEST AND DISEASE

There was no severe incidence of pest and disease observed in the field. Crops were infested with thrips during initial growth period and was controlled by the application of nimbecidine. A few plants were infected with collar rot which was controlled by drenching of copper oxychloride.

3.10 BENEFIT COST RATIO

Benefit cost ratio was calculated using the formula

$$BCR = \frac{\text{Gross income}}{\text{Total expenditure}}$$

3.11 STATISTICAL ANALYSIS

The data obtained from the seed treatment study and field experiment were analyzed statistically using Analysis of Variance (ANOVA) for completely randomized design and factorial randomized block design proposed by Panse and Sukhatme in 1985. It was then tested for its significance using standard statistical tools.

Table 1 Analytical methods followed for soil analysis

Sl.No	Parameters	Method	Reference
1	pH	pH meter	Jackson (1958)
2	EC	Conductivity meter	Jackson (1958)
3	Organic carbon	Chromic acid wet digestion method	Walkley and Black (1934)
4	Bulk density	Undisturbed core sample	Black <i>et al.</i> (1965)
5	Textural analysis	International pipette method	Robinson (1922)
6	Available N	Alkaline permanganate method	Subbiah and Asija (1956)
7	Available P	Bray extraction and photoelectric colorimetry	Jackson (1958)
8	Available K	Flame photometry	Pratt (1965)
9	Available Fe	Atomic absorption spectroscopy	Sims and Johnson (1991)
10	Available Mn	Atomic absorption spectroscopy	Sims and Johnson (1991)
11	Available Zn	Atomic absorption spectroscopy	Emmel <i>et al.</i> (1977)
12	Available Cu	Atomic absorption spectroscopy	Emmel <i>et al.</i> (1977)
13	Available B	Photo electric colorimetry	Bingham (1982)
14	Available Mo	Inductively coupled plasma optical emission spectrometry (ICP-OES)	Soltanpour and Schwab (1977)

Table 2 Properties of initial soil sample

S.No	Parameter	Value
I. Physical properties		
1.	Bulk density (g cm^{-3})	1.32
2.	Particle density (g cm^{-3})	2.55
II. Textural composition		
1.	Sand (%)	77.4
2.	Silt (%)	19.6
3.	Clay (%)	3.75
4.	Textural class	Loamy sand
III. Chemical properties		
1.	pH (1:2.5)	4.36
2.	EC (dS m^{-1})	0.122
3.	Organic carbon (%)	0.65
4.	Organic matter (%)	1.11
5.	CEC ($\text{meq } 100\text{g}^{-1}$)	7.45
6.	Available N (kg ha^{-1})	228.45
7.	Available P (kg ha^{-1})	41.2
8.	Available K (kg ha^{-1})	121.45
9.	Available Zn (mg kg^{-1})	0.98
10.	Available B (mg kg^{-1})	0.20
11.	Available Fe (mg kg^{-1})	65.34
12.	Available Cu (mg kg^{-1})	1.62
13.	Available Mn (mg kg^{-1})	12.30
14.	Available Mo (mg kg^{-1})	0.002

Table 3 Analytical methods followed for leaf and grain analysis

S.No	Parameter	Method	Reference
1.	Total N	Modified kjeldhal digestion method	Jackson (1958)
2.	Total P	Vanadomolybdate yellow colour method	Piper (1966)
3.	Total K	Flame photometry	Jackson (1958)
4.	Total Ca	Atomic absorption spectroscopy	Issac and Kerber (1971)
5.	Total Mg	Atomic absorption spectroscopy	Issac and Kerber (1971)
6.	Total S	Turbidimetric method	Bhargava and Raghupathi (1995)
7.	Total Zn	Atomic absorption spectroscopy	Emmel <i>et al.</i> (1977)
8.	Total B	Azomethane - H colorimetric method	Bingham (1982)
9.	Total Fe	Atomic absorption spectroscopy	Piper (1966)
10.	Total Cu	Atomic absorption spectroscopy	Emmel <i>et al.</i> (1977)
11.	Total Mn	Atomic absorption spectroscopy	Piper (1966)
13.	Total Mo	Inductively coupled plasma optical emission spectrometry (ICP-OES)	Soltanpur and Schwab (1977)



Plate 2 A: Seeds soaked in micronutrient formulation



Plate 2 B: Different stages of growth of seeds

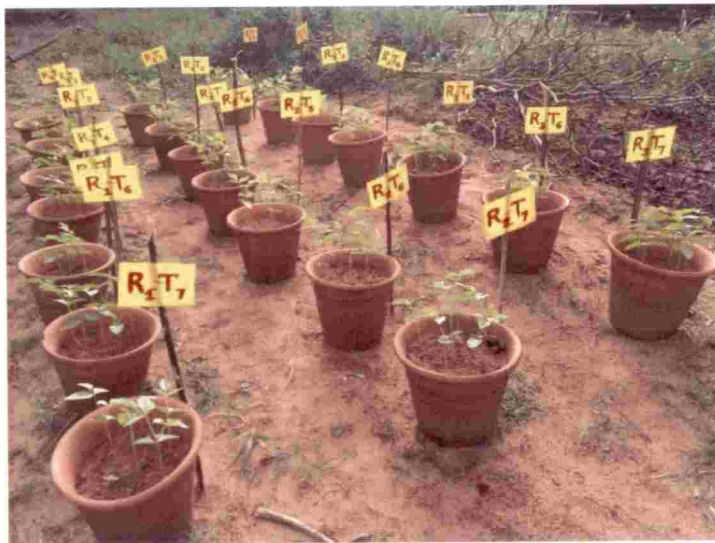


Plate 2 C: Cowpea plants at three leaf stage

Plate 2: Seed treatment study, COA, Padannakkad



Plate 3 A: Field layout



Plate 3 B: Bed preparation



Plate 3 C: Sowing of seeds



Plate 3 D: Germination stage



Plate 3 E: Crop at 15 DAS



Plate 3 F: Crop at 30 DAS

Plate 3: Experimental plot at RARS, Pilicode



Plate 4 A: Flowering and pod formation



Plate 4 B: Harvesting of pods



Plate 4 C: Harvested pods



Plate 4 D: Cowpea grain



Plate 4 E: Field at harvest stage

Plate 4: Experimental plot at harvest stage

Results

4. RESULTS

The results obtained from the experiment entitled “Evaluation of micronutrient formulation in cowpea (*Vigna unguiculata* L. Walp)” was carried out in RARS, Pilicode. The data obtained were subjected to statistical analysis and the results are given in this chapter.

4.1 STANDARDIZATION OF MICRONUTRIENT FORMULATION FOR COWPEA

A micronutrient mixture in liquid form was formulated and standardized for tissue culture banana by Premalatha (2016), an MSc student at COA Padannakkad. Later it was standardized for bhindi by Ashwini (2018) as part of her MSc research work. The mixture is a combination of two solutions A and B in which solution A contains a mixture of micronutrient salts namely zinc sulphate (10 %), boric acid (2 %), copper sulphate (4 %), manganous sulphate (0.1 %), ferrous sulphate (2 %) and ammonium molybdate (0.05 %). Solution B is an organic chelating agent. The objective of this experiment was to standardize the micronutrient formulation for cowpea in seed treatment as well as foliar spray. The micronutrient mixture standardized for vegetables showed less shelf life and forms precipitate when solution A and solution B were mixed together. This was found to be the major drawback of the formulation for banana and vegetables. Several trials were conducted to make a formulation having no precipitate formation while mixing solutions A and B. Thus at a specific combination of solution A and Solution B the mixture was found to be stable and tested for its suitability on cowpea as foliar spray (Plate 4).

Composition of Solution A (1 litre)

ZnSO ₄ .7H ₂ O	- 50 g
CuSO ₄ .5H ₂ O	- 20 g
FeSO ₄ .7H ₂ O	- 10 g

H ₃ BO ₃	- 10 g
MnSO ₄ .H ₂ O	- 0.5 g
(NH ₄) ₆ Mo ₇ O ₂₄ .4H ₂ O	- 0.5 g

4.2 SEED TREATMENT STUDY

4.2.1 Standardization of time and dose of micronutrient formulation for seed treatment of cowpea

Micronutrient mixture at different concentrations at the rate of 0.25, 0.50, 0.75, 1, 1.5 and 2 percentages were used for seed treatment study in cowpea. To standardize the time for seed treatment several trials were conducted. Cowpea seeds were treated with micronutrient mixture both at different concentrations of micronutrient mixture for different durations (1 hr, 2hrs, 4 hrs, 6hrs, 8 hrs, 10 hrs and 12 hrs). Observations on seed germination and its emergence showed that the seeds treated with micronutrient mixture for 6 hours showed better establishment as compared to other duration of seed treatment. In the case of cowpea seeds treated for longer duration, the germination and seedling emergence was adversely affected (Plate 5). Thus time and concentration of micronutrient mixture for cowpea seed treatment were standardized. Based on this a seed treatment study was carried out in cowpea and the following biometric observations were recorded.

4.2.2 Biometric observations from seed treatment study

4.2.2.1 Germination percentage (%)

The data obtained on the germination percentage of cowpea seeds is given in the Table 4. The observations were found to be non significant in all the treatments with 100 per cent germination occurred in T₅ (micronutrient mixture @ 1.5 %) and T₆ (micronutrient mixture @ 2 %).

4.2.2.2 Days taken for germination

Number of days taken for the germination of cowpea seeds are given in the Table 4. Treatments did not show significant difference in the days taken for germination.

4.2.2.3 Seedling length at three leaf stage (cm)

Seedling length at three leaf stage is given in the Table 4. Treatment T₆ (24.23) showed maximum seedling length followed by T₅ (22.31) and there was significant difference between the two treatments. Control recorded the lowest value in T₇ (18.75).

4.2.2.4 Seedling vigour index

Seedling vigour index data obtained from seed treatment study is given in the Table 4. Treatment T₆ (2423) showed highest seedling vigour index which was significantly superior to all other treatments. Treatment T₅ showed the second highest seedling vigour index (2231) and the lowest value was recorded in control (1814.25).

The observations of seed treatment study showed that the treatment T₆ (micronutrient mixture @ 2 %) was identified as the best treatment and T₅ (micronutrient mixture @ 1.5 %) as the second best treatment for evaluating the performance of micronutrient formulation in field trial of cowpea.

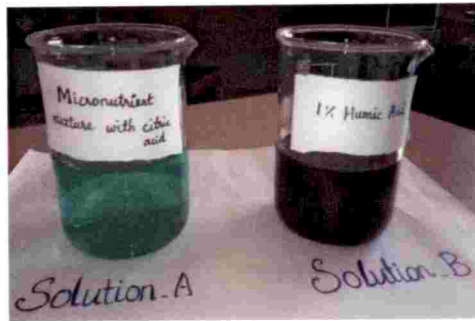


Plate 5 A: Micronutrient mixture for TC banana



Plate 5 B: Precipitate formation on mixing Solution A and B



Plate 5 C: Solution A with Solution B without precipitation

Plate 5: Preparation of micronutrient mixture for cowpea

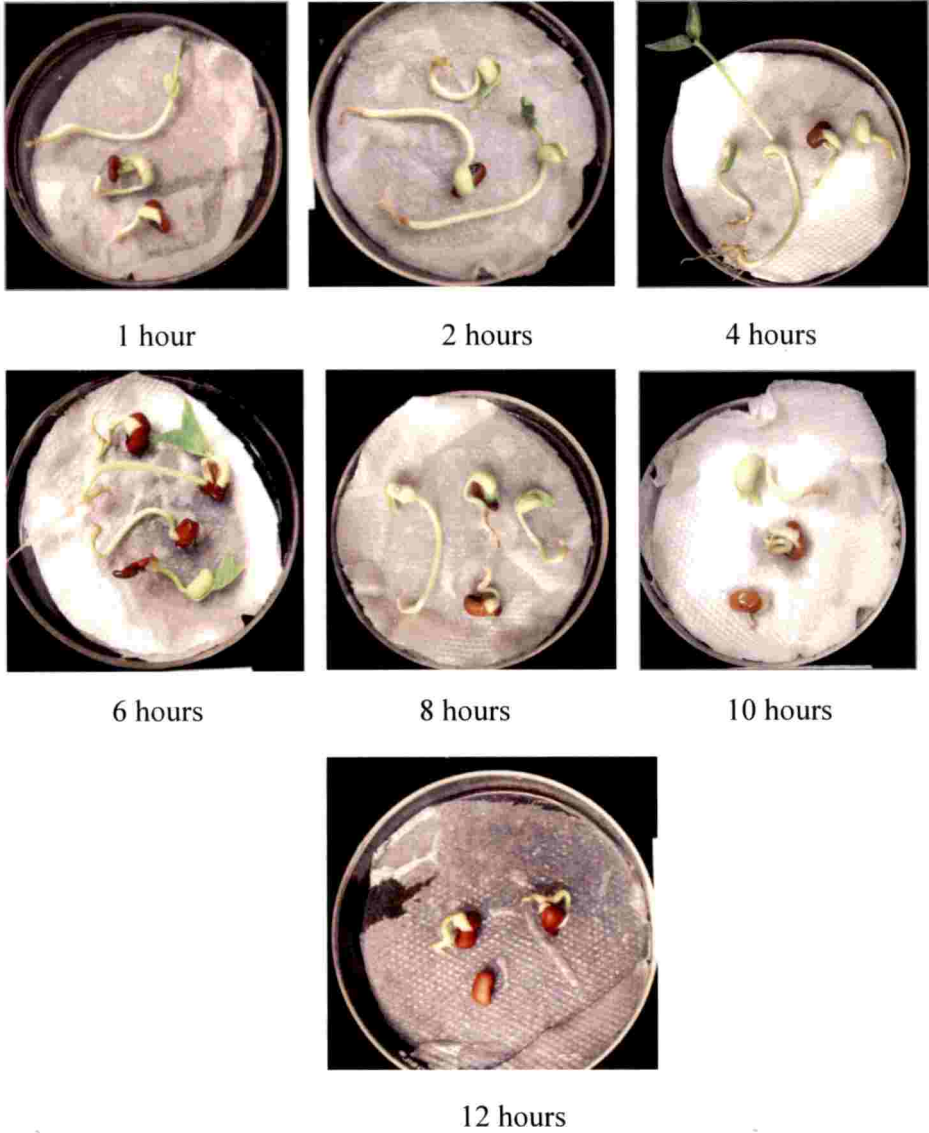


Plate 6: Standardization of time for seed treatment in cowpea using 2% mixture

Table 4: Effect of seed treatment with micronutrient formulation on germination percentage, days taken for germination, seedling length at three leaf stage and seedling vigour index

Treatment	Germination percentage (%)	Days taken for germination	Seedling length at three leaf stage (cm)	Seedling vigour index
T ₁	96.70	1.66	18.80	1817.96
T ₂	96.76	1.66	19.81	1881.95
T ₃	97.77	1.67	19.84	1939.75
T ₄	97.77	1.66	19.94	1949.53
T ₅	100.00	1.33	22.31	2231.00
T ₆	100.00	1.00	24.23	2423.00
T ₇	95	1.77	18.75	1814.25
SE(m)	2.22	0.28	0.40	45.38
C.D.	NS	NS	1.24	138.99

4.3 FIELD EXPERIMENT

4.3.1 Observations on growth and yield characters of cowpea

4.3.1.1 Plant height (cm)

The data on plant height was recorded at the time of flower bud initiation and at harvest. The effect of seed treatment, foliar spray and their interactions on plant height is recorded in the Table 5.

Levels of seed treatment showed significant difference in plant height at flower bud initiation and at harvest. At flower bud initiation S₃ (seed treatment with 2 % micronutrient formulation) recorded maximum plant height (24.31) which was on par with seed treatment with 1.5 % micronutrient formulation, S₄ (23.53). At the time of harvest S₃ recorded maximum height of 39.75 cm which was significantly superior to all other treatments.

Levels of foliar spray of micronutrients showed significant difference in plant height at flower bud initiation and at harvest. Maximum height was observed in F₃ treatment (foliar spray at 15 DAS and 30 DAS) both at flower bud initiation (25.98) and at harvest (41.08).

In the interaction effect maximum plant height was recorded in the treatment S₃F₃ (seed treatment with 2 % micronutrient formulation + two foliar spray) at both the stages given as 29.80 and 44.13, which were significantly different from all other treatments.

4.3.1.2 Number of branches

The data on number branches recorded at flower bud initiation and at harvest with respect to various treatments are recorded in the Table 5.

Significant difference was not observed in the number of branches at flower bud initiation and at harvest in seed treatment, foliar spray and their interaction.

4.3.1.3 Stem diameter (cm)

There was significant difference in stem diameter among the treatments. The results are recorded in the Table 5.

Stem diameter showed significant difference with variation in seed treatment. Maximum diameter was observed in treatment S₄ (2.44) at flower bud initiation which was on par with S₃ (2.43) and seed treatment with rhizobium, S₂ (2.35). At the time of harvest treatment S₃ recorded the highest stem diameter (4.65) which was significantly different from all other treatments.

Levels of foliar spray showed significant difference in stem diameter at flower bud initiation. F₃ recorded maximum stem diameter (2.54) which was on par with single spray of micronutrient formulation, F₂ (2.40). At the time of harvest, F₃ (4.85) recorded maximum stem diameter which was significantly different from all other treatments.

Interaction effect of treatments also showed significant difference among the treatments. At flower bud initiation maximum stem diameter was recorded in seed treatment with rhizobium + two foliar spray, S₂F₃ (2.86) which was on par with seed treatment with 2 % micronutrient formulation + single foliar spray, S₃F₂ (2.66), seed treatment with 1.5 % micronutrient formulation + two foliar sprays, S₄F₃ (2.64) and seed treatment with 1.5 % micronutrient formulation + one foliar spray, S₄F₂ (2.62). At harvest S₂F₃ (5.30) showed maximum stem diameter which showed significant difference from all other treatments.

4.3.1.4 Number of nodules per plant

The data on number of nodules per plant are presented in the Table 6.

Levels of seed treatment showed significant difference in the number of nodules per plant. Maximum nodules per plant was observed in plots treated with rhizobium, S₂ (27.22). The least number of nodules was observed in no seed treatment, S₁ (14.77) which was on par with S₃ (18.88) and S₄ (19.66).

There was no considerable difference in the number of nodules with respect to foliar spray and the interaction effect of seed treatment and foliar spray. The results obtained was found to be non significant.

4.3.1.5 Fresh weight of nodules (g)

The effect of seed treatment, foliar spray and their interaction on fresh weight of nodules are given in the Table 6. Effect of seed treatment, foliar spray and their interaction on fresh weight of nodules was found to be non significant.

4.3.1.6 Dry weight of nodules (g)

The effect of seed treatment, foliar spray and their interaction on dry weight of nodules are given in the Table 6. Effect of seed treatment, foliar spray and their interaction on dry weight of nodules was found to be non significant

Table 5: Effect of seed treatment, foliar spray and their interactions on plant height, number of branches and stem diameter of cowpea (at flower bud initiation stage and harvest)

Treatment	Plant height (cm)		Number of branches plant ⁻¹		Stem diameter plant ⁻¹ (cm)	
	Flower bud initiation	Harvest	Flower bud initiation	Harvest	Flower bud initiation	Harvest
S ₁	20.86	35.97	9.87	9.97	2.11	4.16
S ₂	23.11	38.41	9.89	9.99	2.35	4.58
S ₃	24.31	39.75	10.34	10.45	2.43	4.65
S ₄	23.53	38.63	10.38	10.40	2.44	4.26
SEm (±)	0.27	0.35	0.17	0.16	0.06	0.01
CD (0.05)	0.81	1.05	NS	NS	0.17	0.05
Levels of foliar spray						
F ₁	21.26	36.57	9.85	9.89	2.06	3.91
F ₂	21.61	36.92	10.12	10.15	2.40	4.49
F ₃	25.98	41.08	10.10	10.15	2.54	4.85
SEm (±)	0.23	0.30	0.15	0.15	0.05	0.01
CD (0.05)	0.70	0.90	NS	NS	0.15	0.04
Interactions						
S ₁ F ₁	18.66	34.33	9.87	9.98	1.73	3.58
S ₁ F ₂	21.73	36.40	9.88	9.89	2.32	4.58
S ₁ F ₃	22.20	37.20	9.99	9.98	2.29	4.34
S ₂ F ₁	21.93	36.60	10.11	10.12	2.19	4.38
S ₂ F ₂	21.40	36.63	10.13	10.14	2.02	4.06
S ₂ F ₃	26.00	42.00	10.11	10.12	2.86	5.30
S ₃ F ₁	21.06	37.73	9.98	9.99	2.24	4.38
S ₃ F ₂	22.06	37.40	9.97	9.98	2.66	4.72
S ₃ F ₃	29.80	44.13	10.12	10.13	2.39	4.86
S ₄ F ₁	23.40	37.63	10.12	10.13	2.07	3.30
S ₄ F ₂	21.26	37.26	9.98	9.99	2.62	4.60
S ₄ F ₃	25.93	41.00	9.99	10.00	2.64	4.90
SEm (±)	0.47	0.61	0.30	0.30	0.10	0.02
CD (0.05)	1.40	1.81	NS	NS	0.30	0.08

S₁- no seed treatment; S₂- seed treatment with rhizobium; S₃- seed treatment with micronutrient formulation (best from Exp. 1); S₄- seed treatment with micronutrient formulation (second best from Exp. 1)

F₁- no foliar spray; F₂- one foliar application at 15 DAS; F₃- two foliar application at 15 & 30DAS

4.3.1.7 Number of days taken for first flowering

The data on days taken for first flowering of cowpea as influenced by seed treatment, levels of foliar application of micronutrients and their interactions are given in Table 6. Main treatments as well as their interactions did not show any significant difference in the number of days taken for first flowering. However, the minimum days for flowering in main treatments were observed as 31 days both in S₃ and F₂ treatments. The interaction effect was also non significant with maximum days (32 days) in control.

4.3.1.8 Number of pods per plant

The data on number of pods per plant influenced by seed treatment, levels of foliar application of micronutrients along with their interactions are given in Table 7.

Levels of seed treatment showed significant difference in the number of pods per plant with maximum number recorded in S₃ (21.02) which was on par with S₄ (19.44). The lowest number of pods per plant was observed in S₁ (16.73).

Levels of foliar spray showed significant difference in the number of pods per plant with maximum number recorded in F₃ (20.97). F₁ recorded the lowest number (16.71).

The interaction effect also showed significant difference in number of pods per plant with maximum number recorded in S₃F₃ (25.84) which was on par with seed treatment with 1.5 % micronutrient formulation + two foliar sprays, S₄F₃ (21.01). S₁F₁ (16) recorded the least value.

Table 6: Effect of seed treatment, foliar spray and their interactions on number of nodules per plant, fresh weight of nodules, dry weight of nodules and number of days taken for first flowering of cowpea

Treatment	Number of nodules plant ⁻¹	Fresh weight of nodules (g)	Dry weight of nodules (g)	Number of days taken for first flowering
Levels of seed treatment				
S ₁	14.77	0.21	0.16	32.11
S ₂	27.22	0.39	0.33	31.20
S ₃	18.88	0.40	0.34	31.00
S ₄	19.66	0.47	0.41	31.11
SEm (±)	1.93	0.08	0.08	0.19
CD (0.05)	5.71	NS	NS	NS
Levels of foliar spray				
F ₁	17.29	0.28	0.23	32.00
F ₂	20.37	0.37	0.31	31.25
F ₃	22.75	0.45	0.39	31.41
SEm (±)	1.67	0.07	0.07	0.16
CD (0.05)	NS	NS	NS	NS
Interactions				
S ₁ F ₁	12.83	0.10	0.07	32.33
S ₁ F ₂	15.66	0.27	0.22	31.33
S ₁ F ₃	15.83	0.26	0.20	31.00
S ₂ F ₁	25.66	0.36	0.30	31.00
S ₂ F ₂	24.66	0.33	0.27	32.00
S ₂ F ₃	31.33	0.47	0.41	30.00
S ₃ F ₁	14.00	0.25	0.19	30.66
S ₃ F ₂	20.66	0.41	0.36	31.00
S ₃ F ₃	22.00	0.53	0.48	30.00
S ₄ F ₁	16.66	0.40	0.34	31.00
S ₄ F ₂	20.50	0.46	0.40	31.66
S ₄ F ₃	21.83	0.54	0.48	31.41
SEm (±)	3.35	0.14	0.14	0.33
CD (0.05)	NS	NS	NS	NS

S₁- no seed treatment; S₂- seed treatment with rhizobium; S₃- seed treatment with micronutrient formulation (best from Exp. 1); S₄- seed treatment with micronutrient formulation (second best from Exp. 1)

F₁- no foliar spray; F₂- one foliar application at 15 DAS; F₃- two foliar application at 15 & 30DAS

4.3.1.9 Number of seeds per pod

The effect of seed treatment, foliar spray of micronutrient mixture and their interaction on number of seeds per pod is given in Table 7. Treatments did not show any significant difference in the number of seeds per pod.

4.3.1.10 Pod weight per plant (g)

The effect of seed treatment, micronutrient foliar spray and their interaction on pod weight per plant is given in Table 7.

Levels of seed treatment showed significant difference in pod weight per plant. S₃ recorded highest pod weight per plant (36.14) which was on par with S₄ (31.90). It was then followed by S₂ (29.73) and control recorded the least (24.92).

Levels of foliar spray showed significant difference in pod weight per plant with F₃ (33.49) recorded the highest value which was superior over all other treatments. The treatment F₁ showed lowest value (26.43).

Interaction effect of seed treatment and foliar spray of micronutrient in pod weight was found to be significant with S₃F₃ (38.11) recorded maximum pod weight per plant which was on par with S₄F₃ (31.23). Treatment S₁F₁ (24.64) recorded the lowest value.

4.3.1.11 Length of pod (cm)

Observations obtained on the length of pod per each treatment is given in Table 7. Length of pod did not show any significant difference in the main treatments as well as their interactions.

4.3.1.12 Grain yield (kg ha⁻¹)

Grain yield of cowpea with respect to seed treatment, foliar spray and their interaction is given in the Table 7.

Levels of seed treatment showed significant difference in yield with highest value was obtained in S₃ (2224) superior over all other seed treatments. Lowest yield was obtained in S₁ (1969).

Levels of foliar spray showed significant difference in yield with highest was recorded in F₃ (2173) which was superior over all other treatments and the lowest yield was recorded in F₁ (1979).

Interaction of seed treatment and foliar spray also showed significant difference in the yield with highest was recorded in S₃F₃ (2320) which was superior over other treatment interactions. It was followed by S₃F₂ (2220) which was on par with S₄F₃ (2200). Lowest yield was recorded in S₁F₁ (1820).

4.3.1.13 Dry matter production ($kg\ ha^{-1}$)

Dry matter production per plant as obtained from seed treatment, foliar spray of micronutrient and their interaction is given in the Table 7. Dry matter production recorded was maximum in S₃ (2292.91) which was superior over other seed treatments. Dry matter production was lowest in S₁ (2000.06). Foliar spray F₃ showed maximum dry matter production per plant (2372.92) which was superior over other levels of foliar spray. Interaction of main treatments showed significant difference in dry matter production with S₃F₃ (2613.24) produced highest amount which was superior over other interactions.

4.3.1.14 Root CEC ($cmol\ kg^{-1}$)

Results obtained on root CEC is given in Table 7. Root CEC was found to be significant with respect to different treatments. Among the seed treatment root CEC was found to be highest in S₃ (6.00) and in F₃ (5.87) of foliar spray. In the interaction effect S₃F₃ (6.13) recorded the highest value which was on par with S₂F₃ (5.98).

Table 7: Effect of seed treatment, foliar spray and their interactions on number of pods per plant, number of seeds per pod, pod weight per plant, length of pod, dry matter production and root CEC

Treatment	No of pods plant ⁻¹	No. of seeds pod ⁻¹	Pod weight plant ⁻¹ (g)	Length of pod (cm)	Grain yield (kg ha ⁻¹)	Dry matter (kg ha ⁻¹)	Root CEC (cmol kg ⁻¹)
Levels of seed treatment							
S ₁	16.73	15.46	24.92	17.64	1969	2000.06	5.58
S ₂	17.97	15.73	29.73	17.63	1977	2172.62	5.66
S ₃	21.02	15.13	36.14	18.11	2224	2292.91	6.00
S ₄	19.44	14.80	31.90	17.94	2168	2220.94	5.88
SEm (±)	0.91	0.43	2.28	0.39	16	16.37	0.03
CD (0.05)	2.68	NS	4.26	NS	48	48.33	0.10
Levels of foliar spray							
F ₁	16.71	15.53	26.43	17.85	1979	2042.53	5.72
F ₂	18.63	14.93	27.80	17.85	2101	2099.46	5.76
F ₃	20.97	15.38	33.49	17.80	2173	2372.92	5.87
SEm (±)	0.78	0.37	0.79	0.34	14	14.18	0.03
CD (0.05)	2.32	NS	2.33	NS	41	41.85	0.09
Interactions							
S ₁ F ₁	16.00	15.46	24.64	17.26	1820	1804.07	5.43
S ₁ F ₂	16.67	15.33	25.00	17.50	2000	2068.19	5.60
S ₁ F ₃	15.53	15.60	25.91	18.16	2113	2127.92	5.73
S ₂ F ₁	13.40	15.80	26.00	17.60	1843	2037.39	5.56
S ₂ F ₂	18.53	15.26	26.51	17.93	2000	2193.57	5.67
S ₂ F ₃	20.20	16.13	30.00	17.36	2059	2286.90	5.98
S ₃ F ₁	18.40	15.53	28.32	18.66	2133	2104.28	5.45
S ₃ F ₂	19.67	14.73	29.98	18.00	2220	2161.21	5.82
S ₃ F ₃	25.00	15.13	38.11	17.66	2320	2613.24	6.13
S ₄ F ₁	17.06	15.33	27.24	17.86	2120	2224.37	5.80
S ₄ F ₂	19.93	14.40	28.00	17.96	2186	1974.86	5.86
S ₄ F ₃	21.01	14.66	31.23	18.00	2200	2463.60	5.92
SEm (±)	1.57	0.75	2.38	0.69	28	28.36	0.06
CD (0.05)	4.72	NS	7.15	NS	83	83.71	0.18

S₁- no seed treatment; S₂- seed treatment with rhizobium; S₃- seed treatment with micronutrient formulation (best from Exp. 1); S₄- seed treatment with micronutrient formulation (second best from Exp. 1)

F₁- no foliar spray; F₂- one foliar application at 15 DAS; F₃- two foliar application at 15 & 30DAS

4.4 SOIL NUTRIENT ANALYSIS

4.4.1 Soil pH

The effect of seed treatment, foliar spray and their interaction on soil pH at 45 days after sowing and at harvest is given in the Table 8. Soil pH was found to be non significant in both 45 days after sowing and at harvest. The lowest pH at 45 days after sowing and at harvest was found in S₃ (4.43 and 4.39), F₃ (4.49 and 4.42) and S₂F₁ (4.33 and 4.37).

4.4.2 EC (dSm⁻¹)

Electrical conductivity of soil was not affected by the treatments at both 45 days after sowing and at harvest. The readings are given in the Table 8. Electrical conductivity was found to be higher in S₃ at both stages (0.12 and 0.14), F₃ (0.10 and 0.13) and S₃F₃ (0.12 and 0.14).

4.4.3 Organic carbon (%)

The effect of seed treatment, foliar spray of micronutrient mixture and their interaction is given in the Table 8.

Organic carbon was found to be non significant in seed treatment both at 45 days after sowing and at harvest.

Levels of foliar spray of micronutrient mixture recorded significant difference in organic carbon content at harvest with F₃ (0.63) superior over all other treatments. At 45 days after sowing organic carbon was non significant with respect to different levels of foliar spray.

Interaction effect of treatments were found to be non significant at 45 days after sowing. At harvest organic carbon content of soil was found to be the highest in S₃F₃ (0.69) which was on par with S₂F₃ (0.66), S₄F₃ (0.64), S₄F₁ (0.63), S₄F₂ (0.62) and S₃F₂ (0.62).

Table 8: Effect of seed treatment, foliar spray and their interactions on soil pH, EC and organic carbon at 45 DAS and harvest

Treatment	pH		EC (dSm ⁻¹)		Organic carbon (%)	
	45 DAS	Harvest	45 DAS	Harvest	45 DAS	Harvest
Levels of seed treatment						
S ₁	4.54	4.50	0.11	0.13	1.12	0.60
S ₂	4.44	4.42	0.08	0.11	1.10	0.60
S ₃	4.43	4.39	0.12	0.14	1.11	0.63
S ₄	4.44	4.41	0.09	0.11	1.12	0.63
SEm (±)	0.08	0.08	0.01	0.01	0.009	0.06
CD (0.05)	NS	NS	NS	NS	NS	NS
Levels of foliar spray						
F ₁	4.61	4.66	0.10	0.12	1.11	0.58
F ₂	4.60	4.61	0.10	0.12	1.11	0.59
F ₃	4.49	4.42	0.10	0.13	1.12	0.63
SEm (±)	0.07	0.07	0.01	0.011	0.008	0.014
CD (0.05)	NS	NS	NS	NS	NS	0.04
Interactions						
S ₁ F ₁	4.47	4.51	0.08	0.14	1.12	0.53
S ₁ F ₂	4.62	4.63	0.11	0.14	1.13	0.55
S ₁ F ₃	4.53	4.57	0.0	0.12	1.12	0.54
S ₂ F ₁	4.33	4.37	0.09	0.12	1.10	0.56
S ₂ F ₂	4.51	4.55	0.08	0.11	1.10	0.57
S ₂ F ₃	4.49	4.53	0.08	0.11	1.12	0.66
S ₃ F ₁	4.75	4.80	0.10	0.12	1.13	0.61
S ₃ F ₂	4.62	4.66	0.08	0.10	1.11	0.62
S ₃ F ₃	4.76	4.80	0.12	0.14	1.11	0.69
S ₄ F ₁	4.81	4.86	0.11	0.13	1.10	0.63
S ₄ F ₂	4.67	4.71	0.09	0.11	1.13	0.6
S ₄ F ₃	4.68	4.72	0.12	0.14	1.13	0.64
SEm (±)	0.14	0.14	0.02	0.02	0.015	0.02
CD (0.05)	NS	NS	NS	NS	NS	0.071

S₁- no seed treatment; S₂- seed treatment with rhizobium; S₃- seed treatment with micronutrient formulation (best from Exp. 1); S₄- seed treatment with micronutrient formulation (second best from Exp. 1)

F₁- no foliar spray; F₂- one foliar application at 15 DAS; F₃- two foliar application at 15 & 30DAS

4.4.4 Available nitrogen in soil (kg ha⁻¹)

The effect of treatments and their interaction on the available nitrogen present in the soil is given in the Table 9.

Levels of seed treatment showed considerable difference in the amount of available nitrogen at 45 days after sowing and harvest. At both stages S₂ (seed treatment with rhizobium) showed highest nitrogen content in soil compared to other treatments which were significantly different from other treatments (384.43 and 355.63). S₁ (no seed treatment) showed the lowest value at both stages (362.94 and 346.69).

Levels of foliar spray and the interaction effect on available nitrogen were found to be non significant at 45 days after sowing and at harvest.

4.4.5 Available phosphorus in soil (kg ha⁻¹)

The data obtained on the available phosphorus content affected by seed treatment, foliar spray of micronutrient mixture and their interaction are given in the Table 9.

Levels of seed treatment showed significant difference in the available P content at 45 days after sowing and at harvest with values recorded as 61.67 and 45.11 in S₂ (seed treated with rhizobium) which was superior over all other treatments.

Phosphorus content at both stages were found to be non significant at different levels of foliar spray as well as interactions.

4.4.6 Available potassium in soil (kg ha⁻¹)

Available K content in soil affected by various treatments and their interaction at 45 days after sowing and at harvest is given in the Table 9.

Available K content was highest in S₃ at both 45 days after sowing and at harvest (220.66 and 170.67). F₃ recorded highest available potassium in both stages

(218.16 and 168.33). Interaction effect of main treatments showed maximum value S₃F₃ at 45 days after sowing and at harvest as 229.38 and 179.00 respectively.

Table 9: Effect of seed treatment, foliar spray and their interactions on available N, P, and K in soil

Treatment	Available nitrogen (kg ha ⁻¹)		Available phosphorus (kg ha ⁻¹)		Available potassium (kg ha ⁻¹)	
	45 DAS	Harvest	45 DAS	Harvest	45 DAS	Harvest
Levels of seed treatment						
S ₁	362.94	346.69	54.74	33.33	195.55	145.66
S ₂	384.43	355.63	61.76	45.11	202.66	152.66
S ₃	367.92	349.56	54.54	35.74	220.66	170.67
S ₄	364.70	346.34	54.15	34.26	211.56	161.77
SEm (±)	1.86	1.86	0.41	0.26	0.07	0.05
CD (0.05)	5.51	5.50	1.23	0.79	0.22	0.16
Levels of foliar spray						
F ₁	367.05	356.61	56.29	36.73	200.25	150.25
F ₂	372.72	362.28	56.03	37.36	204.41	154.50
F ₃	370.22	359.78	56.57	37.25	218.16	168.33
SEm (±)	1.61	1.61	0.36	0.23	0.06	0.04
CD (0.05)	NS	NS	NS	NS	0.22	0.14
Interactions						
S ₁ F ₁	359.98	343.73	54.56	33.00	185.00	135.23
S ₁ F ₂	364.48	348.23	54.77	34.00	189.66	140.32
S ₁ F ₃	364.36	348.11	54.90	33.00	212.00	162.44
S ₂ F ₁	378.45	379.65	61.00	44.14	196.09	146.50
S ₂ F ₂	393.08	394.28	62.29	45.21	200.01	150.00
S ₂ F ₃	381.76	372.96	62.00	46.00	212.00	162.30
S ₃ F ₁	366.92	348.56	54.80	35.10	215.09	165.23
S ₃ F ₂	367.69	349.33	53.02	36.12	218.02	168.09
S ₃ F ₃	369.15	350.79	55.80	36.00	229.38	179.00
S ₄ F ₁	362.87	344.51	54.80	34.70	205.02	155.33
S ₄ F ₂	365.63	347.27	54.06	34.10	210.02	160.22
S ₄ F ₃	365.61	347.25	53.60	34.00	219.66	170.33
SEm (±)	3.23	3.22	0.36	0.46	0.13	0.09
CD (0.05)	NS	NS	NS	NS	0.38	0.28

S₁- no seed treatment; S₂- seed treatment with rhizobium; S₃- seed treatment with micronutrient formulation (best from Exp. 1); S₄- seed treatment with micronutrient formulation (second best from Exp. 1)

F₁- no foliar spray; F₂- one foliar application at 15 DAS; F₃- two foliar application at 15 & 30DAS

4.4.7 Available iron in soil (mg kg^{-1})

The data recorded on the available iron content affected by seed treatment, foliar spray of micronutrient mixture and their interaction is given in the Table 10.

At 45 days after sowing available iron content was found to be non significant with respect to various levels of seed treatment. At harvest S_3 recorded highest value (55.22) which was on par with S_4 (54.69).

Levels of seed treatment showed significant difference in available iron content. F_3 showed maximum iron content at 45 days after sowing and at harvest (55.50 and 51.28) which was superior than other treatments.

Interaction effect of seed treatment and foliar spray also showed significant difference in available iron content. Iron content was found to be the highest in S_1F_3 (87.66) at 45 days after sowing and S_3F_3 (60.94) at harvest.

4.4.8 Available manganese in soil (mg kg^{-1})

The data on available manganese at 45 days after sowing and at harvest with respect to seed treatment, foliar spray and their interaction is given in the Table 10.

Seed treatment did not show any significant difference in available manganese content at 45 days after sowing. At harvest manganese content showed significant difference with highest content recorded in S_3 (21.71) which was on par with S_4 (21.37) and S_2 (18.78).

Levels of foliar spray showed significant difference among the treatments with highest value recorded in F_3 (32.72 and 21.83) at both stages and superior over all other treatments.

Interaction between the treatments also showed significant difference with highest value recorded in S_3F_3 with 37.57 and 26.68 at 45 days after sowing and at harvest respectively.

Table 10: Effect of seed treatment, foliar spray and their interactions on available Fe, Mn and Zn in soil

Treatment	Available Fe (mg kg ⁻¹)		Available Mn (mg kg ⁻¹)		Available Zn (mg kg ⁻¹)	
	45 DAS	Harvest	45 DAS	Harvest	45 DAS	Harvest
Levels of seed treatment						
S ₁	82.34	50.34	27.41	16.53	4.16	1.68
S ₂	81.54	49.35	26.67	18.78	4.56	1.73
S ₃	82.36	55.22	32.54	21.71	4.59	1.77
S ₄	82.51	54.69	32.26	21.37	4.56	1.76
SEm (±)	0.63	0.41	0.02	0.97	0.02	0.01
CD (0.05)	NS	1.23	NS	2.94	NS	0.03
Levels of foliar spray						
F ₁	81.96	50.43	27.80	16.92	4.26	1.69
F ₂	81.30	51.28	28.64	17.80	4.32	1.74
F ₃	83.31	55.50	32.72	21.83	5.03	1.76
SEm (±)	0.55	0.36	0.02	0.02	0.02	0.01
CD (0.05)	1.62	1.07	0.07	0.07	0.07	0.03
Interactions						
S ₁ F ₁	78.69	46.69	23.77	12.88	3.46	1.79
S ₁ F ₂	80.68	48.69	25.69	14.80	3.76	1.80
S ₁ F ₃	87.66	55.66	32.78	21.89	5.26	1.60
S ₂ F ₁	83.69	51.69	28.93	18.04	4.46	1.77
S ₂ F ₂	79.69	47.69	24.82	13.93	4.06	1.68
S ₂ F ₃	81.23	48.69	26.26	15.37	4.26	1.88
S ₃ F ₁	82.69	50.68	28.26	17.37	4.66	1.80
S ₃ F ₂	83.72	54.05	31.79	21.07	4.75	1.80
S ₃ F ₃	80.68	60.94	37.57	26.68	5.56	1.70
S ₄ F ₁	82.78	52.69	30.26	19.37	4.46	1.70
S ₄ F ₂	81.10	54.69	32.26	21.37	4.75	1.71
S ₄ F ₃	83.66	56.70	34.26	23.37	5.06	1.88
SEm (±)	1.10	0.72	0.04	0.04	0.01	0.02
CD (0.05)	3.24	2.14	0.14	0.14	0.03	NS

S₁- no seed treatment; S₂- seed treatment with rhizobium; S₃- seed treatment with micronutrient formulation (best from Exp. 1); S₄- seed treatment with micronutrient formulation (second best from Exp. 1)

F₁- no foliar spray; F₂- one foliar application at 15 DAS; F₃- two foliar application at 15 & 30DAS

4.4.9 Available zinc in soil (mg kg⁻¹)

Effect of seed treatment, foliar application of micronutrient and their interaction on available zinc content at 45 days after sowing and at harvest is given in the Table 10.

Available zinc content was found to be non significant in seed treatment at 45 days after sowing. Zinc content showed significant difference at harvest in S₃ (1.77) which recorded the highest value which was on par with S₄ (1.76).

Levels of foliar spray showed significant difference in available zinc content with highest value recorded in treatment F₃ (5.03) at 45 days after sowing which was superior over other treatments and at harvest F₃ recorded the highest value (1.76) which was on par with F₂ (1.74).

Interaction showed significant difference in zinc content at 45 days after sowing with highest value obtained in S₃F₃ (5.56) which was found to be superior over other treatments and at harvest available zinc was non significant with respect to interactions.

4.4.10 Available copper in soil (mg kg⁻¹)

The data obtained on available copper content at 45 days after sowing and at harvest effected by seed treatment, foliar spray and their interaction is given in the Table 11.

Levels of seed treatment showed no significant difference in copper content at 45 days after sowing and at harvest.

Levels of foliar spray showed significant difference in available copper content at 45 days after sowing and at harvest with F₃ superior over other treatments (3.57 and 2.69).

Interaction effect on copper content was found to be non significant at 45 days after sowing but was significant at harvest with higher value obtained in S₄F₃ (3.51).

4.4.11 Available boron in soil (mg kg^{-1})

The data on available boron at 45 days after sowing and at harvest with respect to treatments are given in the Table 11.

Available boron was highest in S_3 (0.59) at 45 days after sowing and non significant at harvest stage. Considering foliar spray of micronutrient mixture available boron was highest in F_3 (0.62) which was superior over other foliar sprays at 45 days after sowing and was non significant at harvest. Interaction effect showed significant difference in boron content with highest value obtained in S_3F_3 (0.71) which was superior over other treatments at 45 days after sowing. At harvest interaction effect did not show significant difference among treatments.

4.4.12 Available molybdenum in soil (mg kg^{-1})

The result obtained on available molybdenum content in soil at different stages is given in the Table 11.

There was no significant difference among the seed treatments, foliar spray and interaction at 45 days after sowing. At harvest S_3 (0.008) and F_3 (0.008) was found to be superior among main treatments. Interaction effect did not show any significant difference in available molybdenum content in soil at harvest.

Table 11: Effect of seed treatment, foliar spray and their interactions on available copper, boron and molybdenum in soil

Treatment	Available Cu (mg kg ⁻¹)		Available B (mg kg ⁻¹)		Available Mo (mg kg ⁻¹)	
	45 DAS	Harvest	45 DAS	Harvest	45 DAS	Harvest
Levels of seed treatment						
S ₁	3.35	1.81	0.51	0.25	0.005	0.004
S ₂	3.48	1.88	0.49	0.24	0.006	0.005
S ₃	3.52	2.93	0.59	0.25	0.005	0.008
S ₄	3.48	2.63	0.57	0.26	0.007	0.006
SEm (±)	0.06	0.02	0.004	0.003	0.00013	0.00004
CD (0.05)	NS	NS	0.013	NS	NS	0.00012
Levels of foliar spray						
F ₁	3.38	2.11	0.50	0.25	0.006	0.005
F ₂	3.42	2.13	0.51	0.25	0.006	0.004
F ₃	3.57	2.69	0.62	0.26	0.009	0.008
SEm (±)	0.05	0.04	0.004	0.002	0.00014	0.00006
CD (0.05)	0.15	0.12	0.011	NS	NS	0.00018
Interactions						
S ₁ F ₁	3.31	1.31	0.44	0.25	0.003	0.002
S ₁ F ₂	3.41	1.61	0.50	0.26	0.004	0.003
S ₁ F ₃	3.33	2.51	0.60	0.25	0.007	0.006
S ₂ F ₁	3.36	2.11	0.52	0.25	0.004	0.003
S ₂ F ₂	3.40	1.51	0.46	0.24	0.005	0.004
S ₂ F ₃	3.69	2.01	0.51	0.25	0.008	0.007
S ₃ F ₁	3.53	2.41	0.52	0.26	0.009	0.008
S ₃ F ₂	3.37	2.87	0.54	0.26	0.008	0.007
S ₃ F ₃	3.66	2.75	0.71	0.25	0.010	0.009
S ₄ F ₁	3.31	2.61	0.53	0.26	0.006	0.005
S ₄ F ₂	3.51	2.55	0.54	0.25	0.005	0.003
S ₄ F ₃	3.61	3.51	0.66	0.26	0.009	0.009
SEm (±)	0.10	0.08	0.008	0.004	0.00003	0.0005
CD (0.05)	NS	0.25	0.023	NS	NS	NS

S₁- no seed treatment; S₂- seed treatment with rhizobium; S₃- seed treatment with micronutrient formulation (best from Exp. 1); S₄- seed treatment with micronutrient formulation (second best from Exp. 1)

F₁- no foliar spray; F₂- one foliar application at 15 DAS; F₃- two foliar application at 15 & 30DAS

4.5 TOTAL NUTRIENT CONTENT IN LEAF

4.5.1 Nitrogen (%)

Nitrogen concentration in the leaf at harvest is given in the Table 12. Effect of seed treatment on nitrogen content was non significant with highest value obtained in S_2 (2.07). Among the foliar spray F_3 (3.08) recorded highest nitrogen content in leaf at harvest which was superior over all other levels of foliar spray. Interaction of main treatments showed highest nitrogen concentration in S_3F_3 (3.70) which was on par with S_4F_3 (3.61).

4.5.2 Phosphorus (%)

Effect of treatments on the phosphorus content of leaf of cowpea is given in the Table 12. Seed treatment showed significant difference in phosphorus content with maximum in S_3 (0.26) which was superior over other treatments. It was significant in foliar spray at harvest with F_3 (0.25) which showed highest phosphorus content in leaf. S_3F_3 (0.28) showed highest value which was superior over other treatments.

4.5.3 Potassium (%)

Potassium content in leaf of cowpea at harvest is given in the Table 12. Seed treatments showed significant difference in potassium content and S_3 (1.39) was superior over other treatments. F_3 (1.29) was superior in potassium content in leaves. Interaction effect showed significant difference in treatments with S_3F_3 (1.46) which was superior over other treatments.

4.5.4 Calcium (%)

The effect of treatments on calcium content in leaf is given in the Table 12. Calcium content was found to be non significant with respect to different levels of seed treatment. Among the foliar spray F_3 (3.19) showed highest calcium content in leaf. Interaction effect did not show considerable difference among treatments.

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Table 12: Effect of seed treatment, foliar spray and their interactions on N, P, K, Ca, Mg and S content in cowpea leaf

Treatment	N (%)	P (%)	K (%)	Ca (%)	Mg (%)	S (%)
Levels of seed treatment						
S ₁	2.00	0.22	1.02	3.06	0.43	0.22
S ₂	2.07	0.22	1.22	3.12	0.63	0.22
S ₃	2.04	0.26	1.39	3.14	0.88	0.23
S ₄	2.04	0.23	1.21	3.07	0.59	0.22
SEm (±)	0.02	0.002	0.01	0.02	0.01	0.002
CD (0.05)	NS	0.006	0.02	NS	NS	NS
Levels of foliar spray						
F ₁	2.15	0.23	1.17	3.05	0.59	0.22
F ₂	2.30	0.24	1.16	3.12	0.66	0.22
F ₃	3.08	0.25	1.29	3.19	0.64	0.22
SEm (±)	0.02	0.002	0.009	0.02	0.01	0.002
CD (0.05)	0.06	0.006	0.025	0.06	NS	NS
Interactions						
S ₁ F ₁	1.21	0.18	0.94	2.99	0.41	0.22
S ₁ F ₂	1.51	0.23	0.98	3.10	0.41	0.23
S ₁ F ₃	3.30	0.24	1.14	3.11	0.46	0.22
S ₂ F ₁	2.50	0.19	1.16	3.11	0.52	0.23
S ₂ F ₂	1.31	0.22	1.17	3.11	0.54	0.22
S ₂ F ₃	1.72	0.24	1.34	3.15	0.82	0.22
S ₃ F ₁	2.60	0.26	1.40	3.13	0.87	0.23
S ₃ F ₂	3.21	0.24	1.31	3.17	1.10	0.23
S ₃ F ₃	3.70	0.28	1.46	3.13	0.66	0.23
S ₄ F ₁	2.31	0.22	1.20	2.99	0.57	0.22
S ₄ F ₂	3.21	0.23	1.21	3.11	0.59	0.23
S ₄ F ₃	3.61	0.25	1.22	3.13	0.60	0.22
SEm (±)	0.04	0.003	0.01	0.04	0.01	0.004
CD (0.05)	0.13	0.010	0.05	NS	0.04	NS

S₁- no seed treatment; S₂- seed treatment with rhizobium; S₃- seed treatment with micronutrient formulation (best from Exp. 1); S₄- seed treatment with micronutrient formulation (second best from Exp. 1)

F₁- no foliar spray; F₂- one foliar application at 15 DAS; F₃- two foliar application at 15 & 30DAS

4.5.5 Magnesium (%)

Magnesium content in cowpea leaves are given in the Table 12. Seed treatment and levels of foliar spray was found to be non significant. Among the interactions S_3F_3 (1.10) showed considerable difference among other interaction effects and was superior over other treatments.

4.5.6 Sulphur (%)

Sulphur content in leaf is presented in the Table 12. It was found to be non significant in main treatments as well as interactions.

4.5.7 Iron (mg kg^{-1})

Iron content in leaf is given in the Table 13. Levels of seed treatment showed significant difference in iron content with S_3 (377.68) recorded the highest value. F_3 (366.03) was significantly different from other foliar sprays. S_3F_3 (390) in the interaction effect showed highest iron content which was superior over other treatments.

4.5.8 Manganese (mg kg^{-1})

Manganese content in the leaf at harvest is given in the Table 13. It showed considerable difference among the treatments with S_3 (275.77), F_3 (293.30) and S_3F_3 (287.33) were superior over other treatments.

4.5.9 Zinc (mg kg^{-1})

Effect of treatments on the zinc content of leaf is presented in the Table 13. It also showed the same trend as in manganese with S_3 (61.44), F_3 (56.83) and S_3F_3 (66.13) showed highest zinc content.

4.5.10 Copper (mg kg^{-1})

Copper content in leaf with respect to different treatments are given in the Table 13. Copper content in leaf was found to be non significant with respect to different treatments and their interactions.

Table 13: Effect of seed treatment, foliar spray and their interactions on Fe, Mn, Zn, Cu, B and Mo content in cowpea leaf

Treatment	Fe (mg kg ⁻¹)	Mn (mg kg ⁻¹)	Zn (mg kg ⁻¹)	Cu (mg kg ⁻¹)	B (mg kg ⁻¹)	Mo (mg kg ⁻¹)
Levels of seed treatment						
S ₁	334.36	212.02	47.16	28.35	25.25	1.02
S ₂	327.02	217.73	49.00	28.09	25.56	1.02
S ₃	377.68	275.77	61.44	27.87	26.83	1.03
S ₄	358.00	235.58	52.92	27.77	26.35	1.00
SEm (±)	2.81	1.99	0.43	0.21	0.19	0.02
CD (0.05)	8.29	5.88	1.27	NS	0.58	NS
Levels of foliar spray						
F ₁	336.51	217.29	49.40	28.30	25.58	1.02
F ₂	345.25	225.25	51.66	28.27	25.94	1.02
F ₃	366.03	293.30	56.83	28.29	26.47	1.02
SEm (±)	2.43	1.73	0.37	0.18	0.17	0.02
CD (0.05)	7.18	5.01	1.10	NS	0.50	NS
Interactions						
S ₁ F ₁	311.00	207.07	42.85	28.32	25.02	1.02
S ₁ F ₂	321.00	217.00	43.92	28.52	25.22	1.02
S ₁ F ₃	371.10	270.00	54.73	28.21	25.52	1.02
S ₂ F ₁	323.12	211.11	46.80	27.25	25.48	1.02
S ₂ F ₂	327.10	218.02	48.60	28.40	25.18	1.02
S ₂ F ₃	331.06	224.01	51.62	28.64	26.02	1.02
S ₃ F ₁	371.06	267.02	56.06	27.69	26.52	1.02
S ₃ F ₂	372.00	273.21	62.13	27.58	26.97	1.02
S ₃ F ₃	390.00	287.33	66.13	28.36	27.02	1.02
S ₄ F ₁	341.01	212.03	51.90	27.15	25.32	1.02
S ₄ F ₂	361.00	223.01	52.00	28.27	26.42	1.02
S ₄ F ₃	372.00	271.04	54.86	28.19	27.32	1.02
SEm (±)	4.86	3.50	0.75	0.36	0.34	0.02
CD (0.05)	14.37	10.20	2.21	NS	NS	NS

S₁- no seed treatment; S₂- seed treatment with rhizobium; S₃- seed treatment with micronutrient formulation (best from Exp. 1); S₄- seed treatment with micronutrient formulation (second best from Exp. 1)

F₁- no foliar spray; F₂- one foliar application at 15 DAS; F₃- two foliar application at 15 & 30DAS

4.5.11 Boron (mg kg^{-1})

Boron content in the leaf of cowpea at harvest is given in the Table 13. Among the seed treatments S_3 (26.83) recorded highest value of boron concentration in leaf which was on par with S_4 (26.35). F_3 (26.47) was found to be superior over other levels of foliar spray. Treatment interactions were found to be non significant with respect to boron concentration in cowpea leaves at harvest.

4.5.12 Molybdenum (mg kg^{-1})

Molybdenum content in cowpea is given in the Table 13. It did not show significant difference in seed treatment, foliar spray and interaction of main treatments.

4.6 NUTRIENT CONTENT IN GRAIN

4.6.1 Nitrogen (%)

Nitrogen content in grain is given in the Table 14. S_2 (3.71) recorded highest nitrogen in grain which was superior over other seed treatments. F_3 (3.76) in foliar spray and S_2F_3 (3.91) in interaction were found to be superior over other treatments.

4.6.2 Phosphorus (%)

Grain phosphorus content in cowpea is recorded in the Table 14. It did not show significant difference in any treatments.

4.6.3 Potassium (%)

Potassium content in grain is presented in the Table 14. S_3 (1.18) in seed treatment and F_3 (1.08) in foliar spray recorded highest potassium content. Interaction effect showed that S_3F_3 (1.25) and S_4F_3 (1.19) were found to be on par with each other.

4.6.4 Calcium (%)

Calcium content in grain of cowpea is given in the Table 14. S₃ (0.59), F₃ (0.52) and S₃F₃ (0.67) recorded maximum calcium content in grain and was superior over other treatments.

4.6.5 Magnesium (%)

Magnesium content in cowpea grain is presented in the Table 14. Magnesium content in grain was found to be non significant in main treatments as well as interactions.

4.6.6 Sulphur (%)

Sulphur content in grain is given in the Table 14. The data were found to be non significant in all the treatments.

Table 14: Effect of seed treatment, foliar spray and their interactions on N, P, K, Ca, Mg and S content in cowpea grain

Treatment	N (%)	P (%)	K (%)	Ca (%)	Mg (%)	S (%)
Levels of seed treatment						
S ₁	3.43	0.32	0.81	0.39	0.17	0.48
S ₂	3.71	0.32	1.01	0.46	0.18	0.48
S ₃	3.50	0.36	1.18	0.59	0.18	0.48
S ₄	3.41	0.36	1.00	0.48	0.18	0.48
SEm (±)	0.02	0.02	0.01	0.001	0.01	0.004
CD (0.05)	0.07	NS	0.04	0.004	NS	NS
Levels of foliar spray						
F ₁	3.38	0.34	0.96	0.44	0.17	0.48
F ₂	3.41	0.33	0.96	0.49	0.18	0.48
F ₃	3.76	0.36	1.08	0.52	0.18	0.48
SEm (±)	0.02	0.02	0.01	0.001	0.014	0.003
CD (0.05)	0.06	NS	0.03	0.003	NS	NS
Interactions						
S ₁ F ₁	3.30	0.27	0.72	0.35	0.17	0.48
S ₁ F ₂	3.40	0.32	0.77	0.40	0.17	0.48
S ₁ F ₃	3.61	0.37	0.93	0.43	0.17	0.48
S ₂ F ₁	3.30	0.35	0.94	0.45	0.18	0.48
S ₂ F ₂	3.31	0.34	0.96	0.47	0.18	0.48
S ₂ F ₃	3.91	0.34	1.13	0.48	0.18	0.48
S ₃ F ₁	3.71	0.35	1.01	0.51	0.17	0.48
S ₃ F ₂	3.62	0.37	1.10	0.61	0.17	0.48
S ₃ F ₃	3.81	0.40	1.25	0.67	0.17	0.48
S ₄ F ₁	3.23	0.37	0.99	0.47	0.17	0.48
S ₄ F ₂	3.31	0.34	1.00	0.48	0.17	0.48
S ₄ F ₃	3.72	0.38	1.19	0.51	0.17	0.48
SEm (±)	0.01	0.02	0.02	0.003	0.021	0.006
CD (0.05)	0.02	NS	0.07	0.010	NS	NS

S₁- no seed treatment; S₂- seed treatment with rhizobium; S₃- seed treatment with micronutrient formulation (best from Exp. 1); S₄- seed treatment with micronutrient formulation (second best from Exp. 1)

F₁- no foliar spray; F₂- one foliar application at 15 DAS; F₃- two foliar application at 15 & 30DAS

4.6.7 Iron (mg kg^{-1})

Percentage of iron in grain is given in the Table 15. Seed treatment S_3 (120.91), foliar spray F_3 (117.65) and interaction S_3F_3 (131.96) were found to be superior in the respective treatments and their combinations.

4.6.8 Manganese (mg kg^{-1})

Manganese content in grain is given in the Table 15. Among the seed treatments, S_3 (14.30) recorded significantly higher value. F_3 (12.71) and S_3F_3 (17.08) showed superior value in the respective treatments and interaction.

4.6.9 Zinc (mg kg^{-1})

Zinc content in grain is given in the Table 15. It also showed the same trend of iron and manganese with S_3 (56.90), F_3 (53.98) and S_3F_3 (59.21) recorded superior values.

4.6.10 Copper (mg kg^{-1})

Copper content in grain also showed significant difference among treatments which is given in the Table 15. S_3 (6.50), F_3 (6.45) and S_3F_3 (7.02) showed copper content superior over other treatments.

4.6.11 Boron (mg kg^{-1})

Boron content in grain is given in the Table 15. S_3 (86.80), F_3 (83.42) and S_3F_3 (89.23) showed highest boron content over other treatments.

4.6.12 Molybdenum (mg kg^{-1})

Molybdenum content in grain is given in the Table 15. It was found to be non significant with respect to seed treatment, foliar spray and their interaction.

Table 15: Effect of seed treatment, foliar spray and their interactions on Fe, Mn, Zn, Cu, B and Mo content in cowpea grain

Treatment	Fe (mg kg ⁻¹)	Mn (mg kg ⁻¹)	Zn (mg kg ⁻¹)	Cu (mg kg ⁻¹)	B (mg kg ⁻¹)	Mo (mg kg ⁻¹)
Levels of seed treatment						
S ₁	103.54	7.74	48.63	5.23	74.80	1.05
S ₂	105.86	8.96	51.13	5.53	75.11	1.06
S ₃	120.91	14.30	56.90	6.50	86.80	1.07
S ₄	109.50	10.74	52.82	6.06	79.13	1.06
SEm (±)	0.03	0.01	0.01	0.001	0.01	0.02
CD (0.05)	0.09	0.03	0.02	0.003	0.04	NS
Levels of foliar spray						
F ₁	105.35	9.21	50.94	5.45	76.20	1.04
F ₂	106.85	9.38	52.19	5.60	77.25	1.05
F ₃	117.65	12.71	53.98	6.45	83.42	1.05
SEm (±)	0.02	0.01	0.009	0.001	0.01	0.02
CD (0.05)	0.07	0.03	0.025	0.002	0.04	NS
Interactions						
S ₁ F ₁	94.83	5.55	45.06	4.50	69.50	0.77
S ₁ F ₂	100.83	5.56	48.10	4.71	70.30	0.69
S ₁ F ₃	114.96	12.11	52.73	6.50	84.61	1.30
S ₂ F ₁	104.90	8.56	50.30	5.21	74.50	1.49
S ₂ F ₂	103.86	9.06	50.90	5.41	75.50	1.54
S ₂ F ₃	108.83	9.26	52.21	6.01	75.31	0.89
S ₃ F ₁	112.86	12.86	55.81	6.20	84.61	1.52
S ₃ F ₂	117.90	12.96	55.92	6.32	86.61	1.25
S ₃ F ₃	131.96	17.08	59.21	7.02	89.23	1.56
S ₄ F ₁	108.83	9.89	52.61	5.91	76.21	1.24
S ₄ F ₂	104.83	9.94	53.86	6.01	76.60	1.35
S ₄ F ₃	114.83	12.39	52.00	6.30	84.61	1.45
SEm (±)	0.05	0.02	0.01	0.002	0.02	0.03
CD (0.05)	0.15	0.06	0.05	0.004	0.08	NS

S₁- no seed treatment; S₂- seed treatment with rhizobium; S₃- seed treatment with micronutrient formulation (best from Exp. 1); S₄- seed treatment with micronutrient formulation (second best from Exp. 1)

F₁- no foliar spray; F₂- one foliar application at 15 DAS; F₃- two foliar application at 15 & 30DAS

4.7 NUTRIENT UPTAKE IN COWPEA GRAIN

4.7.1 Nitrogen uptake (kg ha^{-1})

Nitrogen uptake in cowpea grain is given in Table 16. Among the seed treatment nitrogen uptake in grain was significantly higher in S_3 (82.75). Foliar spray, F_3 (81.87) showed highest uptake among the different levels of foliar spray. Interaction was found to be non significant with respect to nitrogen uptake in grain.

4.7.2 Phosphorus uptake (kg ha^{-1})

Phosphorus uptake in cowpea grain is given in Table 16. Phosphorus uptake in grain was significantly higher in treatments S_3 (8.29) and F_3 (7.87). Interaction S_3F_3 (8.97) showed highest uptake which was significantly different from other interactions.

4.7.3 Potassium uptake (kg ha^{-1})

Potassium uptake in cowpea grain is given in Table 16. Seed treatment S_3 (26.47) showed the highest uptake of potassium in grain which was significantly different from all other levels of seed treatment. among the foliar spray, F_3 (23.70) showed significantly higher uptake of potassium in cowpea grain. S_3F_3 (29.19) recorded highest uptake of potassium compared to other interactions and was found to be superior.

4.7.4 Calcium uptake (kg ha^{-1})

Calcium uptake in cowpea grain is given in Table 16. Among seed treatment and foliar spray calcium uptake by cowpea grain was significantly superior in S_3 (13.43) and F_3 (11.52) respectively. Considering the interaction, S_3F_3 (15.64) recorded significantly higher uptake of calcium compared to other interactions.

4.7.5 Magnesium uptake (kg ha^{-1})

Magnesium uptake in cowpea grain is given in Table 16. Levels of seed treatment showed significant difference in the uptake of magnesium by cowpea grain. S_3 (3.88) showed highest uptake which was on par with S_4 (3.78). Levels of foliar spray also showed significant difference in the uptake of magnesium. F_3

(3.75) showed highest uptake which was on par with F₂ (3.73). interaction was found to be non significant.

4.7.6 Sulphur uptake (kg ha⁻¹)

Sulphur uptake in cowpea grain is given in Table 16. Among the seed treatment sulphur uptake by grain was significantly higher in S₃ (10.79) which was on par with S₄ (10.54). F₃ (10.55) showed highest uptake significantly higher and was found to be on par with F₂ (10.18). interaction effect was found to be non significant.

4.7.7 Iron uptake (kg ha⁻¹)

Iron uptake in cowpea grain is given in Table 17. Iron uptake in grain was significantly higher in S₃ (0.27) compared to other seed treatments and F₃ (0.25) recorded significantly higher uptake of iron among the different levels of foliar spray. Interaction S₃F₃ (0.30) recorded highest uptake of iron in cowpea grain and was found to be significantly higher compared to other interactions.

4.7.8 Manganese uptake (kg ha⁻¹)

Manganese uptake in cowpea grain is given in Table 17. Manganese uptake in grain was found to be non significant with respect to seed treatment and foliar spray. Interaction S₃F₃ (0.04) recorded highest uptake significantly different from all other interactions.

4.7.9 Zinc uptake (kg ha⁻¹)

Zinc uptake in cowpea grain is given in Table 17. Zinc uptake was significantly higher in S₃ (0.12) and F₃ (0.12). Interaction S₃F₃ (0.14) showed significantly higher uptake compared to other interactions.

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4.7.10 Copper uptake (kg ha^{-1})

Copper uptake in cowpea grain is given in Table 17. Copper uptake in cowpea grain was found to be non significant with respect to seed treatment, foliar spray and their interactions.

4.7.11 Boron uptake (kg ha^{-1})

Boron uptake in cowpea grain is given in Table 17. Boron uptake was significantly higher in S_3 (0.19) and F_3 (0.18). Interaction S_3F_3 (0.21) showed significantly higher uptake compared to other interactions.

4.7.12 Molybdenum uptake (kg ha^{-1})

Molybdenum uptake in cowpea grain is given in Table 17. Molybdenum uptake in cowpea grain was found to be non significant with respect to seed treatment, foliar spray and their interactions.

Table 16: Effect of seed treatment, foliar spray and their interactions on uptake of N, P, K, Ca, Mg and S in cowpea grain

Treatment	N (kg ha ⁻¹)	P (kg ha ⁻¹)	K (kg ha ⁻¹)	Ca (kg ha ⁻¹)	Mg (kg ha ⁻¹)	S (kg ha ⁻¹)
Levels of seed treatment						
S ₁	68.20	6.48	16.14	7.87	3.41	9.57
S ₂	69.21	7.13	20.05	9.28	3.56	9.49
S ₃	82.75	8.29	26.47	13.43	3.88	10.79
S ₄	74.29	7.90	21.85	10.64	3.78	10.54
SEm (±)	1.18	0.13	0.36	0.18	0.06	0.16
CD (0.05)	3.48	0.37	1.08	0.53	0.19	0.47
Levels of foliar spray						
F ₁	67.17	6.81	19.35	8.95	3.50	9.56
F ₂	71.80	7.50	20.33	10.45	3.73	10.18
F ₃	81.87	7.87	23.70	11.52	3.75	10.55
SEm (±)	1.10	0.11	0.32	0.15	0.05	0.14
CD (0.05)	3.02	0.32	0.94	0.46	0.16	0.41
Interactions						
S ₁ F ₁	60.09	4.97	13.23	6.37	3.15	8.80
S ₁ F ₂	68.03	6.40	15.40	8.13	3.46	9.67
S ₁ F ₃	76.48	8.05	19.77	9.11	3.60	10.24
S ₂ F ₁	60.91	6.58	17.53	8.36	3.32	8.86
S ₂ F ₂	66.14	7.66	19.20	9.46	3.73	9.66
S ₂ F ₃	80.58	7.14	23.42	10.03	3.64	9.96
S ₃ F ₁	79.24	7.62	25.65	11.04	3.77	10.33
S ₃ F ₂	80.50	8.29	24.59	13.62	3.85	10.74
S ₃ F ₃	88.50	8.97	29.19	15.64	4.02	11.30
S ₄ F ₁	68.43	8.28	20.99	10.04	3.74	10.25
S ₄ F ₂	72.53	7.59	22.13	10.58	3.87	10.66
S ₄ F ₃	81.92	7.62	22.44	11.29	3.74	10.70
SEm (±)	2.04	0.22	0.64	0.31	1.11	0.27
CD (0.05)	NS	0.64	1.90	0.92	NS	NS

S₁- no seed treatment; S₂- seed treatment with rhizobium; S₃- seed treatment with micronutrient formulation (best from Exp. 1); S₄- seed treatment with micronutrient formulation (second best from Exp. 1)

F₁- no foliar spray; F₂- one foliar application at 15 DAS; F₃- two foliar application at 15 & 30DAS

Table 17: Effect of seed treatment, foliar spray and their interactions on uptake of Fe, Mn, Zn, Cu, B and Mo in cowpea grain

Treatment	Fe (kg ha ⁻¹)	Mn (kg ha ⁻¹)	Zn (kg ha ⁻¹)	Cu (kg ha ⁻¹)	B (kg ha ⁻¹)	Mo (kg ha ⁻¹)
Levels of seed treatment						
S ₁	0.20	0.01	0.09	0.01	0.14	0.002
S ₂	0.20	0.01	0.10	0.01	0.14	0.003
S ₃	0.27	0.03	0.12	0.01	0.19	0.003
S ₄	0.23	0.02	0.11	0.01	0.17	0.003
SEm (±)	0.004	-	0.002	0.003	0.003	0.00013
CD (0.05)	0.01	0.001	0.005	NS	0.008	NS
Levels of foliar spray						
F ₁	0.21	0.02	0.10	0.01	0.15	0.003
F ₂	0.23	0.02	0.11	0.01	0.16	0.003
F ₃	0.25	0.03	0.12	0.01	0.18	0.003
SEm (±)	0.003	-	0.002	0.003	0.002	0.00012
CD (0.05)	0.01	0.001	0.005	NS	0.007	NS
Interactions						
S ₁ F ₁	0.17	0.01	0.08	0.008	0.12	0.001
S ₁ F ₂	0.20	0.01	0.09	0.009	0.14	0.001
S ₁ F ₃	0.24	0.03	0.11	0.14	0.17	0.003
S ₂ F ₁	0.19	0.02	0.09	0.01	0.13	0.003
S ₂ F ₂	0.20	0.02	0.10	0.01	0.15	0.003
S ₂ F ₃	0.22	0.02	0.11	0.01	0.15	0.002
S ₃ F ₁	0.24	0.03	0.12	0.01	0.18	0.003
S ₃ F ₂	0.26	0.03	0.12	0.01	0.19	0.003
S ₃ F ₃	0.30	0.04	0.14	0.01	0.20	0.004
S ₄ F ₁	0.23	0.02	0.11	0.01	0.16	0.003
S ₄ F ₂	0.23	0.02	0.12	0.01	0.16	0.003
S ₄ F ₃	0.25	0.03	0.11	0.01	0.18	0.003
SEm (±)	0.007	0.001	0.003	0.003	0.005	0.00013
CD (0.05)	0.02	0.002	0.009	NS	0.014	NS

S₁- no seed treatment; S₂- seed treatment with rhizobium; S₃- seed treatment with micronutrient formulation (best from Exp. 1); S₄- seed treatment with micronutrient formulation (second best from Exp. 1)

F₁- no foliar spray; F₂- one foliar application at 15 DAS; F₃- two foliar application at 15 & 30DAS

4.7 SOIL BIOMASS CARBON (mg kg^{-1} soil)

The data recorded on soil biomass carbon in soil is given in the Table 16. It was found to be non significant with respect to treatments and interactions.

Table 18: Effect of seed treatment, foliar spray and their interactions on soil biomass carbon content

Treatment	Soil biomass carbon (mg kg^{-1} soil)
Levels of seed treatment	
S ₁	168.14
S ₂	167.18
S ₃	169.57
S ₄	168.17
SEm (\pm)	1.271
CD (0.05)	NS
Levels of foliar spray	
F ₁	167.32
F ₂	168.61
F ₃	168.86
SEm (\pm)	1.101
CD (0.05)	NS
Interactions	
S ₁ F ₁	168.45
S ₁ F ₂	167.98
S ₁ F ₃	168.00
S ₂ F ₁	166.87
S ₂ F ₂	167.70
S ₂ F ₃	166.98
S ₃ F ₁	168.32
S ₃ F ₂	170.34
S ₃ F ₃	170.06
S ₄ F ₁	165.65
S ₄ F ₂	168.45
S ₄ F ₃	170.42
SEm (\pm)	2.202
CD (0.05)	NS

S₁- no seed treatment; S₂- seed treatment with rhizobium; S₃- seed treatment with micronutrient formulation (best from Exp. 1); S₄- seed treatment with micronutrient formulation (second best from Exp. 1)

F₁- no foliar spray; F₂- one foliar application at 15 DAS; F₃- two foliar application at 15 & 30DAS

Discussion

5. DISCUSSION

The results obtained from the study "Evaluation of micronutrient formulation on cowpea (*Vigna unguiculata* L. Walp)" are briefly discussed in this chapter. The investigation was conducted in three different steps which includes standardization of micronutrient mixture for cowpea, seed treatment and field experiment.

5.1 STANDARDIZATION OF MICRONUTRIENT FORMULATION FOR COWPEA

A micronutrient formulation developed at college of agriculture, Padannakad has been modified for the foliar application and seed treatment of cowpea. The formulation is a mixture of two solutions, Solution A and Solution B. Solution A contains iron sulphate, manganous sulphate, zinc sulphate, copper sulphate, boric acid and ammonium molybdate at definite proportion. Solution B is an organically chelated solution. Solution A and solution B should be mixed together before foliar spray and after mixing both the solutions the formulation cannot be stored due to its poor shelf life and precipitate formation. Several trials were done to reduce precipitate formation of this formulation when mixed together. A specific combination of solutions A and B which do not form precipitate and could be stored as such for longer duration was formulated after several trials. Thus it could help the farmers to reduce the labour of mixing both solutions A and solution B during foliar spray. The formulation would definitely help in boosting the production and productivity of pulses.

5.2 SEED TREATMENT STUDY

5.2.1 Standardization of time and dose of micronutrient formulation for seed treatment of cowpea

The different concentrations of micronutrient formulation and time required for seed treatment in cowpea was standardized. Cowpea seeds were treated with micronutrient mixture at different concentrations (0.25 %, 0.50 %, 0.75 %, 1 %, 1.5 % and 2 %) for different durations (1 hr, 2hrs, 4 hrs, 6hrs, 8 hrs, 10 hrs and 12 hrs).

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Observations on seed germination and its emergence showed that the seeds treated with micronutrient mixture for 6 hours showed better establishment for the different concentration of micronutrient formulation as compared to other duration of seed treatment. Time for seed treatment was standardized in papaya for boron. Seeds treated with 2 ppm boron solution for six hours showed increased germination and growth of seedlings (Deb *et al.*, 2010).

5.2.2 Effect of seed treatment on seedling characters of cowpea

Cowpea seeds were treated with micronutrient mixture at 0.25 %, 0.50 %, 0.75 %, 1 %, 1.5 % and 2 % for six hours. The biometric observations obtained from seed treatment study are briefly discussed below.

The different concentrations of micronutrient mixture in seed treatment study had no significant influence on germination percentage and days taken for germination whereas seedling length at three leaf stage and seedling vigour index was significantly influenced by the treatments. The treatment T₆ (micronutrient mixture @ 2 %) showed maximum seedling length at three leaf stage and seedling vigour index which might be due to the beneficial effect of micronutrients in enhancing cell elongation and cell division of meristematic tissues. The increased seedling length of cowpea contributed to considerable increase in seedling vigour index in the treatment receiving 2 % concentration of micronutrient mixture. Thus T₆ was considered as the best concentration for seed treatment study. This treatment was followed by T₅ (micronutrient mixture @ 1.5 %) which recorded the second highest values in seedling length at three leaf stage and seedling vigour index. The treatments T₆ (best seed treatment) and T₅ (second best) were selected for the field experiment to evaluate the performance of micronutrient formulation in cowpea. Masuthi *et al.* 2009 had reported that cowpea seeds treated with zinc sulphate showed higher vigour index and Deb *et al.* 2010 reported that papaya seeds treated with boron at 2ppm showed better growth of seedlings.

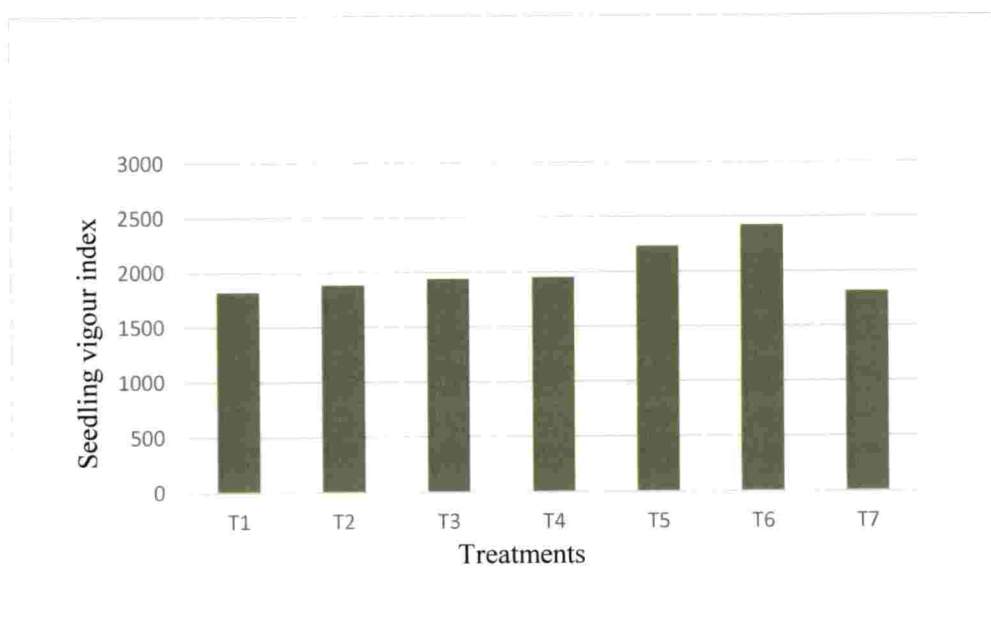


Fig. 1 Effect of seed treatment on seedling vigour index

5.3 FIELD EXPERIMENT

5.3.1 Effect of seed treatment, foliar spray of micronutrient formulation and their interaction on growth attributes

The results obtained from the study revealed that growth attributes like plant height and stem diameter at flower bud initiation and harvest were influenced by seed treatment, foliar spray of micronutrient formulation and their interactions.

Considering the effect of seed treatment, it was evident that plants subjected to micronutrient seed treatment at 2 percentage (S₃) and 1.5 percentage (S₄) showed remarkable variation in plant height at flower bud initiation stage. At harvest seeds treated with 2 percentage micronutrient formulation showed maximum plant height compared to other treatments. With respect to foliar application of micronutrient formulation plants supplied with micronutrient mixture (2 percentage formulation was used for the foliar spray) at 15 DAS and 30 DAS (F₃) showed characteristic difference in plant height compared to single foliar spray as well as no foliar spray

at both flower bud initiation stage and harvest. Among the interactions plants receiving 2 per cent seed treatment along with two foliar spray (S₃F₃) showed maximum plant height at both the stages. Hence the application of micronutrient mixture as seed treatment and foliar spray was found to be the best treatment. The beneficial effect of micronutrients in growth and development of plants might be the reason for increase in plant height compared to control treatment receiving no seed treatment and no foliar spray. Micronutrients has the capacity to increase the auxin content especially zinc and chlorophyll content of tissues thus promoting plant growth and yield. It helps in the growth of cells by promoting cell elongation and cell division of meristematic regions. The micronutrients zinc and boron plays a crucial role in growth and development of new cells in plants. The findings are in accordance with the results obtained by Suryanarayana and Reddy (1978). They reported that plant height of French bean was increased by the application of 0.01 per cent Zn and B. Cowpea seeds treated with borax showed highest plant height at 30 and 60 days after sowing (Masuthi *et al.*, 2009). Application of micronutrients enhanced the plant height of tomato over the control by 30 per cent (Ejaz *et al.* (2011).

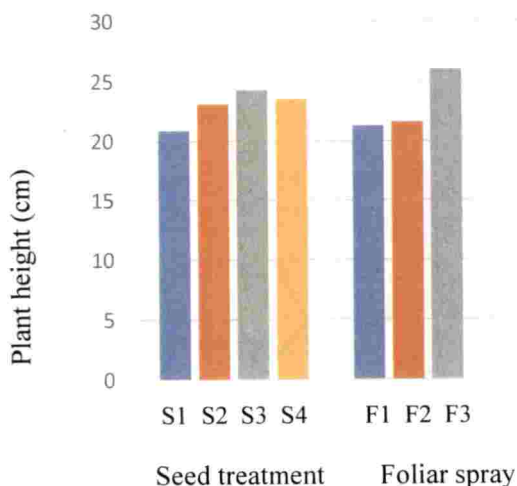


Fig.2 Effect of micronutrient formulation on plant height at flower bud initiation stage

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In the case of stem diameter, effect of seed treatment with micronutrient (S₃ and S₄) and rhizobium (S₂) was significantly evident at flower bud initiation stage. The beneficial effect of micronutrients and rhizobium on growth and development of plants were clearly understood from these observations. Micronutrients helps in plant growth by cell elongation and division. Rhizobium also increases the growth of plants. It stimulates the growth of plants by production of plant hormones. But at the time of harvest seed treatment with micronutrient mixture at 2 percentage showed the highest stem diameter compared to seeds treated with rhizobium, 1.5 percentage micronutrient and no seed treatment. Thus micronutrient enhanced the growth of plants. In treatments receiving foliar spray of micronutrient mixture, at flower bud initiation stage both F₂ and F₃ (one foliar spray and two foliar spray) showed highest stem diameter and at harvest the plants receiving two foliar spray showed highest stem diameter. From this it is evident that treatments receiving two foliar spray showed maximum stem diameter. The beneficial effect of micronutrient spray on cowpea could be concluded from the data. The interaction effect showed that at flower bud initiation stage S₂F₃ (seed treatment with rhizobium + two foliar spray) showed maximum stem diameter which was on par with S₃F₂ (seed treatment with 2 % micronutrient mixture + one foliar spray), S₄F₃ (seed treatment with 1.5 % micronutrient mixture + two foliar spray) and S₄F₂ (seed treatment with 1.5 % micronutrient mixture + one foliar spray). While at harvest treatment receiving rhizobium and two foliar spray showed highest stem diameter. Thus showing the beneficial effect of both micronutrients and rhizobium in plant growth and development. These findings were in accordance with Mathur (2000) reported that seeds of garden pea inoculated with rhizobium showed enhanced growth characters and yield. Suryanarayana and Rao (1981) reported that foliar spray of micronutrient solution containing zinc, copper, manganese, magnesium, boron and molybdenum resulted in enhanced growth of okra. Bukvic *et al.* (2003) reported the similar increasing trend in stem diameter with zinc application in maize.

Dry matter production also showed considerable variations with respect to different treatments.

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Considering the levels of seed treatment dry matter production was found to be highest in seed treatment with micronutrient formulation at two percentage (S₃). It was found to be superior over other seed treatments. In foliar spray dry matter production was highest in plants receiving two foliar spray (F₃) which was also superior compared to other levels of foliar sprays. Hence in the interaction effect S₃F₃ showed considerably higher production of dry matter compared to other interactions. Seed treatment with micronutrients have the capacity to enhance dry matter production. Seed treatment might had resulted in increased seed vigour. Increased seed vigour might be the reason for maximum dry matter production in plants. Similar results were obtained in barley. Seeds of barley treated with micronutrients showed increased dry matter production (Ajouri *et al.*, 2004).

5.3.2 Effect of seed treatment, foliar spray of micronutrient formulation and their interaction on nodulation of cowpea

The results obtained from the study revealed that seed treatment with rhizobium has a positive effect on the number of nodules whereas seed treatment and foliar spray of micronutrient formulation did not result in considerable increase in the number of nodules. The fresh weight and dry weight of nodules were also found to be non significant in seed treatment, foliar spray and their interaction. Rhizobium forms symbiotic association with the roots of legumes and helps fixing nitrogen in the soil. Cowpea, a legume crop has the inherent capacity to fix nitrogen by the formation of root nodules. Inoculation of cowpea seeds with specific rhizobium strains results in increased nodule formation and thus increases the nitrogen fixation in soil. Rhizobia can infect the legume hosts and induce nodule formation. This might be the reason for increased number of nodules in plants inoculated with rhizobium bacteria. Similar results were obtained in cowpea seeds inoculated with rhizobium showed increased nodulation compared to non inoculated seeds (Pant *et al.*, 2000). Yadav (2000) reported that cowpea plants treated with vermicompost and rhizobium showed more number of nodules compared to the control.

5.3.3 Effect of seed treatment, foliar spray of micronutrient formulation and their interaction on yield and yield attributes of cowpea

There was no considerable effect of treatment in the number of days taken for first flowering, number of seeds per pod and length of pod. However, number of pods per plant and pod weight per plant was found to be influenced by the treatments.

Considering the levels of seed treatment, number of pods per plant and pod weight per plant was found to be highest in seed treatment with micronutrient formulation at two percentage (S_3) and was on par with S_4 (seed treatment with 1.5 % micronutrient formulation). While considering foliar spray, number and weight of pods per plant were found to be highest in treatments receiving two foliar spray (F_3). Hence the interaction effect of these three traits followed the same trend with highest values recorded in S_3F_3 (seeds treated with micronutrient formulation at two percentage + two foliar spray) followed by S_4F_3 (seeds treated with micronutrient formulation at 1.5 percentage + two foliar spray). Considering the grain yield of cowpea, highest yield was obtained in S_3 , F_3 and S_3F_3 indicating the essentiality of micronutrients in increasing the yield of cowpea. The role of micronutrients in enhancing the yield and its attributes are evident. Seed treatment with micronutrients had positive influence on yield of cowpea. Seed treatment with micronutrients might had increased the seed vigour resulting in better growth and yield of plants. Micronutrients are involved in active photosynthesis and activates several enzymes by increasing the catalytic activity. It is also involved in hormonal metabolism, pollination and fertilization and enhancement of cell division which ultimately leads to increase in number and weight of pods. By enhancing the cell division and expansion vegetative growth can also be increased. It also helps in increased production of assimilates and its proper partitioning. These findings are in accordance with the results reported by Mallick and Muthukrishnan (1980). Tomato plants treated with micronutrient foliar spray showed increased fruit weight. Pranakrishna (1976) reported that pulse crops treated with sodium molybdate increased the seed yield by 45 percentage compared to the control.

Common beans treated with zinc increased the yield of crops (Kaya *et al.*, 2007). Pod yield in pea was found to be higher in seeds treated with 0.5 per cent boron solution (Kumar *et al.*, 2008). Foliar application of micronutrient mixture (zinc, copper, boron and iron) at two percentage resulted in increased yield in okra by 19 per cent (Datire *et al.*, 2010).

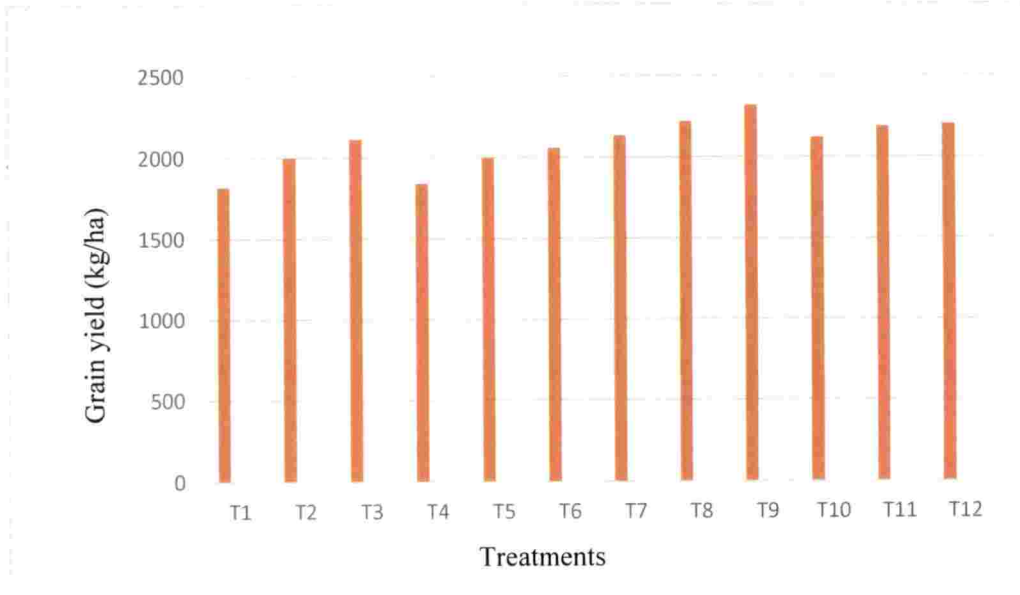


Fig. 3 Effect of treatments on grain yield of cowpea

5.3.4 Effect of seed treatment, foliar spray of micronutrient formulation and their interaction on root cation exchange capacity (root CEC)

Seed treatment and foliar spray of micronutrient formulation showed considerable influence in the cation exchange capacity of roots. Seeds treated with micronutrient mixture at 2 percentage showed higher root CEC. Double spray of micronutrient formulation showed higher root CEC compared to single spray and no foliar spray.

Interaction effect showed higher root CEC in S₃F₃ (seeds treated with micronutrient formulation at two percentage + two foliar spray) which was on par

with S₂F₃ (seed treatment with rhizobium + two foliar spray). Root cation exchange capacity is defined as the capacity of the root to exchange the cations with the soil cations. As root CEC increases the uptake of nutrients also increases which results in better growth and development. Seeds treated with micronutrient solution showed increased seed vigour and this might be the reason for better seedling growth and emergence resulting in shoot and root development. Enhanced growth of plants could have influenced the uptake of nutrients from soil by increasing the exchange of cations between root surface and soil. Rhizobium also enhances the nutrient uptake of plants, which might be the reason for higher root CEC in seeds treated with rhizobium. This results goes in line with the findings of Pattanayak *et al.* 2000. He observed that green gram seeds treated with sodium molybdate along with rhizobium showed increased nutrient uptake, plant growth and yield of crops. El-Fouly and EL-Sayed in 1997 reported that foliar spray of micronutrients increases the uptake of nutrients through soil by promoting root growth.

5.3.5 Effect of seed treatment, foliar spray of micronutrient formulation and their interaction on soil nutrient status

The effect of treatments and their interaction on the soil nutrient status is briefly discussed below.

5.3.5.1 Soil pH and EC

Soil pH and electrical conductivity were found to be non significant with respect to different treatments and their interaction. The plots treated with micronutrient solution and double foliar spray showed the lowest pH values compared to other plots. Also seeds treated with rhizobium and single foliar spray exhibited lowest pH in the interaction at 45 days after sowing and harvest. Electrical conductivity was found to be highest in seed treatment with two percentage micronutrient mixture, double foliar spray and their interaction at both 45 days after sowing and at harvest. Electrical conductivity of the soil solution was the controlled sum of cation and anion concentrations (Lipman *et al.*, 1926).

5.3.5.2 Organic carbon

Organic carbon content in the soil was found to be non significant at 45 days after sowing and harvest in treatments receiving different levels of seed treatment. Treatments receiving micronutrient foliar spray at 15 days after sowing and 30 days after sowing showed increased organic carbon content in the soil at the time of harvest. While organic carbon at 45 days after sowing was found to be non significant. Interaction effect was found to be non significant at 45 days after sowing. However, at harvest S₃F₃ also showed highest organic carbon content which was on par with seed treatment with rhizobium + two foliar spray (S₂F₃), seed treatment with 1.5 percentage micronutrient formulation + one foliar spray, two foliar spray and no foliar spray (S₄F₂, S₄F₃ and S₄F₁) and seed treatment with 2 percentage micronutrient formulation + one foliar spray (S₃F₂). Foliar spray of micronutrient formulation increased the organic carbon content in soil (Premalatha, 2016).

5.3.5.3 Available nitrogen

Available nitrogen in the soil was highest in the soil where cowpea seeds treated with rhizobium were planted (S₂) at both 45 days after sowing and harvest. Levels of foliar spray and the interaction effect was non significant in the amount of nitrogen available in the soil at both stages. Nitrogen content of soils treated with rhizobium showed higher nitrogen content compared to others. The enhanced nitrogen fixation of legumes in symbiotic association with rhizobium might be the reason for increase in the nitrogen content of soil at the time of harvest. Similar results were reported by Carsky *et al.* 2002.

5.3.5.4 Available phosphorus

Available phosphorus content in the soil was found to be the highest in treatment S₂ (seeds treated with rhizobium) at both 45 days after sowing and harvest. The reason for this might be the inoculation of rhizobium. Rhizobium has the capacity to solubilize the phosphorus present in the unavailable form to available form and make it available for plants. Nyoki and Ndakidemi (2018)

reported that soybean seeds treated with rhizobium showed increased uptake of phosphorus. Available phosphorus was found to be non significant at 45 days after sowing and at harvest for both foliar spray and interactions.

5.3.5.5 Available potassium

Available potassium in the soil was found to be highest in treatment S₃ (seed treatment with 2 percentage micronutrient mixture), F₃ (double foliar spray of micronutrient mixture) and S₃F₃ (micronutrient seed treatment at two percentage and double foliar spray) at both 45 DAS and harvest. From this it is evident that the micronutrients have capacity to increase the availability of potassium for plants. This fall in line with the findings of Sekhon and Singh (2013). They concluded that availability of potassium for the crop can be enhanced by the application of micronutrients along with macronutrients. Enhanced availability of potassium may be due to the fact that micronutrients ensure the efficient use of macronutrients (Swati *et al.*, 2011).

5.3.5.6 Available micronutrients (Fe, Mn, Zn, Cu, B and Mo)

Different seed treatments did not show any significant difference in the status of these micronutrients in soil at 45 days after sowing except in the case of boron. Boron was found to be superior in S₃ (seed treatment with micronutrient mixture at 2 %). Significant difference in the concentration of iron, manganese, zinc and molybdenum in soil were recorded at harvest. Content of iron, zinc and manganese in soil was superior in S₃ which was on par with S₄ (seed treatment with micronutrient mixture at 2% and 1.5% respectively). Molybdenum content at harvest recorded highest value in S₃. This indicates that seed treatment had pronounced effect on micronutrient concentration in soil. Pattanayak *et al.* 2000; Nyoki and Ndakidemi, 2018 reported similar results.

Considering different levels of foliar spray, F₃ (double foliar spray of micronutrient mixture) recorded the highest status of iron, manganese, zinc, boron and copper at 45 days after sowing. At harvest iron, manganese and copper were

found to be superior over other treatments and zinc content was highest in F₃ which was on par with F₂ (single foliar spray). Boron content was found to be non significant at harvest stage. Molybdenum was significant only at harvest receiving treatment F₃. Thus foliar spray of micronutrient mixture provided a significant role in increasing the micronutrient concentration in soil also. Similar results were reported by Alva (2009), Thiyageshwari and Ramanathan (2001), Ravi *et al.* (2008).

Treatment interactions showed significant changes in the availability of micronutrients. Manganese, zinc, copper and boron were found to be higher in treatments receiving both seed treatment and two foliar spray of micronutrient mixture (S₃F₃) at 45 days after sowing. Iron showed significantly higher values in S₁F₃ (no seed treatment and double foliar spray). At harvest iron and manganese status were found to be higher in treatment receiving both micronutrient seed treatment (S₃) and two foliar spray. Zinc, copper, boron and molybdenum were non significant with respect to treatment interactions at harvest.

5.3.6 Effect of seed treatment, foliar spray of micronutrient formulation and their interaction on nutrient content of cowpea leaves

5.3.6.1 Nitrogen content in leaf

Nitrogen content in the leaf was found to be non significant with respect to different levels of seed treatment. Among the foliar spray, treatments receiving two foliar spray of micronutrient mixture at 15 days after sowing and 30 days after sowing showed considerably higher nitrogen content in leaves compared to other levels of foliar spray. In treatment interactions, treatments receiving micronutrient seed treatment along with two foliar spray of micronutrient mixture showed highest nitrogen content in cowpea leaves while the lowest concentration was obtained in control. The essentiality of micronutrients in plant is evident in the result of the study. Similar results were obtained by Premalatha (2016) in banana and Ashwini (2018) in okra.

5.3.6.2 Phosphorus content in leaf

Phosphorus content in leaf showed considerable difference with respect to treatments and their interactions. Seed treatment with 2 % micronutrient mixture showed highest phosphorus content in cowpea leaves. Among foliar spray phosphorus was found to be highest in two foliar sprays of micronutrient mixture. In the interaction effect combination of these two treatments (seed treatment and foliar spray of micronutrient mixture) showed highest value which was significantly different from other interactions. Hence it is evident that micronutrients help in the uptake of phosphorus by plants. These nutrients also promote phosphate metabolism and translocation to different parts of plants. Similar results were reported by Ashwini (2018). She reported an increase in the phosphorus content of bhindi leaves by foliar application of micronutrient mixture.

5.3.6.3 Potassium content in leaf

Average potassium concentration in cowpea leaves followed the similar trend as in leaf phosphorus content. Potassium concentration was found to be highest in both seed treatment and double foliar spray of micronutrient mixture as well as their interaction. Thus the beneficial effect of micronutrients in enhancing potassium concentration in leaves of cowpea is revealed in this study. The presence of boron in the micronutrient formulation might be the reason for the enhanced potassium uptake in plants. Similar results were obtained by Ashwini (2018) and Premalatha (2016).

5.3.6.4 Calcium, magnesium and sulphur concentration in leaf

Among the secondary nutrients, calcium was found to be significant only with the application of micronutrient mixture as foliar spray. Magnesium was showed considerable difference among the interaction effects and was found to be superior in treatment receiving micronutrient seed treatment (S₃) and two sprays of micronutrient formulation. Sulphur was non significant among the treatments. Premalatha (2016) reported an increase in calcium content of banana leaves sprayed with 3 % micronutrient formulation.

5.3.6.5 Micronutrient concentration in leaf at harvest

The concentration of iron, manganese, zinc and boron in the leaf were influenced by different seed treatments, levels of foliar sprays and their interactions. Fe, Mn and Zn were recorded maximum in seed treatment receiving 2 % micronutrient formulation (S₃). While B was recorded maximum in S₃ and was on par with S₄ (seed treatment with 1.5 % micronutrient formulation). Similar results were reported by Pattanayak *et al.*, 2000.

Iron, manganese, zinc and boron concentration of cowpea leaf was significantly influenced by the application of micronutrient mixture at 15 days after sowing and harvest. Similar results were recorded by Premalatha (2016) in tissue culture banana and Ashwini (2018) in bhindi. Application of zinc sulphate along with borax was found to increase the nutrient concentration in plants (Ejaz *et al.*, 2011). Foliar application of 0.5 % borax in potato at 35 days after sowing increased the nutrient concentration in plants (Das, 1977).

Iron, manganese and zinc concentration was found to be significant in the interaction S₃F₃ (seed treatment receiving 2 % micronutrient formulation + two foliar sprays). The application of micronutrients as seed treatment and foliar spray resulted in increased concentrations of these nutrients in the leaves.

5.3.7 Effect of seed treatment, foliar spray of micronutrient formulation and their interaction on nutrient content of cowpea grain

5.3.7.1 Nitrogen, phosphorus, potassium concentration and uptake in grain

Seed treatment of cowpea showed significant difference in the concentration of nitrogen and potassium in grain while phosphorus concentration was found to be non significant. Nitrogen content in grain was maximum in S₂ (seed treatment with rhizobium). This might be due to the positive effect of rhizobia on increasing the nitrogen content in grain. Thilakaratna *et al.* (2019) reported that in common bean, seed treatment with rhizobium showed increased nitrogen content in grain. Considering the potassium concentration in grain, seed treatment with micronutrient mixture at 2 % showed highest concentration. The presence of boron

in the micronutrient mixture might be the reason for increase in potassium content of grain as it enhances the potassium uptake by plants.

Among the foliar spray nitrogen and potassium concentration were found to be highest in treatments receiving two foliar sprays of micronutrient mixture. Thus application of micronutrient spray could be beneficial in increasing the grain concentration of nitrogen and potassium. Since it increases the uptake of nutrients, the concentration of nutrients in plant tissues might be increased. Gad (2012) reported that application of molybdenum increased the concentration of nitrogen and potassium in groundnut seeds.

Interaction effect of treatments also showed significant difference in the concentration of nitrogen and potassium in grain. Nitrogen concentration was found to be highest in S₂F₃ (seed treatment with rhizobium + two foliar sprays). Seed treatment with rhizobium might be the reason for increase in nitrogen content which is enhanced by the foliar spray of micronutrient mixture. Potassium content in grain was found to be highest in S₃F₃ which was on par with S₄F₃. Micronutrients, especially boron help in the uptake and translocation of potassium in plants. This might be the reason for increased potassium content in grains supplied with micronutrient seed treatment and foliar spray. Pattanayak *et al.* 2000, Thiyaeswari and Ramanathan, 2001 and Hanwate *et al.*, 2018 reported similar results.

Nitrogen, phosphorus and potassium uptake by grains of cowpea showed significant difference among different treatments and their interactions. The uptake of these nutrients were found to be higher in treatments receiving seed treatment with 2% micronutrient and two foliar sprays. This indicates the essentiality of micronutrients in nutrient uptake of grains. Seed treatment with micronutrients increased the uptake of nutrients (Pattanayak *et al.*, 2000). Foliar spray of micronutrients increases the uptake of nitrogen, phosphorus and potassium (Choudhary *et al.*, 2017). In the interaction effect phosphorus and potassium uptake were found to be highest in treatment receiving the best micronutrient seed treatment (S₃) and two foliar sprays. The interaction effect of this two main treatments resulted in highest nutrient uptake.

5.3.7.2 Calcium, magnesium, sulphur concentration and uptake in grain

Treatments and their interaction showed significant difference in the concentration of calcium in cowpea grain, while magnesium and sulphur concentration were found to be non significant in main treatments and their interactions. Calcium content in grain was higher in seeds receiving micronutrient treatment (2 %) and two foliar sprays. In the interaction effect treatment S₃F₃ resulted in higher calcium content. Foliar application of micronutrients increased the nutrient concentration in potato (Das, 1977).

Considering the nutrient uptake, calcium uptake was found to be higher in seed treatment receiving 2 % micronutrient mixture and two foliar sprays and their interactions. Magnesium and sulphur uptake were higher in S₃ which was on par with S₄ (seed treatment with 1.5 % micronutrient formulation). Foliar spray F₃ recorded maximum uptake of magnesium and sulphur in grains which was on par with F₂ (single foliar spray). Interaction was found to be non significant regarding uptake of magnesium and sulphur.

5.3.7.3 Micronutrient concentration and uptake in grain

Seed treatment showed significant difference in the concentration of iron, manganese, zinc, copper and boron content in grain and were found to be highest in seeds receiving 2 % micronutrient formulation. Hence it is revealed that micronutrient seed treatment has the capacity to increase the nutrient concentration in grain. Similar results were obtained by Pattanayak *et al.*, 2000. Maize seeds treated with micronutrient increased the concentration of iron, manganese and zinc content in grains (Harris *et al.*, 2007).

Foliar application of micronutrient mixture also showed significant difference in the amount of iron, manganese, zinc, copper and boron concentration in grain. Treatments receiving foliar spray of micronutrient mixture at 15 and 30 days after sowing showed higher concentration of these micronutrients. Divyasree, 2018 reported that foliar application of micronutrient mixture containing Fe, Mn,

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Zn, Cu and B significantly increased the respective nutrient concentrations in grains of mung bean.

Similar results were obtained in the interaction effects. Seed treatment with the best micronutrient formulation and two foliar spray recorded highest value of micronutrients in grain except in the case of molybdenum. The combined effect of the individual treatments might be the reason for this.

Considering the trends of nutrient uptake, uptake of iron, zinc and boron was found to be highest in seed treatment receiving 2 % micronutrient mixture, two foliar sprays and their interactions. Uptake of manganese was found to be highest in the interaction receiving seed treatment with 2 % micronutrient formulation and two foliar spray. Thus micronutrient application resulted in better uptake of these nutrients in the grains of cowpea. These findings are supported by the results obtained by Pattanayak *et al.*, 2000 and Ravi *et al.*, 2008. Hanwante *et al.* 2018 reported that application of micronutrients as seed treatment along with foliar spray increased the uptake of micronutrients in soybean.

5.3.8 Incidence of pest and disease

There was no severe incidence of pest and disease observed in the field. Crops were infested with thrips during initial growth period and was controlled by the application of nimbecidine. A few plants were infected with collar rot which was controlled by drenching of copper oxychloride.

Summary

6. SUMMARY

The salient findings of the study entitled “Evaluation of micronutrient formulation in cowpea (*Vigna unguiculata* L. Walp)” are summarized in this chapter.

The investigation was carried out at College of Agriculture, Padannakkad and Regional Agricultural Research Station (RARS), Pilicode during 2017 to 2019. The objectives of the study were to evaluate the effect of micronutrient formulation through seed treatment and foliar nutrition on growth, nodulation and yield of cowpea (*Vigna unguiculata* L. Walp) and to study its effect on plant nutrient uptake and residual soil nutrient status. The study was conducted in three parts - Standardization of micronutrient formulation for cowpea, study of time and dose of micronutrient formulation for seed treatment in cowpea and field experiment to evaluate the performance of micronutrient formulation in cowpea.

Micronutrient formulation was standardized by two research workers earlier. Micronutrient formulation is a mixture of two solutions *viz.* solution A and solution B. Solution A is a mixture of zinc sulphate, boric acid, copper sulphate, manganous sulphate, ferrous sulphate and ammonium molybdate whereas solution B is an organic acid which act as a chelate. These two solutions should be mixed in a specific ratio at the time of application because of the poor shelf life of micronutrient formulation. A combination of Solution A and Solution B were prepared so that it can be supplied to farmers as micronutrient formulation which reduces the difficulty of mixing solutions during field application. The micronutrient formulation can be diluted to different concentrations for seed treatment and foliar spray.

Laboratory study was carried out at COA, Padannakkad with different concentrations of micronutrient formulations. The objective of the study was to standardize the time and dose for seed treatment in cowpea using micronutrient formulation. The experiment was carried out in completely randomized design with 7 treatments and 3 replications which included seed treatment with micronutrient

formulation @ 0.25, 0.50, 0.75, 1, 1.5, 2 % as T₁, T₂, T₃, T₄, T₅ and T₆ respectively. T₇ (seed treatment with water) was the control. Seeds treated with different concentrations of micronutrient formulation were kept in petri plates for germination studies which was then planted in pots and observations were recorded upto three leaf stage. Observations on germination percentage, number of days taken for germination, seedling length at three leaf stage and seedling vigour index were recorded. Treatment T₆ (micronutrient formulation @ 2 %) showed highest seedling length at three leaf stage and seedling vigour index which was followed by treatment T₅ (micronutrient formulation @ 1.5 %). Thus the best and second best treatment from seed treatment study was concluded as T₆ and T₅ respectively.

The field experiment was carried out at Regional Agricultural Research Station (RARS), Pilicode to evaluate the effect of micronutrient formulation on cowpea. The experiment was conducted in randomized block design with 12 treatments replicated three times. The treatment consisting of combination of four levels of seed treatment (no seed treatment, seed treatment with rhizobium, seed treatment with best concentration from experiment 1 and second best seed treatment from experiment 1) and three levels of foliar application of micronutrient (no foliar application, one foliar application at 15 DAS and two foliar applications at 15 and 30 DAS). Manures and fertilizers application and other cultural practices were followed as per POP, KAU (2016) for all the treatments uniformly.

The salient findings from field experiment are summarized below

1. Different levels of seed treatment showed significant improvement on growth characters like plant height and stem diameter at flower bud initiation and harvest. Seed treatment with micronutrient formulation at 2 % (S₃) and 1.5 % (S₄) showed maximum plant height at flower bud initiation. At harvest S₃ showed highest plant height compared to others. Stem diameter was found to be higher in S₃, S₄ and S₂ (seed treatment with rhizobium) at flower bud initiation stage. At harvest S₃ recorded the

maximum stem diameter. Whereas number of branches was non significant with respect to seed treatment.

2. Levels of foliar spray showed that foliar spray of micronutrient formulation at 15 DAS and 30 DAS (F₃) produced maximum plant height at flower bud initiation stage and harvest. Stem diameter was found to be highest in treatments receiving F₃ and F₂ (foliar spray at 15 DAS) during flower bud initiation stage while at harvest F₃ recorded the highest value.
3. Interaction effect of treatments showed that plants receiving 2 percent seed treatment along with two foliar spray (S₃F₃) produced maximum plant height at both the stages. Seed treatment with rhizobium + two foliar spray (S₂F₃) recorded maximum stem diameter at flower bud initiation stage and harvest.
4. Dry matter production was found to be highest in plants receiving seed treatment with micronutrient formulation at two percentage (S₃) among the seed treatments. Foliar spray with micronutrient formulation at 15 DAS and 30 DAS produced maximum dry matter among the different levels of foliar spray. Plants receiving seed treatment with 2 percent micronutrient formulation along with two foliar spray produced highest dry matter in the interactions compared to control.
5. Seed treatment with rhizobium produced more number of nodules per plant compared to other seed treatments. Effect of foliar spray and interaction was not significant with respect to number of nodules per plant. Also fresh weight and dry weight of nodules were not affected by different treatments.
6. Among the seed treatment S₃ (seed treatment with micronutrient formulation at two percentage) produced maximum number of pods per plant, pod weight per plant and grain yield. Similarly, treatments receiving two foliar spray and seeds treated with micronutrient formulation at two percentage + two foliar spray showed maximum yield.
7. Seed treatment and foliar spray of micronutrient formulation showed considerable influence in the cation exchange capacity of roots. Seeds treated with micronutrient formulation at 2 percentage showed higher root

- CEC. Double spray of micronutrient formulation showed higher root CEC compared to single spray and no foliar spray. In the interaction effect seeds treated with micronutrient formulation at two percentage + two foliar spray recorded highest CEC which was on par with rhizobium + two foliar spray.
8. The effects of treatment application on soil nutrient status were studied at 45 DAS and at harvest stage. The pH and EC were found to be no significant. While organic carbon was found to be superior in F₃ and interaction S₃F₃ at harvest.
 9. Treatments significantly influenced the available nitrogen, phosphorus and potassium in soil at 45 DAS and at harvest. S₂ showed highest available nitrogen and phosphorus at both stages. Available potassium was found to be highest in S₃, F₃ and S₃F₃ at both the stages.
 10. Treatments significantly influenced the micronutrient concentration in soil at 45 DAS and at harvest. At 45 DAS boron content was highest in S₃. At harvest iron, zinc and manganese was highest in S₃ and was on par with S₄ while Mo was highest in S₃. Considering foliar spray of micronutrient formulation, iron, manganese, zinc, boron and copper in soil were in F₃ at 45 DAS. At harvest iron, manganese, copper, molybdenum and zinc were highest in F₃. In the interaction S₃F₃, manganese, zinc, copper and boron were higher at 45 days after sowing. At harvest iron and manganese recorded highest value in S₃F₃.
 11. Seed treatment influenced the phosphorus and potassium content in leaf which was higher in seed treatment receiving micronutrient formulation at 2%. Whereas, nitrogen content in leaf was not influenced by seed treatment.
 12. Two foliar spray of micronutrient formulation increased the nitrogen, phosphorus and potassium content in cowpea leaves. In the interaction S₃F₃ recorded highest NPK content in cowpea leaves.
 13. Treatments also influences the concentration of calcium and magnesium content in leaf. Foliar spray of micronutrient formulation increased the calcium content in leaf. Magnesium was found to be higher in treatment S₃F₃. However, sulphur content in leaf was found to be non significant.

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14. Seed treatment with 2 % micronutrient formulation recorded maximum concentrations of iron, manganese and zinc in cowpea leaves. Boron was highest in S₃. Double foliar spray of micronutrient formulation showed highest concentrations of Fe, Mn, Zn and B concentrations in cowpea leaf. In the interaction seed treatment receiving 2 % micronutrient formulation + two foliar sprays showed highest content of Fe, Mn and Zn in leaves.
 15. Nitrogen content in grain was found to be highest in S₂ (seed treatment with rhizobium) while potassium was highest in S₃. Phosphorus concentration in grain was found to be non significant with respect to seed treatment. Foliar spray of micronutrient formulation at 15 DAS and 30 DAS showed highest concentrations of nitrogen and potassium in grain. In the treatment interaction nitrogen was higher in S₂F₃ (seed treatment with rhizobium + two foliar sprays) and potassium in S₃F₃. Similarly, the uptake of N, P and K was higher in seed treatment with 2 % micronutrient formulation and two foliar sprays. P and K uptake in the interaction was higher in S₃F₃.
 16. Calcium content in grain was higher in S₃, F₃ and S₃F₃ and its uptake also followed the same trend. Magnesium and sulphur content in grain was found to be non significant with respect to different treatments while its uptake was higher in S₃ and F₃.
 17. The concentration of micronutrients viz., Fe, Mn, Zn, Cu and B were found to be higher in seed treatment with micronutrient formulation at 2 %, double foliar spray of micronutrient formulation and their interactions. Uptake of Fe, Zn and B was higher in seed treatment with micronutrient formulation at 2 %, double foliar spray of micronutrient formulation and their interactions. While Mn uptake was higher only at interaction S₃F₃.

Conclusion

The results obtained from the investigation revealed that the application of micronutrient formulation as foliar spray (15 and 30 DAS) along with seed treatment (2 % micronutrient formulation) was found to be highly effective in increasing the growth and yield characters of cowpea. The uptake of nutrients in grain was also found to increase with the application of micronutrient formulation enhancing the nutritional quality of grain. Considering the benefit cost ratio treatment T₉ (seeds treated with micronutrient formulation at two percentage along with two foliar spray) was found to be the most effective and profitable method.

Future line of work

- Micronutrient seed treatment along with rhizobium inoculation can be tried in cowpea
- Effect of other varieties and hybrids are to be studied for knowing crop performance to micronutrient formulation

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References

7. REFERENCES

- Afria, B.S., Pareek, C.S., Garg, D.K., and Singh, K. 1999. Effect of foliar spray of micronutrients and their combinations on yield of Pomegranate. *Ann. Arid Zone* 38 (2):189–190.
- Agrios, N.G. 2005. *Plant pathology*. Elseiver-Academic press p. 635.
- Ajouri, A., Asgedom, H., and Becker, M. 2004. Seed priming enhances germination and seedling growth of barley under conditions of P and Zn deficiency. *J. Plant Nutr. Soil Sci.* 167: 630-636.
- Alam, S.S., Moslehuddin, A.Z.M., Islam, M.R., and Kamal, A.M. 2010. Soil and foliar application of nitrogen for Boro rice (BRRIadhan 29). *J. Bangladesh Agril.Univ.* 8 (2): 199–202.
- Ali, S., Shah, A., Arif, M., Mirja, G., Ali, I., Khan, M.F.A., and Moula, N. 2009. An Introduction of Plant Nutrients and Foliar Fertilization: A Review. *J. Agric.* 25: 1–10.
- Alloway, W.H. 1986. Soil-plant-animal and human interrelationships in trace element nutrition. In: Mertz, W.(ed.), *Trace elements in Human and Animal nutrition*. Academic press, Oriando, San Diego, New York, Austin, London, Montreal, Sydeney, Tokyo, Toronto. pp. 465–488.
- Alva, A.K. 2009. Effects of various pre-plant and in-season nitrogen management practices for potatoes on plant and soil nitrogen status. *Commun. Soil Sci. Plant Ann.* 40(16): 649-659.
- Amrutrao, M.V. 2018. Effect of nitrogen and phosphorus levels with and without rhizobium on forage yield and quality of summer cowpea (*Vigna unguiculata* (L.) Walp.) under middle Gujarat conditions. M.Sc. (Ag) thesis, Anand Agricultural University, Gujarat. 88 p.

- Anitha, S., Sreenivasan, E., and Purushothaman. 2005. Response of cowpea (*Vigna unguiculata* L.) to foliar nutrition of zinc and iron in the oxisols of Kerala. *Legume Res.* 28 (4): 294–296.
- Arnon, D.I. and Stout, P.R. 1939. Molybdenum as an essential element for higher plants. *Plant Physiol.* 14: 599–602.
- Ashwini, B.N. 2018. Response of bhindi (*Abelmoschus esculentus* L.Moench) to fertigation and foliar nutrition in red loam soil of Kasaragod. M.Sc.(Ag) thesis, Kerala Agricultural University, Thrissur, 207p.
- Babaeian, M., Tavassoli, A., Ghanbari, A., Esmailian, Y., and Fahimifard, M. 2011. Effects of foliar micro-nutrient application on osmotic adjustments, grain yield and yield components in sunflower (Alstar cultivar) under water stress at three stages. *Afr. J. Agric. Res.* 6 (5): 1204–1208.
- Babaeva, E.Y., Volobueva, V.F., Yagodin, B.A., and Klimakhin, G.I. 1999. Sowing quality and productivity of *Echinacea purpurea* in relation to soaking the seed in manganese and zinc solutions. *Izvestiya Timiryazevskoi Sel'skokhozyaistvennoi Akademii.* 4: 73-80.
- Baligar, V.C., Fageria, N.K., and Elrashidi, M.A. 1998. Toxicity and nutrient constraints on root growth. *Hortic. Sci.* 33: 960–965.
- Baloch, Q.B., Chachar, Q.I., and Tareen, M.N. 2008. Effect of foliar application of macro and micro nutrients on production of green chilies (*Capsicum annuum* L.). *J. Agric. Tech.* 4 (2): 177–184.
- Barber, S.A. 1995. *Soil Nutrient Bioavailability: A Mechanistic Approach* (2nd ed.) Wiley, New York. 414p.
- Battacharya, B., Chakraborty, A., Bandyopadhyay, S., and Samanta, D. 1997. Effect of S, Zn and Mo on groundnut (*Arachis hypogaea* L.) grown with saline water irrigation in coastal saline soil of West Bengal. *Indian Agric.* 41(2): 145-153.

- Bell, R.W. and Dell, B. 2008. *Micronutrients for Sustainable Food, Feed, Fibre and Bioenergy Production* (1st Ed.). IFA, Paris, France.
- Benepal, P.S. 1967. Influence of micronutrients on growth and yield of potatoes. *Am. Potato J.* 44: 363.
- Bernal, M., Cases, R., Picorel, R., and Yruela, I. 2007. Foliar and root Cu supply affect differently Fe and Zn-uptake and photosynthetic activity in soybean plants. *Environ. Exp. Botany* 60: 145–150.
- Bhagiya, S.R., Polara, K.B., and Polara, J.V., 2005. Effect of B and Mo on yield, quality and nutrient absorption by groundnut. *Adv. Plant Sci.* 18(2): 803.
- Bhargava, B. S. and Raghupathi, H. B. 1995. Analysis of plant material for macro and micronutrients. In: Tandon, H.L.S. (ed.), *Methods of analysis of soils, plants, water and fertilizers*. Malhotra publishing house, New Delhi, pp. 61-62.
- Bingham, F. T. 1982. Boron. In: Page, A. L. (ed.), *Methods of soil analysis* (2nd ed.). *Am. Soc. of Agron.* Madison, USA, p.438.
- Boorboori, M.R., Asli, E.D., and Tehrani, M. 2012. Effect of micronutrient application by different methods on yield, morphological traits and grain protein percentage of barley (*Hordeum vulgare* L.). *J. Adv. Environ. Biol.* 6 (2): 740.
- Black, C. A., Evans, D. D., Ensminger, L. E., White, J. L., and Clark, F. E. 1965. *Methods of soil analysis*. *Am. Soc. Agron.* Madison, USA, p.156.
- Bressani, R., 1985. Nutritive value of cowpea. In: Singh, S.R. and Rachie, K.O. (eds), *Cowpea research, production and utilization*. Wiley, New York, pp. 353-359.
- Bukvic, G., Antunovic, M., Popovic, S., and Rastija1, M. 2003. Effect of P and Zn fertilization on biomass yield and its uptake by maize lines (*Zea mays* L.). *Plant Soil Environ.* 49 (11): 505–510.

- 123
- Carsky, R.J., Vanlauwe, B., and Lyasse, O. 2002. Cowpea rotation as a resource management technology for cereal-based systems in the savannas of West Africa. In: Fatokun, C.A., Tarawali, S.A., Singh, B.B., Kormawa, P.M., and Tamo, M. (eds) *Challenges and opportunities for enhancing sustainable cowpea production*. International Institute of Tropical Agriculture, Ibadan, Nigeria, pp 252–266.
- Chatterjee, R. and Bandyopadhyay, S. 2017. Effect of boron, molybdenum and biofertilizers on growth and yield of cowpea (*Vigna unguiculata* L. Walp.) in acid soil of eastern himalayan region. *J. Saudi Soc. Agri Sci.* 16: 332-336.
- Chen, L.S., Figueredo, A., Villani, H., Michajluk, J., and Hungria, M. 2002. Diversity and symbiotic effectiveness of rhizobia isolated from field-grown soybean nodules in Paraguay. *Biol. Fertil. Soils* 35: 448-457.
- Choudhary, S.K., Jat, M.K., and Mathur, A.K. J. 2017. Effect of micronutrient on yield and nutrient uptake in sorghum. *Pharmacognosy Phytochemistry* 6(2): 105-108.
- Clark, R.B. and Zeto, S.K. 2000. Mineral acquisition by *Arbuscular mycorrhizal* plants. *J. Plant Nutr.* 23: 867–902.
- Dadhich, S.K. and Somani, L.L. 2007. Effect of integrated nutrient management in Soybean-Wheat crop sequence on the yield, micronutrient uptake and post-harvest availability of micronutrients on Typic Ustochrepts soil. *Acta Agronomica Hungarica* 55 (2): 205–216.
- Das, C.R. 1977. Study of the effect of foliar application of B, Mo, Ca and planofix on growth, yield and quality of potato (*Solanum tuberosum*). *Seminar on recent advances in plant science*, Calcutta, India 135 p. *Field Crop Abstr.* 30 (1): 629.
- Datire, R.B., Laware, S.L., and Apparao, B.J. 2010. Effect of Organically Chelated Micronutrients on Growth and Productivity in Okra. *Asian J. Exp. Biol. Sci. Spec.* 200: 115–117.

- De, N. and Rai, M. 2005. Micronutrient disorders and management in vegetable crop: Review. *Veg. Sci.* 32 (1): 1-10.
- Deb, P., Das, A., Ghosh, S.K., and Suresh, C.P. 2010. Improvement of seed germination and seedling growth of papaya (*Carica papaya* L.) through different pre-sowing seed treatments. *Acta Hort.* 851: 313-316.
- Deshmukh, M.C. and Joshi, R.N. 1973. Effect of rhizobial inoculation on the extraction of protein from the leaves of cowpea. *Indian J. Agric. Sci.* 43(6): 539-542.
- Divyashree, K.S. 2018. Effect of different methods of micronutrients mixture application on growth and yield of mungbean in Cauvery command area. *J. Pharmacognosy Phytochemistry* 7(3): 3578-3582.
- Dobereiner, J. 1997. Biological nitrogen fixation in the tropics: Social and economic contributions. *Soil Bid. Biochem.* 29(36): 771- 774.
- Donald, C.M., and Spencer, D. 1951. The control of molybdenum deficiency in subterranean clover by pre-soaking the seed in sodium molybdate solution. *Australian J. Agric. Res.* 2: 295-301.
- Durairaj, S. 1993. Effect of lignite humic acid and zinc on the performance of rice in a coastal saline soil. M.Sc (Agri) thesis, Annamalai University, Tamilnadu. 89p.
- Edmond, J.B., Sen, T.L., Andrews, F.S., and Holfacre, R.G. 1997. *Fundamentals of Horticulture* (4th Ed.). McGraw Hill Book Co., New York.
- Ejaz, M., Rehman, U.S., Waqas, R., Manan, A., Imran, M., and Bukhari, A.M. 2011. Combined efficacy of macronutrients and micronutrients as foliar application in tomato grown by vegetable forcing. *Int. J. Vet. Med. Sci.* 5: 327-335.

- El-Fouly, M.M. and El-Sayed A.A. 1997. *Foliar fertilization: An environmentally friendly application of fertilizers*. Dahlia Greidinger International Symposium on "Fertilization and Environment" 24-27 March, pp. 346–357.
- El-Magid, A.A.A., Knany, R.E., and El-Fotoh, H.G.A. 2000. Effect of foliar application of some micronutrients on wheat yield and quality. *J. Ann. Agric. Sci.* 1: 301–313.
- Emmel, R. H., Solera, J. J., and Stux, R. L. 1977. *Atomic absorption methods manual*. Instrumentation Laboratory Inc., Wilmington, pp. 67-190.
- Engelhard, A.W. 1990. *Soil borne Plant Pathogen, Management of Disease with Macro and Micro Elements*. American Phytopathological Society Press, St. Paul, MN. 217pp.
- Evans, I., Solberg, E., and Huber, D.M. 2007. Copper and plant diseases. In: Dantnoff, L.E., Wade, H.E., and Huber, D.M. (eds.). Mineral nutrition and plant diseases. *Am. Phytopathol. Soc.* pp. 177–188.
- Fageria N.K., Baligar V.C., and Clark R.B. 2002. Micronutrients in crop production. *Adv. Agron.* 77: 185 – 268.
- Fageria, N.K., Valigar, V.C., and Wright, R.J. 1997. Soil environment and root growth dynamics of field crops. *Recent Res. Dev. Agron.* 1: 15–58.
- Farooq, M., Wahid, A., and Siddique, K. H. M. 2012. Micronutrient application through seed treatments - a review. *J. Soil Sci.Plant Nutr.* 12 (1): 125-142.
- Fernandez, V. and Eichert, T. 2009. Uptake of hydrophilic solutes through plant leaves: current state of knowledge and perspectives of foliar fertilization. *Crit. Rev. Plant Sci.* 28: 36–68.
- Francois, L.E. 1989. Boron tolerance of snap bean and cowpea. *J. Am. Soc. Hortic. Sci.* 114(4): 615-619.

- Gad, N. 2012. Influence on molybdenum on groundnut production under different nitrogen levels. *World J. Chem.* 7(2): 64-70.
- Ganjir, H., Abed, I.A., and Rutherford, W. 1973. Status of micronutrients in Indian soils. *Plant Soil* 57 (4): 208–215.
- Giller, K. E. and Cadisch, G. 1995. Future benefits from biological nitrogen fixation: An ecological approach to agriculture. *Plant Soil* 174: 255-277.
- GOI [Government of India]. 2016. *Agricultural Statistics at a glance 2016* [online]. Available: <http://eands.dacnet.nic.in> [11 January 2018].
- GOK [Government of Kerala]. 2016-2017. *Agricultural Statistics 2016-2017* [online]. Available: www.ecostat.kerala.gov.in/images/pdf/publications/Agriculture/.../rep_agristat_1516.pdf [11 January 2018].
- Graham, D.R. 1983. Effects of nutrients stress on susceptibility of plants to disease with particular reference to the trace elements. *Adv. Bot. Res.* 10: 221–276.
- Graham, R.B. and Webb, M.J. 1991. Micronutrients and disease resistance and tolerance in plants. In: Mortvedt, J.J., Cox, F.R., Shuman, L.M., and Welch, R. M. (eds), *Micronutrients in Agriculture*, (2nd Ed.). SSSA, Madison, WI. pp. 329– 370.
- Grewal, H.S., Graham, R.D., and Rengel, D. 1996. Genotypic variation in zinc efficiency and resistance to crown rot disease (*Fusarium graminearum*) in wheat. *Plant Soil* 186: 219–226.
- Gris, E. 1843. Memoir relatif a l' action des composes soluble ferrugineux sur la vegetation. *Comptes Rendus de l'Acad. Sci.* 17: 679–686.
- Gupta, N.K. and Gupta, S. 2005. *Plant physiology*. Oxford & IBH publishing co.pvt.ltd. 580p.
- Gupta, U.C. 1993. Factors affecting boron uptake by plants. In: Gupta, U.C. (ed.), *Boron and its role in crop production*, CRC Press, Boca Raton, FL. Pp. 87–123.

- 129
- Gupta, V.K. 1995. Micronutrients research. In: Tandon, H.L.S. (ed.), *Micronutrient Research and Agricultural Production*. Fertilizer Development and Consultation Organisation, New Delhi, India, pp. 132–160.
- Gupta, V.K. and Gupta, S.P. 1985. Influence of zinc sources on the yield and zinc nutrition on sodic soils. *Ann. Arid Zones* 24: 63–67.
- Hall, A.E. 2004. Breeding for adaptation to drought and heat in cowpea. *Eur J Agron* 21:447–454.
- Hall, A.E. and Ehlers, J.D. 1997. Cowpea (*Vigna unguiculata* L. Walp.). *Field Crops Res.* 53: 187-204.
- Hall, A.E., Ismail, A.M., Ehlers, J.D., Marfo, K.O., Cisse, N., Thiaw, S., and Close, T.J. 2002. Breeding cowpeas for tolerance to temperature extremes and adaptation to drought. In: Fatokun, C.A., Tarawali, S.A., Singh, B.B., Kormawa, P.M., and Tamo, M. (eds) *Challenges and opportunities for enhancing sustainable cowpea production*. International Institute of Tropical Agriculture, Ibadan, Nigeria, pp 14–21.
- Hanwate, G. R., Giri, S. N., and Yelvikar, N. V. 2018. Effect of foliar application of micronutrients on nutrient uptake by soybean crop. *Int. J. Pure App. Biosci.* 6 (5): 261-265.
- Harris, D., Rashid, A., Miraj, G., Arif, M., and Yunas, M. 2008. A 'On-farm' seed priming with zinc in chickpea and wheat in Pakistan. *Plant Soil* 306: 3-10.
- Harris, D., Rashid, A., Miraj, G., Arif, M., Shah, H. 2007. A On-farm seed priming with zinc sulphate solution - A cost-effective way to increase the maize yields of resource-poor farmers. *Field Crops Res.* 10: 119-127.
- Hashemy, M.K., Malakoty, M.J., and Tabatabaey, V.S.J. 1998. Effect of foliar application of micronutrients in quality and quantity potato crop in Eastern Azarbyjan Province. *J. Sci. Res. Soil Water* 12: 44–55.

- Hasslett, B.S., Reid, R.J., and Rengel, Z. 2001. Zinc mobility in wheat: uptake and distribution of zinc applied to leaves or roots. *Ann. Bot.* 87 (3): 379–386.
- House, W.A. and Welch, R.M. 1989. Bioavailability of and interaction between zinc and selenium in rats fed wheat grain intrinsically labeled with ⁶⁵Zn and ⁷⁵Se. *J. Nutr.* 119: 916–921.
- Hristozkova, M., Geneva M., and Stancheva, I. 2006. Response of pea plants (*Pisum sativum* L.) to reduced supply with molybdenum and copper. *Int. J. Agric. Biol.* 8(2): 218-220.
- Huber, D.M. 1980. The role of mineral nutrition in defense. In: Hosfall, J.G. and Cowling, E.B. (eds), *Plant Pathology: An Advanced Treatise*, AP, NY. pp. 381–406.
- Huber, D.M. and Graham, R.D. 1999. The role of nutrition in crop resistance and tolerance to disease. In: Rengel, Z. (ed.), *Mineral nutrition of crops fundamental mechanisms and implications*. Food product press, New York, pp. 205–226.
- Huber, D.M. and Wilhelm, N.S. 1988. The role of manganese in resistance to plant diseases. In: Graham, R.D., Hannam, R., and Uren, N.C. (eds), *Manganese in Soils and Plant*. Kluwer Academic, Dordrecht, The Netherlands. pp.155–173.
- Hussanin, M., Ayaz Khani, M., Khan, M.B., Farooq, M., and Farooq, S. 2012. Precision Farming: A New Approach. *J. Rice Sci.* 19: 24–30.
- Issac, R. A. and Kerber, J. D. 1971. Atomic absorption and flame photometry techniques and uses in soil, plant and water analysis. In: Walsh, L. M. (ed.). *Instrumental methods for analysis of soil and plant tissue*. Soil Science Society America, Madison, USA, pp. 17-37.
- Jackson, M. L. 1958. Soil chemical analysis. In Cliffs, E. N. J. (ed.). *Soil Science* University of Wisconsin, USA, Madison, pp. 89-102.

- Jamal, Z., Hamayun, M., Ahmad, N., and Chaudhary, M.F. 2006. Effect of soil and foliar application of different concentrations of NPK and foliar application of $(\text{NH}_4)_2\text{SO}_4$ on different parameters in wheat. *J. Agron.* 5 (2): 251–256.
- Jenkinson, D.S. and Poulson, D.S. 1976. The effect of biocidal treatments on metabolism in soil: A method for measuring soil biomass. *Soil Biol. Biochem.* 8: 209–213.
- Johansen, C., Musa, A.M., Kumar rao, J.V.D.K., Harris, D., Ali, M.Y., Shahidullah, A.K.M., and Lauren, J.G. 2006. Correcting molybdenum deficiency of chickpea in the high barind tract of Bangladesh. *J. Plant Nutr. Soil Sci.* 170: 752-761.
- Johnson, S.E., Lauren, J.G., Welch, R.M., and Duxbury, J.M. 2005. A comparison of the effects of micronutrient seed priming and soil fertilization on the mineral nutrition of chickpea (*Cicer arietinum*), lentil (*Lens culinaris*), rice (*Oryza sativa*) and wheat (*Triticum aestivum*) in Nepal. *Exper. Agric.* 41: 427-448.
- Ju, W., Liu, L., Fang, L., Cui, Y., Duan, C., and Wu, H. 2019. Impact of co-inoculation with plant-growth-promoting rhizobacteria and rhizobium on the biochemical responses of alfalfa-soil system in copper contaminated soil. *Ecotoxicology Environ. Saf.* 167: 218-226.
- Kannan, S. 1986. Foliar absorption and transport inorganic nutrients. *Plant Sci. J.* 341–375.
- Katiyar, P., Chaturvedi, O.P., and Katiyar, D. 2012. Effect of foliar spray of zinc, calcium and boron on spike production of gladiolus cv. Eurovision. *Hort. Flora. Res. Spectrum* 1 (4): 334–338.
- Katkar, R.N. 2005. Foliar nutrition in cotton: A review. *Crop Res. J.* 29: 186–188.
- Katyal, J.C. 2004. *Role of micronutrients in ensuring optimum use of macronutrients*. IFA International Symposium on Micronutrients, New Delhi, India. pp 3 – 17.

- Katyal, J.C. and Sharma, B.D. 1991. DTPA-extractable and total zinc, copper, manganese and iron in Indian soils and their association with soil properties. *Geoderma* 49: 165–179.
- Katyal, J.C. and Vlek, P.L.G. 1985. Micronutrient problem in tropical Asia. *Fertil. Res.* 7: 69–94.
- Kavitha, C. and Sujatha, M.P. 2015. Evaluation of soil fertility status in various agro ecosystems of Thrissur District, Kerala, India. *Int. J. Agric. Crop Sci.* 8 (3): 328-338.
- Kaya, M., Atak, M., Khawar, K.M., Ciftci, C.Y., and Ozcan, S. 2007. Effect of pre-sowing seed treatment with zinc and foliar spray of humic acids on yield of common bean (*Phaseolus vulgaris* L.). *Int. J. Agric. Biol.* 7: 875-878.
- Kaya, M., Atak, M., Mahmood, K.K., Ciftci, C.Y., and Ozcan, S. 2005. Effect of pre-sowing seed treatment with zinc and foliar spray of humic acids on yield of common bean (*Phaseolus vulgaris* L.). *Int. J. Agric. Biol.* 6 (7): 875–878.
- Kazi, S.S., Ismail, S., and Joshi, K.G. 2012. Effect of multi-micro nutrient on yield and quality attributes of the sweet orange. *Afr. J. Agric. Res.* 7 (29): 4118 – 4123.
- Khalid, B.Y. and Malik, N.S.A. 1982. Presowing soaking of wheat seeds in copper and manganese solutions. *Commun. Soil Sci. Plant Anal.* 13: 981-986.
- Khanal, N., Joshi, K.D., Harris D., and Chand, S.P. 2005. Effect of micronutrient loading, soil application and foliar sprays of organic extracts on grain legumes and vegetable crops under marginal farmers' conditions in Nepal. *Micronutrients in South and South East Asia: Proceedings of an International Workshop held in Kathmandu, Nepal, 8-11 September*, 121-132.
- Kinaci, E. and Gulmezoglu, N. 2007. Grain yield and yield components of triticale upon application of different foliar fertilizers. *Interciencia* 32 (9): 624–628.

Kizilgoz, I. and Sakin, E. 2010. The effects of increased phosphorus application on shoot dry matter, shoot P and Zn concentrations in wheat (*Triticum durum* L.) and maize (*Zea mays* L.) grown in a calcareous soil. *Afr. J. Biotechnol.* 9 (36): 5893–5896.

Krishnappa, M., Srinivasan, C.N., Basarkar P.W., and Sastry, J.A. 1992. Effects of iron, zinc and molybdenum on protein content of groundnut varieties. *J. Maharashtra Agric. Univ.* 17(2): 232-235.

Kumar, R., Kumar, S.S., and Pandey, A.C. 2008. Effect of seed soaking in nitrogen, phosphorus, potassium and boron on growth yield of garden pea (*Pisum sativum* L.). *Ecol. Environ. Conserv.* 14: 383-386.

Kumar, J.V.D.K., Harris, D., Johansen, C., Musa, A.M. 2004. Low cost provision of molybdenum (Mo) to chickpeas grown in acid soils. In: *Proceedings of the Zvarevashe, CARE Stakeholder Workshop, Report IDG/00/18, Silsoe Research Institute, Bedford, UK.*

Lake, D.L., Kisk, P.W.W., and Lester, J.N. 1984. Fractionation, characterization and speciation of heavy metals in sewage sludge and sludge-amended soils: A reviews. *J. Environ. Qual.* 13: 175–183.

Ledgard, S. F. and Steele, K. W. 1992. Biological nitrogen fixation in mixed legume/grass pastures. *Plant Soil* 141(1-2): 137-153.

Lehri, L. K., Gangwar, B.R., and Mehrotra, C.L. 1974. Bacterisation experiments with rhizobium. *J. Indian Soc. Soil Sci.* 22(1): 66-69.

Liew, C.S. 1988. Foliar fertilizers from Uniroyal and their potential in Pakistan. la vegetation. *Comptes Rendus de l'Acad. Sci.* 17: 679–686.

Lindsay, W.L. 1991. Inorganic equilibria affecting micronutrients in soil. In: Mortvedt, J.J., Cox, F.R., Shuman, L.M., and Welch, R.M. (eds), *Micronutrients in Agriculture* (2nd Ed). SSSA, Madison, WI, pp. 89–112.

- Lipman, C. B., Davis, A. R., and West, E. S. 1926. The tolerance of plants for NaCl. *Soil Sci.* 22: 303-322.
- Lipman, C.B. and MacKinney, G. 1931. Proof of the essential nature of copper for higher green plants. *Plant Physiol.* 6: 593-599.
- Madani, M.C. and Singh, K.P. 1977. Effect of three different isolates of rhizobium on the growth of moong (*P.aureua*). *Allahabad Far.* 48(7): 483-485.
- Malakouti, M. J. and Tehrani, M.H. 1999. *Effect of micronutrients in yield increase and improvement of crops quality*. Tarbiat Modarres University Press.
- Malhi, S.S. 2009. Effectiveness of seed-soaked Cu, autumn- versus spring-applied Cu, and Cu-treated P fertilizer on seed yield of wheat and residual nitrate-N for a Cu-deficient soil. *Can. J. Plant Sci.* 89: 1017-1030.
- Mallick, M.F.R. and Muthukrishnan, C.R. 1980. Effect of micronutrients on the quality of tomato (*Lycopersicon esculentum* Mill.). *Veg. Sci.* 7(1): 6-13.
- Manasa, V., Hebsur, N.S., Malligawad, L.H., Kumar, L.S., and Ramakrishna, B. 2015. Effect of water soluble fertilizers on uptake of major and micronutrients by groundnut and post-harvest nutrient status in a vertisol of Northern transition zone of Karnataka. *Ecoscan* 9(1-2): 1-5.
- Marschner, H. 1995. *Mineral Nutrition of Higher Plants*. Academic Press, San Diego, 889p.
- Masuthi, D.A., Vyakaranahal, B. S., and Deshpande, V. K. 2009. Influence of pelleting with micronutrients and botanical on growth, seed yield and quality of vegetable cowpea. *Karnataka J. Agric. Sci.* 22(4): 898-900.
- Mathur, K. 2000. Effect of Rhizobium inoculation and various sources of nitrogen on growth and yield of summer greengram *Vigna radiata* (L.) Wilczek. M. Sc. (Ag) thesis, CCS HAU, Hisar, India, 132p.
- Mchargue, J.S. 1922. Role of manganese in plants. *J. Amer. Chem. Soc.* 44: 1592-1598.

Mengel, K., Kirkby, E.A., Kosegarten, H., and Appel, T. 2001. Principles of Plant Nutrition. Kluwer Academic Publishers, Dordrecht, The Netherlands, p.849.

Mini, V. 2015. Assessment and management of micronutrient deficiencies in onattukara. Ph.D thesis, Kerala Agricultural University, Thrissur, p.267.

Mishra, B.K., Sahoo, C.R., and Bhol, R. 2012. Effect of foliar application of micronutrients on growth, yield and quality of tomato cv. Utkal Urbasi. *Environ. Ecol.* 30 (3): 856–859.

Mishra, H.P., Singh, K.P., and Yadav, J.P. 1990. Influence of Zn, Fe, B and Mn and their uptake on onion grown in calcareous soil. *Haryana J. Hortic. Sci.* 19(1-2): 153-159.

Mitsui, S. and Ueda, M. 1963. Cation Exchange Capacity of Crop Roots in Relation with Ion Uptake: Part 1. Method of determining the cation exchange capacity and intensity of plant roots. *Soil Sci. Plant Nutr.* 9(1): 6-12.

Mona, E., Eleiwa, Ibrahim, S.A., and Mohamed, F. 2012. Combined effect of NPK levels and foliar nutritional compounds on growth and yield parameters of potato plants (*Solanum tuberosum* L.) *Afr. J. Micro. Res.* 6 (24): 5100–5109.

Moosavi, A.A. and Ronaghi, A. 2011. Influence of foliar and soil applications of iron and manganese on soybean dry matter yield and iron-manganese relationship in a Calcareous soil. *Asian J. Crop Sci.* 5 (12): 1550–1556.

Mori, S. 1999. Iron Acquisition by Plants. *Curr Opin Plant Biol.* 2: 250–253.

Mortvedt, J.J. 2000. Bioavailability of micronutrients. In: Sumner, M.E. (ed.), *Handbook of Soil Science.* pp. 71-88. CRC Press, Boca Raton, Florida.

Nasiri, Y., Zehtabe-Salmasi, S., Nasrollahzade, S., Najafi, N., and Ghassemi-Golezani, K. 2010. Effect of foliar application of micronutrient (Fe and Zn) on flower yield and essential oil of chamomile (*Matricaria chamomila* L.). *J. Med. Plants Res.* 4 (17): 1733–1737.

Niranjana, H.G., Prakash, S.S., Basavegowda, Yelladhalli N.A., and Chandranath, H.T. 2005. Effect of micronutrient seed treatment on growth and yield of groundnut. *Seed Res.* 33(2): 138-141.

Noor, S., Hannan M.A., and Islam, M.S. 1997. Effect of molybdenum and boron on the growth and yield of groundnut. *Indian J.Agric. Res.* 31(1): 51-58.

Nyoki, D. and Ndakidemi, P.A. 2014. Influence of *Bradyrhizobium japonicum* and phosphorus on micronutrient uptake in cowpea. a case study of zinc (Zn), iron (Fe), copper (Cu) and manganese (Mn). *Am. J. Plant Sci.* 5: 427-435.

Nyoki, D. and Ndakidemi, P.A. 2018. Rhizobium inoculation reduces P and K fertilization requirement in corn-soybean intercropping. *Rhizosphere* 5: 51-56.

Panse, V.G. and Sukhatme, P.V. 1985. *Statistical methods for agricultural workers* (4th Ed.). Indian Council of Agricultural Research, New Delhi, p.347.

Pant, L.M., Dwivedi, G.K., and Katiyar, A.K. 2000. Survival and response to inoculation of rhizobium leguminosarum Br. Phaseoli on french bean grown under temperate conditions. *J. Indian Soc. Soil Sci.* 47(3): 558-560.

Pasquet, R. S. 1998. Morphological study of cultivated cowpea *Vigna unguiculata* (L.) Walp. Importance of ovule number and definition of cv gr Melanophthalmus. *Agronomie* 18: 61-70.

Patel, K.C., Patel, K.P., Ramani, V.P., and George, V. 2009. Effect of multi-micronutrient mixture on yield of okra (*Abelmoschus esculentus* L.) grown on *goradu* soils of kheda district. *Veg. Sci.* 36 (3): 393–394.

Patel, K.P., Patel, A.K., Patel, A.M., Patel, K.C., and Patel, V.P. 2008. Effect of multi-micronutrient mixture on yield and uptake of micronutrients by Wheat (*Triticum aestivum* L.) growth on sandy loam soils of North Gujrat. *Asian J. Soil Sci.* 3 (1): 84–87.

- Patel, M.M., Patel, I.C., Patel R.I., and Acharya S. 2011. Effect of zinc and iron on yield and yield attributes of rainfed cowpea (*Vigna unguiculata* L. Walp). *Ann. Arid Zone* 50(1): 17-19.
- Patel, T.S. 1994. Effect of biofertilizers and chemical fertilizers on vegetative growth and yield of garden pea (*Pisum sativum* L.) M.Sc. (Ag) Thesis. JNKVV, Jabalpur (MP). 136 p.
- Patil, J. 1981. Cultural practices and infectious crop diseases. *Adv. Ser. Agric. sci.* 9: 73–189.
- Pattanayak, S.K., Dash, V., Jena, M.K., and Nayak, R.K. 2000. Seed treatment of green gram with molybdenum and cobalt: Effect on nodulation, biomass production and N uptake in an acid soil. *J. Indian Soc. Soil Sci.* 48, 769-773.
- Peoples, M.B., Herridge, D.E., and Ladha, J.K. 1995. Biological nitrogen fixation: An efficient source of nitrogen for sustainable agricultural production. *Plant Soil* 174: 3-28.
- Piper. 1966. Aging of crystalline precipitates. *Analyst* 77: 1000-1001.
- Prakash, J.S. 1998. Growth and mineral nutrient composition of six cultivars of greengram grown at normal and deficient levels of iron supply. *Madras Agric. J.* 85: 96-99.
- Pranakrishna, S. 1976. Effect of nitrogen micronutrients and rhizobium culture on cowpea and blackgram. M.Sc.(Ag) thesis, Orissa University of Agriculture and Technology, 56p.
- Prasad, R., Kumar, D., Shivay, Y.S., and Rana, D.S. 2014. Boron in Indian agriculture – A review. *Indian J. Agron.* 59 (4): 511–517.
- Pratt, P. F. 1965. *Potassium in methods of soil analysis.* (2nd Ed.). American Society of Agronomy, Madison, USA, pp. 1019-1021.
- Premalatha, A. 2016. Formulation and evaluation of micronutrient mixture for foliar application in TC banana (*Musa* sp var. Nendran) M.Sc.(Ag) thesis,

Kerala Agricultural University, Thrissur, 195p.

- Rasmussen, P.E. and Boawn, L.C. 1969. Zinc seed treatment as a source of zinc for beans. *Agron. J.* 61: 674-676.
- Rattan, R.K. and Sharma, P.D. 2004. Main micronutrients available and their method of use. *Proceedings IFA International Symposium on Micronutrients.* 1-10.
- Ravi, S., Channal, T., Hebsur, N.S., and Dharamtti. 2008. Effect of sulphur, zinc and iron nutrition on growth, yield, nutrient uptake and quality of safflower (*Carthamus tinctorius* L.). *Karnataka J. Agric. Sci.* 21: 3.
- Reddy, C.H. 1981. Response of cowpea to phosphatic fertilizer, rhizobium culture and growth regulators. M.Sc. (Ag) thesis, Anand Agricultural University, Gujarat. 214 p.
- Reddy, Y.T. and Reddi, S.G.H. 2002. *Principles of Agronomy.* Kalyani Publishers, Ludhiana. pp. 204-206.
- Rehman, A., Farooq, M., Cheema, Z.A., and Wahid, A. 2012. Role of boron in leaf elongation and tillering dynamics in fine grain aromatic rice. *J. Plant Nutr.* 32: 12-13.
- Rehman, H., Aziz, T., Farooq, M., Wakeel, A., Rengel, Z. 2012. Zinc nutrition in rice production systems. *Plant Soil* 307: 56-57.
- Reuveni, M., Oppenheim, D., and Reuveni, R. 1998. Integrated control of powdery mildew on apple trees by foliar sprays of mono-potassium phosphate fertilizer and sterol inhibiting fungicides. *Crop Prot.* 17: 563-568.
- Rhoden, E.G. and Allen, J.R. 1982. Effect of B, Mn and Zn on nodulation and N₂-fixation in southern peas. *J. Commun. Soil Sci. Plant Anal.* 13(4): 43-46.
- Robinson, G. W. 1922. A new method for the mechanical analysis of soils and other dispersions. *J. Agric. Sci.* 12: 306-321.

- Roemheld, V. and El-Fouly, M.M. 1999. Foliar nutrient application Challenge and limits in crop production. *Proceedings of the 2nd International Workshop on Foliar Fertilization*, Bangkok, Thailand, 4-10 April 1999.
- Romheld, V. and Marschner, H. 1986. Mobilization of iron in the rhizosphere of different plant species. *Adv. Plant Nutr.* 2: 155–204.
- Salih, H.O. 2013. Effect of foliar fertilization of Fe, B and Zn on nutrient concentration and seed protein of cowpea. *ISOR J. Agric. Vet. Sci.* 6(3): 42-46.
- Sanginga, N., Dashiell, K.E., Diels, J., Vanlauwe, B., Lyasse, O., Carsky, R.J., Tarawali, S., Asafo-Adjei, B., Menkir, A., Schulz, S., Singh, B.B., Chikoye, D., Keatinge, D., and Ortiz, R. 2003. Sustainable resource management coupled to resilient germplasm to provide new intensive cereal-grain-legume-livestock systems in the dry savanna. *Agric. Ecosyst. Environ.* 100: 305–314.
- Saric, T. and Saciragic, B. 1969. Effect of oat seed treatment with microelements. *Plant Soil* 31: 185-187.
- Sarkar, D., Mandal, B., and Kundu, M.C. 2007. Increasing use efficiency of boron fertilisers by rescheduling the time and methods of application for crops in India. *Plant Soil* 301: 77–85.
- Sathya, S., Pitchai, G.J., and Indirani, R. 2009. Boron nutrition of crops in relation to yield and quality—a review. *Agric. Rev.* 30 (2): 139–44.
- Sekhon, N. K. and Singh, C. B. 2013. Plant nutrient status during boll development and seed cotton yield as affected by foliar application of different sources of potassium. *Am. J. Plant Sci.* 4: 1409-1417.
- Selvi, D. and Thiageshwari, S. 2002. Effect of integrated nutrient management on yield of brinjal and bhendi in a mixed black soil (Vertic ustropept). *Madras Agric. J.* 89 (7–9): 378–382.

Shitole, S.M. and Dhumal, K.N. 2012. Influence of foliar applications of micronutrients on photosynthetic pigments and organic constituents of medicinal plant *Cassia angustifolia*. *An. Bio. Res.* 3 (1): 520–526.

Shueadshen, A.K.H. 1991. Foliar application of trace elements to rice. *Khimizatsiya-Sel's Kogo- Khozaistva* 3: 46–50.

Shukla, A.K., Behera, S.K., Shivay, Y.S., Singh, P., and Singh, A.K. 2012. Micronutrients and field crop production in India: A review. *Indian J. Agron.* 57 (3): 123–130.

Shukla, A.K., Dwivedi, B.S., Singh, V.K., and Gill, M.S. 2009. Macro Role of Micronutrients. *Indian J. Fertil.* 5 (5): 11–12, 15–18, 21–24 & 27–30

Silberbush, M. 2002. Simulation of ion uptake from the soil. In: Waisel, Y., Eshel, A., and Kafkafi, U. (eds), *Plant Roots: The Hidden Half* (3rd Ed.). Marcel, Dekker: New York, 651–661.

Simmonds, N.W. 1966. Planting and management. In : *Bananas* (2nd Ed.). Longman Group Limited, London and New York. pp. 156–204.

Simoglou, K. and Dordas, C. 2006. Effect of foliar applied boron, manganese and zinc on tan spot in winter durum wheat. *Crop Prot.* 25: 657–663.

Sims, J. R. and Johnson, G. V. 1991. Micronutrient soil tests. In: Mortvedt, J. J. (ed.), *Micronutrients in Agriculture* (2nd ed.). Soil Science Society of America, Madison, USA, pp. 427-476.

Singaram, P. and Prabu, P.C. 2001. Effect of zinc and boron on growth and quality of grapes cv. Muscat. *Madras Agric. J.* 88 (4–6): 233–236.

Singh, B., Natesan, S.K.A., Singh, B.K., and Usha, K. 2003. Improving zinc efficiency of cereals under zinc deficiency. *Curr. Sci.* 88: 36-14.

Singh, D.K., Singh A.K., Singh, M., Bordoloi, L.J., and Srivastava, O.P. 2012. Production potential and nutrient uptake efficiency of pea (*Pisum sativum* L)

as influenced by different fertility levels and micronutrients. *J. Indian Soc. Soil Sci.* 60(2): 10-11.

Singh, M.V. 1999. Experiences of AICRP on micro and secondary nutrients and pollutant elements in soils and plants. *Fertil. News* 44 (4): 63–82.

Singh, M.V. 2003. Issues and challenges in promoting safe use of micronutrients in agriculture during new millennium: Interface on micronutrients, MCEAR, Pune. pp.241.

Singh, M.V. 2006. *Micro and secondary nutrients and pollutant elements research in India*. Progress report, IISS, Bhopal, pp. 1–110.

Singh, M.V. 2007. Efficiency of seed treatment for ameliorating zinc deficiency in crops. *Proceeding of Zinc Crop Conference, Istanbul, Turkey*.

Singh, M.V. 2009. Micronutrient nutritional problems in soils of India and improvement for human and animals. *Indian J. Fertil.* 5 (4): 11–16, 19–26 & 56.

Singh, M.V. 2012. *Spread of micro-nutrient deficiencies specially boron in India and response of field crops*. “Brain storming workshop on soil test based nutrients including boron and other micro nutrients” Organized by ICRISAT – Agriculture Directorate, Karnataka -Rio Tinto India at Bangaluru, India.

Sivaiah, N.K., Swain, S.K., Raju, B., and Varma, S.V. 2013. Influence of micronutrients application on growth and seed quality in tomato (*Lycopersicon esculentum* MILL.) *Int. J. Dev. Res.* 3 (11): 191–195.

Slaton, N.A., Wilson-Jr, C.E., Ntamatungiro, S., Norman, R.J., and Boothe, D.L. 2001. Evaluation of zinc seed treatments for rice. *Agron. J.* 93: 152-157.

Soltanpur, P.N. and Schwab, A.P. 1977. A new soil test for simultaneous extraction of macro and micro-nutrients in alkaline soils. *Comm. Soil Sci. Plant Anal.* 8: 195-207.

Sommer, A.L. and Lipman, C.B. 1926. Evidence on the indispensable nature of zinc and boron for higher green plants. *Plant Physiol.* 12 (3):231–249.

Srivastava, A.K. and Singh. 2003. *Citrus nutrition*. International books distributing Co., Lucknow. 11–17.

SSO [Soil Survey Organization]. 2007. *Benchmark Soils of Kerala*. Agriculture (SC Unit) Department, Govt. of Kerala, Thiruvananthapuram, 623p.

Subbiah, B. V. and Asija, G. L. A. 1956. A rapid procedure for the estimation of available nitrogen in soil. *Curr. Sci.* 32: 325-327.

Suresh, P.R., Govindan, M., Gladis, R., and Joseph, B. 2014. *Soil Fertility status of Kasargod district*. College of Agriculture, Padannakad. 32p.

Suryanarayana, V. and Rao, S.K.V. 1981. Effect of growth regulators and nutrient sprays on the yield of okra. *Veg. Sci.* 8: 12–14.

Suryanarayana, V. and Reddy, P.E. 1978. Effect of growth regulators and micronutrients on French bean. *Veg. Sci.* 6(1): 130-134.

Swati, B., Singh, P., Mahasagar, H., and Singh, D.B. 2011. Response of foliar application of micronutrients on tomato variety Rashmi. *Indian J. Hortic.* 68(2): 278-279.

Takkar, P.N., Chibba, I.M., and Mehta, S. 1989. Twenty years of coordinated research on micronutrients in soils and plants, IISS, Bhopal.

Tarawali, S.A., Singh, B.B., Gupta, S.C., Tabo, R., Harris, F., Nokoe, S., Fernández-Rivera, S., Bationo, A., Manyong, V.M., Makinde, K., and Odion, E.C. 2002. Cowpea as a key factor for a new approach to integrated crop-livestock systems research in the dry savannas of West Africa. In: Fatokun, C.A., Tarawali, S.A., Singh, B.B., Kormawa, P.M., and Tamo, M. (eds) *Challenges and opportunities for enhancing sustainable cowpea production*. International Institute of Tropical Agriculture, Ibadan, Nigeria, pp 233–251.

- Terpolilli, J.J., Hood, G.A., and Poole, P.S. 2012. What Determines the Efficiency of N₂-Fixing *Rhizobium*-Legume Symbioses? *Adv. Microbial Physiol.* 60: 325-386.
- Thakur, R. N., Arya, P. S., and Thakur, S. K. 1999. Response of french bean (*Phaseolus vulgaris* L.) varieties to fertilizer levels, rhizobium inoculation and their residual effect on onion (*Allium cepa*) in mid hills of northwestern Himalayas. *Indian J. Agron.* 44: 416-418.
- Thampatti, K.C.M., Cherian, S., and Iyer, M.S. 2005. Managing iron toxicity in acid sulphate soils by integrating genetic tolerance and nutrition. *Int. Rice Res. Notes* 30: 37-39.
- Thilakarathna, M.S., Chapagain, T., Ghimire, B., Pudasaini, R., Tamang, B.B., Gurung, K., Choi, K., Rai, L., Magar, S., Bishnu, B.K., Gaire, S., and Raizada, M.N. 2019. Evaluating the effectiveness of rhizobium inoculants and micronutrients as technologies for Nepalese common bean smallholder farmers in the real-world context of highly variable hillside environments and indigenous farming practices. *Agric.* 9(20): 23-26.
- Thiyageshwari, S. and Ramanathan, G. 2001. Uptake of nutrients as influenced by application of micronutrients and cytozyme to soybean in inceptisol. *J. Soil Crops* 11(1): 1-6.
- Tilak, K.V.B.R., Singh, C.S., and Rana, J.P.S. 1981. Effects of combined inoculation of *Azospirillum brasilense* with *Rhizobium trifolii*, *Rhizobium meliloti*, and *Rhizobium* sp. (*Cowpea miscellany*) on nodulation and yield of clover (*Trifolium repens*), lucerne (*Medicago sativa*), and chickpea (*Cicer arietinum*). *Zentralblatt* 136 (2): 117- 120.
- Tisdale, S.L., Nelson, W.L., and Beaton, J.D. 1985. Soil Fertility and Fertilizers (4th Ed.). Mc Millan, New York. P.754.
- Torun, A., Itekin, I.G.A., Kalayci, M., Yilmaz, A., Eker, S., and Cakmak, I. 2001. Effects of zinc fertilization on grain yield and shoot concentrations of zinc,

boron and phosphorus of 25 wheat cultivars grown on a zinc-deficient and boron-toxic soil. *J. Plant Nutr.* 2: 1817–1829.

Udode-Haes, H.A., Voortman, R.L., Bastein, T., Bussink, D.W., Rougoor, C.W., and Weijden, V.W.J. 2012. Scarcity of micronutrients in soil, feed, food, and mineral reserves - urgency and policy options. *Platform Agric. Innovation Soc.* 2: 546–554.

Vargas, M.A.T. and Hungria, M. 1997. Fixaco Biologia, do N₂ na culture da soja. In: Vargas, M.A.T. and Hungria, M. (eds.), *Biologia dos Solaos de Cerrados, Planatina*. pp 279-360.

Vargas, R. and C. Ramirez, 1989. Response of soyabeans and groundnuts to rhizobium and to N, P and Mo fertilizer on a typic pellustert from Canas, Guanacaste. *Agronomia Costarricense* 13(2): 175-18.

Verma, A.N., Ram, K., and Sharma, R.K. 1973. Growth, yield and quality of tomato as affected by foliar application of boron in sand culture. *Mysore J. Agric. Sci.* 7(1): 130-132.

Vieira, R. F., Cardoso, E. J. B. N., Vieira, C., and Cassini, S. T. A. 1998. Foliar application of molybdenum in common beans. I. Nitrogenase and reductase activities in a soil of high fertility. *J. Plant Nutr.* 21(1): 169-180.

Walkley, A. J. and Black, I. A. 1934. Estimation of soil organic carbon by chromic acid and titration method. *Soil Sci.* 31: 21-38.

Wallihan, E.F., Embleton, T.W., and Printy, W. 1958. Zinc deficiency in avocado. *Calif. Agric.* 12 (6): 4–5.

Wankhade, S.Z., Dabre, W.M., Lanjewar, B.K., Sontakey P.Y., and Takzure, S.C. 1992. Effect of seed inoculation with rhizobium culture and molybdenum on yield of groundnut (*Arachis hypogaea*). *Indian J. Agron.* 37(2): 384-385.

Warrington, K. 1923. The effect of boric acid and borax on the board bean and certain other plants. *Ann. Bot.* 37 (4): 629–672.

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- Wiley-VCH and Wiley-VCH, L. 2007. *Ullmann's Agrochemicals: liquid fertilizers*. pp. 36.
- Wolde-meskel, E., Heerwaarden, J., Abdulkadir, B., Kassa, S., Aliyi, I., Degefu, T., Wakweya, K., Kanampiu, F., and Giller, K.E. 2018. Additive yield response of chickpea (*Cicer arietinum* L.) to rhizobium inoculation and phosphorus fertilizer across smallholder farms in Ethiopia. *Agric. Ecosyst. Environ.* 261: 144-152.
- Yadav, R.D. 2000. Effect of INM on nodulation, yield, quality and available nutrient status in soil after harvest of green gram. *Legume Res.* 3(4): 122-124.
- Yadav, R. D. and Malik, C.V. S. 2005. Effect of rhizobium inoculation and various sources of nitrogen on growth and yield of cowpea (*Vigna unguiculata* L. Walp). *Legume Res.* 28: 38-41.
- Yousefi, M. and Zandi, P. 2012. Effect of foliar application of zinc and manganese on yield of pumpkin (*Cucurbita pepo* L.) under two irrigation patterns. *Electr. J. Polish Agric. Univ.* 15 (4): 2-6.
- Zahran, H.H. 1999. Rhizobium-legume symbiosis and nitrogen fixation under severe conditions and in an arid climate. *Microbiol. Mol. Biol. Rev.* 63(4): 968-989.

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EVALUATION OF MICRONUTRIENT FORMULATION IN COWPEA
(Vigna unguiculata L. Walp)

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ABSTRACT

The project "Evaluation of micronutrient formulation in cowpea (*Vigna unguiculata* L. Walp)" was carried out at College of Agriculture, Padannakkad and Regional Agricultural Research Station (RARS), Pilicode during 2017 to 2019. The objectives of the study were to evaluate the effect of micronutrient formulation through seed treatment and foliar nutrition on growth, nodulation and yield of cowpea and to study its effect on plant nutrient uptake and residual soil nutrient status.

Laboratory study was carried out at COA, Padannakkad to formulate and standardize the micronutrient solution for cowpea. Different concentrations of the standardized micronutrient formulation were used for seed treatment study (Experiment 1). The experiment was carried out in completely randomized design with 7 treatments and 3 replications which included seed treatment with micronutrient formulation @ 0.25, 0.50, 0.75, 1, 1.5 and 2 per cent as T₁, T₂, T₃, T₄, T₅ and T₆ respectively. T₇ (seed treatment with water) was the control. Observations on germination percentage, number of days taken for germination, seedling length at three leaf stage and seedling vigour index were recorded. Treatment T₆ (micronutrient formulation @ 2 %) showed highest seedling length (24.23 cm) and seedling vigour index (2423) which was followed by T₅ (micronutrient formulation @ 1.5 %). Thus the best and second best treatment from seed treatment study was concluded as T₆ and T₅ respectively and was selected for the field experiment (Experiment 2).

The field experiment was conducted in randomized block design with 12 treatments replicated three times. The treatment consisting of combination of four levels of seed treatment (no seed treatment, seed treatment with rhizobium, seed treatment with best concentration from experiment 1 and second best seed treatment from experiment 1) and three levels of foliar application of micronutrient (no foliar application, one foliar application at 15 DAS and two foliar applications at 15 and 30 DAS). Biometric observations were recorded at flower bud initiation and at harvest.

Plant height at flower bud initiation and at harvest was superior in S₃ (Seed treatment with micronutrient formulation at 2 %) while stem diameter was highest in S₄ (seed treatment at 1.5 %) and S₃ respectively in both stages. Double foliar spray was superior in plant height and stem diameter at both stages. Interaction S₃F₃ (2 per cent seed treatment plus two foliar spray) recorded highest plant height at both stages and stem diameter was highest in S₂F₃ (treatment with rhizobium plus two foliar spray). Number of nodules was highest in S₂ (seed

treatment with rhizobium). Number of pods per plant (21.02 & 20.97), pod weight per plant (36.14 g & 33.49 g), grain yield (2224 kg ha⁻¹) and dry matter production (2292.91 & 2372.92 kg ha⁻¹) was superior in S₃ and F₃ (two foliar spray). Interaction S₃F₃ was superior in number of pods per plant (25), pod weight per plant (38.11 g), grain yield (2320 kg ha⁻¹) and dry matter production (2613.24 kg ha⁻¹).

Leaf analysis revealed that P and K content in leaf was highest in S₃. N, P, K and Ca content in leaf was highest in F₃. Interaction S₃F₃ recorded highest N, P and K content in cowpea leaf. Mg content was highest in S₂F₃ (treatment with rhizobium plus two sprays). Fe, Mn, Zn and B content in leaf was highest in S₃ and F₃. Interaction S₃F₃ (2 per cent seed treatment plus two foliar spray) recorded highest Fe, Mn and Zn content in cowpea leaves. The uptake of N, P, K, Ca, S, Fe, Mn, Zn and B were highest in both S₃ and F₃ whereas Mg uptake was highest in S₃ and F₂ (one foliar spray). Cu uptake was highest in F₃. Interaction S₃F₃ recorded highest uptake of N, P, Mn and Zn. N content in cowpea grain was superior in S₂ and F₃. While K, Ca, Fe, Mn, Zn, Cu and B content in cowpea grain was highest in both S₃, F₃ and interaction S₃F₃. In case of interaction, N was highest in S₂F₃. The uptake of N, P, K, Ca, Mg, S, Fe, Zn and B in cowpea grain was highest in both treatments S₃ and F₃ and the interaction, S₃F₃ recorded highest uptake of P, K, Ca, Fe, Mn, Zn and B in grain.

Studies on residual soil nutrient status revealed that available N and P in soil was highest in S₂ at both 45 DAS and at harvest. Available K was highest in S₃, F₃ and S₃F₃ at both stages. Organic carbon content was found to increase in plots receiving two foliar spray and interaction S₃F₃. Available Fe, Mn and Zn was highest in S₃ at harvest and in F₃ at both stages. Available B was highest in S₃ and F₃ at 45 DAS while Mo at harvest. B and Zn were highest in S₃F₃ at 45 DAS while at harvest available Fe was found to be higher in S₃F₃. Available Fe at 45 DAS was highest in S₁F₃ (no seed treatment plus two foliar spray). Root CEC was superior in S₃, F₃ and in interaction S₃F₃.

The results obtained from the investigation revealed that the application of micronutrient formulation as two foliar spray (15 and 30 DAS) along with seed treatment (2 per cent) was found to be highly effective in increasing the growth and yield characters of cowpea. The uptake of nutrients in cowpea leaf and grain was also found to increase with the application of micronutrient formulation enhancing the nutritional quality of grain.

APPENDIX I

Daily average weather parameters of RARS, Pilicode

Date	Temperature		Relative humidity		Rainfall (mm)
	Max	Min	I	II	
19-10-2018	30	25	92	70	1.8
20-10-2018	30	24	96	81	21.8
21-10-2018	28.9	23.9	88	69	0
22-10-2018	30.5	24	92	70	0
23-10-2018	30.5	24.5	91	63	0
24-10-2018	31.5	23.5	91	57	0
25-10-2018	32	23.5	88	59	0
26-10-2018	32	24	80	49	0
27-10-2018	33	24	91	56	0
28-10-2018	32.5	22.5	83	47	0
29-10-2018	33	22	91	54	0
30-10-2018	32.5	21	85	48	0
31-10-2018	32	21.5	88	53	0
01-11-2018	32	24	91	64	0
02-11-2018	31.5	25	92	58	0
03-11-2018	32.5	25	93	71	0
04-11-2018	31	25	92	63	8.2
05-11-2018	32	25	95	69	0
06-11-2018	31.5	25	90	58	0
07-11-2018	32.5	25	88	63	0
08-11-2018	32	23	87	54	0
09-11-2018	31.5	23	84	52	0
10-11-2018	32.5	24	87	61	0
11-11-2018	32	23	91	66	0
12-11-2018	31.5	23	87	60	0
13-11-2018	31.5	23.5	86	60	0
14-11-2018	32	21.5	91	56	0
15-11-2018	31.9	19.5	91	47	0
16-11-2018	32.5	23.5	83	63	0
17-11-2018	31.5	23.5	91	58	0
18-11-2018	32	25	88	63	0
19-11-2018	31.5	25.5	96	73	0
20-11-2018	30.9	24.5	92	68	0
21-11-2018	30.5	25.5	92	65	0
22-11-2018	32.5	25.5	88	60	0
23-11-2018	32.5	25	93	63	0

24-11-2018	32	24	94	69	6.8
25-11-2018	31.5	24	92	69	40
26-11-2018	31.5	24	96	66	0
27-11-2018	31.5	24	85	63	0
28-11-2018	32	21.5	87	46	0
29-11-2018	32	20.5	93	55	0
30-11-2018	32.5	25	88	66	0
01-12-2018	32	24	92	67	0
02-12-2018	31.5	24	91	58	0
03-12-2018	32.5	21.5	92	64	0
04-12-2018	31	22	87	63	0
05-12-2018	31	23.5	90	63	0
06-12-2018	31.5	20.5	93	70	0
07-12-2018	31.6	21.8	93	66	0
08-12-2018	32.2	23	91	63	0
09-12-2018	32	23	91	60	0
10-12-2018	32	23.5	91	73	4.2
11-12-2018	30	22	92	66	0
12-12-2018	31.4	23.5	91	66	0
13-12-2018	30.3	21	91	60	0
14-12-2018	31.5	21.2	91	66	0
15-12-2018	31.5	20.5	91	66	0
16-12-2018	32	21	95	66	0
17-12-2018	31	19	90	53	0
18-12-2018	30.5	17.7	93	70	0
19-12-2018	32.5	18.5	89	70	0
20-12-2018	31.5	23	93	61	0
21-12-2018	31	22	93	63	0
22-12-2018	31.2	21.8	88	59	0
23-12-2018	32	23.5	91	60	0
24-12-2018	32	24	96	66	0
25-12-2018	31	22.5	92	63	13.8
26-12-2018	32	23	91	66	0
27-12-2018	31.5	23	92	66	0
28-12-2018	31	22.5	91	66	0
29-12-2018	31	23	93	69	0
30-12-2018	30.5	23.2	92	66	0
31-12-2018	30.5	21	94	60	0

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APPENDIX II

Interaction effect of seed treatment and foliar spray of micronutrient formulation on
B : C Ratio

Treatments	Cost of cultivation	Gross return	B:C Ratio
T ₁	83947	127600	1.52
T ₂	100000	160000	1.60
T ₃	107988	180340	1.67
T ₄	86671	131740	1.52
T ₅	100000	160000	1.60
T ₆	104036	170620	1.64
T ₇	105712	183940	1.74
T ₈	110333	198600	1.80
T ₉	110441	199900	1.81
T ₁₀	104971	181600	1.73
T ₁₁	106198	181600	1.71
T ₁₂	113953	196000	1.72

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