GROWTH, FLOWERING, FLORAL BIOLOGY AND SPIKE SHEDDING IN PEPPER (Diper nigrum L.)

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THESIS

Submitted in partial fulfilment of the requirements for the degree of

Master of Science in Horticulture

Faculty of Agriculture Kerala Agricultural University

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DECLARATION

I, hereby declare that this thesis entitled "Growth, flowering, floral biology and spike shedding in pepper (<u>Piper nigrum L.</u>)" is a bonafide record of research work done by me during the course of research work and the thesis has not previously formed the basis for the award to me of any degree, diploma, associateship, fellowship or other similar title of any other University or Society.

Vellenikkara, 7⁷⁶April, 1981.

Pina

Rema Menon

CERTIFICATE

Certified that this thesis entitled "Growth, flowering, floral biology and spike shedding in pepper (<u>Piper nigrum L.)</u>" is a record of research work done independently by Smt. Rema Menon under my guidance and supervision and that it has not previously formed the basis for the award of any degree, fellowship or associateship to her.

Vellanikkara, 7% April, 1981.

Dr. P.C. Sivaraman Nair, Director of Research.

CERTIFICATE

We, the undersigned members of the Advisory Committee of Smt. Rema Menon, a candidate for the degree of Master of Science in Horticulture agree that the thesis entitled "Growth, flowering, floral biology and spike shedding in pepper (<u>Piper nigrum</u> L.)" may be submitted by Smt. Rema Menon in partial fulfilment of the requirement for the degree.

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Lenna Henow REMA MENON

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INTRODUCTION

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INTRODUCTION

Pepper (Piper nigrum L.) is the most popular among the spices. India's share in world production of pepper used to be substantial. But when other producing countries took up pepper cultivation extensively, with scientific management, their production increased considerably and India's share in export came down drastically. The world trade in black pepper was estimated to be about 125.34 thousand tonnes during 1977. During that year. India had 55.5 per cent of the total world area under pepper, which accounted for 20.5 per cent of the world production. On the other hand Brazil and Malaysia with 5.9 per cent and 5.2 per cent of the area shared 28.6 and 22.4 per cent production respectively. Indonesia with 27.5 per cent of the area accounted for 24 per cent of the total production.

During 1978-79, India produced 26.10 thousand tonnes in an area of 111.97 thousand hectares. Pepper is essentially an export oriented crop in India, accounting fr about 50 per cent of the total export earnings from all spices. During the year 1978-79*.

*Source: Directorate of Cococ., Arecanut & Spices Development, Calicut. the export earnings was to the tune of 5.280 million.

Among the pepper producing states in India, Kerala ranks first both in area and production. During 1978-79, the area under pepper in Kerala was 108.26 thousand hectares and production was 25.12 thousand tonnes.

Considering the importance of this spice in the economy of India and Kerala, very little work has been carried out during the past years. The production in a perennial crop depends upon the growth and flowering behaviour, which in turn is influenced by the interaction of genetic and environmental factors. Pepper is not an exception to this. Hence, in order to exploit the full potential of crop production in pepper, the detailed understanding of the growth and flowering pattern is quite important. A knowledge on the mode of pollination and pollen viability are also important for successful crop production. Though certain work on these aspects have been carried out both in India and abroad, they are neither specific nor exhaustive (Hasan, Iljas, 1960; Martin and Gregory, 1962; De Waard, 1967; Nambiar et al., 1978). Another important factor that hinders the production of pepper is spike shedding, which is

found to vary from 14.5 to 65.2 per cent in different cultivars (Pillai et al., 1977). The factors responsible for spike shedding are yet to be clearly understood. An understanding of the pattern of spike shedding will be of use to identify the factors responsible for the same which will in turn help to arrive at effective control measures.

In view of the above facts, the present studies have been undertaken in pepper at the College of Horticulture, with the following objectives.

To study (1) the pattern of growth and flowering

(11) the floral biology

(111) the pattern of spike shedding.

REVIEW OF LITERATURE

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REVIEW OF LITERATURE

<u>Piper nigrum</u>, the black pepper of commerce is systematically placed under the family Piperaceae of the order Piperales.

Engler and Prantle as quoted by Benson (1970) regarded the Piperales as representing one of the most primitive type of dicotyledons. Rendle (1971) related Piperales to Polygonales in the trimerous relation of the flower and the orthotropous solitary ovule. Contrary to the views of Rendle and Engler, it is now considered that the family Piperaceae, though of undeterminate origin, is not one of the most primitive dicots, but an independent and terminal offshoot of direct Ranalian ancestry (Shukla and Misra, 1979).

1. GROWTH STUDIES

Growth studies are important for understanding the cropping behaviour of a plant which is evident from the works of Gustafson (1926), Reed (1929), Barnard (1932) and McNunn: (1939).

In pepper, growth studies have not been reported so far. But the relation between vegetative growth and

fruitfulness has been studied in various other crops, such as apple (Barnard and Reed, 1933), mango (Naik and Rao, 1942; Spencer and Kennard, 1955; Krishnamurthi <u>et al.</u>, 1961; Teotia <u>et al.</u>, 1970), citrus (Raimund, 1949; Krishnamurthi <u>et al.</u>, 1960; Randhawa and Sinha, 1963; Singh and Ghose, 1965; Hariano and Marioko, 1975), sapota (Sundararajan, 1961) and guava (Aravindakshan, 1964).

In mango, growth in a particular season influences greatly the capacity for flower production in the succeeding season. The nature of shoot growth in mango is cyclic as described by Naik and Rao (1942) and Krishnamurthi <u>et al.</u> (1961). They observed that a period of growth alternated with a period of quiescence. During the course of one year, five cycles of growth were recorded. Teotia <u>et al.</u> (1970) concluded that tree vigour could be taken as a criterion for yield in mango.

Halma and Compton (1936) studied the importance of growth flushes in citrus. Krishnamurthy <u>et al.</u> (1960) distinguished five cycles of shoots growth for one year. The maximum total growth was observed to be in the month of March, followed by that in July, August and September

in the descending order. Similar observations were made by Randhawa and Sinha (1963) and Singh and Ghose (1965).

In citrus, Saur (1951) elassified flowering shoots as (1) shoots bearing flowers but no leaves (11) shoots bearing flowers and few leaves (111) shoots with leaves and solitary flowers (iv) shoots with several flowers and several large leaves and (v) vegetative shoots with no flowers. In mango, Naik and Rao (1942) classified them as shoots with flowered leaders, non-flowered leaders and current year's leaders. In guava Dasarathy (1951) distinguished the current season shoots as flowering and vegetative types. They were classified into three as shoots which produced flowers and ceased growth, shoots which continued growth producing flowers and shoots purely vegetative (Aravindakshan, 1964).

In pepper the inflorescence is produced on the fruiting branches also known as the laterals. The lateral branches are plagiotropic with a sympodial growth habit and showing a periodical growth (De Waard and Zeven, 1969).

2. SEX AND SEX RATIO

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Koorders (1908) reported that most wild <u>Piper</u> species and some wild forms of <u>Piper nigrum</u> in the Western Ghats were dioscious. But surveys conducted in the forests of the Western Ghats revealed the frequent occurance of hermaphroditism among the wild types, as pointed out by Nambiar <u>et al.</u> (1978).

In pepper, majority of the cultivated varieties are reported to be hermaphrodite (Hasan Iljas, 1960; Nambiar and Sayeed 1962; Martin and Gregory, 1962), but there are few exceptions like Uthirankotta, Karuvilanchi and Mundi, which are predominantly pistillate.

Hagan Iljas (1960) observed that stamens may be present in a rudimentary form embedded in the tissues below the surface, which provided on explanation for the restricted hermaphroditism in some of the female cultivars. Male plants though rare could be easily recognised by their vigorous vegetative growth (Koorders, 1908; Marinet, 1953; Hasan Iljas, 1960; Cobley and Steele, 1976).

Nambiar <u>et al</u>. (1978) found wide variation among the cultivars with respect to the proportion of male,

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female and hermaphrodite flowers in a spike. The male flowers in a spike varied from 0 to 19 per cent, while bisexual flowers showed a wider variation of 2 to 93 per cent. The proportion of female flowers has been found to increase with an increase in the intensity of shade. Spikes produced during the off-season were also characterised by more number of female flowers than in those produced during the normal flowering season.

Shanmugavelu and Rao (1977) have also reported the influence of season on the sex of pepper vines. According to them, in the Travancore cultivars the heavy rains during March to May provide a favourable atmosphere for exhibiting hermaphroditism. On the other hand, lack of rain for six months from October have an unfavourable influence on the Malabar cultivars.

Cramer (1907), Anandan (1924), BlackLock (1954) and Nambier <u>et al</u>. (1978) stressed that a high ratio of hermaphrodite flowers was essential for increased production.

3. FLOWER PRODUCTION AND BLOSSOM STUDIES

The inflorescence in pepper develops simultaneously on the current season growth, opposite to a leaf. De Waard

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and Zeven (1969) reported the presence of two successive primordials of a raceme within a single bud giving rise to abundant flowering.

The inflorescence of pepper has been variously described as a catkin supporting 50 to 150 small secsile flowers by De Waard and Zeven (1969), a long slender pendulous spike by Cobley and Steele (1976).

De Waard and Zeven (1969) studied the development of the inflorescence in detail. They observed that the spike exhibited positive geotropism several days after emergence. After about 15 days when the immature raceme has increased in length, flowers appeared from the basal portion. A protogynic stage developed and existed for five days, subsequently followed by the appearance of the stamens from the base. Four or five days later each stigme was accompanied by one or two stamens. The development was fundamentally centripetal, but an irregular appearance or dominancy was frequently observed.

From studies conducted at the Pepper Research Station, Panniyur, by Nambiar <u>et al.</u> (1978) the following observations were made. The spike emerged covered in a sheath, the colour of which varied from green to pinkish

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or even violet. It took about 20 to 25 days for the full emergence of the spike from the sheath.
The flowers opened 8 to 14 days after the emergence of the spike, starting from the base and progressively advancing towards the tip. The complete opening of the flowers on a spike took about 6 to 9 days.

The period of existence of the protogynic stage was found to vary. Anandan (1924) and Cobley and Steele (1976) reported that in India protogyny extended over a period of 7 to 8 days. In Fuerto Rice 3 to 8 days passed before anther dehiscence was observed (Martin and Gregory, 1962). According to Hassan Iljas (1960) protogyny was a varietal character, which some times did not exist. From recent reports it is seen that though protogyny is seen in majority of the cases, the simultaneous opening of the male and female flowers and protandry are found as exceptions (Nambiar <u>et al.</u>, 1978).

In pepper, the naked flowers are more or less sunk in the fleshy axis of the spike on which they closely occur (Benson, 1970; Rendle, 1971). They described the flowers as minute, bracteate, usually

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bisexual, sometimes unisexual with no perianth. Purseglove (1977) described the bracts as ovate and fleshy.

According to Rendle (1971), in Piperaceae, the number of stamens varied from 1 to 10, but most of the flowers may be derived from a trimerous type with two whorls, each of 3 stamens, as occurs in <u>Piper amalago</u>; <u>Piper nigrum</u> has only two stamens the posterior one of the inner whorl having aborted. Cobley and Steele (1976) and Purseglove (1977) reported the number of stamens as 2 to 4, occuring on either side of the ovary in hermaphrodite flowers. De Waard (1967) observed that the stamen pushed its way through the catkin tissue and appeared as a white spherical body on the top of a short thick filament.

The ovary has been described as ovate, unilocular and superior (Cobley and Steele, 1976; Purseglove, 1977; Shukla and Misra, 1979). The number of carpels varied from 1 to 4 as reported by Benson (1970) and 1 to 5 reported by Shukla and Misra (1979). The ovule is single, solitary and orthotropous with two or sometimes one integument (Benson, 1970; Shukla and Misra, 1979). Shukla and Misra (1979) also described the placentation as basal.

Benson (1970) reported the number of stigma as 2 to 5, while De Waard (1967) and Purseglove (1977) found the number as 3 to 5. Cobley and Steele (1976) described the stigma as star shaped and sessile, the number corresponding to the number of carpels. Martin and Gregory (1962) reported that the succulent papillae of 10 μ in diameter were extremely sensitive to mechanical damage.

3.1 Anthesis

Nambiar <u>et al</u>. (1978) reported that anthesis in pepper commenced from 19.30 hours. Flower opening started from the base of the spike and continued towards the tip.

3.2 Anther dehisoence

In Sarawak, works by De Waard (1967) indicated that dehiscence of anther usually took place between 12.00 and 14.00 hours on days when a relative humidity approximately 60 per cent was attained at a temperature of 32°C and under conditions of bright sunshine. The mass of pollen spilled freely over adjascent stigmas and other parts of the spike. He also observed that

dehiscence within pairs was not simultaneous as a rule. Nambiar <u>et al</u>. (1978) suggested that in case of protogyny the anthers dehisced at anytime, within four days after the stigma became receptive. It was also reported that temperature and relative humidity controlled partially the longitudinal dehiscence of the pollen sac (Hasan Iljas, 1960; Martin and Gregory, 1962).

3.3 Receptivity of atigma

De Waard (1967) associated a viscous condition of the stigma with receptivity. He also found that increased relative humidity extended the receptive period. Nambiar <u>et al</u>. (1978) observed that the period of receptivity of stigma varied based on the position of flowers on the spike. The flowers at the base of the spike had a receptive period of 7 to 9 days, while it was only 3 to 5 days for those towards the tip.

4. POLLINATION

4.1 Mode of pollination

In pepper several modes of pollination have been - suggested.

Hasan Iljas (1960) has pointed out that the flower structure of pepper does not facilitate insect pollination. But Martin and Gregory (1962) had viewed that wingless insects occassionally occuring on the recemes may be potential pollinators, which was supported by De Waard (1967) from similar observations in Sarawak.

Wind pollination was considered to be as ineffective as insect pollination by Hasan Iljas (1960). He found that pollen transportation by Wind was negligible. Contrary to this, studies in Puerto Rico by Martin and Gregory (1962) indicated that 32 to 64 per cent of the pollen on the spike may be dispersed to the air within 24 hours of dehiscence and on release the small grains may be subject to wind transportion. Insect and wind pollination can evidently be considered to be accidental in majority of the cases.

Geitonogamy has been suggested as an effective mode of pollination by Hasan Iljas (1960) and De Waard (1967). This is a composite mode of self pollination involving the combined effects of rainwater alternating with prolonged periods of sun and wind. Dispersed

pollen grains move along the spike by gravity. Free hanging recemes inside polyethylene isolation bags displayed an unrestricted fruitset irrespective of insects or rainwater supporting the occurance of geitonogamy. According to both the authors, the factors stimulating geitonogamic fertilization are positive geotropism, spiral arrangement of the flowers, sequential ripening of the stigma and non-chronological dehiscence of anthers.

De Weard (1967) had also suggested that true self pollination or autogamy could be effective in hermaphrodite cultivars, wherein stigma branches curved very close to the fresh cluster of pollen during dehiscence. Autogamy was favoured particularly when environmental conditions promoted the extended period of receptivity of the stigma.

Nambiar <u>et al</u>. (1978), who could find large number of pollen grains in dripping water suggested that rain water was the chief pollinating agent in pepper. The evidence for this was drawn from an observational trial, wherein a few vines were protected from rain, half of which were sprayed with water and the other half left

unsprayed. Normal setting was found in vines sprayed with water, while the vines left unsprayed gave only a very low fruit set.

4.2 Artificial Pollination

Martin and Gregory (1962) described two different methods of hand pollination in pepper. In one method ripe anthers were opened by means of a scalpel and the pollen mass scooped up, which was applied to the appropriate stigma. The efficacy of this method was low. The other method consisted of brushing both donor and receptive spikes using a camel hair brush where fertilization was successful.

De Waard (1967) developed a method of hand pollination, in the oultivar Kutching, making use of the extended period of protogyny. Prior to hand pollination all spikes present on the receiving vine were removed to prevent geitonogamy between neighbouring spikes. At three or more locations branches which exhibited actively growing apical buds were selected and isolated in a bag of cheese cloth streched around a strong wire frame. No other spikes were allowed to develop. As soon as the stigma appeared on the proximal portion of the spike, a number of these, usually two or three per spike were marked. From the donor vine ripe anthers were selected and placed at the end of a long pin. Subsequently the entire pollen cluster was gently brought into contact with the young stigma. This method was found to be successful in 50 to 75 per cent of routine pollinations.

Nambiar et al. (1978) suggested that it was convenient to restrict pollination to a few flowers in a spike which would open within the next one or two days. In the method developed by them, anthers from the flowers of the selected portion of the spike. were removed with a dissecting needle without causing injury to the ovary and the remaining portion of the spike cut off with a pair of scissors. The emasculated flowers were covered with a paper bag, preferably with paraffin coated paper bag. Follen grains from selected spikes were collected by washing them with distilled water and collecting the washings or by extracting mature anthers and crushing them in distilled water. The pollen suspension thus obtained was used to pollinate flowers using a fountain pen filler, which was carried out at 20,00 hours. For ensuring maximum set the process was repeated for 3 to 5 successive evenings.

The pollinated spikes were kept covered till fruits started developing.

5. POLLEN STUDIES

Study of pollen grains is of great significance in floral biology and in interpreting taxonomic relationships and the origin of plants. The storage and germination of pollen grains play an important role in assisted pollination and hybridisation programmes.

5.1 Pollen production

Oberle and Geortzen (1952) and Rao and Khader (1962) suggested the use of a haemocytometer for estimating the pollen produced by a flower. In pepper, the number of pollen per anther is found to vary with cultivars. In Indian cultivars, the pollen yield per spike was estimated to be 5,00,000 to 7,00,000 by Marinet (1955) and 1,00,000 to 3,00,000 by Martin and Gregory (1962).

The influence of atmospheric conditions on pollen production has been reported by Brooks and Puri (1963), which was supported by results obtained by Sharma and Singh (1970) in mango. Higher temperature and drier climate appeared to be associated with increased pollen production per anther. According to Stanley and Linskens (1974) it is possible to derive a regression coefficient to predict the day of maximum pollen shed for any one species at a given location, from the degree hour heat sums if sufficient data is available.

5.2 Pollen morphology

Pollen morphology as an expression of phylogeny and evolution of plants has been stressed by various workers (Wodehouse, 1935; Rao, 1961; Nair, 1965). According to them the form of pollen grains was as useful as any other characteristic in the classification of plants. As a general rule, they served best in distinguishing between higher groups of plants such as tribes, families, genera and some species. Thie view was supported by the works of Fogle (1977) and Maas (1977) who have suggested pollen ultrastructure as a means to identify the tree fruit species and even clones within species. Nair (1965) classed the pollen grains of the family Piperaceae as monocolpate. Several investigators (Hasan Iljas, 1960; Martin and Gregory, 1962) reported that pollen grains of pepper were small with a mean diameter of 10 + irrespective of cultivars.

5.3 Pollen viability

Stanley and Linskens (1974) have classified the methods for testing the viability of pollen grains as germination and non-germination assays. In pepper, pollen viability studies have not been reported so far. However, in many other crops this has been pursued in detail.

5.3.1 Stain tests.

Zirkle (1937) suggested a method for testing the viability of pollen grains, by mounting them in acetocarmine. The grains which stained well, looked plumpy and well shaped were taken by him as fertile and the unstained shrivelled ones as nonviable or sterile. This method have been adopted by Balasubramanyam (1959) in guava, Nirmalendunath and Randhawa (1959) in pomegranate, Singh (1961) in mango, Singh (1962) in litchi, Nalawadi et al. (1977) in sapota and Thankammapillai et al. (1978) in gingerito find out the percentage of fertility.

The staining properties of various other compounds, suggestive of pollon fertility have been reported. They are iodine (Barnett and Carver, 1964; Brooks and Brooks, 1967), tetrasolium salts (Aslam <u>et al.</u>, 1964) and propinocarmine (Deshmukh <u>et al.</u>, 1978).

The different stains possess specific staining properties as explained by Stanley and Linskens (1974). According to them acetocarmine preferentially stained chromosomes, iodine stained starch and tetrazolium salts changed their colour in the presence of enzymes occuring in the viable pollen. They further observed that the use of stains was not sufficiently accurate when compared to germination tests because the immature and aborted pollen grains contained levels of constitutive chemicals enough to yield positive results in the stain tests.

5.3.2 Pollen germination

Compared to stain tests, in vitro germination of pollen can give a more accurate index of pollen viability. The level of germinability determined in vitro is some what inacurate according to Nair (1977). Addicott (1943) and Visser (1955) viewed pollen germination and pollen tube growth as two distinct physiological processes independent of each other.

5.3.2.1 Role of sucrose in pollen germination.

The role of sucrose in pollon germination and tube growth have been much debated. Many workers believed

that the externally supplied sugars had only an osmotic role and were not utilized by the tube for any nutritional purpose (Jost, 1905; Martin, 1913; Anthony and Harlan, 1920; Visser, 1955). This view was contradicted by others (Brink, 1924; O'Kelly, 1955; Vasil, 1958), who pointed out that apart from having an osmotic role the externally supplied sugars in the medium or in the style definitely served as a nutrient material for the growing tubes. However the work of O'Kelly (1955) has shown that germination was neither an osmotic or turgor phenomenon nor did sugars serve merely as a source of nutrition.

Adams (1916) obtained good pollen germination at various concentrations of cane augar for different crops such as 2.5 to 10 per cent for apple, 4 to 8 per cent for pear and 8 per cent for black currants. Successful pollen germination in many other crops have been reported. This include 20 per cent sucrose and 1.5 per cent agar for plum (Randhawa and Nair, 1960), 16 per cent sucrose and 0.7 per cent agar for sapota (Rao and Khader, 1960), 25 per cent sucrose and 0.5 per cent agar for mango (Singh, 1961), 30 per cent sucrose for cashew (Damodaran <u>et al.</u>, 1966) and 15 per cent sucrose for cocca (Ravindran, 1977).

Brink (1924) observed that when pollen was cultured in sugar or sugar agar medium the pollen tubes were as long or even longer than those formed in nature.

5.3.2.2 Effect of boric acid in pollen germination.

Schumucker (1935) discovered the stimulative role of boric acid in pollen germination and tube growth. In <u>Nymphaea</u> and in many other species studied by him, the element was found to occur in the pistillate tissues. Since then the role of boric acid in germination and tube growth had been studied by various workers in many plants. Thompson and Batjer (1950) in their studies on the pollen of different species of fruit trees, found that boron or boric acid in low concentrations of 25 to 40 ppm stimulated both germination and tube growth. Resnik (1956) observed that the addition of borio acid at concentrations ranging from 10 to 100 ppm improved germination by 10 to 15 per cent in 'Mayer' lemon. He obtained similar results with many other varieties and species of citrus.

Studies by Munzer (1960) revealed that 1 to 10 per cent boric acid stimulated pollon germination and tube growth in more than 60 species of angiosperms. Singh (1961) obtained increased pollen germination and tube elongation in mango with 20 ppm boron or boric acid. Rao and Khader (1960) found that germination of sapota pollen could be enhanced appreciably by the addition of 100 ppm boric acid to the sucrose-ager medium. Jose and Magoon (1972) could improve germination and tube growth with 200 ppm boric acid added to 5 per cent sucrose medium. Ravindran (1977) also emphasised the need of boric acid (100 ppm) for proper germination and tube growth of cocce pollen.

5.3.2.3 Effect of calcium nitrate in pollen germination.

The influence of calcium nitrate on pollen germination and tube growth, though not as effective as boric acid, is reported by various workers. It has been found that the addition of electrolytes to pollen culture hinder growth or inhibits it entirely. Lidforss (1896) and Brink (1924) observed that calcium nitrate even in small amounts were toxic to pollen. Contrary to this, works by Brewbaker and Rwack (1963), Kwack and Brewbaker (1963), Kwack (1965), Jose and Magoon (1972) and Ravindran (1977) have revealed the essential role of calcium in pollen germination and tube growth. The action of calcium

appeared to be based on the non-metabolic incorporation of calcium into pectic substances of the pollen wall. It has also been suggested that the presence of calcium resulted in increased resistance against the bursting of pollen tubes.

5.4 Pollen storage

Follen being a very delicate material, its handling requires great care. Various workers have described the methods of collecting pollen in detail (Fletcher, 1906; Barrett and Arisumi, 1952; Stanley and Linskens, 1974).

The term "pollinicuration" has been proposed by King (1962) to refer to procedures as collection, drying, testing viability, storage and shipment, particularly as those inclusive in the techniques of plant breeding.

A proper combination of factors such as low temperature, relation humidity and light have great bearing on pollen storage, as is evident from various reviews made by Pfundt (1910), Knowlton (1922), Doroshonko (1928), Nebel and Ruttle (1937), Maheshwari (1944), Visser (1955) and Singh et al. (1961).

5.4.1 Storage by controlling temperature and humidity.

King and Hesse (1938) studied the pollen storage requirements of 16 deciduous fruits and found the optimum temperature for storing pollen to be about 30°F. Nebel (1939) could successfully store the pollen of apple, pear, plum, peach and apricot for 2 to 5½ years in desiccator over sulphuric acid with 50 per cent R.H. at 28°C. Gollmick (1942) was able to extend the viability of grape pollen for a year at 1°C and 40 to 50 per cent R.H. Similar successful pollen storage studies by controlling temperature and humidity were conducted in papaya (Traub and O'Rork, 1936), coconut palm (Liyanage, 1949), olive (Nicolaissen, 1953), stone fruits (Remy, 1953), grapes (Nagarajan <u>et al.</u>, 1965) jack (Sinha, 1972) and lime (Shukla and Misra, 1975).

5.4.2 Storage by freezing.

Griggs <u>et al</u>. (1953) were able to store the pollen of plum, peach, almond, apple, pear, cherry and olive for 1 to 3 years in a home freezer at - 18°C. They observed that there was no difference in the germination percentage of pollen at the time of collection and almost after one year of storage. Singh (1962 a) reported the viability of mango pollen to be 14 months, when stored in deep freeze condition in a desiccator. In citrus, 50 per cent viability after 90 days storage in deep freeze was recorded by Sachan and Patro (1970). In Kagzi lime, Shukla and Misra (1975) reported 40 to 64 per cent fruit set with pollen stored in deep freeze for 15 days. Lyophilization or freeze drying of pollen has been reported to be one of the efficient method of pollen storage (Stanley and Linskens, 1974; Nair, 1977).

5.4.3 Storage by drying and dehydration.

Sedov (1955) made a comparative study of the shade dried and sun dried apple pollen and found that shade dried pollen gave better germination. Pollen dried in shade and stored in a desiccator: over calcium chlorite in darkness was found to be most viable. Good storage life over calcium chloride had been reported in various fruit crops by Tatarineev and Ostrowhova (1950, 1956), Soost and Cameroon (1954), Singh (1960, 1961, 1962) and Satjan and Kleeva (1964). In apple Satjan and Kleeva (1964) observed that pollen stored in desiccator for one year failed to germinate in artificial media, but showed germination on the stignatic surface.

6. FRUIT DROP

Fruit drop is a universe phenomenon exhibited by a wide variety of crops. This problem which is severe in many fruit crops has been much investigated and reviewed as by Coit and Hodgson (1916, 1918), Murneek (1933), Harrold (1935), Srivastava (1938), Naurial (1955), Korrigs and Kester (1959), Singh (1960, 1961), Chadha and Singh (1963, 1964), Singh (1965), Palmer <u>et al.</u> (1968), Pollard and Biggs (1969), Wilson (1969), Rogers (1971), Jawanda <u>et al.</u> (1972) and Hariom <u>et al.</u> (1975).

In pepper, spike shedding has so far been considered as a natural phenomenon. Recently Fillai <u>et al</u>. (1977) have reported that loss of crop due to spike shedding may be as high as 40 per cent, especially during unfavourable years.

6.1 Factors influencing fruit drop

Several theories have been put forward in an attempt to explain the mechanism of fruit drop. Lewis (1946) had postulated that the relative post fertilization production of hormones by the developing embryo determined the further retainment. If embryo failed to develop, the ovulary tissues abscised. If they developed, the ovulary tissues remained attached until maturity. In mange degeneration and abortion of ovule as early as at the four nucleate stage of embryo sac was responsible for the drop of 40 to 50 per cent of fruits (Singh, 1965).

Addicott and Lynch (1955) explained the possible influence of an abscission mechanism, which was supported by the works of Chadha and Singh (1963) and Randhawa (1971). A relation between endosperm and embryo development has been suggested by Leuty and Bukovac (1968). They observed that fruits about to abscise and those considered as potential drops were characterised by a smaller pericarp or no embryo or an aborting embryo or more than one of these factors.

Hormones are important in that they have a decisive role in bringing about abscision of flowers and fruits. Auxin-abscission relationship has been supported by several workers (Addicott and Lynch, 1955; Coyne and Al-Yasuri, 1964; Bardwaj, 1975; Hariom <u>et al.</u>, 1975; Varma 1976; Addicott and Wiater, 1977). The influence of gibberellins, abscisin, cytokinin and ethylene on abscission had also been explained by various workers (Gustafson, 1960; Cooper and Henry, 1971; Varma, 1976; Addicott and Wiater, 1977).

According to Bardwaj (1975), the interaction between various growth regulators determine the ultimate abscission. The auxin and gibberellins produced in the seed and the abscision in the pericarp may be transported to and interact at the abscission zone located at the base of the pedicel. If auxin and gibberellin were not available in sufficient amounts so as to neutralize the effect of abscisin, the flower or fruit would be shed. Thus ultimately, the relative balance between various plant growth regulators determines the retention and abscission.

Various external factors are also involved in flower and fruit drop of which temperature (Yamagutchi, 1954) and moisture status of the soil (Carns, 1951) are most important.

6.2 <u>Waves of drop</u>

The abscission of flowers and partially developed fruits is not ordinarily continuous but proceeds in more or less definite waves. The waves of drop have been worke out in different crops. The interval between and the intensity of different waves depend upon several factors such as species, variety, occurance of fertilization.

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position of flowers and prevailing weather conditions as reported by Randhawa (1971).

In apples, Howlett (1927) recognised two periods off abscission. The 'first drop' which was heavy, began shortly after petal fall and lasted for two to three weeks, while the so called 'June drop' which began a few days after the completion of the first drop continued for two to four weeks. This view was shared by Mc Cown (1938), Vyvyan (1946) and Randhawa (1971).

Chadha and Singh (1964) distinguished three waves of drop in mango as pinhead drop, post setting or 'April drop' and unripe fruit drop or 'May drop'. In citrus Randhawa (1971) recorded three waves of drop which were during the month following full bloom. 'June drop' and preharvest drop.

In pepper, Pillay <u>et al</u>. (1977) studied the various aspects of spike shedding and made the following observations. The spikes dropped from the plant at various stages of their maturity beginning from the time of emergence to the date of harvest. But the intensity of shedding was found to be high at two definite stages of maturity of spikes, that is, after

the fertilization of flowers and then in the advanced stages of fruit development. They found that the percentage of setting in spikes dropped at the early stages of development was very low, varying from 0 to 25 per cent. Hence they associated poor setting in the spikes with the drop observed at this stage. Shedding in the advanced stages of fruit development was thought to be due to physiological disturbances in the plant, which was caused by prolonged spells of drought or heavy rains or the sharp and sudden alteration of the two. This view was supported by the observation that intensive shedding occured during years in which heavy North-East monsoon showers were obtained after a spell of dry period following the South-West monsoon.

They concluded that, four major factors, that is, genetic character of the plant, imbalanced nutrient status of the soil, climatic factors and diseases or pests were responsible for spike shedding. They further observed that the influence of these factors were interrelated and complimentary. In one season one or more of these factors may have an upper hand, though the effects of others cannot be ignored. The extent of shedding in different varieties studied by them varied from 14.02 per cent to 65.62 per cent. In the variety Panniyur-I shedding was observed to be 26.24 per cent.

MATERIALS AND METHODS

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MATERIALS AND METHODS

The studies on the growth, flowering, floral biology and spike shedding were carried out at the Pepper Research Scheme, Vellanikkara (attached to the College of Horticulture) for a period of 16 months from May, 1979 to August, 1980. The vines were 4 years old and were under uniform cultural and manurial treatments as per the package of practices of the Kerala Agricultural University.

The details of the materials and mothods adopted are as given below:

1. SHOOT GROWTH

Eight standards of the cultivar Panniyur-I were selected for studying the growth of shoots. Twenty lateral shoots were selected at random on each standard. The shoots were tagged and numbered serially. The extension in shoot length was measured in om at weekly intervals commencing from 20th May, 1979 to 30th May, 1980.

2. FLOWER CHARACTERS

For studying flower characters, six vines of

Panniyur-I were selected. The various aspects studied were the pattern of flowering, development of the spike, sex-ratio and floral biology which included anthesis, anther dehiscence, stigmatic receptivity and mode of pollination.

2.1 Pattern of flowering

Twenty lateral shoots of each standard were selected at random, tagged and numbered serially. Observations were made for the number of shoots flowered and number of spikes per shoot.

2.2 Spike development

Fifty spikes on each of the six standards were observed for studying the developmental stages of inflorescence. Measurements of spike length in on were taken at an interval of 2 days from the day of emergence to the day of attaining full length. Number of days required for the completion of anthesis and anther dehiscence in individual spikes were also recorded by daily observation of mature spikes. Similar observations were taken from the cultivar Karimunda.

2.3 <u>Sex ratio</u>

A random selection of fifty spikes on each of the

six standards were used for determining the sex-ratio. From the observation on the total number of flowers in a spike and number of hermaphrodite, pistillate and staminate flower per spike, sex-ratio was worked out. The sex-ratio of Panniyur-I was compared with that of Karimunda.

2.4 Floral biology

To study the floral biology, spikes were collected and flowers were described. Drawings were made of spikes, pistillate and herraphrodite flowers.

2.4.1 Anthesis.

To know the exact time of anthesis, 25 mature spikes were tagged and observed twice daily in the morning and evening. Preliminary observations indicated that anthesis took place during the late evening. Thereafter mature spikes along with the shoot were plucked at 18.00 hours and kept with their basal portion immersed in water. The spikes were observed at bi-hourly intervals to determine the actual time of anthesis.

2.4.2 Anther dehiecence.

The spikes tagged for determining the time of

anthesis were used for acertaining the time of anther dehiscence. The maturity of the anthers was evident from their brownish white colour. From initial observations it was noted that anthers dehisced during the afternoon hours. To find out the actual time, spikes with mature anthers were plucked along with the shoot at 13.30 hours, kept with their basal portion immersed in water and observed at hourly intervals.

2.4.3 Receptivity of stigma.

The receptivity of stigma was judged by the fresh, white colour and the shiny appearance. The confirmation of this was made by pollination studies. Nature spikes were selected and bagged using polythene bags. The spikes were observed daily and 15 to 20 flowers of each spike that would open the next day were emasculated using a needle. The remaining portion of the spike was cut off using a pair of scissors, after which the bags were replaced carefully. A pollen suspension prepared from mature anthers (pollen suspension was prepared in water by gentle crushing of anthers) was used to pollinate the flowers using a fountain pen filler. Pollination was done between 09.00 and 10.30 hours on 25 spikes each at different stages starting from one day prior to anthesis to eight days after anthesis. Twenty emasculated spikes were left bagged without pollination to observe if fruit set took place in pepper in the absence of pollination.

2.4.5 Mode of pollination.

To find out the role of rainwater in pollination, the following method was adopted. Fifty spikes were selected and labelled on each of the six standards used for the study. Twentyfive spikes of each standard were covered using polythene bags well in advance of anthesis. The remaining twentyfive were left open. After fruitset the bage were removed. The number of flowers which had set fruit were counted on covered as well as open spikes and the percentage fruitset worked out.

3. POLLEN STUDIES

The pollen grains of pepper being microscopic cannot be collected as such. Therefore the method adopted was to collect mature whole anthers from spikes plucked from individual standards. The maturity of anthers was judged from the change of colour from white

to brownish white before dehiscence. A pollen suspension prepared by gentle crushing of such anthers was used for studying the different aspects of pollen such as morphology, fertility, number of pollen per flower, pollen germination and pollen viability.

3.1 Morphology and fertility

A drop of the pollen suspension prepared was transferred to acctocarmine and glycerine kept on a clean slide using the base of a dissection needle. After covering with a clean cover glass, the slides were kept for about 30 minutes to allow the pollen grains to take stain properly before examining under a The diameter of pollen grain was measured microscope, using an occular micrometer and their average was taken as the diameter of each pollen grain. The diameter of 50 normal, well shaped and well stained pollen grains at random were recorded from each slide. From the total number of well stained pollen grains fertility was calculated. The experiment was repeated using iodine-potassium iodide (0.1% iodine in 2% potassium iodide) as stain.

3.2 Estimation of pollen production

A hasmocytometer as adopted by Rao and Khader

(1962) was used for estimating the number of pollen grains per flower. Spikes with mature anthers about to dehisce were collected. Hundred anthers were · generated with a dissection needle and taken in a small beaker. The anthers were crushed gently and 2.5 ml of water containing 0.25 per cent Calgon (Oberle and Geortzen, 1952) was added and the contents thoroughly stirred in order to obtain an even dispersion of the graine in the suspension. A drop of this suspencion drawn in a fine pipette was transferred to each of the two counting chambers of a hacmocytometer. Each chamber had an area of nine square millimeter ruled into square millimeter areas. Each of the four corner square millimeter areas were ruled into 16 while other five square milimeter areas were ruled into smaller divisions. The counting chambers were 0.1 mm in depth so that the volume of solution over 0.1 mm² was 0.1 mm³. The number of pollen per flower was calculated as,

> If N = average number of pollen counted per corner square. X = number of grains per anther. N : X= 0.1 : 25. 0.1 = 25 N X = 250 N

The pollen grains in each of the four corner squares of each counting chamber were counted with the help of a hand tally counter and by using the low power objective of the microscope.

For each standard, ten such estimates were made and the total flowers examined per standard was 250 numbers.

3.3 Effect of sucrose in pollen germination

The experiment was conducted initially using varying concentrations of sucrose (5, 10, 20, 25 and 30%) with 1 per cent agar and incubating in a moist chamber. There was no germination even after 48 hours. Repeating the experiment with lower concentrations did not better the results. Thereafter the concentration of agar was lowered to 0.5 per cent and the experiment carried out with 0, 2, 4, 6, 8 and 10 per cent sucrose where good germination was noticed.

3.4 Effect of boric acid and calcium nitrate in pollen germination

Effects of boric acid and calcium nitrate were determined at 0, 2, 4, 6, 8 and 10 per cent sucrose with 0.5 per cent agar. After incubating in a moist chamber for 4 hours, the growth was arrested with

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Carnoy's fluid and observations were made. The concentrations of boric acid and calcium nitrate tried were 10, 20, 30 and 40 ppm. The independent as well as the combined effects were studied.

For estimating germination, on an average, 400 pollen grains were counted from twenty microscopic fields and for tube length 100 pollen tubes were measured using an occular micrometer. Germination was expressed in percentage and tube length in μ . The average value for the experiment repeated thrice was taken.

3.5 Pollen storage

Mature anthers were extracted from spikes with a dissection needle. The anthers were crushed gently in distilled water and tap water so as to get a pollen suspension. Both suspensions were stored as follows:

1. In a deciccator at room temperature.

2. At 4°C.

The viability of pollen under each treatment was determined at daily intervals by staining with acetocarmine and percentage fertility worked out.

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4. SPIKE SHEDDING

For studying the pattern of spike drop, twenty standards of the cultivar Panniyur-I were used. The base of these standards were kept clean and daily counts were made of the spikes shed by each till harvest. The yield of each standard at harvest was determined. The total number of spikes shed by each standard was estimated. The percentage shed during different months was also worked out.

Another trial was conducted to study the different stages of spike drop starting from emergence to harvest. For this, twenty shoots were labelled on each of the six standards. The spikes produced on each shoot were observed at 3 day intervals from emergence.

5. STATISTICAL ANALYSIS

The data on the different characters studied were subjected to statistical analysis, following the methods suggested by Snedecor and Cochran (1967). Transformations were done wherever needed and the data analysed by the analysis of variance technique. Significant results were compared after finding out the critical differences.

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RESULTS

RESULTS

The results of the study on growth, flowering, floral biology and spike shedding are presented below. The analysis of variance tables for the different aspects are given in appendices II to IX.

1. SHOOT GROWTH

The extension growth of pepper shoots at weekly intervals from 20-5-1979 to 19-8-1979 is presented in Table 1a. The data showed that shoot growth started at the end of May and continued upto the middle of August. Statistical analysis showed significant variation in the mean extension growth and the percentage of shoots showing growth between weeks at one per cent level.

The maximum mean growth was recorded during the period between the third weeks of June and July (5.55 cm) which accounted to 63.68 per cent of the total growth for the year. The percentage of shoots that showed growth was also maximum during this period, which ranged from 71.25 per cent to 86.25 per cent. This was followed by growth during the period between the third week of May and second week of June (2.37 cm) which accounted to 26.28 per cent of the total growth and when 7.5 to

Week	Mean extens- ion in cm	Mean growth expressed on percentage of the total	Percentage of shoots showing growth		
20-5-79	0.16	1.39	7.50	(15.89)*	
27-5-79	0.30	2.97	18.75	(25.70)	
3 -6-7 9	` 0. 50	5.51	37.50	(37.76)	
10-6-79	0.61	6.87	50.00	(45.00)	
17-6-7 9	0.60	9.54	62.50	(52.24)	
24-6-79	1.01	11.70	71.25	(57.61)	
1-7-79	1.05	12.11	82.50	(65.27)	
8-7-79	1.11	13.19	85.00	(67.21)	
15-7-79	1.25	. 14.51	86.25	(68.28)	
22-7-79	1.11	12.17	78,75	(62.58)	
2 9-7-7 9	0.64	6.83	56.25	(48.62)	
5-8-79	0.32	2.96	37.50	(37.76)	
12-8-79	0.06	0.51	12.50	(20.70)	
19-8-79	0	0	0		
F value	32 .25* *		35.00*	·*	
CD (0.05)	0.20	9.81			

Table 1a. Weekly growth of pepper shoots (c.v. Panniyur-I) (Mean of 160 shoots)

** Significant at one per cent level

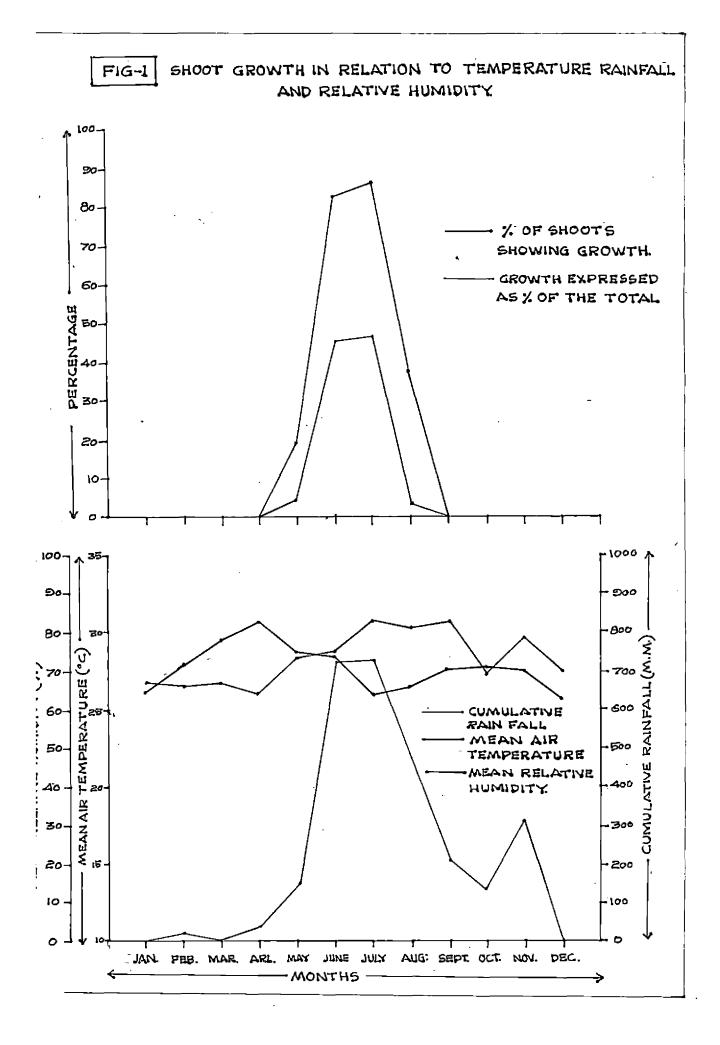
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* Values in parenthesis indicate angular transformed ones

to 62.5 per cent of the shoots showed growth. Minimum growth (1.02 cm), accounting to 10.30 per cent of the total was recorded for the period between the fourth week of July to the second week of August. During this period growth was observed in 12.50 to 56.25 per cent of the shoots. Thereafter no growth was observed. The total extension growth for the year was 10.27 om.

Considering the months, maximum growth occured in July (46.70%) followed by June (35.73%), May (4.35%) and August (3.47%). The percentage of shoots that showed growth followed the same trend with the maximum in July (76.32%) and minimum in August (50.00%). Shoot growth in relation to temperature rainfall and relative humidity is illustrated in Fig.1.

During the months of June and July, when shoot growth was maximum, the average temperature was 28.55°C and 26.05°C respectively. The total rainfall for these periods ranged between 722.7 mm and 729.8 mm, while the relative humidity varied between 75.0 and 85.0 per cent (Appendix I). During May and August, when growth was low, the temperature was 28.75°C and 26.5°C respectively. The rainfall ranged from 155.1 mm to 462.6 mm and the



relative humidity from 74.5 per cent to 81.0 per cent. There was no growth from September to November, when rainfall varied between 127.3 mm and 317.4 mm. There was not much variation in temperature (27.7 to 27.55°C) and relative humidity (70.0 to 82.5%).

The mean growth of the individual standards is presented in Table 1b. Statistical analysis showed that the mean shoot extension and the mean percentage of shoots that showed growth in each standard varied significantly at one per cent level. The extension in shoot growth varied between 5.28 cm and 12.04 cm and the percentage of shoots in which growth was observed varied from 65 per cent to 100 per cent.

2. FLOWER CHARACTERS

The results of the studies on flower characters are presented as follows.

2.1 <u>Plowering pattern</u>

The observations on flowering of pepper vines during the different months are presented in Table 2a. The data indicated that in pepper, flowering was confined to four months of the year, 1979, that is, May, June,

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Vines	Mean extens- ion in cm	Percente showing	age of shoots growth
s ₁	5.28	70.00	(56,79)
s ₂	8.79	90.00	(71.56)
^S 3	12.04	100.00	(90.00)
s ₄	12.0 0	90.00	(71.56)
s ₅	7.80	75.00	(60.00)
s ₆	11.73	85.00	(67.21)
^s 7	7.09	70.00	(56.79)
s ₈	6 .58	65.00	(53 .73)
P value	13•75**	20.27**	•
CD (0.05)	0.20	7.70	

Table 1b. Mean extension growth of pepper shoots in different standards (c.v. Panniyur-1)

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** Significant at one per cent level

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* Values in parenthesis indicate angular transformed ones

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	No. of	Percentage of shoots flowered		No. of spikes per shoo		
Months	shoots studied			Mean	% of the total	
May	120	35.00	(36,06)*	0.53	11.80	
14 2 y	120	99400	()0:00)-	0.00	11.00	
June	120	60.00	(50 . 83)	1.33	29.62	
July	120	79.17	(63.13)	2.26	50.33	
August	120	25.00	(30.39)	0.38	8.46	
F value 3	1.93**		95.49**			
CD (0.05)0.47		4.56				

Table 2. Pattern of monthly flowering in pepper (c.v. Panniyur-1)

** Significant at one per cent level

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* Values in parenthesis indicate angular transformed ones.

July and August, Maximum percentage of shoots flowered in July (79.17%) followed by June (60.00%), May (35.00%) and August (25.00%). No off season flowering was observed. Statistical analysis showed significant variation between months with respect to the percentage of shoots flowered. The mean number of spikes per shoot also followed the same pattern with the maximum in July (50.33%) and minimum in August (8.46%). There was significant difference in the mean number of spikes per shoot among the months from May to August.

The data on flowering of individual standards are presented in Table 2b. Statistical analysis showed that significant variation existed between standards with respect to the percentage of flowering. S_4 (95%) recorded significantly higher flowering followed by S_5 (85%), S_6 (85%), S_1 (85%), S_2 (85%) and S_3 (80%). No significant difference was noted for the mean number of spikes per shoot between standards. The percentage of aborted spikes varied from 4021 to 26.31 per cent in different standards.

2.2 Spike development

The extension growth of spikes from emergence is

Details of standard	No. of shoots studied	Percenta- ge of shoots flowered	Mean number of spikes per shoot	
s ₁	120	85	3.65	13.69
^S 2	120	85	5.15	4.85
⁸ 3	120	80	2.85	26.31
s ₄	120	95	4.75	4.21
S ₅ .	120	8 5	4.55	7.69
^S 6	120	85	б.00	0.00
1997-19-17-19-19-19-19-19-19-19-19-19-19-19-19-19-				
F value		5.72**	2.21 ^{NS}	
CD (0.05)		5.58		

Table 2b. Pattern of flowering in different pepper standards (c.v. Panniyur-T)

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** Significant at one per cent level

NS Not significant

presented in Table 3a, Fig.2 and Plates I and II.

The spikes of both cultivars showed a linear growth pattern. The length of spike at emergence was 1.34 cm in Panniyur-I, which showed regular increase till the twenty-second day. At this stage the length was 10.2 cm, after which the growth decreased. The spike attained the maximum length of 12.5 cm in 31.67 days.

The spike of Karimunda had a length of 0.9 cm at the time of emergence, which increased steadily till the twenty-fifth day. Thereafter the growth decreased, reaching the maximum length of 6.20 cm in 29.25 days.

The period of spike development, anthesis and anther dehiscence of Panniyur-I and Karimunda are given in Table 3b.

In Panniyur-I, the spike attained maximum length in 31.67 days after its emergence. Anthesis started from the base of the spike and progressed towards the tip. The anthesis of the first few flowers (3 to 12) commenced 17.33 days after emergence. The anthesis of all the flowers in a spike was completed in 25.44 days after emergence. The interval between anthesis of the

Table 3a.	Extension growth	of spil	kes from	emergence	n in	pepper
	(o.vs. Panniyur-I	and Ka	rimunda)	(Mean of	100	spikes)

		Panniyu	<u>r-1</u>	Kari	Karimunda			
	Extension (cm)	Increase in growth(cm)	Growth ex- pressed as percentage of total	Exten- sion (cm)	Increa- se in growth (om)	Growth expressed as per- centage of total		
1	1.34	1.34	10 .7 2	0.90	0.90	14.52		
4	2.70	1.36	10,88	1.40	0.50	80.6		
7	3.60	0.90	7. 20	1.80	0.40	6.45		
10	4.80	1.20	9.60	2.40	0.60	9.68		
13	6.30	1.50	12.00	3.00	0.60	9.68		
16	7.60	1.30	10.40	3.70	0.70	11.90		
19	8.90	1.30	10.40	4.30	0.60	9,68		
22	10.20	1.30	1 0.40	4.90	0.60	9.68		
25	11.00	0.80	6.40	5.50	0.60	9.68		
28	11.60	0.60	4.80	5.90	0.40	6.45		
31	12.20	0.60	4.80	6.20,	0.30	4.84		
34	12.50	0.30	2.40	0	0	0		

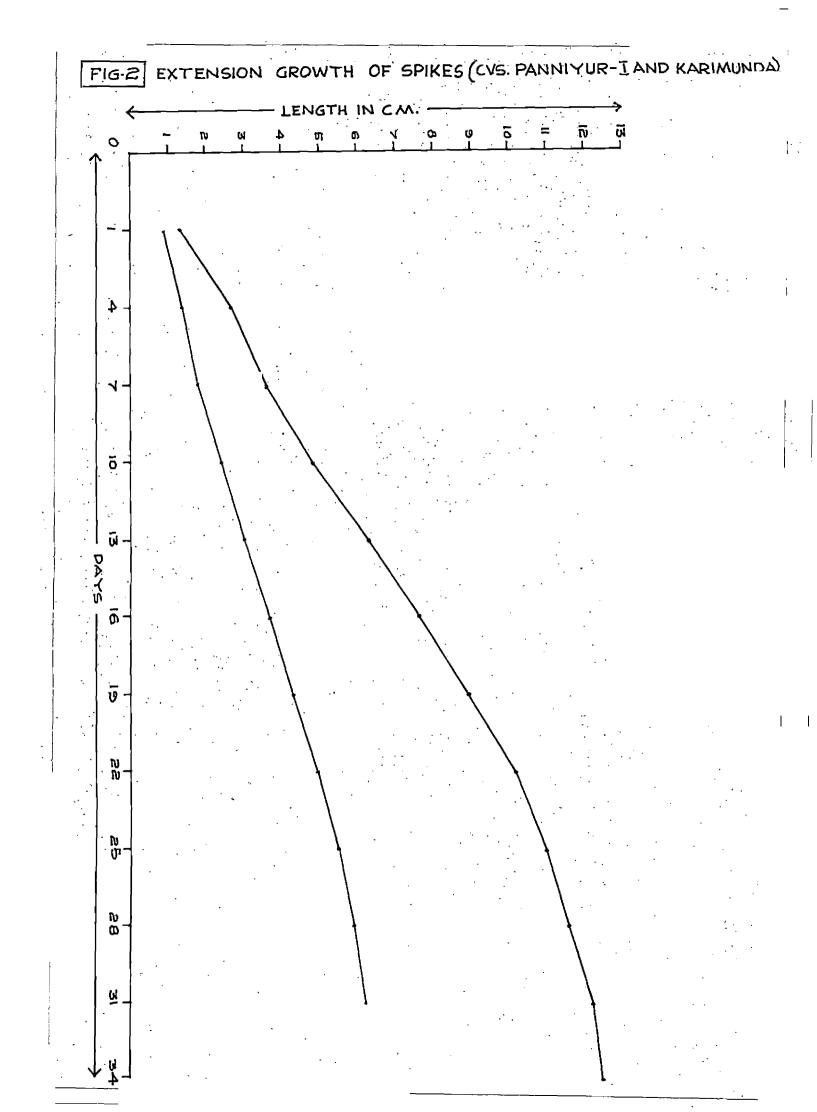


Plate I Stages of spike development in cv. Panniyur-I

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Plate II Stages of spike development in cv. Karimunda

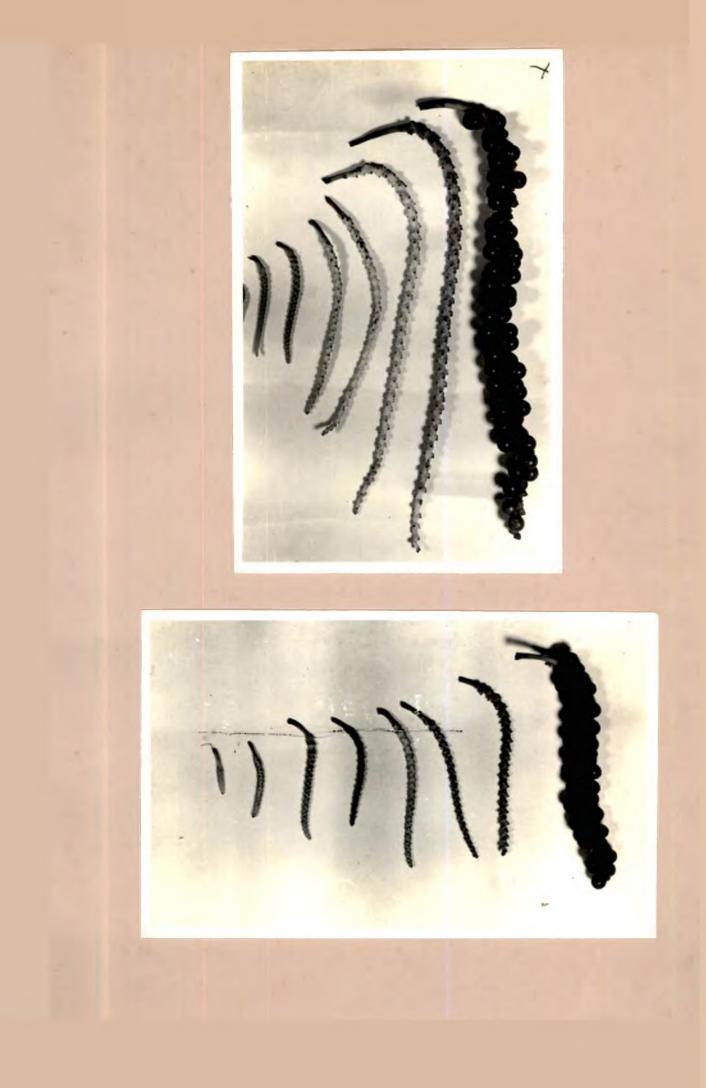


Table 3b. Pattern of spike development, anthesis and anther dehiscence in paper (orvs. Panniyur-I and Karimunda)

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Cultivar		Mean number of days taken from emergence to					
· . ·	Attain max. length	First flower opening		ther de-	ther de-	Interval between first and last flower opening	Interval between first and last anther dehiscence
Panniyur-1	31.67	17.33	25.44	23.85	.31.57	8.12 (range 6 - 10)	7.72 (range 4 - 9)
Karicunda	29 .2 6	19.01	28,22	24.23	29 .2 8	9.21 (range 4 - 11)	5.0 (range 3 - 8)
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first and the last flower was 8.12 days. The immature spike was green in colour, the colour being retained for the first 4 to 5 days of anthesis. Thereafter a greenish yellow colour developed from the base and continued towards the tip. The colour deepened as the days advanced and the whole spike attained a yellow colour by the time all flowers were open. The stigma had a white, shiny appearance during the receptive period. The flowers were protogynous and the protogynic stage existed for 6.52 days. The anther dehiscence usually commenced from the base of the spike, progressing towards the tip. But a non-chronological dehiscence was observed occassionally. The first anthers (1 to 6) dehisced 23.85 days after spike emergence. The dehiscence was longitudinal, which within anther pairs was not simultaneous. The anthers which were white in colour initially, turned brownish white before dehiscence. The dehiscence of all the anthers in a spike was completed in 31.57 days and the period of dehiscence was 7.72 days.

The spike development pattern in Karimunda was similar to that in Panniyur-I, differing in the mean number of days taken for attaining the various stages. The spike attained maximum length in 29.26 days. Anthesis

started 19.01 days after emergence and was completed in 28.22 days after emergence, the period being 9.21 days. Anther dehiscence commenced 24.28 days after emergence and lasted for a period of 5.00 days. The protogynic condition existed for 5.27 days.

2.3 Floral biology

The different espects of floral biology of pepper are described below.

2.3.1 Sex ratio.

The sex ratio pattern of Panniyur-I and Karimunda are given in Table 4. In Panniyur-I, two types of flowers that is, hermaphrodite and pistillate, were observed. The total number of flowers per spike ranged between 99 and 116.83 with a mean of 107.74. Hermaphrodite flowers varied from 96.0 to 112.6 with a mean of 104.7 and accounted to 97.18 per cent of the total flowers per spike. The rest which were pistillate flowers ranged between 1 to 4 with a mean of 3.13 (2.82%).

In Karlmunda, the total number of flowers observed per spike varied from 42.50 to 48.61 with a mean reaching 45.23, which were all hermaphrodite.

Variety	Mean nuaber of flowers/ spike	Nean number of hermaphro- dite flowers	Percen- tage	Mean number of pistil- late flowers	Percen- tage	Mean number stami- nate flowers	Fercen-
Panniyu r-1	107.74	104 •7	97 .1 8	3.13	2.82	0	Ο.
Karinunda	46.23	46.23	100.00	0	0	0	0
			, 				

Table 4. Pattern of sex-ratio in pepper (CAVS, Panniyur-I and Karimunda)

- 18

2.3.2 Hermaphrodite flower.

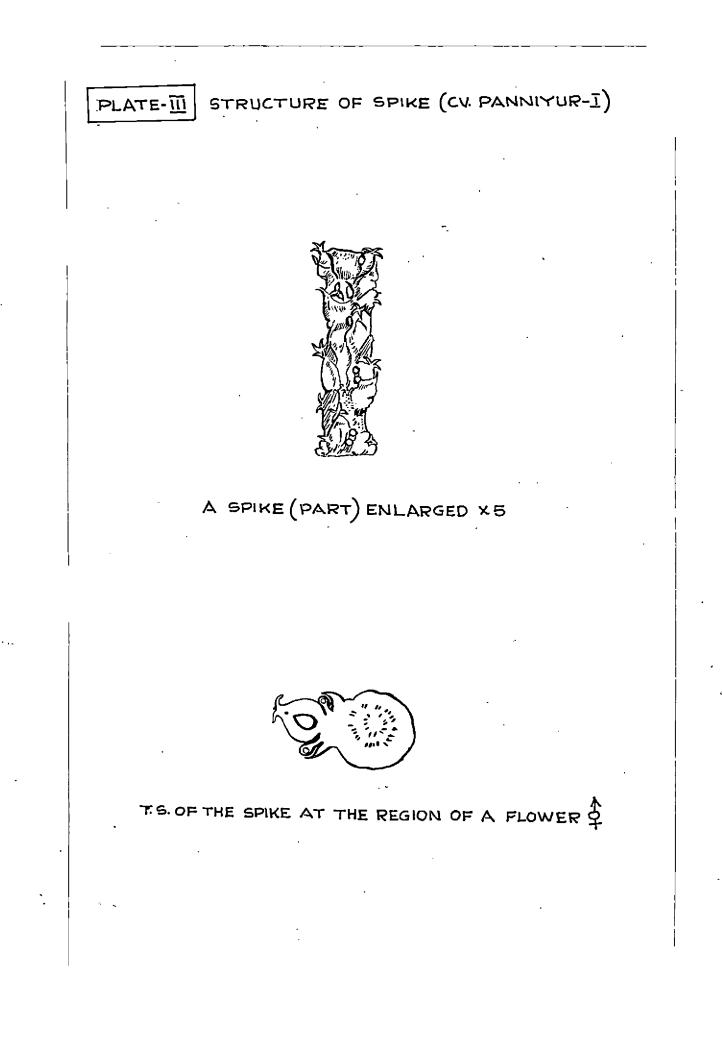
The flowers occured closely, more or less sunk in the fleshy axis of pendant spikes (Plate III & IV). Individual flowers were small, measuring about 2 mm, bracteate and without perianth. The bracts appeared, ovate, fleshy and sheild like. The stamens, which were persistant varied from 1 to 2, each consisting of two anthers, borne on a very short filament. The anthers were spherical, white before and brownish black after dehiscence. The stigma was sessile, star shaped and deciduous with 3 to 4 lobes. The stigma was shiny white when receptive, turning black after fertilization. The ovary was globose, inilocular and superior, with a single orthotropous ovule.

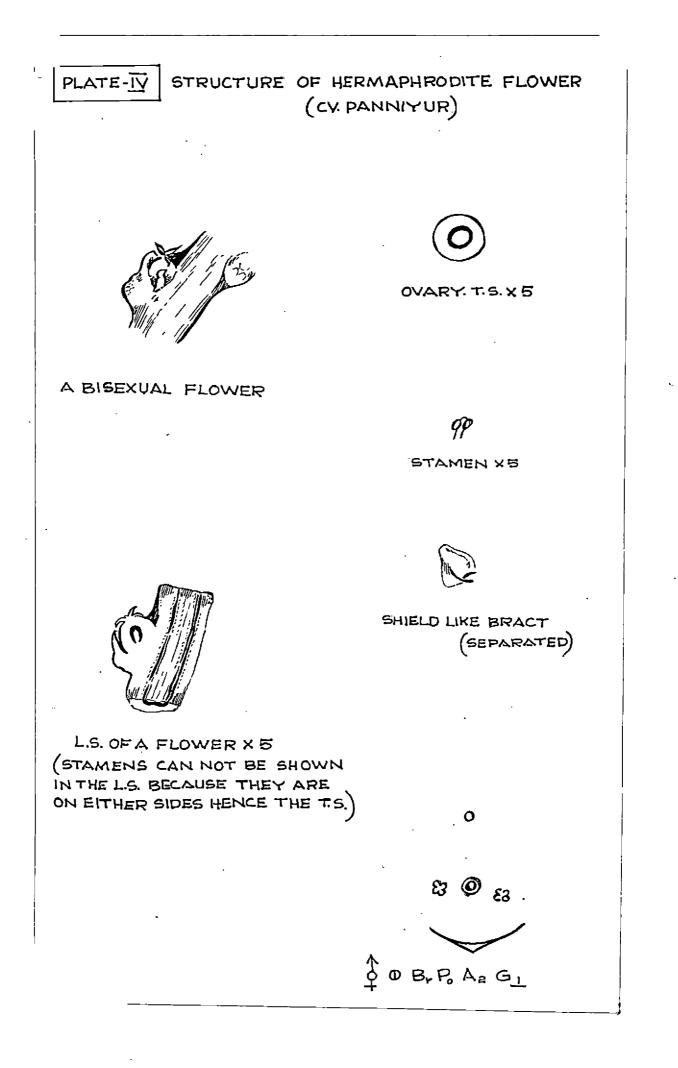
2.3.3. Pistillate flower.

Pistillate flowers resembled hermaphrodite flower except for the absence of stamens.

2.3.4 Anthesis.

The data in Table 5 give the anthesis time of pepper flowers. The anthesis started between 18.00 and 18.30 hours and continued up to 02.30 hours of the next day. Maximum flowers opened between 18.30 and





Tie9 (hours)	Number of flowers obscryed	Numbe r opened	Percentage of the total
18.00	84	. 0	
18.30		12	14.28
20.30		36	42.87
22.30		18	21.42
24.30	,	11	13.10
02 .30	,	7	8.33
04.30		0	0.00
Total	، 1999 - میں باریک کر	84	100.00

Table 5. Period of anthesis in papper (c.v. Panniyur-I)

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23.30 hours. The study showed that anthesis in pepper was for a period of 8.30 hours between 18.00 and 02.30 hours.

2.3.5 Anther dehiscence.

Observations on anther dehiscence at hourly intervals starting from 13.30 hours are presented in Table 6. The data indicated that dehiscence started by 14.30 hours and continued up to 16.30 hours. Maximum dehiscence (56%) occured between 14.30 and 15.30 hours, followed by 24 per cent between 15.30 and 16.30 hours.

2.3.6 Stigmatic receptivity.

The white, shiny stigmatic surface was found to be retained up to 5 to 8 days after anthesis. Pollination studies presented in Table 7 showed that fruit set occured when pollinated from the day of anthesis up to seven days after anthesis. Maximum fruit set was recorded during the first three days, where the set ranged from 77.25 to 85.00 per cent. The percentage of set on the fourth day was 72.19. Thereafter the set decreased progressively till the eight day, when it was nil.

<u>59</u>

Time (hours)	Number of anther observed	Number dehis- ced	Percentage of the total
13.30	25	O	
14.30		5.	20.00
15.30	· 、	14	56.00
16.30		6	24.00
17.30		0	0.00
Total	9 1997 - 1997 - 1997 - 1997 - 1 997 - 1997 - 1997 - 1997 - 1997 - 1997 - 1997 - 1997 - 1997 - 1997 - 1997 - 1997	25	100.00

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Table 6. Period of anther dehiscence in pepper (c:v. Panniyur-I)

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Pollinated	Number of spikes	of flow-		Percen- tage set
One day before anthesis	25	20	0	0
First day of anthesis	25	20	15.45	77.25
Second day of anthesis	25	20.47	16.60	81.09
Third day of anthesis	25	20.00	17.00	85.00
Fourth day of anthesis	25	20.10	14.50	72.14
Fifth day of anthesis	25	20.05	13.65	68.07
Sixth day of anthesis	25	20.10	8.00	39.80
Seventh day of anthesis	25	22.00	2.51	11.40
Eighth day of anthesis	25	20.00	0	0
No pollination	25	20.00	0	0

Table 7. Percentage of fruit set on hand pollination at different intervals in pepper (c.v. Panniyur-T) (Time 9 AM - 10.30 AM)

3. MODE OF POLLINATION

The result of the study on the mode of pollination in pepper is presented in Table 8.

The data revealed that spikes protected from rain registered a lower fruit set compared to spikes that were left open. Under bagged conditions, the set ranged from 61.42 per cent to 72.57 per cent with a mean of 67.35 per cent in the different standards. Under open conditions, the set recorded varied from 81.05 per cent to 86.50 per cent with a mean reaching 83.42 per cent. Statistical analysis showed significant difference between treatments at one per cent level.

The effects of different pollination treatments (no pollination, open pollination and hand pollination) are presented in Table 8b. No set was recorded in the absence of pollination. Under open pollination, a set of 83.44 per cent was noticed. When hand pollinated, the set observed was only 80.00 per cent. This indicated that under open pollination, a higher percentage of set was possible than under hand pollination.

4. POLLEN STUDIES

The results of the different aspects of pollen studies are described below.

Treatments	No. of spikes obser- ved	Mean number of flo- wers/ spike	Number sot	Porcen- tage set
Spikes bagged	150	100.07	67.13	67.35
Spikes open	150	106.70	89.02	83.42
			t =	2,228**

Table 8a. Fruit set in pepper (c.v. Panniyur-I) under bagged and open conditions

** Significant at one per cent level

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	<u> </u>		
Treatments	Number of flowers observed	Number set	Percentage set
	<u></u>		`
No pollination	250	0	0
Open pollination	640	534	83.44
Hand pollination	150	120	80.00

Table 8b. Fruit set in Pepper (o, v. Panniyur-I) under different pollination treatments

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4.1 Morphology and fertility

Pollen grains of pepper were not visible to the naked eye. Microscopic examination showed that the pollen grains were light brown spherical structures with a well demarcated exine and a single nucleus. A single pollen grain measured 9.52 to 10.20 μ in diameter with a mean reaching 9.86 μ . The fertility recorded by the stain test was 98.47 per cent in acetocarmine and 97.61 per cent in iodine with a mean of 98.04 per cent (Table 9).

4.2 Pollen production

The recorded number of pollen per flower ranged from 7000 to 13000 (Table 10), with a mean of 9542. Statistical analysis showed significant variation between standards at 1 per cent level in the number of pollen grains per flower.

4.3 Role of sucrose in pollen germination

Results of pollon germination in different sucrose concentrations are given in Table 11. Maximum germination was observed in four per cent sucrose followed by six, two, eight, ten and zero. There was no germination in twelve per cent sucrose media. Statistical analysis showed significant difference between treatments at one

Stain used	Number of pollen obser- ved	Number	Number of non- viable pol- len	Percen- tage of ferti- lity	Average size of pollen (~)
Acetocarmine	655	6 45	10	98.47	10.20
Iod 1 n 0 (0.1%)	630	615	15	97.61	9.52
Total	1285	1260	25	196.08	19.72
Meen	,	630	12,50	98.04	9.86
			والمراق متريك فالمراجع المراجع		,

Table 9. Pollen morphology and fertility in pepper (c.v. Panniyur-I)

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Number of	Number of pollen/flower		
TICACLE OPSCIAST	Mean	Range	
250	9090	7000 - 12000	
250	9389	7300 - 11000	
250	10148	7200 - 13000	
7 50	28627		
250	9 542 .3 3		
	5 .93 *	; •	
	1709.34	、	
	flowers observed 250 250 250 750	flowers observed Mean 250 9090 250 9389 250 10148 750 28627 250 9542.33 5.93*	

Table	10.	Pollen	production	in	different	standards	in
	•	pepper	(c v. Panni	yw	r→])		

** Significant at one per cent level

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Sucrose concentration (%)	Interval between incubation and observation hrs.	Percentage germina- tion	Mean tuba length (µ)
0	18	3.20	18.70
2	18	69.55	80.86
4	18	87.29	99.82
6	18	73.10	90.50
8	18	45.13	59.67
10	18	20 .2 8	40 .21
12	18	0	0
F value 2007.4	ļ0 * *		in en andre son andre gebeer om
CD 1.5	53		

Table	11.	Pollen germination in pepper (c.v. Panniyur-I)
		in sucrose agar media

** Significant at one per cent level

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per cent level. The data in Table 12 showed that the percentage germination in four per cent sucrose-agar media reached the maximum of 87.29 per cent in four hours at room temperature in humid chamber. There was no increase in tube growth after four hours.

4.4 Effect of boric acid and calcium nitrate in pollon germination

The effects of boric acid and calcium nitrate at different sucrose concentrations with 0.5 per cent agar are presented in Table 13 and Table 14 respectively.

Table 13 revealed that boric acid at 10, 20 and 30 ppm increased pollen germination at all levels of sucrose. The maximum germination percentage (94.68%) was recorded in four per cont sucrose with 20 ppm boric acid.

Data in Table 14 showed that at all levels of sucrose, calcium nitrate at 10 and 20 ppm increased germination percentage. But the percentage germination was reduced in the presence of 30 and 40 ppm calcium nitrate. Maximum germination (89.97%) was noted at four per cent sucrose with 20 ppm calcium nitrate.

However, the maximum percentage of pollen

Sl. No.	Hours after pollen planting	Percentage germination	Tube length
1	1	25.62	28.11
2	2	59.83	44.00
3	3	75 °32	69.00
4	4	87.29	99.82
5	5	87.25	99.00
6	6	87.26	99.32
7	7	87.11	99.51
8	8	87.25	99,50

Table 12. Duration of optimum incubation for maximum germination in four per cent sucrose agar media

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Sucrose	Concentrations of boric acid (ppm)					
(%)	0	10	20	30	40	
0	3,20	3.44	4.31	o	0	
2	69 .55	77.37	83.52	77.64	58.58	
4	87.29	90.38	94.68	89.20	71.52	
6	73.10	76.47	80.22	76.69	62.00	
8	45 .13	52.15	58.79	54 . 67	38 .1 0	
10	20.28	26.21	3 3•49	2 7. 67	14.60	
value	643.01**	809	4.42**	***************************************		
D	0.74		0.70			

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Table 13. Effect of boric acid on pollen germination percentage at different sucrose concentrations in pepper (c.v. Panniyur-I)

** Significant at one per cent level

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Table 14. Effect of calcium nitrate on pollen germination percentage at different sucrose concentrations in pepper (c.v. Panniyur-1)

Sucrose	Concentr	ations of	calcium	nitrate	(ppm)
(%)	0	10	20	30	40
0	3.20	1.37	O	0	0
2	69.55	71.80	73.72	69.00	49 .3 6
4	67.29	89.15	89.97	84.57	61.15
6	73.10	75.29	7 7. 66	64.88	44.18
8	45 .13	47.13	49.43	41 .7 8	25.31
10	20.28	22.12	23.85	14•29	4.53
F value	1173.58**	5278.1	7**		\$ <u></u>
CD	0.92	0.8	2		

** Significant at one per cent level

germination (95.77%) was observed for 20 ppm boric acid and 10 ppm calcium nitrate in four per cent sucrose and 0.5 per cent agar media (Table 15).

Statistical analysis of the data for pollen germination showed that the treatments varied significantly. The individual effects and the combined effects of all treatments were found to be significant at one per cent level. Normal pollen grains and tube growth in sucrose agar media are illustrated in Plates V and VI respectively.

4.5 Pollen storage

The percentage viability of pollen grains in acetocarmine under different treatments, recorded at daily intervals are presented in Table 16. The data indicated that pollen grains of pepper could be stored for 1 to 4 days in water at room temperature and at 4°C.

Pollen suspension in distilled water stored at 4°C remained viable for four days. The viability ranged from 86.88 per cent on the first day to 12.10 per cent on the fourth day after collection, with a maximum of 86.88 per cent on the first day. At room temperature and kept in a deciccator, pollen in distilled water lost

Plate V Normal pollen grains of pepper, cv. Panniyur-I (X 120)

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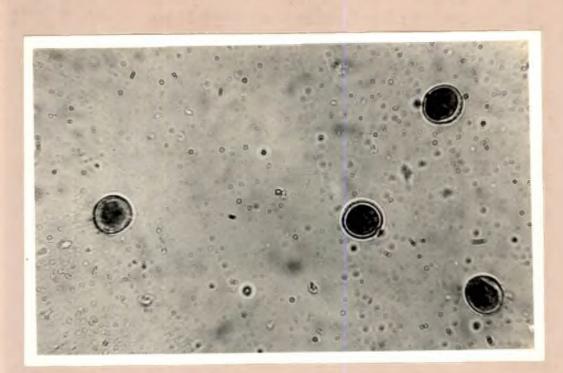
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Plate VI Pollen germinated in sucrose agar media (X 120)





Concentra-	Concentration of boric acid (ppm)					
tion of calcium nitrato (ppm)	0	10	20	30	40	
0	87 .29	90.38	94.68	89.20	71.52	
10	89.15	93.19	95.77	93.48	71.94	
20	89.97	89.95	90.27	88.39	68.42	
30	84.57	86.30	88.35	79.63	60.70	
40	61.15	62.33	60 .3 3	5 0 •44	45.53	
P value	301.36**	609.68*			,	
JD	1.85	1.53				

Table 15. Effect of borie acid and calcium nitrate on pollon germination percentage at 4 per cent sucrose in pepper (c.v. Panniyur-1)

** Significant at one per cent level

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			Days after collection					
		Treatments	1	2	3	4	5	
1.		len suspension in tilled water					 -	
	a.	Kept in dec ict ator at room temperature	65.50	27.17	0	0	0	
	b.	Kept at 4°C	86:88	69.23	33.00	12.10	0	
2.		len suspension in inary (tap) water		·	,			
	а.	Kept in decicator at room temperature	21.96	Ō	0	0	0	
	b.	Kept at 4°C	79.26	48.67	17.38	0	0	

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Percentage pollen fertility in acetocarmine under different storage treatments in pepper (c.v. Panniyur-I) Table 16.

viability on the third day. Here the percentage viability was 65.50 and 27.17 per cent on the first and second day after collection respectively.

In tap water, pollen lost viability on the fourth day when kept at 4°C and on the second day at room temperature. At 4°C, the viability ranged from 79.26 per cent to 17.38 per cent on the first and third day after collection respectively. At room temperature, the viability was only 21.96 per cent after storing for one day.

5. SPIKE SHEDDING

The observations on spike shedding during the different months are presented in Table 17 and in Fig.3. Shedding was observed in almost all the months starting from flowering till harvest. Maximum shed was noticed in June (51.31%) followed by July (23.00%), November (13.94%), December (4.81%), August (2.69%) and October (2.63%). Shedding was minimum in May (1.69%), while in September it was nil. Statistical analysis showed that months differed significantly for the percentage of spikes shed.

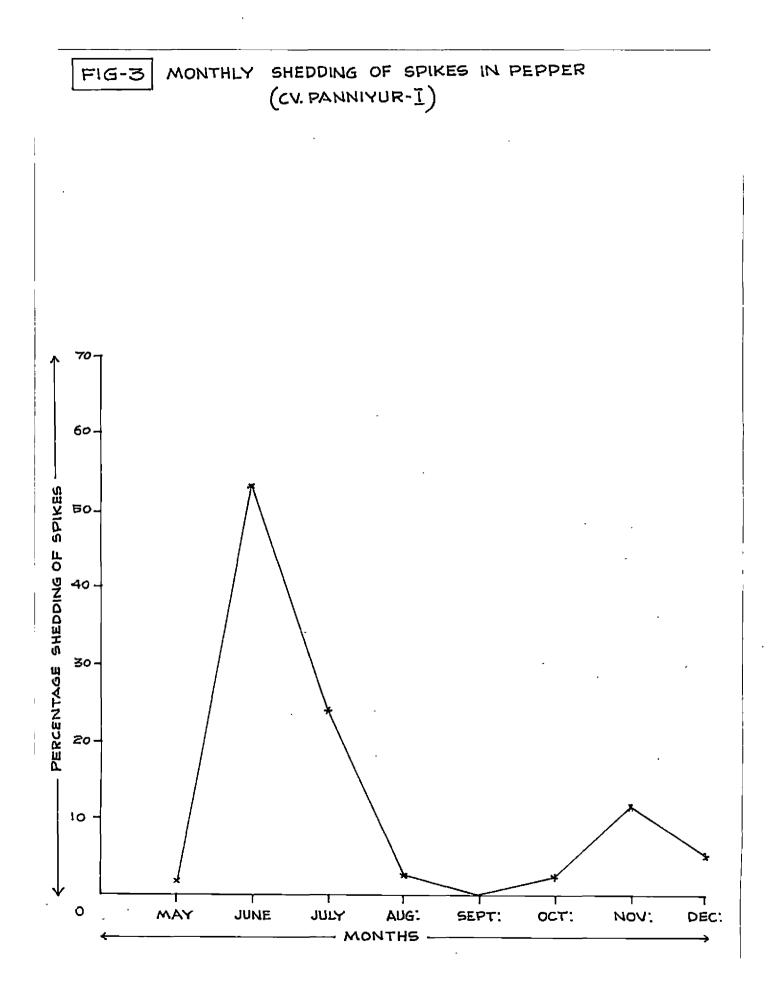
The extent of shedding shown by the different

Monthe 1979	Mean number of spikes shed	Percentage shed	of the total
May	1,69	1.69	(7.49)*
June	51.31	51.31	(45.75)
July	23.00	23.00	(28,66)
August	2.69	2.69	(9.46)
September	0.00	0.00	(0.00)
October	2 .63	2.63	(9.28)
November	13.94	13.94	(21.89)
December	4.81	4.81	(12.66)
F value	45.17**		nan-datanan ang antang kan nanan ing tan'na ¢
CD	6.53		

Table 17. Spike shedding during different months in pepper (c. v. Panniyur-I)

** Significant at one per cent level

* Values in parenthesis indicate angular transformed ones



standards is presented in Table 18. The number of spikes shed by a standard varied from 66 to 168 with a mean of 100.06 spikes per standard. This accounted to a mean shed of 23.82 per cent of the total spikes produced per standard. There was no significant difference between standards at 1 per cent level.

The data on the percentage of spikes dropped during the various stages of development is given in Table 19. The data indicated that spikes were shed at all stages of development starting from emergence till harvest. It was possible to recognise eight stages of shedding. Maximum percentage of shedding (53.15%) was noticed during the first month of fruit set. This was followed by shedding before anthesis (24.86%) and that during anthesis (16.57). The shed showed a decline during the second (5.52%), third (2.76%) and fourth (1.66%) months of fruit set which increased to 5.52 per cent and 9.94 per cent during the fifth and sixth month of fruit set.

			_
No.	Number of spikes produced	Number of spikes shed	Shed expressed as percentage of total spikes
s,	278	158	56.83
^S 2	508	168	33.07
s _z	231	144	62.34
^S 4	350	120	34.29
⁸ 5	614	81	13.19
^S 6	462	102	22.08
\$ ₇	388	101	26.03
^S 8	371	66	17 .7 9
s ₉	624	113	18,11
^S 10	347	42	12.10
8 ₁₁	556	45	8.09
^S 12	463	49	10.58
^S 13	79 7	91	11.42
^S 14	883	7 6	8.61
^S 15	362	92	25.41
^S 16	825	153	21.10
Total	1 8059	1601	381.04
Mean F val	503.69 Lue	100.06 _{NS} 0.26 ^{NS}	23.82

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Table 18. Extent of shedding in different standards in pepper (c.v. Panniyur_1)

NS Not Significant

11. 10.	Stages	Percentage shed
	Before anthesis	24.86
?	During anthesis	16.57
5	First month of fruit set	33.15
	Second month of fruit set	5.52
5	Third month of fruit set	2.76
5	Fourth month of fruit set	1.66
?	Fifth month of fruit set	5.52
	Sixth month of fruit set	9.94

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Table 19. Shedding of spikes at different stages of development in pepper (c.v. Panniyur-1)

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DISCUSSION

DISCUSSION

The results of the various studies conducted with respect to growth, flowering, floral biology and spike shedding are discussed below.

1. SHOOT GROWTH

The maximum extension growth of shoots (82.43%) and the percentage of shoots that showed growth (71.25% to 86.25%) in the cultivar Panniyur-1 corresponded to the months of June and July. Growth ceased by the middle of August and no further extension was observed. The extension growth in the different standards varied between 5.28 cm and 12.04 cm, while the percentage of shoots showing growth varied between 65 per cent and 100 per cent.

The maximum growth observed during the months of June and July is natural in pepper considering the high soil moisture level consequent to rain in June (722.7 mm) and July (729.8 mm). The maximum and minimum temperature during that period was 35.7°C and 22°C respectively with a mean of 28.5°C. The relative humidity during the above two months ranged between 53 per cent and 98 per cent with a mean of 85 per cent.

The optimum temperature and rain may be responsible for the higher growth during these months and the lower rainfall during May (155.1 mm) and August (462.6 mm) for the poor growth during that period.

In addition to the main growth, an offseason growth is also noticed in pepper during certain years when a dry spell is followed by high rainfall. But this was not observed in Panniyur-I during the year of study. The absence of growth during September and October may be attributed to the continuous rains experienced during August (462.6 mm) and September (208.7 mm). This indicates that continuous rain is not conducive for a second flush, but more detailed studies are required to confirm the above indications.

2. FLOWER CHARACTERS

2.1 Flowering pattern

The maximum flowering with respect to the percentage of shoots spiked and the mean number of spikes per shoots was observed during the months of June and July. The standards differed significantly for the number of shoots spiked and the mean number of spikes per shoot. The data clearly follows the pattern of shoot growth which is quite natural because of the fact that spikes are produced on the current season's growth opposite to a leaf.

The percentage of aborted spikes varied from O to 26.31 per cent. The reason for the occurance of spike abortion are not clear. It can be partly explained as due to, physiological and genetic causes. More detailed studies are required in this line.

2.2 Spike developement

The growth of spikes in both the cultivars followed a linear pattern. In Fanniyur-I, the spike attained the maximum length of 12.5 cm in 31.67 days after emergence. In Karimunda it took 29.26 days for attaining the maximum length (6.2 cm). The period of anthesis ranged between 6 to 10 days (Mean 8.12) and 4 to 11 days (Mean 9.21) in Fanniyur-I and Karimunda respectively. The period of anther dehiscence was 4 to 9 days (Mean 7.72) in Fanniyur-I and 3 to 8 days (Mean 5.00) in Karimunda. These observations are in agreement with the findings of Nambiar <u>et al.</u> (1978) except in the number of days taken for attaining maximum length, which according to them was 20 to 25 days. The flowers were protogynous and the protogynic condition existed for 5 to 8 days (Mean 6.52) in Panniyur-I and 3 to 7 days (Mean 5.27) in Karimunda. Anandan (1924) and Cobley and Steele (1976) observed that protogyny existed for 7 to 8 days under Indian conditions. Martin and Gregory (1962) observed protogyny for 3 to 8 days in Puerto Rico. Nambiar <u>et al.</u> (1978) found the protogynic stage to last for 2 to 10 days in different cultivars studied. Therefore the results of the present studies are in confirmity with the findings of the above authors.

Protandry and the simultaneous occurence of anthesis and dehiscence were not observed during this study as was reported by Nambiar <u>et al.</u> (1978).

The dehiscence of anthers was longitudinal, which usually commenced from the base of the spike, though occassionally a non-chronological dehiscence was noticed. Similar observations were made by De Waard (1967).

2.3 Floral biology

In Panniyur-1, the mean number of flowers

in a spike was 107.74, of which 104.7 were hermaphrodite (97.18%) and 3.13 were pistillate (2.82%). In Karimunda, a spike had a mean of 46.23 flowers, which were all hermaphrodite. The preponderance of hermaphrodite flowers in these cultivars is responsible for the high percentage of berry set.

The flowers were bracteate without perianth as recorded by Benson (1970) and Rendle (1971). The bract appeared fleshly and shield like as described by Purseglove (1977). In hermaphrodite flowers, the stamens were found on either side of the overy and the number veried from 1 to 2. Rendle (1971) found the stamen number as 2, while Cobley and Steele (1976) and Purseglove (1977) observed the number as ranging from 2 to 4. The number of stignatic lobes were observed to vary from 3 to 4. Benson (1970) recorded the number of stigmatic lobes as 2 to 5, while De Waard (1967) and Purseglove (1977) reported the number as varying from 3 to 5. The elight variation in the number of stamens and stigmatic lobes in Panniyur-1 is a varietal attribute. The climate and physiological condition of the vine are also likely to contribute to such variation. The stigna was sessile and star shaped as observed by Cobley and Steele (1976). The ovary

appeared ovate, unilocular and superior with a single orthotropous ovule. Similar observations were made by Benson (1970), Purseglove (1977) and Shukla and Misra (1979).

2.3.1 Anthesis

In pepper flowers, anthesis was observed for a period of 8.30 hours between 18.00 hours and 02.30 hours on the next day, with a maximum between 18.30 hours and 24.30 hours. These observations are in confirmity with the findings of Nambiar <u>et al</u>. (1978) who found the anthesis to commence from 19.30 hours under Panniyur conditions.

2.3.2 Anther dehiscence

Anther dehiscence started by 14.30 hours and continued up to 16.30 hours with the maximum between 14.30 and 15.30 hours. The mean temperature during the period of study was 31.1°C and relative humidity ranged between 68.0 and 98.0 per cent.

De Waard (1967) reported that anther dehiscence took place between 12.00 and 14.00 hours at Sarawak, when 60 per cent RH and 32°C temperature prevailed and under conditions of bright sunshine. Hasan Iljas (1960) and Martin and Gregory (1962) have also reported the influence of relative humidity and temperature on anther dehiscence. Slight variation in the time of dehiscence observed during the present studies may be attributed to variation in temperature, humidity and sunshine.

2.3.3 Stigmatic receptivity

stigmatic receptivity commenced from the day of anthesis and lasted upto seven days after anthesis. Receptivity was not observed during the preanthesis stage. The results are in agreement with those obtained by Nambiar <u>et al.</u> (1978), who observed that flowers at the base of the spike had a receptive period of 7 to 9 days. De Waard (1967) found the peak period of receptivity to be three to five days after anthesis, while in the present studies the peak receptive period was observed to be on the first three days. The difference observed may be due to climatic variations.

2.3.4 Pollen studies

Pollon grains of pepper were microscopic. Pollon grains were spherical, measuring within a range of 9.52 to 10.2 μ in diameter with a mean of 9.86 μ .

Stain tests recorded 98.04 per cent fertility. Pollen number per flower (in four anthers) ranged from 7000 to 13000 with a mean of 9542.

Hasan Iljas (1960) and Martin and Gregory (1962) found the mean diameter of pollen grains as 10 irrespective of cultivars which is almost in agreement with the present observations.

The pollen yield per spike has been estimated to be 5 to 7 lakhs by Marinet (1955) and 1 to 3 lakhs by Martin and Gregory (1962) in Indian cultivars. According to the present findings, the mean number of flowers per spike is 107 in Panniyur-1 with 97.18 per cent hermaphrodite flowers. Therefore in the case of this cultivar under Kerala conditions 7 to 13 lakhs pollen grains are estimated per spike which is higher than the estimation of the above authors. Even among their findings there is wide difference. Therefore the higher pollen production may be a varietal character which in turn is affected by climatic variations.

Follen germination studies showed that sucrose, boric acid and calcium nitrate influenced germination of pepper pollen. Germination was observed in zero to ten per cent sucrose with 0.5 per cent agar with a

maximum (87.29%) in four per cent sucrose agar. Optimum time for incubation was found to be four hours. In the presence of sucrose and boric acid, maximum germination (94.68%) was recorded in four per cent sucrose with 20 ppm boric acid. With calcium nitrate, maximum (89.97%) was recorded for 20 ppm calcium nitrate in four per cent sucrose agar media. With regard to the combined effects of the three, maximum was observed for 20 ppm boric acid and 10 ppm calcium nitrate in four per cent sucrose and 0.5 per cent agar (95.77%).

Pollen germination in sucrose agar media has been reported in several crops, with varied concentrations such as 25 per cent sucrose and 0.5 per cent agar for mango (Singh, 1961), 16 per cent sucrose and 0.7 per cent agar for sapota (Rao and Khader, 1962) and 15 per cent sucrose for cocca (Ravindran, 1977). The effect of sucrose in pollen germination may either be nutritive or due to the comotic or turgor phenomena of sugar as postulated by 0'Kelly (1955).

The stimulative influence of boric acid on pollen germination has been reported by various workers which include 25 to 40 ppm in various fruit crops

(Thompson and Batjer, 1950), 10 to 100 ppm in citrus (Resnik, 1956), 20 ppm in mango (Singh, 1961), 200 ppm in sapota (Jose and Magoon, 1972) and 100 ppm in occoa (Ravindran, 1977). The results of the present studies are in agreement with the above workers, except in the concentration. The beneficial effect of boron is due to the fact that it is found to occur in the stigmatic fluids and the pistillate tissues as suggested by Schumucker (1935).

Calcium nitrate has been reported to inhibit (Lidforss, 1896; Brink, 1924) as well as promote (Kwack and Brewhaker 1965; Jose and Magoon, 1972; Ravindran, 1977) pollen germination. The promotive action of calcium nitrate in the present studies may be attributed to the non-metabolic incorporation of calcium with pectic substances of the pollen wall and the increased resistance of the pollen tubes against bursting in the presence of calcium as explained by Jose and Magoon (1972).

Pollen storage studies revealed the possibility of storing pollen grains of pepper as a suspension in water. Distilled water proved to be a better storage medium than tap water. In distilled water at 4°C.

the pollen had a storage life upto 4 days, while in tap water it was 3 days. The shorter storage life in tap water can be due to the possible impurities present in it. The fact that pollen retains its viability in water also suggests the role of rain water for proper pollination in pepper.

3. MODE OF POLLIMATION

Pollination studies showed that under protected conditions in polythene bags, spikes registered a lower fruit set (67.35%) than under open conditions. Hand pollination of spikes showed a lower fruit set (80.00%) than open pollination (83.44%).

Nambiar <u>et al</u>. (1978) observed a very low fruit set in vines protected from rains. They considered rain water as the chief pollinating agent. During the present studies, though the spikes were protected with polythene bags it was not possible to exclude water droplets completely. Water droplets resulting from high humid conditions within the bag and also due to transpiration of the spike were retained inspite of the holes provided at the bottom. These droplets may have

assisted the movement of pollen grains down the spike, thus effecting pollination. In The pendulous nature of the spike and the dehiscence of anther from the bage of the spike would have helped the transport of pollen by gravity. The result of the present study confirmed the view of De Waard (1967) who found geitonogamy under the influence of gravity and dew drops to be effective in hermaphrodite cultivars.

Sreekunary Anna and Vijayagopal (1977) reported that rain water did not have any role in pollination of pepper, as they did not find any difference in fruit-set between protected and unprotected spikes. But they did not consider the possible effect of water drop; within the bags on pollination. The significantly lower fruit set observed in bagged spikes during the present study indicates the role of rain water in the natural pollination of pepper.

For the present it is reasonable to assume that rain water in combination with other factors such as gravity does have a role in the natural pollination of pepper.

The lower fruit set under hand pollination may have been due to the injury caused to the ovary during emasculation. De Waard (1967) obtained a set of 50 to 75 per cent with hand pollination which is lower than that recorded during the present study. The variation in the results may be due to differences in the method adopted.

4.SPIKE SHEDDING

Spike shedding was observed in almost all the months starting from flowering till harvest. The maximum shed was noticed in June (51.31%) and minimum in Nay (1.695). In September the shedding was nil. Spike shedding was observed to the tune of 23.82 per cent of the spikes produced per standard. Spike shedding occured during the various stages of development in varying intensities. The shed which occured during the period between spike emergence and first month of fruit set accounted to 74.58 per cent. Shed during the second, third and fourth months worked out to 9.94 per cent. During the fifth and sixth month of fruit set a shed of 15.46 per cent was noticed.

Two waves of spike shedding were observed in the cultivar Panniyur-1. The first wave started from

the spike emergence to the first month of fruit set, which corresponded to the period from May to August. The second wave, which can be considered as preharvest drop corresponded to the months of November and December. Pillai <u>et al.</u> (1977) could identify two peaks of spike shedding, that is, in the post fertilization period and in the advanced stages of spike development. Dropping of fruits in waves have been reported in different crops such as apple (Me Cown, 1938; Randhawa, 1971) and mengo (Singh, 1960; Singh, 1963).

The shed noticed during the period between emergence and anthesis may be due to an abscission mechanism as reported by Addicott and Lynch (1955), Ohadha and Singh (1963) and Randhawa (1971) in different crops. The drop during anthesis can be attributed to lack of pollination. Poor berry setting can be considered as a factor responsible for the shedding noticed during the first month of fruit set. Pillai <u>et al</u>. (1977) found very low percentage of setting (0 to 25%) in the spikes dropped during the post fertilization period. The drop at this stage may also be related to the relative production of

hormones by the developing embryo as explained by Lewis (1946).

The drop observed from the second month of fruit set till harvest can be explained as due to nutritional or hormonal imbalance. Bardwaj (1975) have suggested that the interaction of the various plant growth regulators determines the ultimate absoission. Pillai <u>et al</u>. (1977) attributed moisture stress as one of the factors for spike shedding in the advanced stages of fruit development. But during the year of study, moisture cannot be taken as a limiting factor during the pre-harvest shedding, because there was rain during October (127.3 mm) and November (317.4 mm). The fact that fungus attack (<u>Colletotrichum</u>s) was noticed during the second wave of shedding indicates the possible association of pathogens.

During the year of study, shedding observed in Panniyur-1 was 25.82 per cent Pillai <u>et al</u>. (1977) observed a shedding of 26.24 per cent in this cultivar. The variation observed clearly indicates that environmental conditions nutritional factors, and physiological condition of the vine also play a role in spike shedding.

SUMMARY

SUMMARY

The present studies were carried out at the Pepper Research Scheme attached the College of Horticulture during a period of 16 months from May, 1979 to August, 1980, with the following objectives.

To study (i) the pattern of growth and flowering (ii) the floral biology and (iii) the pattern of spike shedding.

The following conclusions were made based on the present studies.

1. Shoot growth in pepper (c.v. Panniyur-I) was observed for a period of four months from May to August.

2. The maximum growth occurred during July (46.70%) followed by that in June (35.73%), May (4.36%) and August (3.47%). The percentage of shoot that showed growth also followed the same trend. Significant variation was observed for the mean extension growth and the percentage of shoots showing growth in the different months.

3. The extension of shoots varied between 5.28 and 12.04 cm with a mean of 10.27 cm. Significant

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difference was noted for the mean growth in the different standards. No off season growth was noticed during the year of study.

4. Flowering followed a pattern similar to growth with maximum in July (50.33%) followed by that in June (29.62%), May (11.80%) and August (8.46%). Significant variation existed between standards with respect to the percentage of flowering. The percentage of aborted spikes varied from 40% to 26.31 per cent in the different standards.

5. The spike development followed a linear pattern. In Panniyur-I the maximum length (12.5 cm) was attained in 31.67 days, while in Karimunda (6.20 cm) it took 29.26 days.

6. The period of anthesis in a spike varied from 6 to 10 days (Mean 8.12) in Panniyur-I and 4 to 11 days (Mean 9.21) in Karimunda. The anthesis of the first few flowers commenced 17.33 days after emergence of spike in Panniyur-1. The anthesis of all the flowers in a spike was completed in 25.44 days. In Karimunda, anthesis started 19.01 days after emergence and was completed in 28.22 days. 7. The flowers were protogynous. The protogynic stages existed for 5 to 8 days (Mean 6.52) in Panniyur-1 and 3 to 7 (Mean 5.27) in Karimunda.

8. Anther dehiscence which commenced 23.85 days after spike emergence, extended for a period of 4 to 9 days (Mean 7.72) in Panniyur-T. In Karimunda it started 19.01 days after emergence and lasted for a period of 3 to 8 days (Mean 5.00). The dehiscence which was longitudinal, usually commenced from the base of the spike and progressed towards the tip. A nonchronological dehiscence was observed occasionally.

9. The mean number of flowers per spike in Panniyur-1 was 107.74, with 97.18 per cent hermaphrodite and 2.82 per cent pistillate flowers. In Karimunda, the spike had a mean of 46.23 flowers, which were all hermaphrodite.

10. Anthesis started between 18.00 and 18.30 hours and continued upto 02.30 hours of the next day. The peak was recorded between 18.30 and 24.30 hours.

11. Anther dehiscence which started from 14.30 hours continued upto 16.30 hours, with the maximum between 14.30 and 15.30 hours.

12. The flowers were found to be receptive from the day of anthesis upto seven days after anthesis with the maximum set (77.25 to 85.00%) for the first three days. Receptivity decreased thereafter to 11.40 per cent on the 7th day. The flowers were not receptive during the pre-anthesis period.

13. Pollen was spherical with a mean diameter
of 9.86 p. Stain test recorded 98.04 per cent fertility. Pollen production per flower (7000 to 13000)
showed significant variation among the standards.

14. Sucrose, boric acid and calcium nitrate were found to influence pollen germination at specific concentrations. Without boric acid and calcium nitrate, four per cent sucross gave maximum germination (87.29%). A germination of 94.68 per cent was recorded in four per cent sucrose with 20 ppm boric acid. With calcium nitrate, maximum germination (89.97%) was obtained at 20 ppm in four per cent sucrose. However, a combination of the three was found to be the bost, wherein maximum germination of 95.77 per cent was recorded in the media containing four per cent sucrose, 20 ppm boric acid and 10 ppm calcium nitrate.

15. The pollen was found to be viable for 7 to 4

days in water. Maximum viability in acetocarmine (85.88%) was recorded on the first day after collection when stored in distilled water at 4°C. The viability decreased to 12.10 per cent on the fourth day.

16. Significantly higher percentage of set (83.42%) was obtained under open conditions than in bagged conditions (67.35%). The result favoured the view that rain water played a major role in the natural pollination of pepper.

17. Under open pollination, a higher set (83.44%) was possible than under hand pollination (80.00%). No set was recorded in the absence of pollination.

18. Spike shedding was observed in almost all the months starting from spike emergence till harvest. Maximum (51.31%) was noticed in June and minimum in May (1.69%). Shedding was nil in September. There was significant difference in the percentage of shedding between months.

19. Shedding in the different standards accounted for a mean of 23.82 per cent of the spikes produced per standard. There was significant variation between standards for the percentage of spikes shed. 20. Spikes were shed at various stages of development, starting from emergence to harvest. It was possible to recognize eight stages. Maximum shedding (33.15%) was noticed during the first month of berry set, followed by that which occurred before anthesis (24.86%) and that during anthesis (16.57%). Minimum shed (1.66%) was observed in the fourth month of set.

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REFERENCES

REFERENCES

Adams, J. (1916). On the germination of pollen of apple and other fruit trees. <u>Bot</u>. <u>Gaz. 61</u>: 131-147.

Addicott, F.T. (1943). Pollen germination and pollen tube growth as influenced by plant growth substances. <u>Plant Physiol.</u> 18: 270-279.

- Addicott, F.T., and Lynch, R.S. (1955). Physiology of abscission. Ann. Rev. Pl. Physiol. 6: 78-82.
- Addicott, F.T., and Wiater. (1977). Hormonial control of abscission. <u>Plant growth and regulation</u>. Spring Verlag, New York: 22-40.
- Anandan, M. (1924). Observations on the habit of the pepper vine with special reference of the reproductive phase. <u>Madras Agric. Dept. Yearbook</u>: 49-69.
- Anthony and Harlan (1920). Physiology of pollen. The Bot. Rev. 27(3): 325-381.
- Aravindakshan, M. (1964). Studies on certain aspects of growth and flowering in some varieties of guava with special reference to influence of pruning on flowering and fruit set. <u>Madras agric.</u> J. 51(11): 482.
- Aslam, M., Brown, M.S., and Kohel, R.J. (1964). Evaluation of seven tetrazolium salts as vital pollen stains in cotton, <u>Gossypium hirsutum</u>. <u>Crop Sci</u>. 4(5): 508.
- Balasubramanyam, V.R. (1959). Studies on the blossom biology of guava (<u>Psidum guajava</u> L.). <u>The Indian</u> J. <u>Hort.</u> 16: 69-75.
- Bardwaj, S.N. (1975). Boll shedding in cotton. Indian J. Plant Physiol. 18(2): 9-15.

- * Barnard, C. (1932). Studies on growth and fruit bud formation. 1. A year's observation on Victoria apples. J. Dep. Agric. Victoria (Australia). 30: 349-361.
- *Barnard, C., and Reed, F.M. (1933). Studies on growth and fruit bud formation. J. Dep. Agric. Victoria (Australia). 31: 57-29.
 - Barnett, L.F.L., and Carver, R.F. (1967). Meiosis and pollen stainability in Switchgrass, <u>Panicum</u> <u>virgatum</u>. <u>Grop. Sci.</u> 7(4): 301-313.
 - Barrett, H.C., and Arisumi, T. (1952). Methods of pollen collection, emasculation and pollimation in fruit breeding. <u>Proc. Amer. Soc. Hort. Sci. 59</u>: 259-263.
 - Benson, L. (1970). <u>Plant classification</u>. Oxford and IBH Publishing Co., Bombay: 189-190.
 - Blacklock, J.S. (1954). A short history of pepper culture with special reference to Sarawak. <u>Trop. Agric</u> <u>Trin. 31</u>: 40-56.
 - Brewbaker, J.L., and Kwack, B.H. (1963). The essential role of Ca in pollen germination and pollen tube growth. <u>Aver. J. Bot. 50</u>: 859-65.
 - Brink, J. (1924). Germination of pollon grains of apple and other fruit trees. <u>Bot. Gaz. 6</u>: 131-147.
 - Brooks, J.S., and Puri, Y.P. (1963). Atmospheric conditions influencing pollon shedding in hops, <u>Humulus</u> <u>lupulus</u>. <u>Crop. Sci.</u> 3(6): 56-57.
 - Brooks, J.S., and Brooks, N.H. (1967). Pollon abortion in relation to cytoplasmic male starility in sorghum. <u>Crop. Soi.</u> 7(1): 47-49.
 - Carns (1951). Some effects of water and oxygen in absicission in vitro. <u>Plant Physiol</u>. <u>26</u>: 212-214.

Chadha, K.L., and Singh, K.K. (1963). Studies on fruit drop in Mango 1. The Indian J. Hort. 20: 17-85.

Chadha, K.L., and Singh, K.K. (1964). Fruit drop and its relation to fruit growth. <u>The Indian J. Hort.</u> 21(3 & 4): 197-201.

- Cobley, L.S., and Steele, W.M. (1976). <u>An introduction</u> to the botany of tropical crops. The ELBC and Longman, London: 235-238.
- *Coit, J.E., and Hodgson, R.W. (1916). The causes of June drop of Naval oranges. <u>Calif. Univ. J. egric.</u> 4(8-10): 27-29.
- *Colt, J.E., and Hodgeon, R.W. (1918). The June drop of Washington Naval oranges. <u>College of Agri. Expt.</u> <u>Sta. Berkley Bull</u>: 290.
- *Cooper, W.C., and Henry, W.H. (1971). Absission chemicals in relation to citrus fruit harvest. <u>J. Agric.</u> <u>Fd. Chem. 19</u>: 559-563.
- Coyne, D.P., and Al-Yasuri, S. (1964). Effect of growth regulators in delaying pod abscission and embryo abortion in the interspecific cross. Crop. Sci. 4(4): 435.
- *Cramer, P.J.S. (1907). The yielding capacity of pepper vines. <u>Teyemannia</u>. <u>18</u>: 343.
 - Damodaran, V.K., Abraham, J., and Alexander, K.H. (1966). The morphology and biology of cashew flower. <u>Ag. Res. J. Herala</u>. <u>4</u>(2): 78-85.
 - Dasarathy, T.B. (1951). The guava (<u>Psidium guajava</u>, L.). <u>Madras agric. J.</u> 38: 521-527.
 - Deshnukh, J.N., Rautt, R.S., and Manekar, A.G. (1978). Studies on pollen viability in diploid and colchicine induced tetraploid sponge gourd. <u>Sci. and Cult.</u> 44(3): 126-129.

- *De Waard, P.W.F. (1967). A method of artificial pollination in pepper cv. Kuching. Unpublished.
- De Waard, P.W.F., and Zeven, A.C. (1969). Pepper (<u>Pepper nigrum L.</u>). <u>Outlines of Perennial crop</u> <u>breeding in the tropics</u>. Veenman, Wageningen:409-423.
- *Doroshenko, A.V. (1928). Physiology of pollen. Bull. Appl. Bot. Pl. Breed. 18: 217-344.
 - Fletcher, S.W. (1906). Methods of crossing fruits. <u>Proc. Amer. Soc. Hort. Sci.</u> 40: 29-40.
 - Fogle, H.W. (1977). Identification of clones within tree fruit species by pollen exine pattern. J. Amer. Soc. Hort. Sci. 102(5): 552-560.
 - *Gollmick, F. (1942). The longivity of grape pollen. Agnew. Bot. 24: 221-232.
 - Griggs, W.H., Vansell, G.H., and Iwakiri, B.T. (1953). The storage of hand collected and bee collected pollen in a home freezer. <u>Proc. Amer. Soc. Hort.</u> <u>Sci. 62</u>: 304-305.
 - Gustafson, F.G. (1926). Growth studies on fruits. <u>Plant Physiol. 1</u>: 265-272.
 - Gustafson, F.G. (1960). Influence of Gibberellic acid on setting and development of fruits in tomato. <u>Plant Physiol. 25</u>: 116-120.
 - Halma, F.F., and Compton, C. (1936). Growth of citrus trees. <u>Proc. Amer. Soc. Hort. Sci. 34</u>: 80-83.
 - Hariano, S., and Marioko, S. (1975). Influence of fruit load on the growth and fruit yield in citrus trees. <u>J. Jap. Soc. Hort. Soi.</u> 44(2): 99-106.
 - Hariom, Srivastava, R.D., Misra, R.S., and Bana, D.S. (1975). Effect of growth substances on drop, further retention and quality of apple. <u>Prog.</u> <u>Hort.</u> 2(1): 57-60.

- Marrold, J.J. (1935). Comparative study of the developing and aborting fruits of <u>Prunus persica</u>. <u>Bot</u>. <u>Gas. 26</u>: 505-520.
- *Hasan Iljas, B. (1960). Some notes on the floral biology of black pepper (<u>Piper nigrum L.</u>). <u>Pemb. Balai</u> <u>Besar Penj. Pert. Bogor.</u> (157): 1-22.
 - Howlett. F.S. (1927). Some factors of importance in fruit setting studies with apple varieties. <u>Proc. Amer. Soc. Hort. Sci. 23</u>: 307-315.
 - Jawanda, J.S., Sinha, M.K., and Hippal, D.K. (1972). Studies on nature and periodicity of fruit drop in sweet orange. 29(3 & 4): 269-277.
 - Jose, J.S., and Magoon, M.L. (1972). Studies on pollen germination in dioscorea. <u>The Indian J. Hort.</u> 29(1): 85-93.
 - Jost, B.A. (1905). Physiology of pollen. The Bot. Rev. 27(5): 325-381.
 - King, J.R. (1962). Pollinicuration A proposed new term. South Indian Hort. 10: 164.
 - King, J.R., and Hease, C.O. (1938). Pollen longivity studies with deciduous fruits. <u>Proc. Amer. Soc.</u> <u>Hort. Sci. 36</u>: 310-313.
- *Knowlton, H.B. (1922). Studies in pollen with special reference to longivity. <u>Mem. Cornell. Agric.</u> <u>Exp. Sta. 52</u>: 751-793.
- *Koorders, S.H. (1908). Die Piperaceae Von Java. <u>Verhandelingen Kon. Akad. Wetenschappen</u>, <u>Amsterdam 2e Sectie 14(4): 1-75.</u>
- Korrigs, M.H., and Kester, D.E. (1959). Fruit setting in almond. The pattern of flower and fruit drop. <u>Proc. Amer. Soc. Hort. Soi. 24</u>: 214.

Krishnamurthi, S., Randhawa, G.S., and Nair, P.C.S. (1960). Growth studies in some citrus species under subtropical conditions. <u>The Indian J.</u> <u>Hort. 17</u>(3 & 4): 171-184.

- Krishnamurthi, S., Randhawa, G.S., and Nair, P.C.S. (1961). Growth studies in mango (<u>Mangifera indica</u>) under Delhi.conditions. <u>The Indian J. Hort.18(2)</u>: 106-118.
- Ewack, B.H. (1965). The effect of Ca on pollen germination. Proc. Amer. Soc. Hort. Sci.86: 818-823.
- Kwack, B.H., and Brewbaker, J.L. (1965). The protective action of calcium in relation to survival and growth inhibition of pollen. <u>Plant Physicl.</u> <u>28</u>(22): 32-34.
- Leuty, S.J., and Bukovae, H.J. (1968). A comparison of the growth and anatomical development of naturally abscising, non absolving and naphthalene acetic acid treated peach fruits. <u>Phytomorphology</u>. 18: 372-379.
- Lewis, D. (1946). Chemical control of fruit formation. J. Pomol. Hort. Sci. 22: 175-183.
- Lidforss, B. (1896). Zur Biologic des pollens. <u>Jahrb.</u> <u>Wiss. Bot. 29</u>: 1-38.
- Liyanage, D.V. (1949). Preliminary studies on the floral biology of the coconut palm. <u>Trop. Agric. 171</u>: 75-83.
- Maas, J.L. (1977). Pollen ultra structure of straw berry and other small fruit crops. <u>J. Amer. Soc. Hort.</u> <u>Sci. 102(5): 560-571.</u>
- Maheshwari, P. (1944). On the longivity of pollen. Indian J. Hort. 2: 82-87.

- Marinet, J. (1953). La culture du poivre dans l'Inde. Rapport de mission (14 Janvier an 7 fevrier, 1953).
- Marinet, J. (1955). 'Etude' economique et oulturale due poivre du Cambodge. <u>Agron. Trop. 10</u>: 279-360.
- Martin, O. (1913). Physiology of pollen. <u>The Bot. Rev.</u> <u>27(3): 325-387.</u>
- Martin, F.W., and Gregory, L.E. (1962). Mode of pollination and factors affecting fruit set in <u>Piper</u> <u>nigrum L. Crop Sci. 2</u>: 295-299.
- McCown, M. (1938). Abscission of flowers and fruits of the apple. Proc. Amer. Soc. Nort. Sci. 36: 320-321.
- McMunn, R.L. (1939). Growth increment in apples. <u>Proc.</u> <u>Amer. Soc. Hort. Soi. 26</u>: 379-383.
- Munser, R. (1960). Investigations on the physiology of pollen germination with special reference to the effect of boric acid. <u>Biol. Series</u>. 79: 59-84.
- Murneek, A.E. (1933). The nature of shedding immature apples. <u>Mo. Agric. Exp. Stn. Res. Bull</u>. 201.
- Nagarajan, C.R., Krishnamurthy, S., and Rao, V.N.M. (1965). Storage studies with grape pollen. South Indian Hort. <u>13</u>(1 & 2): 1 - 14.
- Naik, K.C., and Rao, N.M. (1942). Some factors governing fruit bud formation in mango (<u>Mangifera</u> <u>indica</u> L.) II. Relation between growth and flowering. <u>Madras agric. J.</u> 30(11): 365-374.
- 'Nair, F.K.K. (1965). Pollen grains in some families of Indian Monochlamydeae. <u>Bot. Notiser</u>. 118: 281-288.
- Nair, P.K.K. (1977). <u>Advances in pollen spore research</u>. Today and Tomarrows printers and publishers, New Delhi.

Nalawadi, V.G., Dasappa, and Sulikeri, G.S. (1977). Floral biology of some variaties of sapota. <u>Prog. Hort. 9(1): 17-21.</u>

Nambiar, P.K.V., and Sayeed, P.M. (1962). Progress report for the year 1951-1952. Pepper Res. Scheme, Pepper Res. Sta., Panniyur, Taliparamba.

Nambiar, P.K.V., Pillai, V.S., Sasikumaran, S., and Chandy, K.S. (1978). Pepper research at Panniyur. A resume. J. <u>Pln. Crops 6(1)</u>: 4-11.

Nauriyal, J.P. (1955). A study of premature fruit drop in citrus. <u>The Indian J. Hort. 12</u>(2): 39-52.

Nebel, B.R. (1939). Longevity of pollen in apple, pear, plum, peach, apricot and sour cherry. <u>Proc.</u> <u>Amer. Soc. Hort. Sci. 37</u>: 130-132.

Nebel, B.R., and Ruttle, M.L. (1937). Storage experiments with pollen of cultivated fruit trees. <u>J. Fomol.</u> 14: 347-359.

Nicolaissen, N. (1953). Pollen - the preservation of its viability by cold storage. <u>Industr. Refrig.</u> (Oct.): 24-27.

Nirmalendunath and Randhawa, G.S. (1959). Studies on floral biology in pomegranate, <u>Punica granatum</u>. <u>The Indian J. Hort. 16(3): 121-141.</u>

Oberle, G.D., and Geortzen, K.L. (1952). A method for evaluating pollen production of fruit varieties. <u>Proc. Amer. Soc. Hort. Sci. 59</u>: 263-266.

O'kelly (1955). Physiology of pollen. The Bot. Rev. 27(3): 325-381.

Palmer, R.L., Hield, H.Z., and Lewis, L.W. (1968). Citrus leaf and fruit abscission. <u>Proc. 1st</u> <u>Int. Citrus. Symp.</u> 3: 1135-1143.

- * Pfundt, M. (1910). Einfluse der Luflfewcht igkeit and die Lebensdauer des Elutenstaubes. <u>Jber</u>. <u>Wiss. Bot. 47</u>: 1-40.
 - *Pillai, V.S., Sasikumaran, S., and Mambiar, P.K.V. (1977). A note on preliminary observation of spike shedding in pepper. (<u>Piper nigrum L.</u>). <u>Arecanut and Spices Bulletin. 2(4): 93-94</u>;
 - Pollard, J.E., and Biggs, R.H. (1969). Relation of cellulose to abscission of citrus. <u>Plant Physiol</u>. <u>44</u>: 31-35.
 - Purseglove, J.W. (1977). <u>Propical crops dicotyledons</u>. Vol. I & II. The ELBS and Longman, London: 443.
 - Raimund, H.M. (1949). Citrus growth studies. J. Hort. Sci. 25: 50-59.
 - Randhawa, G.S. (1971). <u>Use of plant growth regulators</u> and gibberellins in Horticulture. I.C.A.R., New Delhi.
 - Randhawa, G.S., and Nair, P.K.K. (1960). Studies on floral biology of plum grown under sub tropical conditions. <u>II</u>. <u>The Indian</u>. <u>J. Hort</u>. <u>17</u>: 83-95.
 - Randhawa, G.S., and Sinha, R.B. (1965). Fruiting in relation to growth in citrus. <u>The Indian J.</u> <u>Hort. 20(1): 1-12.</u>
 - Rao, C.V. (1961). Pollen types in Epacridaceae. J. Indian bot. Soc. 40: 409-423.
 - Rao, V.N.M., and Khader, J.B.M. (1960). Pollination and pollen germination studies in some fruit crops. <u>Paper presented at the Academy of Agrl.</u> <u>Soiences, Coimbatore</u>.
 - Rao, V.N.M., and Khader, J.B.M. (1962). Estimation of pollen production in fruit crops. <u>Madras agric</u>. J. 40(5): 152-155.

- Ravindran, P.N. (1977). In vitro germination and growth of cocca pollen. J. Pln. Crops. 5(2): 109-111.
- Reed, H.S. (1929). Correlation between growth and fruit production of apricot. <u>Proc. Amer. Soc.</u> <u>Hort. Sci. 25</u>: 247-249.
- *Remy, P. (1953). A contribution to the study of pollen of stone fruit trees. <u>Ann. Amol.plantos</u>. <u>J</u>: 351-388.
- Rendle, A.B. (1971). <u>The classification of flowering</u> <u>plants</u>. <u>II</u>. Cambridge AI the University Press: 89-91.
- *Resnik, M.E. (1956). Effect of growth substances on the germination of citrus pollen. <u>Bull. Res.</u> <u>Conac. Israel Bot. 5</u>: 223-224.
 - *Rogers, B.J. (1971). Scanning electron microscopy of abscission zone surfaces of Valencia orange fruit. <u>Planta 97</u>: 358.
 - Sachan, B.P., and Patro, K.D. (1970). Studies on pollen storage of some citrus species. <u>Prog. Hort</u>. 1(4): 13-16.
- *Satjan, I.M., and Kleeva, R.F. (1964). The viability of pollen of some fruit crops (Russian). <u>Bjull</u>. <u>cloro. Bot. Sada</u>. (55): 38-41.
- * Saur, H.R. (1951). Growth of orange shoots. <u>Aust. J.</u> of <u>Agric. Res. 2</u>: 105-117.
 - *Schumucker (1935). Uber dem Einfluss von Borsaure any Pflanson insbesondere Kcimole pollen Korner. <u>Planta</u>. 23: 264-283.
- *Sedov, E.N. (1955). The viability of the apple pollen as affected by methods after ripening and storage. <u>Agrobiologia</u>. 3: 134-136.
 - Shanmugavelu, K.G., and Rao, V.N.M. (1977). Spices and plantation crops. Popular Book Depot, Madras.

- Sharma, D.K., and Singh, R.N. (1970). Studies in some pollination problems in mango. <u>The Indian J.</u> <u>Hort. 27</u>: 1-15.
- Shukle, K.S., and Misra, R.N. (1975). Studies on the effectiveness of stored pollen and fresh pollen grains in the fruit set of Kagzi lime <u>C. aurantifolia</u>. <u>Prog. Hort. 7(3): 77-80.</u>
- Shukla, P., and Misra, P.S. (1979). <u>An introduction to</u> <u>taxonomy of anglosperus</u>. Vikas Publishing House Pvt. Ltd., New Delhi. 392-394.
- Singh, S.N. (1961). Studies on the morphology and viability of the pollen grains of mango. <u>Hort. Adv.</u> <u>V</u>: 121-124.
- Singh, S.N. (1962). Storage of litchi pollen. Hort. Adv. VI: 78-80.
- Singh, S.N. (1962a). Studies on the pollen storage of mango. <u>The Indian J. Hort</u>. 19(3 & 4): 92-103.
- Singh, R.N., Randhawa, G.S., and Sharma, D.K. (1961). Pollen storage and pollen germination in fruit crops. A review. <u>The Indian J. Hort.</u> 18(1):
- Singh, V.R. (1960). Studies on the fruit drop of mango. (<u>Mangifera indica</u>). II. Nature and extent of fruit drop. <u>Hort. Adv. IV</u>: 143.
- Singh, V.R. (1961). Studies on the fruit drop of mango. IV. Embryo development, its degeneration and studies of fruit pedicel and abscission zone. <u>Hort. Adv. V</u>: 218-224.
- Singh, J.P., and Ghose, S.F. (1965). Studies on the growth behaviour of different citrus. spp. and cultivars in nursery. <u>The Indian J. Hort. 22</u>(3): 266-277.
- Singh, L.B. (1965). Mango in India. Problems and achievements. <u>Advances in agricultural sciences</u> and their applications. Agricultural College and Research Institute, Coimbatore: 312-314.

- Sinha, N.M. (1972). Follen viability and pollen storage studies in jack fruit (<u>Artocarpus heterophylla</u>). <u>Prog. Hort.</u> 4 (3 & 4): 45-53.
- Snedecor, G.W., and Cochran, W.G. (1967). <u>Statistical</u> <u>methods</u>. Oxford and IBH Publishing Co., New Delhi.
- Soost, R.K., and Cameron, J.W. (1954). Production of hybrids by the use of stored trifoliata orange pollen. Proc. Amer. Soc. Hort. Soi. 63: 234-238.
- Spencer and Kennard, W.L. (1955). Studies on mango fruit set in Puerto Rico. <u>Trop. Agric. 32</u>: 323.
- Sreekumary Amma and Vijayagopal, P.D. (1977). The role of rain water in the pollination of pepper (Piper nigrum L.). <u>Agri. Res. J. Kerala. 15</u>(2): 188-189.
- Srivastava, D.N. (1938). Studies on the non setting of pears. 1. J. Pom. and Hort. Sol. 16: 39-62.
- Stanley, R.G., and Linskens, M.F. (1974). <u>Pollen biology</u>, <u>biochemistry and management</u>. Springer Verlag Bulin Hiedelburg, New York.
- Sundararajan, S. (1961). Studies on certain aspects of growth, flowering and fruiting in some variaties of sapota (<u>Achras sapota</u>). <u>Madras agric</u>. J. 51(11): 483-488.
- *Tatarincev, A.S., and Ostrowhova, N.V. (1950). Storing pollen of tree and bush fruits. <u>Saddi ogorod</u>. <u>3</u>: 23-28.
- *Tatarincev, A.S., and Ostrowhova, N.V. (1956). Prolonged pollen storage of fruit trees. <u>Priroda</u>. <u>Leningrad</u>. 2: 103-111.
- Teotia, S.S., Bhati, D.R., and Phogat, K.P.S. (1970). Use of tree vigour as a criteria for yield in mango var. Dashehri. <u>Prog. Hort. 2(1): 5-13</u>.

e.

- Thankamma Pillai, P.K., Vijayakumar, G., and Nambiar, M.C. (1978). Flowering behaviour, cytology and pollen germination in ginger (Zingiber officinale). J. Pln. Crops. 5(1): 12-13.
- Thompson, A.H., and Batjer, L.P.(1950). The effect of boron in the germinating medium on pollon germination and tube growth for several deciduous tree fruits. <u>Proc. Amer. Soc. Hort. Sci. 56</u>: 227-305.
- Traub, H.P., and O'Rork, C.T. Jr. (1936). Papaya pollen germination and storage. <u>Proc. Amer. Soc.</u> Hort. <u>Sci. 34</u>: 18.
- Varma, S.K. (1976). Role of GA in the phenomenon of abscission in flower buds and bolls of cotton. <u>Indian J. Plant. Physicl. XIX(1): 12-16.</u>
- Vasil, J.K. (1958). Storage experiments with pollen of cultivated fruit trees and vegetables. <u>Sci. Cult.</u> 24(5): 253-236.
- *Visser, T. (1955). Germination and storage of pollen. Meded. Land Hogersch. Mageningen. 55(1): 1-68.
- *Vyvyan, M.C. (1946). Fruit fall and its control by synthetic growth substances. <u>Tech. Comm.</u>(18). <u>Imp. Bur. Hort. & Plentation Crops.</u> 62-70.
- *Wilson, W.C. (1969). Four years of abscission studies on oranges. <u>Proc. Flo. St. Hort. Soc. 82</u>: 75-81.
 - Wodehouse, R.P. (1935). Pollen grains. Mc. Graw Hill Book Co., New York.
 - *Yamagutchi, S. (1954). Some inter-relations of oxygen, carbondioxide, sucrose and ethylene in abscission. <u>Doctorate Thesis</u>. Univ. Calif.
- *Zirkle, C. (1937). Acetocarmine mounting media. Solence

*Originals not seen

APPENDICES

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APPENDIX - I

Weather data (monthly average) for the period from January 1979 to January 1980.

Nonth	Tempe- rature	Rola- tive humidity %	Total rain- fall mm	No. of rainy days
January	26.35	67.0	Nil	N11
February	28.20	66.5	22.0	4
March	29.50	67.0	3.2	1
Apr i 1	30 . 70	64.0	46.5	- 4
May	28.75	74.5	155.1	10
June	28 .55	75.0	722.7	22
July	26.05	83.0	729.8	28
August	26.50	81.0	462.6	19
September	27.70	82.5	208.7	18
October	27.70	7 0.0	127.3	16
November	27.55	78 .5	317.4	1 8
December	25,80	70.0	N11	Hil
1980				,
Jennery	25 .70	61.5	N1 1	N11

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Source: 'B' class Observatory, Mannuthy.

APPENDIX - II

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Source	âf	Mean squares		
		Mean growth	Percentage of shoots showing growth	
Total	103			
Weoks	12	1.29	3406.95	
Standards	7	0.55	1972.42	
Error	84	0.04	97.33	
	·		-	
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Analysis of variance for weekly growth of different standards

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APPENDIX - III

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Analysis of variance for spiking of pepper standards

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Source	d£ .	Percentage of shoots spiked	Number of spikes per shoot	
Total	23			
Months	3	1311.98	4.47	
Standards	5	78.57	0.31	
Error	15	13.74	0.14	

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APPENDIX - IV

Source	đf	Mean squares
Total	29	
Standards	2	10278500.00
Error	2 7	1734750.00

Analysis of variance for pollen number per flower

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APPENDIX - V

Analysis of variance for pollen germination in sucrose agar medium

Source	dſ	Mean squares
Totel	17	
Sucrose	5	1465.18
Error	12	0.73

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APPENDIX - VI

Analysis of variance for pollen germination in agar medium with varied concentrations of sucrose and boric acid

dſ	Mean squares
89	
4	643.01
5	8894.42
20	33-93
60	1.08
	89 4 5 20

APPENDIX - VII

Analysis of variance for pollen germination in agar medium with varied concentrations of sucrose and calcium nitrate

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Source	df	Mean squares
Total	89	
Sucrose (S)	4	1830.79
Calcium nitrate (C)	5	8233.94
SxC	20	91.69
Error	60	1.56

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APPENDIX - VIII

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Analysis of variance for pollen germination in four per cent sucrose - agar medium with varied concentrations of boric acid and calcium nitrate

df	Nean squares
74	
4	635.87
4	1286.43
16	18.94
50	2.11
	74 4 4 16

GROWTH, FLOWERING, FLORAL BIOLOGY AND SPIKE SHEDDING IN PEPPER (Diper nigrum L.)

BY REMA MENON

ABSTRACT OF A THESIS Submitted in partial fulfilment of the requirements for the degree of

Master of Science in Horticulture

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Department of Horticulture (Plantation Crops) COLLEGE OF HORTICULTURE Vellanikkara :: Trichur

APPENDIX - IX

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Analysis of variance for monthly shedding of spikes

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Source	df	Mean squares
Total	111	
Nonths	6	3958.17
Standards	15	22.23
Error	90	86.72
• • • • • • • • • • • • • • • • • • •		۲۵ (۱۹۰۰ - ۲۰۰۹) «۲۰۰۹ - ۲۰۰۹ - ۲۰۰۹ - ۲۰۰۹ - ۲۰۰۹ - ۲۰۰۹ - ۲۰۰۹ - ۲۰۰۹ - ۲۰۰۹ - ۲۰۰۹ - ۲۰۰۹ - ۲۰۰۹ - ۲۰۰۹ - ۲

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ABSTRACT

The present studies were carried out in the College of Horticulture, Kerala Agricultural University, during the year 1979-80. The object was to study the pattern of growth and flowering, floral biology and epike shedding in pepper (<u>Piper nigrum L.</u>). The studies were conducted on four year old pepper vines (c.v. Panniyur-1 and Karimunda) receiving uniform cultural and manurial practices as per the package of practices of the Kerala Agricultural University.

Shoot growth in pepper was observed during four months of the year, from May to August, with maximum in July and minimum in August. The mean growth and the percentage of shoots that showed growth varied significantly from month to month and standard to standard. The flowering followed a pattern similar to growth. The percentage of aborted spikes ranged from 4.011 to 26.31 per cent.

The spike development followed a linear pattern. The spike attained maximum length in 31.67 and 29.26 days in Panniyur-1 and Karimunda, respectively. The period of anthesis (from the first to the last flower in a spike) was for 6 to 10 days (Mean 8.12) in Panniyur-1 and 4 to 11 days (Mean 9.21) in Karimunda. Anther dehiscence extended for a period of 4 to 9 (Mean 7.72) and 3 to 8 days (Mean 5.00). The protogynic stage existed for 5 to 8 and 3 to 7 days (Mean 6.52 and 5.27) in the two cultivars.

In Panniyur-1, the spike had hermaphrodite (97.18%) and pistillate flowers (2.82%), while in Karimunda all the flowers were hermaphrodite (100%).

Anthesis started between 18.00 and 18.30 hours and attained a peak between 18.30 and 24.30 hours. Anther dehiscence which commenced from 14.30 hours was maximum between 14.30 and 15.30 hours. Stignatic receptivity lasted for seven days after anthesis, with the maximum for the first three days. Rainwater was found to be a major factor in pollination.

Pollen grains had a mean diameter of 9.86 . Pollen production per flower (7000 to 13000) varied from standard to standard. Sucrose at concentrations of 2, 4, 5, 8 and 10 per cent, boric acid and calcium nitrate at concentrations of 10, 20, 30 and 40 ppm each promoted pollen germination. A combination of the three (4% sucrose, 20 ppm boric acid and 10 ppm calcium nitrate) gave maximum germination (95.77%). Pollen remained viable for 1 to 3 days in water under the different treatments. Spike shedding which occurred during almost all the months from spike emergence to harvest, was maximum in June and minimum in May. No shedding was observed in September. Eight stages of shedding could be observed, with the maximum shed during the first month of set. The percentage of shedding varied significantly between months and between standards.