DEVELOPMENT AND QUALITY EVALUATION OF MICROENCAPSULATED BANANA PSEUDOSTEM JUICE POWDER

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by

SARANYA S (2014-18-110)

THESIS

Submitted in partial fulfilment of the requirement for the degree of MASTER OF TECHNOLOGY

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AGRICULTURAL ENGINEERING (Agricultural Processing and Food Engineering) Faculty of Agricultural Engineering & Technology Kerala Agricultural University



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DECLARATION

I hereby declare that this thesis entitled "Development and quality evaluation of microencapsulated banana pseudostem juice powder" is a *bonafide* record of research work done by me during the course of research and the thesis has not previously formed the basis for the award of any degree, diploma, associateship, fellowship or other similar title of any other University or Society.

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Dedicated to Beloved parents I My Profession

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SYMBOLS AND ABBREVIATIONS

| et al. | : | and others |
|--------|---|-----------------------------------|
| % | : | per cent |
| & | : | and |
| / | : | per |
| < | : | less than |
| > | • | greater than |
| ± | : | Plus or minus sign |
| o | : | degree |
| °B | : | Degree brix |
| °C | : | degree celsius |
| a* | : | Greenness or redness |
| Al | : | Aluminium |
| ALPE | : | Aluminium laminated polyethylene |
| AOAC | : | Association of analytical chemist |
| b* | : | Blueness or yellowness |
| C.V. | : | Coefficient of variation |
| Ca | : | Calcium |
| CRD | : | Completely randomized design |
| Cu | : | Copper |
| df | : | Degree of freedom |
| etc. | : | etcetera |
| F | : | F value |

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| Fe | : Iron |
|--------------------|---|
| Fig. | : Figure |
| g | : gram |
| g/ml | : Gram per mililiters |
| h | : Hour |
| H_2SO_4 | : Sulphuric acid |
| HCL | : Hydrochloric acid |
| HDPE | : High density polyethylene |
| HG | : Horse gram extract |
| К | : Potassium |
| КСАЕТ | : Kelappaji College of Agricultural Engineering and Technology |
| kg | : kilogram |
| Kg/cm ² | : Kilogram per square centimetre |
| L* | : Lightness or darkness |
| LBD | : Loose bulk density |
| LDPE | : Low density polyethylene |
| MD | : Maltodextrin |
| mg | : milli gram |
| Mg | : magnesium |
| min | : minute |
| ML | : milk |
| ml | : milliliter |
| Mn | : Manganese |
| Na | : Sodium |

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| Ni | : | Nickel |
|------|---|------------------------------|
| No. | : | Number |
| Р | : | Pseudostem juice |
| р | : | probability |
| pН | : | percentage of H+ ions |
| РР | : | Poly propylene |
| rpm | : | revolution per minute |
| S | : | second |
| S | : | Significant |
| SEM | : | Scanning electron microscopy |
| Si | : | Silicon |
| SI. | : | Serial |
| SD | : | Standard deviation |
| Т | : | treatment |
| TBD | : | Tapped bulk density |
| viz | • | namely |
| w.b. | : | wet basis |
| w/w | : | weight by weight |
| WAI | : | Water absorption index |
| WSI | • | Water solubility index |
| Zn | : | Zinc |
| μm | : | micro metre |

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Introduction

CHAPTER I

INTRODUCTION

Banana (Musa sp.) commonly known by the name plantain is a large perennial herb with leaf sheaths that form trunk like pseudostem. Different parts of the plantain tree have several medicinal uses. Due to the enhanced food value and versatile medicinal value, it is one of the important low cost food crops in India. World production of banana is estimated at 48.9 million tones out of which 10.4 million tonnes, is contributed by India. Because of higher return as compared to other crops, the area under banana is increasing steadily. After harvest of the fruit, the pseudostems are cut and deserted in the fields to become organic waste because it cannot be used for the second time. The huge quantity of biomass thus produced mostly in the form of pseudostem is an absolute waste. For each ton of bananas harvested, around four tons of wastes are generated, among which 75% consists of banana plant pseudostem. About 60-65% pseudostem is produced from a whole plantain tree (NAIP, 2011). Disposal of these huge wastes creates environmental pollution and also the farmers need to spend more money for its disposal. Even though this pseudostem having high medicinal values like anti-urolithiatic activities, value- added products from banana pseudostem is mostly restricted to non food products like fiber extraction and paper products only. Also, it takes extreme amount of energy and time to be processed into biogas, pulp and paper. High perishability and its immediate browning might be the reason for its restricted application in food industry. In order to add value to banana plantation and exploitation of waste produced, the pseudostem could be processed into products (NAIP, 2011).

The selection of proper processing technique plays a significant role in better utilization of raw materials. The major problem faced during pseudostem processing is its browning (Priya *et al.*, 2014) and preservation of its nutritional value along with prolonged shelf life. Thermal processing is found to be the best method to inactivate enzyme activity and further browning of pseudostem. Among thermal processing

technique, steam blanching technique is best applicable for pseudostem processing. Steam blanching of pseudostem arrest enzyme activity and further browning. Innovative technology like microencapsulation can improve the retention time of the nutrient in pseudostem. It offers protection of sensitive food components against nutritional loss and also to preserve flavors by coating tiny droplets with a suitable wall material. Key function of microencapsulation is the controlled release of sustenance fixings at correct time at ideal spot. Spray drying is the most widely recognized and monetarily feasible procedure to deliver microencapsulated nourishment materials economically. Compared to other micro encapsulation techniques, it is promptly accessible and production expenses are lower than most other methods. The process has usually demonstrated financial viability as well as productivities. During this drying process, the evaporation of solvent, that is most frequently water, is quick and the entrapment of the interest compound occurs quasiinstantaneously. Microencapsulation by spray drying process should preferably be considered as a workmanship than a science in view of the numerous variables to streamline and the many sided quality of the heat and mass exchange wonders that happen during the microcapsule development (Gharsallaoui et al., 2007).

Development of a process technology for pseudostem powder from the blanched pseudostem juice itself will reduce browning, storage and handling problems and make its use in convenient households. The development of thermal processing, micro encapsulation technologies like blanching and spray drying respectively, facilitates the exploitation of pseudostem, the waste created after harvest of banana. This will add value to the banana plantation and provide an additional income to farmers. Microencapsulating the healthy banana pseudostem juice with suitable wall material can be suggested as a healthy ready to drink mix.

Consumer's concerns about diet and health have changed a lot. Nowadays people are moving towards healthy life style with diet including nutrient rich fortified foods. Incorporating additional ingredients like horse gram extract and milk during microencapsulation of pseudostem will improve the nutritional and medicinal value of final product.

Horse gram (*Macrotyloma uniflorum Lam.*) is one of the lesser known legumes also known as Kulthi (Mishra and Pathan, 2011). Horse gram is an underutilized legume, famous for its medicinal uses like treatment of asthma, bronchitis, tumor urinary discharge and treatment of kidney stones etc (Ghani, 2003). It also cures menstrual disorder, leucorrhoea, indigestion and breathing problems.

Milk's primary offering point is calcium, and milk-drinking is touted for building solid bones in children and avoiding osteoporosis in more adults. Youngsters require a lot of milk to manufacture bones and teeth, and will supplies vital fat and proteins and will also provides nourishment for grown-ups with crucial vitamins and minerals (Henriksen, 2009).

Blending pseudostem juice with horse gram extract and milk will enrich the resulting blend with vitamins, proteins and minerals. Adding suitable natural flavorings like ginger extract and cardamom will give better sensory qualities to the product and boost the consumer acceptance.

Considering the above cited facts, a study was undertaken to develop a process protocol for banana pseudostem powder with the following objectives:

- 1) Development of process protocol for microencapsulated banana pseudostem juice powder
- 2) Standardisation of spray drying parameters for the development of banana pseudostem juice powder
- 3) Quality analysis and storage studies of banana pseudostem juice powder

Review of Literature

CHAPTER II

REVIEW OF LITERATURE

In present investigation, an attempt has been made to develop microencapsulated banana pseudostem juice powder by spray drying followed by its quality analysis and shelf life studies. This chapter deals with a brief account of literature, which has direct and indirect bearing on particular objectives of the investigation.

2.1 Banana pseudostem

Pseudostem is a part of plantain which is an under utilised vegetative component disposed, most often as a waste. Its massive accumulation leads to unhygienic surroundings and pollution. However, its curative properties are high. Few researchers have explored its medicinal properties and approached to utilise it (Ravi *et al.*, 2011).

2.1.1 Medicinal uses

Kailash and Varalakshmi (1993) uncovered an investigation on antiurolithiatic action of banana stem juice and its effect in albino rats. The study revealed that alcoholic extricate produced from banana stem juice lessened oxalate, calcium and phosphate in rat's urine. Furthermore, it increased urine volume thereby diminishing the tendency to crystallize.

Feriotti and Igutti (2008) quantified pseudostem sap proximate composition *viz.* total solid- 0.308%, protein- 0.0141%, lipid- 0.005%, total sugar - 0.191%, ash - 0.104% and mineral composition of sap (*viz.* Sodium- 88mg/l, Potassium - 874 mg/l, Calcium - 130 mg/l, Magnesium - 116 mg/l). Investigations were also performed to estimate the pesticides connected with banana plantations to check its deposits in debasing sap. It was confirmed that out of the thirty-four pesticides examined, none exceeded the cutoff limit expressed by Brazilian direction.

Prasobh and Revikumar (2011) carried out Ultra Violet analysis (UV), chemical test and *in vitro* studies to find the presence of potassium nitrate and magnesium nitrate in pseudostem extract and proved that the juice extract possess an amazing capacity to dissolve kidney stones.

Saravanan and Aradya (2011) analysed the antioxidant properties of banana stem extract. Proto catechize acid, gentisic acid, cinnamic acid catechin, caffeic acid and Ferulic acid, *etc.* which comes under phenolic acids were present in banana stem extract. These acids inhibit the initiation of oxidative chain reactions. The antioxidant properties of polyphenols reduced the risk of cancer and cardiovascular diseases and may prevent or repair cell damage caused by reactive oxygen species.

Sampath *et al.* (2012) studied the importance and medicinal value of banana plant and its traditional use as an ayurvedic medicine. This study revealed that pseudostem juice can improve the functional efficiency of kidney and liver. It helps in dissolution of calcium oxalate, a cause for the formation of kidney stones. Diuretic character of pseudostem juice assisted in detoxification of the body and the presence of vitamin B_6 promoted in haemoglobin and insulin production.

Gopinathan (2013) conducted studies on anti-ulcer activity of aloe vera juice, banana stem and flower juice combinations in rats. It evidenced that banana stem juice inhibited the alcohol induced ulcer congestion, haemorrhage, necrosis and reduced gastric ailments in rats.

Spray-dried banana stem juice powder was estimated by Ponnambalam and Sellapan (2014). The juice powder was examined using inductively coupled plasma mass spectrometry (ICP –MS). Various elements necessary to human health (*viz.* Al, Ca, Cu, K, Fe, Mg, Mn, Ni, P, Si and Zn) were identified which varied from 375.445 µg/l to 901.382 mg/l. Concentration of three elements (K-901.382 mg/l, Mg-18.634 mg/l, and Na-3.82 mg/l) were found to be high in samples that aided in diuretic and urolithiatic activity.

2.1.2 Value added products from pseudostem

For better utilisation of banana pseudostem, a project was certified by National Agricultural Innovation Project (NAIP). Various value added products *viz.*, fibre, fabrics, yarn, vermi compost, liquid fertilizer, papers, pickles and candy were developed from banana pseudostem (NAIP, 2011).

Bornare and Sumaiya (2015) developed a Ready-To-Serve (RTS) healthy beverage by blending equal ratio of pseudostem juice and papaya pulp. It showed a good score during sensory analysis.

Present study on development and quality evaluation of banana pseudostem juice powder includes fortification of banana pseudostem by incorporating diverse healthy components like horse gram and milk. Natural flavourings like ginger extract and cardamom powder were used for enhancing oraganoleptic properties.

2.2 Horse gram (Macrotyloma uniflorum Lam.)

Horse gram (*Macrotyloma uniflorum Lam. or Dolichos biflorus Linn.*) is a popular pulse rich in protein, iron, calcium and phenols mostly used for the treatment of diseases like heart diseases, bronchitis, asthma, urinary discharges and kidney stones. Apart from this, its inherent property to deal with obesity was significant (Mehra and Upadhaya, 2013).

Bhuvaneshwari *et al.* (2014 a) studied the anti-obesity activity of *Dolichos biflorus* in Albino rats. During their experiment, rats were grouped into four groups with three male and female rats in each group. Hot and cold extracts were fed with different proportions to each group. The rats fed with a dose of 5 mg/kg showed best result. The result was best prominent in males than females.

Bhuvaneshwari *et al.* (2014 b) conducted studies on the anti-obesity activity of horse gram in a group of human volunteers. Males and females were grouped with 10 members in each group. Horse gram extracts were given for each one. Reduction in

body weight was noted. A significant response of horse gram was observed in males than females.

Verma *et al.* (2014) developed food products from horse gram bovine milk blends. Horse gram extract were taken for blending with bovine milk in different ratios (40:60, 50:50 and 60:40). The results revealed that horse gram extract contained 91.99 g moisture, 3.64 g protein, 0.25 g fat, 0.80 g crude fiber, 0.13 g ash and 3.19 g carbohydrates per 100 ml. All the variants of developed products were acceptable in sensory evaluation conducted at 9-point hedonic scale.

2.3 Milk

As proposed in Charaka samhitha, benefits of cow milk has innumerable benefits. It nourishes the body tissues, helps in rebuilding tissues, increases life expectancy, improves intelligence, strength, assists in easy peristalsis and relieves tiredness *etc*. Diverse products were already developed by incorporating milk with other components. Enrichment of components with milk will give additional health benefits and increased nutritional value of products.

Laxminarayana *et al.* (1997) described the method of preparation of milk shake powder from cow milk blended with *Musa cavendish* variety of banana (5:1). Banana pieces were heated with water and carboxy methyl cellulose was added. The mix was spray dried and ground sugar was blended with dried mix to obtain final sugar content of 42.5% in banana milk shake powder.

Rehman *et al.* (2007) developed ready to drink soy-cow milk blend. The astringency of soy milk was reduced by blending skim milk powder and it also improved the fat and sugar content of final product.

Takami *et al.* (2014) developed low lactose flavoured milk powder which promoted increased milk consumption and adequate calcium intake. It could be suggested as a nutritive alternative for lactose intolerant people to maintain their milk-drinking habits.

2.4 Flavourings

Flavourings are added to foods to impart taste and aroma. There are certain natural flavours which are derived from herbs and spices. These natural food flavors are not included in the definition of flavorings for regulatory purposes. The ginger and cardamom were the selected flavours for the pseudostem juice powder.

2.4.1 Ginger extract

Zingiber officinale Ros. commonly known as ginger is a spice with wide range of medicinal applications. From earlier times, it was used for most of the digestive disorders. Apart from this, it has anti-inflammatory and chemo-protective effects (Malhotra and Singh, 2003).

Iheagwara (2013) studied the stability and sensorial quality of mackerel fish by addition of ginger extract and its effect on it. Peroxide value, mould count *etc.*, were analysed for over 20 days storage and noted that samples with ginger extract showed low mould count and samples with 5% ginger extract showed higher acceptance than control without ginger extract.

2.4.2 Cardamom powder.

Cardamom is an ancient spice known as the "Queen of spices". Sharma *et al.* (2011) had reported its uses as anti-venom drug, lower blood pressure, analgesic activity, anti-oxidant activity *etc.*

Agaoglu *et al.* (2005) conducted studies on anti-microbial effect of cardamom seed. Seed extracts were tested for bacteria and fungus and determined inhibitory activity on some of the strains and it was least in case of *Escherichia coli*.

A major problem associated with the pseudostem is its perishability. Immediate browning reactions lead to its reduced shelf life. Pretreatment of pseudostem juice is essential prior to product development.

2.5 Pre-treatments of banana pseudo stem

Priya *et al.* (2014) studied the quick browning reaction of banana central core. Various chemicals like citric acid, potassium meta-bi-sulfate (KMS) and ascorbic acid (AA) with varying combinations (0.05%, 0.1% and 0.2%) were used for standardising the pre-treatment method based on colour, pH and microbial load. Combination of (0.1% KMS + 0.1% AA) gave good result under 3 days in ambient and 5 days in refrigerated condition.

Nithya *et al.* (2014) developed banana pseudostem powder by means of spray drying method. Prior to drying fresh pseudostem slices were treated with 0.3% citric acid solution and steam blanched at 100°C for 30 seconds and one minute respectively. Samples blanched at one minute interval were standardised based on peroxidase test.

2.6 Microencapsulation by spray drying

Microencapsulation in food refers to protecting sensitive food components against nutritional loss and preserves its flavor by coating tiny droplets with a suitable wall material. Key function of microencapsulation is the controlled release of food ingredients at right time at right place. This will improve the application and effectiveness of food ingredient delivery as well as resolve micronutrient deficiencies in the diet (Claude and Fustier, 2007).

Studies were conducted by Inglett *et al.* (1987) on encapsulation of orange oil using oligosaccharides prepared by the action of alpha-amylase on ordinary corn, waxy corn, amylo-maize, corn, rice, cassava, wheat and potato starches. The study supported the hypothesis that presence of high maltodextrins and syrup solids permit the formation of encapsulated products with excellent oxidative stability. Amylo-maize and potato starches exhibited poorer stabilities while normal corn, waxy corn, cassava, rice and wheat glucose syrup solids yielded the best and approximately equivalent shelf-lives.

Dolinsky *et al.* (2000) conducted studies on fruit drying and suggested that 30 - 55% maltodextrin should be added to the fruit and vegetable juices in order to obtain the microencapsulated fruit juice powder. Adhikari *et al.* (2003) also had reported that the addition of maltodextrins significantly reduced the stickiness of fructose solutions, proving it to be an effective drying aid.

Liu *et al.* (2007) investigated the encapsulated *Agaricus bisporus* flavor. He illustrated that depending on the properties of core material to be encapsulated and the purpose of micro-encapsulation, encapsulant materials are generally selected from a range of proteins, carbohydrates, lipids and waxes which may be used alone or in combination. The materials chosen as encapsulants must be typically film forming, pliable, odorless, tasteless and non-hygroscopic.

Kausadikar *et al.* (2015) microencapsulated lemon oil by spray drying to inspect its application on flour tea. Wall materials like maltodextrin, gum arabic, modified starches and its blends were used for drying process. Results suggested that maximum encapsulation efficiency was observed with wall material concentration (30%), core material concentration (10%) and the inlet temperature (175° C). Encapsulation efficiency found to be maximum for gum arabic. Blend of gum arabic to maltodextrin in the ratio of 50:50 gave the best combination for maximum encapsulation efficiency (82.60%) than other blends and found to be stable for 6 months in ice tea premix containing 1.5% encapsulated lemon oil.

2.7 Optimisation and standardisation of spray drying parameters

Spray drying system has been generally utilised for drying heat-sensitive foods, pharmaceuticals and substances, on account of the rapid evaporation of solvent from the droplets. Although it is considered as a dehydration process, which can also be used as an encapsulation method when it entraps 'active' material within a protective matrix. Unlike other microencapsulation methods, it offers an alluring point of preference in delivering microcapsules in simple continuous processes (Shahidi and Han, 1993).

Mainly microencapsulation by spray - drying involved four stages: preparation of dispersion or emulsion; homogenization of dispersion; atomisation of feed emulsion; and dehydration of atomised particles (Shahidi and Han, 1993). The main factors in spray-drying that must be optimised were feed temperature, air inlet temperature, and air outlet temperature (Liu *et al.*, 2007).

Chegini and Ghobadian, (2007) conducted studies on spray drying of orange juice at 65% concentration. Investigations were done on parameters like wall material, inlet and outlet temperature, feed flow rate and sticky point temperature. The experimental data showed that product yield and wall deposition are influenced by inlet temperature and feed flow rate. Optimum conditions for drying of orange juice feed flow rate of 15 ml /min , inlet air temperature of 130°C and outlet air temperature of 85°C and sticky point temperature was determined as 44°C for powder containing 2% moisture.

Fazaeli *et al.* (2012) studied the effect of processing conditions and carrier concentration for improving drying yield and other quality attributes of spray dried mulberry (*Morus nigra*) juice. Independent variables such as, inlet air temperature (110, 130 and 150°C), compressed air flow rate (400, 600 and 800 l/h), and maltodextrin concentration (8, 12 and 16%) were monitored. The optimal conditions for drying yield and total anthocyanin content corresponded to temperature (130°C), maltodextrin concentration (8%) and compressed air flow rate of 800 l/h.

Koc *et al.* (2012) studied the optimisation of spray drying conditions for producing yoghurt powder. At each condition, yoghurt powder was subjected to the measurement of physical attributes and the values of tapped and un tapped bulk densities of yogurt powder were 746 and 538 kg.m⁻³ respectively.

Sabhadinde (2014) conducted a study on orange juice powder with the aid of pilot plant spray drier with concurrent air flow. Five concentrations of maltodextrin (10, 12.5, 15, 17.5 and 20%) as wall material and inlet/outlet temperatures of 200°C and 120°C were selected for the study. The result demonstrated as physicochemical properties of powders produced depended on some process variables such as characteristics of liquid feed and drying air, as well that type of atomiser and carrier agents used.

Avila *et al.* (2015) evaluated the effect of maltodextrin (MD) and spray drying process conditions on sugarcane juice powder. Four factors such as maltodextrin, inlet air temperature, outlet air temperature and atomisation speed were optimised by a central composite design with a response surface analysis *i.e.*, MD- 20%, inlet air temperature- 130°C, Outlet air temperature-75°C, atomisation speed- 22,000 rpm.

2.8 Physico-chemical properties of powder

Gaoula and Adamopoulos (2010) conducted experiments on tomato soup powder and its moisture content under spray drying conditions and observed that increasing compressed airflow rate decreases the moisture content. That means moisture content of the powder decreased with inlet air temperature and decreased with air flow rate.

Influence of process conditions on properties of microencapsulated rosemary essential oil by spray drying was evaluated by employing a central composite rotatable experimental design. By the analysis, it was noted that bulk density increased with wall material concentration and decreased with inlet air temperature but the particle density decreased with both wall material concentration and the inlet air temperature variable. Moderate wall material concentration (24%), moderate feed flow rate (0.7 l/h) and low inlet air temperature (135°C) were standardised as the best spray drying conditions (Fernandes *et al.*, 2013).

Silva *et al.* (2013) developed propolis powder using spray drying and evaluated its properties like water activity, particle size, hygroscopicity *etc.* Stable powder with particle size of 15-24 μ m obtained with low hygroscopicity.

Patil *et al.* (2014) analysed the physical and nutritional properties of spray dried guava powder and noted a decreasing trend in moisture content with increased inlet temperature. A negative influence on bulk density was observed because of the increased porosity of the powder. Higher solubility was observed at low bulk density. For obtaining least hygroscopic, powder process conditions were optimised *i.e.*, inlet temperature above 185°C.

The physicochemical properties of grape juice powder under different spray drying conditions were evaluated by Sarabandi *et al.* (2014). Reduction in moisture content, bulk density, water activity and hygroscopicity values with high inlet air temperature were reported while, solubility and wettability decreased with temperature. Overall, increasing inlet air temperature and maltodextrin concentration led to reduced stickiness and enhances powder yield.

Shishir *et al.* (2015) carried out research to assess physical attributes of spray dried pink guava powder with different concentrations of maltodextrin (MD) at several inlet temperatures to preserve unique physical characteristics of final product. The results showed that both inlet temperatures and maltodextrin concentration have a significant influence on the powder properties. Samples spray dried at 150°C inlet temperature and 15% maltodextrin concentration exhibited better properties *viz*. lower moisture content, higher yield, better bulk density, good flowability and moderate colour characteristics.

Wang *et al.* (2015) conducted studies on "Impacts of spray-drying conditions on the physicochemical properties of soy sauce powders using maltodextrin as auxiliary drying carrier". Powder properties like bulk density, cohesiveness, particle size *etc.* showed a significant effect on spray drying conditions.

Zhou *et al.* (2004) conducted studies on spray drying of fermented liquor and analysed the effect of spray drying parameters on the developed product by establishing a mathematical model. The results were validated and optimised spray drying conditions as liquid feed rate of 60 ml/min, atomizing air pressure of 0.1 MPa and inlet air temperature of 180°C.

Spray drying operations were optimised to preserve *Lactobacillus paracasei* SD1 in skim milk powder and the air outlet temperature was standardised as 80°C. It was safe after six month of storage at Storage at 4°C (108 cfu/g). At elevated temperature the level of survivability was found to be decreasing. The overall study revealed that the production of probiotic *L. paracase.i* SD1 in large scale could be achieved by spray drying and it would be one of the cost effective method for the same (Teanpaisan *et al.*, 2012).

Carranza *et al.* (2014) evaluated microencapsulated *Lactobacillus casei* for its quality and survival during spray drying process with different vegetable extracts like asparagus, orange or grape peel *etc.* The quality of encapsulated powder produced at an inlet temperature of 145°C was analysed after 60 days of storage study at 25°C, 10^7 cfu/g of *L. casei* were survived, which guaranteed its use to develop functional food. Powder with less moisture content (<2%wb), water activity (<0.30 a_w) and reduced stickiness were obtained by using maltodextrin as adjunct.

Koc *et al.* (2014) evaluated spray-dried maltodextrin and studied its physical properties. The study revealed that atomising air flow and inlet air temperature had more effect than feed temperature and feed flow rate on the physical properties of maltodextrin powder. The results showed that the mean diameter was between 3.503 and 6.045 μ m for maltodextrin powders.

Chegini and Taheri (2014) studied the physicochemical properties of spray dried whey powder with an aid of pilot-scale co-current spray dryer. Independent variables and response variables were inspected. The result illustrated that powder

with 15% solid had low pH than powder with 30% solid. In addition, the pH of powder was found to be lower at an inlet temperature of 180°C and higher at 145°C. Besides, the electrical conductivity was more in high acidic powder. By scanning electron microscopy analysis, the particle size of powder was in range of 11.26–18.23 μ m.

Ishiwu *et al.* (2014) studied the effect of inlet-air temperature on physico chemical and sensory properties of spray-dried soy milk using a co-current spray drier with a constant feed rate of 20.5 ml/sec at air inlet temperatures of 204°C and 240°C. Developed samples were examined for proximate composition and physico-chemical properties. Protein content of $62.05 \pm 0.23\%$, ash (1.41 $\pm 0.02\%$), fat (19.92 \pm 0.08%), lysine (5.02 \pm 0.29%) and carbohydrate content (12.85 \pm 0.01 %) were quantified. Physical properties *viz*. mean total solid (10.33 \pm 0.33%) and pack bulk density (0.57 \pm 0.00 g/ml) were computed. Lower solubility was found at 240°C with reconstitution temperature of 40°C and 60°C. The value of wettability for samples dried at 204°C was in the range of 36-22 sec and that for 240°C was 29-18 sec. By sensory evaluation, sample dried at 204°C scored high.

Morinda citrifolia L. fruit was spray dried with maltodextrin as wall material. Temperature and core to wall ratio were selected as independent variables. Optimisation of the product was done by response surface methodology (RSM) by analysing the effect of total phenolic content (TPC), 2, 2-diphenyl picryl hydrazyl (DPHH), and total flavonoid (TF). Optimised value for the core to wall material ratio was found to be 1:1.5 and inlet temperature of 95°C (Krishnaiah *et al.*, 2015).

Reddy *et al.* (2014) conducted studies on Osmanabadi goat milk and influence of spray drying conditions on developed powder. Proximate composition of powder was quantified (Moisture content-4.08%, Titrable acidity-0.14%, Protein-25.48%, Carbohydrate-36.99%, Ash -6.60% and Fat- 26.85%). Elevated temperature resulted in reduced loose and tapped bulk density. By Hausner ratio and Carr index analysis flowability was found to be possible and fair respectively.

2.9 Packaging and storage studies of spray dried products

Mary *et al.* (2007) conducted studies on the packaging and storage studies of spray dried ripe banana powder under ambient conditions and concluded that banana powder could be successfully stored under ambient conditions for one year by packing in nitrogen flushed aluminium foil laminated pouches with minimum changes in colour, flavour, texture, microbial load and organoleptic qualities.

Yu *et al.* (2013) conducted shelf life studies and storage stability of spraydried bovine colostrum powder under different storage conditions. Aluminiumlaminated polyethylene (ALPE) and polyethylene terephthalate (PET) pouches were taken as packaging material for storage study under different temperature and relative humidity. The shelf life of powder was evaluated as 425.5 and 86.5 days in ALPE and PET pouches at 25°C temperature and 50% relative humidity (RH). Storage stability of the powder in terms of colour change, moisture content, hydroxymethyl furfural, IgG concentration, and thiobarbituric acid were analysed. From the analysis, ALPE pouches were found to be the best packaging material for storage of bovine colostrum powder under storage condition of 4°C temperature and 40–70% RH.

Sagar and Kumar (2014) evaluated the effect of drying treatments and storage stability on quality characteristics of bael powder. To arrest caking, maltodextrin and tricalcium phosphate were added and dried in mechanical drier inorder to get dehydrated powder. Moisture content was between 4% - 5%. Sieved powders were packed in 400 gauge and 200 gauge HDPE, 150 gauge PP, LDPE and pouches and kept for storage at low temperature (7°C) and at ambient condition (18–35°C) up to 6 months. Quality characteristics of powder was evaluated before and after storage. The 200 gauge HDPE is exhibited as best packaging material for storage at low temperature with least changes in stored product.

2.10 Cost estimation of spray dried powders

Sunitha (2012) calculated the cost of production for *Garcinia cambogia* powder based on the data available from the market. Total fixed cost of production was estimated to be Rs.297840/- and variable cost calculated as Rs. 201,600/-. The cost incurred for the production of 100g of *Garcinia cambogia* powder was Rs.45/-.

Reddy *et al.* (2014) estimated the production cost for spray dried agglomerated goat milk. The equipment used for production of agglomerated goat milk powder was homogenizer, rotary vacuum flash evaporator, spray dryer and fluidized bed dryer. The operating cost of process equipment was calculated as 0.17, 18.4, 46.4 and 4.80 kWh/8h, respectively. The total production cost for 1 kg of agglomerated goat milk powder was estimated to be Rs. 475.04/-. The B: C ratio of agglomerated goat milk powder was determined (1.47:1). The B: C ratio shows that the developed process technology for agglomerated goat milk powder is economically feasible.

Material and Methods

CHAPTER III

MATERIALS AND METHODS

This chapter deals with the materials used and methodology followed for the development and quality evaluation of microencapsulated pseudostem juice powder and are discussed in this chapter under the following sections.

Table 3.1 Raw materials

| | Raw materials | | | | | | |
|----------------------|---|--|--|--|--|--|--|
| Banana pseudostem | | | | | | | |
| Horse gram | Raw cleaned horse gram available in local markets were used for the preparation of extract | | | | | | |
| Milk | Pasteurized cow milk procured from local markets were used for sample preparation | | | | | | |
| Wall materials | Maltodextrin and corn starch which were procured from local markets were used for the microencapsulation process | | | | | | |
| Flavourings | Natural flavourings like ginger and cardamom extracts were extracted from fresh ginger and cardamom procured from local markets | | | | | | |

3.1 Pretreatments of banana pseudostem

Pre-treatment of banana pseudostems were carried out to arrest the discolouration of samples. Pretreatments of banana pseudostem included steam blanching along with citric acid treatment. According to Ioannou and Ghoul, 2013 and Mazzeo *et al.* (2011) steam blanching needed less blanching time and less leaching of water soluble nutrients compared to water blanching. Hence sliced

pseudostems treated with different citric acid concentrations (0.1%, 0.2% and 0.3%) were steam blanched at 100°C.

The steam blancher unit (Plate 3.1) in the Department of Food and Agricultural Process Engineering, KCAET, Tavanur was used for standardising the blanching time of pseudostem. Blanching was performed for various durations *viz.* 30 s, one, two and three minutes.

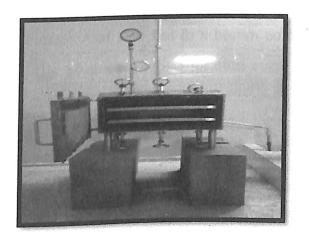


Plate 3.1 Steam blancher

A batch type steam blancher (600×500 mm) comprised of a blanching chamber, pressure discharging valve, inlet and outlet valve with pressure gage was used for this investigation. An arrangement of perforated stainless steel trays on which the citric acid treated slices were held and were arranged parallel inside the blanching chamber. Then, the food grade steam from a steam blancher was supplied to the trays to maintain the chamber temperature approximately at 100°C and was controlled by inlet valves. The blanching trials were carried for 30 s, one and two minute durations (Sotome *et al.*, 2009).

3.1.1 Standardisation of pretreatment

Standardisation of blanching time for the citric acid treated pseudostem samples were estimated by peroxidase test and catalase test (Ndiaye *et al.*, 2009).

3.1.1.1 Peroxidase test

Peroxidase test was carried out to verify the enzyme inactivation (*i.e.*, peroxidase inactivation) in steam blanched pseudostem samples. The test procedure included crushing of steam blanched pseudostem samples which was taken immediately after blanching. Two grams of crushed pseudostem samples were taken in a test tube with 20 ml distilled water. About one ml of 1% guicol solution and 1.6 ml of 0.3% hydrogen peroxide were added to this and thoroughly mixed. Immediate discoloration of samples to a brown colour was the indication of peroxidase activity. The absence of discoloration even after 5 min indicated enzyme inactivation (Shivhare *et al.*, 2009)

3.1.1.2 Catalase test

Catalase test was carried out for standardisation of blanching time of pretreated pseudostem samples by analysing the inactivation of catalase enzyme present in the pseudostem samples. Approximately two grams of crushed pseudostem samples were taken in a test tube immediately after blanching and was mixed with 20 ml of distilled water. It was kept undisturbed for 15 min. After 15 min, 0.5 ml of 1% hydrogen peroxide was added. Strong gas release in the form of bubbles in samples was the indication of catalase enzyme and its absence (Shivhare *et al.*, 2007 and Grimm *et al.*, 2012). Standardisation of blanching of pre treated pseudostem samples was done by considering the factors like enzyme inactivation, colour with different blanching preservatives and time combinations (Prajapaty *et al.*, 2011).

3.2 Experiment I

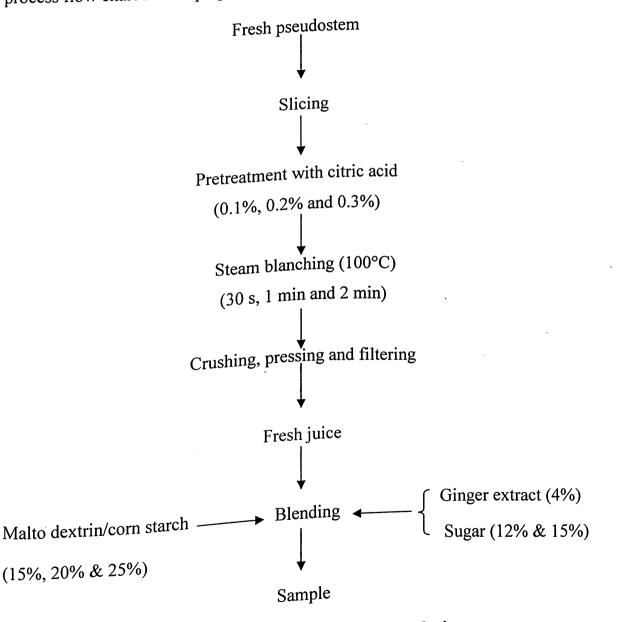
3.2.1 Process technology for production and optimisation of spray dried banana pseudostem juice powder (Product-I)

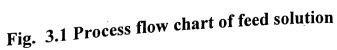
Microencapsulated nutraceutical powders were developed from banana pseudostem juice with maltodextrin and corn starch as wall materials and are discussed under subsequent sections.

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3.2.2 Preparation of feed solution

Pre-treated banana pseudostem samples were crushed by a mixer and pressed manually to squeeze out the juice. Squeezed out juice was filtered in a clean muslin cloth and it was mixed with wall materials like maltodextrin and corn starch along with sugar at various concentrations. Ginger extract was added as a natural flavouring in all the samples. Various treatment combinations for the development of pseudostem juice powder is given in Table.3.1 and the detailed process flow chart for the preparation of feed solution is shown in Fig.3.1.





3.2.3 Raw material concentration and combination

Raw materials like juice from pretreated pseudostem, sugar and wall materials such as maltodextrin and corn starch were taken in various concentrations in each treatment. Equal proportion of ginger extract (4%) was added in all the treatments during the study. Pseudostem based nutraceutical powder was developed by microencapsulating with the aid of spray drying under different operating conditions.

| Combi | nations | |
|--------------|-------------|--|
| Maltodextrin | Corn starch | Raw materials (%) |
| T1 | T7 | P ₆₉ :S ₁₂ :M ₁₅ /C ₁₅ |
| T2 | Т8 | P ₆₆ :S ₁₅ :M ₁₅ /C ₁₅ |
| T3 | Т9 | P ₆₄ :S ₁₂ :M ₂₀ /C ₂₀ |
| T4 | T10 | $P_{61}:S_{15}:M_{20}/C_{20}$ |
| T5 | T11 | P ₅₉ :S ₁₂ :M ₂₅ /C ₂₅ |
| Т6 | T12 | P ₅₆ :S ₁₅ :M ₂₅ /C ₂₅ |

P=Pseudostem juice, S=Sugar, M=Maltodextrin, C=Corn starch

3.3 Experiment II

3.3.1 Spray drying of banana pseudostem juice-horse gram extract blend (Product-II)

Ginger flavoured banana pseudostem juice powders were developed by blending horse gram extract in different proportions. Microencapsulation by spray drying technology was adopted for the product development. Detailed methodology for the development of banana pseudostem juice-horse gram extract blend is discussed under following sections.

3.3.2 Preparation of feed solution

Feed solution was prepared by blending freshly prepared pseudostem juice from the pretreated banana pseudostems, horse gram extract and the adjunct. Maltodextrin and corn starch were used as adjunct and equal proportion of ginger extract was added in all treatment combinations for improved sensory quality of final powder. Various treatment combinations for the development of pseudostem juice powder is given in Table 3.3.

Horse gram extract was prepared by the method explained by Verma *et al.* (2014). Raw horse gram was weighed, cleaned and soaked in water for 12 h. Soaked horse gram was pressure cooked for about 10 min and crushed to get the pulp. Pulp which was extracted from the cooked horse gram was allowed to pass through a 125μ sieve to get the clear extract. The procedure for extracting horse gram extract is presented in Fig. 3.2.

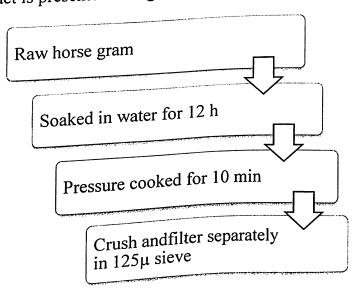


Fig. 3.2 Preparation of horse gram extract

3.3.3 Raw material concentration and combination

Raw materials like juice from pretreated pseudostem, horse gram extract and wall materials such as maltodextrin and corn starch were taken in various concentrations in each treatment. Equal proportions of ginger extract (2%) were added in all the treatments during the study. Pseudostem based nutraceutical powder was developed by micro encapsulating each treatment with the aid of spray drying under different operating conditions. Treatments with raw material combinations were listed in Table 3.3 given below.

| Combin | nations | |
|--------------|-------------|---|
| Maltodextrin | Corn starch | Raw materials (%) |
| T1 | T7 | P ₆₈ :HG ₁₀ :M ₂₀ /C ₂₀ |
| T2 | Т8 | P ₅₈ :HG ₂₀ :M ₂₀ /C ₂₀ |
| Т3 | Т9 | P48:HG30:M20/C20 |
| T4 | T10 | P ₆₃ :HG ₁₀ :M ₂₅ /C ₂₅ |
| T5 | T11 | P53:HG20:M25/C25 |
| T6 | T12 | P43:HG30:M25/C25 |
| | | |

 Table 3.3 Treatment combinations

P=Pseudostem juice, HG=Horse gram extract, M=Maltodextrin, C=Corn starch

3.4 Experiment III

3.4.1 Development and quality evaluation of milk fortified banana pseudostem juice powder (Product-III)

Fortified banana pseudostem juice powder was developed by incorporating milk and horse gram in various proportions. The detailed methodology adopted for its development is discussed in subsequent sections.

3.4.2 Feed preparation

Freshly prepared pseudostem juice squeezed out from the pre-treated pseudostem slices and horse gram extract along with milk were blended together during feed preparation. Horse gram extract which was prepared from the pressure cooked horse gram and the pasteurised milk were blended at different concentrations. Various treatment combinations and their concentrations in each treatment is tabulated and given in the Table 3.4.Cardamom powder made from dried cardamom seeds were added in equal proportions among all treatments. Process flow chart for the preparation of feed solution is shown in Fig. 3.3.

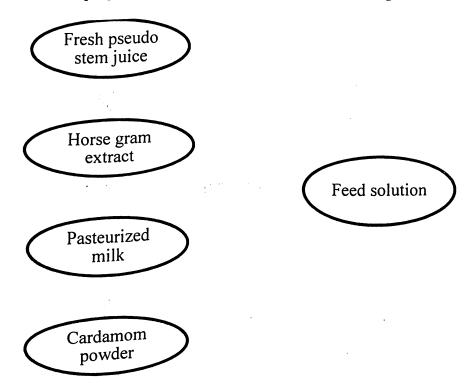


Fig. 3.3 Process flow chart of feed solution

3.4.3 Raw material concentration and combination

Different concentrations of raw materials such as, pretreated pseudostem juice, horse gram extract and milk were blended in each treatment. Since milk is a good carrier, wall materials such as maltodextrin and corn starch were not added for the development of product-III. An equal proportion of cardamom powder (2 pinches, *i.e.*, 0.625 g) was added in all the treatments during the study for enhanced sensorial quality. Pseudostem based nutraceutical powder was developed by micro encapsulating each treatment with the aid of spray drying under different operating conditions. Treatments with raw material combinations were listed in Table 3.4 given below.

| Table 3.4 Treatmen | t combinations |
|--------------------|----------------|
|--------------------|----------------|

| Combinations | Raw materials (%) |
|--------------|---|
| T1 | P ₇₅ :HG ₁₀ :ML ₁₅ |
| T2 | P ₆₅ :HG ₂₀ :ML ₁₅ |
| Т3 | P55:HG30:ML15 |
| T4 | P ₆₅ :HG ₁₀ :ML ₂₅ |
| T5 | P55:HG20:ML25 |
| T6 | P45:HG30:ML25 |
| T7 | P ₆₀ :HG ₁₀ :ML ₃₀ |
| T8 | P ₅₀ :HG ₂₀ :ML ₃₀ |
| T9 | P ₄₀ :HG ₃₀ :ML ₃₀ |
| T10 | P40:HG10:ML50 |
| T11 | P ₃₀ :HG ₂₀ :ML ₅₀ |
| T12 | P20:HG30:ML50 |
| | |

P=Pseudostem juice, HG=Horse gram extract, ML=milk, CP=Cardamom powder

3.5 Microencapsulation by spray drying

Spray drying method was used to obtain pseudostem based nutraceutical powders which were made from different composition (feed solutions prepared in experiment I, II and III). A lab model vertical co-current SMST tall type spray dryer (Plate 3.4) with an evaporation rate of 1000 ml.h⁻¹ was used during the research work. It comprised of air filter, air heater, air distributor, fluid nozzle, drying chamber, collection glass bottles, cyclone separator and an air compressor.

The main factors influencing the powder quality included the spray drying parameters like inlet air temperature, outlet air temperature, feed rate and atomizer speed. For present study distilled water was pumped into the drying chamber in order to adjust the spray drying temperature. This was done about 30 min prior to feeding. The feed was introduced into the spray dryer through feed pipes when it reached the pre-set inlet temperature. Drying was carried out at different inlet air temperature and feed pump speeds to develop microencapsulated banana pseudostem juice powders.



Plate 3.2 Spray dryer

3.5.1 Standardisation of spray dryer parameters

The spray drying parameters were optimised based on the yield and external appearance of the powder. It is having a significant role in producing higher powder efficiency and yield. These response variables were selected as they had significant influence on quality characteristics of the final product (Chegini and Ghobadian, 2007). Spray dryer parameters like feed pump rpm, main blower rpm, Inlet temperature, outlet temperature were optimised from preliminary studies. Spray drying parameters to be standardised are presented in Table 3.5 given below.

Table 3.5 Spray dryer parameters

| SI. No. | Parameter | Experiment I | Experiment II | Experiment III |
|------------|--------------------------------|--------------|---------------|----------------|
| 1 | Main blower rpm | 1800 | 1800 | 1800 |
| 2 | Inlet temperature(°C) | 180 - 190 | 180 - 190 | 185 - 200 |
| 3 | Feed pump rpm | 10-20 | 10-20 | 10-20 |
| 4 | Pressure (Kg/cm ²) | 2 | 2 | 2 |

Based on a thorough review of literature and the preliminary studies conducted, the process parameters which would influence the product quality characteristics, utilisation potential and storage stability were chosen as independent variables. The product quality characteristics which are characteristics of these parameters were selected as dependent variables.

3.6 Physicochemical properties of microencapsulated pseudostem powder

Pseudostem juice powder samples were analysed for physicochemical properties as described in AOAC and from the literature collected.

3.6.1 Moisture content

Hot air oven method described in AOAC (No. 990.20; AOAC, 2005) was carried out for moisture content analysis. The moisture content of pseudostem juice powder was determined by weighing one gram powder sample in a preweighed petri dish and dried in an oven at 100°C for 4 h. Weight differences of the samples were noted after cooling it in desiccators. Procedure was repeated until two similar successive readings were obtained. Determination of moisture content of samples was carried out three times and the average value was considered as moisture content of spray pseudostem juice powder. The moisture content of the sample on percent (w.b) was calculated by using the following formula:

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Moisture content (%) =
$$\frac{W_1 - W_2}{W_1} \times 100$$
 3.1

Where, $W_1(g) =$ Initial weight of sample, $W_2(g) =$ Final weight of sample

3.6.2 Total soluble solids

Total soluble solid (TSS) in fresh pseudostem juice and the diluted pseudostem juice powder was measured using a hand refractometer (Erma inc, Tokyo). Pseudostem powder was mixed with water and allows the sample to settle. One or two drops of the prepared sample were placed on the hand refractometer for TSS measurement. It was expressed in degree Brix (AOAC, 1990).

3.6.4 pH

The pH of the reconstituted pseudostem juice powder was measured using a digital pH meter (Systonics, Naroda) available at Dept. FAPE, Tavanur. Distilled water of pH 7 was used to calibrate the pH meter before determination of pH of powder. Ten milliliters of diluted powder sample was taken in a beaker and the electrode of pH meter was immersed in the sample to determine pH. The reading was directly recorded from the pH meter and was repeated thrice for precision. Average value was considered as pH of powder (AOAC, 1990).

3.6.5 Titrable acidity

Acidity of pseudostem samples was determined by the titration method (No. 947.05 AOAC 2005). Ten ml of diluted powder sample (one gram powder in 10 ml distilled water) was taken in a conical flask and two to three drops of phenolphthalein indicator were added to it. The sample containing indicator was titrated against 0.1N NaOH until light pink end point appeared for few seconds. Volume of 0.1N NaOH used was recorded. Determination of titratable acidity of samples was carried out three times and the average value was considered as titratable acidity of the sample. The acidity of pseudostem powder was computed by using the following expressions:

29

Percentage acidity of powder =
$$\frac{9 \times \text{ml of } 0.1 \text{ N NaOH} \times 0.1}{\text{Weight of powder (g)}} \qquad \dots 3.2$$

3.6.6 Water activity

The water activity of pseudostem juice powder was carried out using Aqua lab water activity meter (M/s. Aqua Lab, U.S.A; model: Series 3TE). Water activity (a_w) is a measure of the amount of water available (Troller and Christian, 1978). For determining the water activity, the pseudostem powder was filled in the disposable cups of the water activity meter and the sample drawer knob is turned to OPEN position. After opening the drawer, the disposable cup with powder was then placed in the drawer and closed. The sample drawer knob was then turned to the READ position and the water activity of powder was noted from the LCD display of the water activity meter. Experiment repeated three times and the average value taken as the water activity.

3.6.7 Total ash

The total ash content of pseudostem powder was determined by muffle furnace method described in AOAC, 2005 (Method No.930.30). One gram of powder samples weighed into a pre weighed crucible of 7-8 cm diameter. Crucibles along with weighed samples were loaded into muffle furnace. Samples were heated about 5 h at 550°C. It was then cooled in desiccator and weighed. The difference in weights taken as the total ash content and is expressed in percentage.

Total ash (%) =
$$\frac{\text{Weight of ash (g)}}{\text{Weight of sample (g)}} \times 100$$
 3.3

3.6.8 Bulk density

The bulk density of spray dried powder obtained from different treatments was measured according to the procedure described by Gong et al. (2008) and Lebrun et al. (2012). Approximately, one gram of powder was freely poured into a 10 ml graduated cylinder without tapping. The level of samples in cylinder was noted for measuring loose bulk density of spray dried powder. Same samples were

repeatedly tapped manually by lifting and dropping the cylinder under its own weight at a vertical distance. This was done until negligible difference in volume between succeeding measurements was observed and is used for the measurement of tapped bulk density of powder. Experiment was repeated for accuracy and average values were considered as loose bulk density and tapped bulk density of samples. The bulk density of powder was computed using the following expression

Loose bulk density
$$(g/ml) = \frac{\text{Weight of sample}(g)}{\text{Bulk sample volume}(ml)}$$
 3.4

Tapped bulk density $(g/cm^3) = \frac{\text{Weight of sample}(g)}{\text{Tapped power volume}(cm^3)}$ 3.5

3.6.9 Colour characteristics

Colour of product is an important parameter that will be valued during product marketing. Colour of the pseudostem based nutraceutical powders were measured using Hunter lab colour flex meter (Hunter Associates Laboratory, Reston, Virginia, USA). The colour was measured by using CIELAB scale at 10° observer at D₆₅ illuminant. It works on the principle of focusing the light and measuring the energy reflected from the sample across the entire visible spectrum. The three dimensional scale L*, a* and b* values were used for colour measurement. The luminance (L*) forms the vertical axis, which indicates light dark spectrum with a range from 0 (black) to 100 (white). In the same way, a* indicates the green - red spectrum with a range of - 60 (green) to + 60 (red) and b* indicates the blue - yellow spectrum with a range from - 60 (blue) to + 60 (yellow) dimensions respectively (Reddy *et al.*, 2014).

The instrument was standardised before placing the sample by placing black and white tile provided with the instrument. Once the instrument was standardised, it was ready to measure the colour. It can also be cross checked by placing the white tile which was provided by the L^* , a^* and b^* values. The sample was filled in the sample cup. The deviation of the colour of the sample to

standard was also observed and recorded in the computer interface. The experiment was repeated thrice for each sample and average was taken as color range.

3.6.10 Particle size analysis by Scanning Electron Microscopy (SEM)

The morphology of powder was determined using scanning electron microscope (SEM) in NIIST, Trivandrum. The scanning electron microscope (SEM) determines the particle size of a powder by using a beam of high energy electrons and electromagnet.

Scanning electron microscopy analysis of the samples was carried out using, JSM-6400 scanning electron microscope (JEOL, Tokyo, Japan). Prior to examination, the samples were uniformly spread on a sample holding stub made of aluminium. A carbon tape was stuck to the sample holding side of the stub and then a thin uniform layer of samples was coated. These samples being nonconductive were sputter coated with gold to render them electrically conductive by using HUMMLE VII Sputter Coating Device (Anatech Electronics, Garfield, N.J., USA). The sputter coated samples were examined at 15 kV. The micrographs were taken at different magnifications.

3.7 Reconstitution properties of spray dried pseudostem juice powder

Reconstituted properties of food powders are important for its market quality and consumer acceptability. Reconstitution properties such as solubility, wettability, water solubility index and water absorption index of spray dried pseudostem juice powder were determined using standard procedures as explained below.

3.7.1 Wettability

Time in seconds necessary to achieve complete wetting of the pseudostem juice powder when it is poured into water at room temperature is noted as wettability. For wettability determination, a glass funnel held on a stand and was set over the beaker containing 100 ml of distilled water at room temperature. A glass rod was kept inside the funnel to block its lower opening. To this setup, one gram sample was placed around the glass rod and then the glass rod was lifted. The time taken for complete wetting of powder particles were noted using a stop watch. Determination of wettability was carried out thrice for spray dried powder and the average value was considered wettability of powder (Jinapong *et al.*, 2008, Desousa *et al.*, 2008, Falade and Omojola, 2010).

3.7.2 Solubility

Solubility of pseudostem powder was carried by the method explained by Chauca *et al.* (2005). One gram pseudostem powder was mixed with 100 ml of water at room temperature for 30 min. A 10 ml aliquot of the supernatant solution was transferred to a 15 ml centrifuge tube and centrifuged for 15 min at 15,000 rpm. The aliquot of the supernatant was then taken in a pre-weighed aluminum moisture dish, evaporated on a steam bath and dried in an oven at 110°C overnight. The solubility was calculated as per equation 3.7.

Solubility(%) =
$$\frac{10 \times \text{Solid in supernatant (g)}}{\text{sample weight (g)}} \times 100$$
 3.6

3.7.3 Water solubility index

The water solubility index of pseudostem juice powder was determined by mixing 2.5 g powder sample and distilled water (30 ml) vigorously in a 100 ml centrifuge tube, incubated in a 37°C water bath for 30 min and then centrifuged for 20 min at 10,000 rpm in a centrifuge (Rotek, 50Cps). The supernatant was carefully collected in a pre-weighed beaker and oven dried at a temperature of 103 \pm 2°C (Anderson, 1969 and Sabhadinde, 2014). Water solubility index was calculated as follows

Water solubility index (%) =
$$\frac{\text{Weight of supernatant}}{\text{Weight of sample}} \times 100$$
 3.7

3.7.4 Water absorption index

Water absorption index of the pseudostem juice powder was determined by agitating suspension of 2.5 g powder in 25 ml distilled water for one hour followed by centrifugation at 3000 rpm for 10 min. The free water was removed from the wet residue, which was then drained for 10 min. The wet residue was then weighed which was taken as the water absorption index of pseudostem juice powder (Sabhadinde, 2014).

Water absorption index (%) =
$$\frac{\text{Weight of residual}}{\text{Weight of sample}} \times 100$$
 3.8

3.8 Proximate analysis

3.8.1 Determination of total carbohydrate

Total carbohydrate present in pseudostem juice powder was determined by Anthrone method. The sample (100 mg) was taken in a boiling tube. It was hydrolyzed by keeping it in a boiling water bath for three hours with 5 ml of 2.5 N hydro chloric acid and cool to room temperature. Followed by neutralisation of the sample with solid sodium carbonate until the effervescence ceases. Again the volume was made up to 100 ml and centrifuged. Supernatant was collected and 0.5 and one milliliter aliquots were taken for analysis. Standards (100 mg glucose + 100 ml distilled water) were prepared by taking 0, 0.2, 0.4, 0.6, 0.8 and one milliliter of the working standard in which, '0' serves as blank. By adding distilled water, volume was made up to one milliliter in all the tubes including the sample tubes. Then 4 ml of anthrone reagent was added and is heated for 8 minutes in a boiling water bath. The readings were taken using spectrometer (*i.e.*, Green to dark green colour at 630 nm was read after cooling) and concentration of the standard versus, absorbance graph was plotted. From the graph, amount of carbohydrate present in the sample tube was calculated by the equation given below (Nithya et al., 2014).

Carbohydra te (%) =
$$\frac{\text{mg of glucose}}{\text{Volume of test sample}} \times 100$$
 3.9

3.8.2 Mineral analysis

3.8.2.1 Atomic absorption spectrometry (AAS)

Pseudostem juice and horse gram extract both are rich in minerals and vitamins. To quantify the minerals present in the spray dried pseudostem juice atomic absorption sprectrophotometer present in Radio tracer laboratory, Vellanikkara campus, Thrissur was used. Atomic absorption spectrometry (AAS) is used as an analytical method to measure the concentrations of elements present in the sample. Atomic absorption is so sensitive that it can measure down to parts per billion of a gram (μ g dm⁻³) in a sample (Ponnambalam and Sellappan, 2014).

3.9 Protein

The nitrogen content in pseudostem powder sample was estimated by using Kjeldahl instrument by Micro-Kjeldahl method (AOAC 2005). Seven grams of Potassium sulfate (K_2SO_4), 0.8 g of cupper (II) sulphate penta hydroxide (CuSO₄.5H₂O) and one gram of powder sample were added to digestion tubes and then 12 ml of hydrochloric acid (H₂SO₄) was added slowly. The digestion tubes were placed on fume ejection system until the digest cleared (clear with light blue–green colour) and was then cooled to room temperature. Digested sample was distilled with 40% sodium hydroxide (NaOH) solution. The distillate was collected in 30 ml of 4% boric acid solution and sample after distillation were titrated against 0.1N hydro chloric acid (HCl) until get the light pink colour was obtained. Methyl red indicator solution and bromocresol green indicator solutions were used as an indicator during titration. The protein content was calculated as:

Protein (%) = $\frac{(14 \times \text{titrate value} \times \text{Normality of alkali} \times 6.25)}{\text{Sample weight}} \times 100 \dots 3.10$

3.10 Flowability and Cohesiveness

3.10.1 Carr's index (CI)

The compressibility index or the Carr's index can be measured from the predetermined bulk density and tapped bulk density values. The index of pseudostem samples was determined by equation 3.11 given below and its powder characteristics was determined by referring the values tabulated by Lebrun *et al.* (2012).

Carr's index (%) =
$$\frac{\text{Tapped bulk density (g/ml)} - \text{Loose bulk density (g/ml)}}{\text{Tapped bulk density (g/ml)}} \dots 3.11$$

3.10.2 Hausner ratio (HR)

Hausner ratio of the pseudostem juice powder was determined by the method described by Shisir *et al.* (2015). HR value for the powder was calculated by the formula given by:

Hausner ratio (HR) =
$$\frac{\text{Tapped bulk density (g/ml)}}{\text{Loose bulk density (g/ml)}} \dots 3.12$$

| Table 3.0 | Specifications for | Carr's Index (%) | Hausner Ratio |
|-----------|--------------------|------------------|---------------|
| Sl. No. | Flowability | 0-10 | 1-1.11 |
| 1 | Excellent | 11-15 | 1.12-1.18 |
| 2 | Good | 16-20 | 1.19-1.25 |
| 3 | Fair | 21-25 | 1.26-1.34 |
| 4 | Possible | 26-31 | 1.35-1.45 |
| 5 | Poor | 32-37 | 1.46-1.59 |
| 6 | Very poor | >38 | >1.60 |
| 7 | Very, very poor | | |

Table 3.6 Specifications for Carr's Index and Hausner Ratio

Source: Lebrun et al. (2012)

3.11 Microbiological Analysis

Microbiological analysis of prepared samples included determination of total viable count. Nutrient agar media was used for the microbial analysis. For carrying out microbial analysis, agar media and glass wares were autoclaved at 121°C for 15 min to make them sterile. One gram of the pseudostem juice powder was taken and added to 10 ml of sterile water blank. Emulsion was shaken well for 10 to 15 min to obtain homogenised suspension of microorganisms and this gave a dilution of 10^{-1} . One ml from (10^{-1}) this dilution was transferred to 9 ml of sterile water blank with a sterile one ml pipette, which gave a dilution of 10^{-2} . The process was repeated up to 10^{-4} dilutions with the sterile water blank. Sterile one ml aliquots from all dilutions were transferred to the sterile petri dishes with nutrient agar for the enumeration of microbes. The experiments were carried out in triplicate for greater accuracy. About 15-20 ml of growth media were poured to plates at temperature (45-50°C) and the plates were rotated clockwise and anticlockwise directions on the flat surface to have a uniform distribution of colonies. Plates were kept undisturbed until the agar gets solidify. After solidification, the plates were inverted and incubated at room temperature for 2-5 days (bacteria one day, yeast and fungi three days). Total plate counts (TPC) were determined on plate count agar pour plates and enumerated after an incubation period of 48-72 h at 30°C (Stillings et al., 1998). The colonies were counted after the incubation period and the number of colony forming units per ml of sample were calculated by applying the following formula:

Number of colony forming units (CFU's) per gram of the sample

$$CFU = \frac{Mean number of CFU's \times Dilution factor}{Ouality of sample on weight basis} \dots 3.13$$

37

3.12 Product yield

Pseudostem powder recovered after feeding was estimated by taking the ratio of weight of sample collected to the juice fed to the drier (Arslan et al., 2015 and Fazaeli et al., 2012) and is given by:

. . . .

Product yield (%) = $\frac{\text{Solid content in product}}{\text{Solid content in feed}} \times 100$

3.14

3.13 Packaging and storage study

Pseudostem juice powders developed by each experiment were packed in LDPE zip pouches and were inserted in aluminium pouches and sealed using a hand sealing machine (Sharma et al., 2003). Sealed pouches were stored in ambient condition for a period of six months and quality parameters viz. moisture content, water activity, TSS, pH, titrable acidity, solubility, wettability, bulk density, solid content and colour values of the powders were recorded in each month (Liu et al., 2010 and Cristina et al., 2008).

3.14 Sensory evaluation

The drink prepared from powder was assessed for their sensory attributes like appearance, flavour, taste and overall acceptability by a 9-point hedonic scale test (Appendix H). Optimised pseudostem powders developed by spray drying method were reconstituted with normal water at 1:10 ratio during sensory analysis. The prepared products were evaluated for sensory characteristics by a panel of 12 judges. Fresh fortified pseudostem juice which was made prior to drying was kept as control for comparison (Neto et al., 2015).

3.15 Statistical analysis

3.15.1 Experimental design

All the experiments in the study were conducted in triplicate and mean values were reported. Completely randomised design (CRD) by Design expert software 7.0 was used to analyse the data. After proper analysis, data were accommodated in the tables as per the needs of objectives for interpretation of results. Statistical significance was examined by analysis of variance (ANOVA) for each response. The p-values were used as a tool to check the significance of each of the coefficients, which, in turn were necessary to understand the pattern of the mutual interactions between the test variables. The smaller the magnitude of the p, the more significant is the corresponding coefficient.



CHAPTER IV

RESULTS AND DISCUSSION

This chapter enunciates the experiments conducted to standardise various parameters for developing spray dried pseudostem juice powder. The chapter also discusses in detail on quality changes that occurred during storage period of optimised product.

4.1 Pretreatment standardisation

The pretreatment (blanching) was done to inactivate enzymes and browning of pseudostem. Pretreatments were carried out by dipping the sliced pseudostem pieces in different concentrations (0.1, 0.2 and 0.3%) of citric acid solution. Samples treated with citric acid were steam blanched at 100°C for 30 s, one and two minutes. The pretreatment was standardised using peroxidase and catalase test. The samples treated with 0.3% citric acid solution and steam blanched at one minute showed negative signals in both test indicating enzyme inactivation. Thus, the blanching time for pseudostem was standardised as one minute and 0.3% citric acid concentration for pre-treatment was fixed and similar results were reported by Nithya *et al.* (2014).

4.2 Physicochemical characteristics of fresh pseudostem juice

Physico-chemical characteristics of fresh pseudostem juice were Quantified prior to spray drying process and are presented in Table 4.1.

| Characteristics of fresh participation Characteristics | Mean ± SD |
|--|-----------|
| Moisture content (% w.b.) | 94.8±0.65 |
| Moisture content (Moisture content (Brix) | 3±0.360 |
| Total soluble solids (°Brix) | 6.5±0.060 |
| рН | |

Table 4.1 Characteristics of fresh pseudostem juice

SD - Standard deviation

The present study on quality evaluation of three products at its fresh form and during storage were investigated and given in subsequent sessions.

4.3 Experiment I: Process technology for production and optimisation of spray dried banana pseudostem juice powder

4.4 Experiment II: Spray drying of banana pseudostem juice-horse gram extract blend

4.5 Experiment III: Development and quality evaluation of milk fortified banana pseudostem juice powder

4.3 Experiment I

4.3.1 Process protocol for production and optimisation of spray dried banana pseudostem juice powder (Product-I)

The detailed processing methodology for the production of spray dried pseudostem juice powder was explained in Chapter III. Different concentrations of wall materials viz., maltodextrin (MD) and corn starch (15, 20 and 25%) and sugar (12 and 15%) were blended with pre-treated pseudostem juice as presented in earlier chapter. These blends were dried in spray dryer at inlet temperatures of 180, 185 and 190°C with varying feed pump speed of 10, 15 and 20 rpm to produce pseudostem juice powder. The main blower rpm of dryer was kept constant at 1800 rpm throughout the study. Different raw material combinations and spray drying parameters were fixed according to the preliminary trials and were statistically analysed using analysis of variance (ANOVA) for product optimisation. From preliminary trials conducted at different feed pump rpm 10, 15 and 20, it was noted that at higher rpm, the outlet temperature was lowered to 48°C. This drastic difference between inlet and outlet temperature led to high moisture (>7%) product. At lower feed pump speed of 10 rpm, it took long time for processing the juice to powder leading to increase in product cost and time of processing (Suzihaque et al., 2015). Thus a medium feed pump rpm of 15 was selected for further studies while low and high feed pump speed were omitted. In

case of wall material, very less yield (<10%) was obtained with corn starch compared with MD. Therefore corn starch was not considered for further studies.

Treatments with different combination of MD, sugar and pseudostem juice mentioned in Chapter III at temperature of 180, 185 and 190°C were considered for development of spray dried pseudostem juice powder.

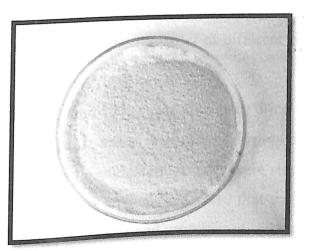


Plate 4.1 Spray dried banana pseudostem juice powder

The effect of processing conditions viz., raw material concentration and inlet air temperature on physico-chemical properties, reconstitution properties, flow properties, microbial analysis, proximate composition, yield and storage stability of spray dried pseudostem powder were determined and are explained in following sections.

4.3.2 Physico-chemical properties of spray dried banana pseudostem powder 4.3.2.1 Effect of spray drying parameters on moisture content

Moisture content of spray dried pseudostem juice powder was in the range of 2.99-4.60% (w.b.) (Appendix A1). As expected, it was observed that at elevated inlet air temperature (190°C) moisture content of sample decreased due to rapid evaporation. Similar trend in moisture content was observed in tomato pulp powder (Gaoula and Adamapoulos, 2010), pineapple (Abadio et al., 2004) and orange juice powder (Chegini and Ghobadian, 2007). Considering the effect of wall material, higher MD concentrations showed a positive effect in reducing the moisture content of spray-dried powder. This phenomenon was explained by

Kha *et al.* (2010) and Shishira *et al.* (2014). According to their study on spray dried gac aril powder and pink guava powder addition of MD increased the total solids of feed and reduced the amount of water for evaporation. This led to reduced moisture content (2.99%) with increase in MD concentration (25%). It was observed that the moisture content of spray dried pseudostem powder increased slightly with increase in sugar concentration. The variation in moisture content of spray dried banana pseudostem powder at different process parameter is shown in Fig. 4.1.

Treatment with 25% MD and 12% sugar and spray dried at 190°C inlet air temperature scored the best moisture value. The effect of spray drying parameters on moisture content of spray dried pseudostem powder were statistically analysed and given in Appendix I1. Statistical analysis indicated that various spray drying parameters and MD concentration showed significant effect (p<0.0001) on moisture content.

4.3.2.2 Effect of spray drying parameters on water activity

Water activity is an important parameter for determining the product stability. Stability of food usually decreases with increase in water activity. The water activity of spray dried pseudostem juice powder varied from 0.295 to 0.430 (Appendix A2). The effects of independent variables on water activity were in agreement with results reported by Gaoula and Adamopoulos (2010) and Chegini and Ghobadian (2007) in orange and tomato juice powder, respectively. Powder with low moisture content of 3.10% and high sugar concentration of 15% showed low water activity of 0.295. Similar findings were reported by Nithya *et al.* (2014). At low moisture content and high sugar concentration, the available water for microbial growth gets reduced and thus powder stability increases.

It is evident from Fig. 4.2 that the water activity is maximum of 0.430 for T1 treatment and minimum of 0.295 for T6 treatment. In general, the water activity for all the treatments was found to be at safe level, *i.e.*, below 0.6 as suggested by Fennema (1996). The results from statistical analysis illustrates significant effect of various spray drying parameters and MD concentration on water activity

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(p<0.0001). The ANOVA associated with completely randomized design was implemented and presented in Appendix 12.

4.3.2.3 Effect of spray drying parameters on bulk density

Loose and tapped bulk density values of developed pseudostem juice powders with various raw material concentrations at different spray drying parameters were investigated. Tapped bulk density (TBD) of banana pseudostem juice powder samples ranged between 0.410-0.510 g/ml (Appendix A3) and loose bulk density (LBD) ranged from 0.258 to 0.457 g/ml (Appendix A4). It was observed that the bulk densities of spray dried banana pseudostem juice powder decreased with increase in inlet air temperature. As a result of rapid evaporation, vapour droplets were formed and gets expanded at high inlet air temperature of 190°C. Thus at elevated temperature micro particles become hollowed and results in low bulk density (TBD - 0.410 g/ml and LBD - 0.258 g/ml). Similar results were reported by Samborska and Bienkowska, (2013) for spray dried honey powder and Chegini and Ghobadian (2007) for orange juice powder.

The changes in bulk density pertaining to MD concentration were analysed for all the six treatments. The results confirmed that both loose and tapped bulk density of final product decreased with increase in MD concentration. This might be due to high porosity of powder observed with increase in wall material concentration (Caparino *et al.*, 2012). The study on spray dried tomato pulp by Shrestha *et al.* (2007) illustrated an increase in MD concentration with a decrease in Shrestha *et al.* (2007) illustrated an increase in MD concentration with a decrease in the bulk density. Lower bulk density values were observed at 25% MD concentration.

Figures 4.3 and 4.4 represent the changes in bulk density values (*i.e.*, TBD and LBD) of spray dried banana pseudostem juice powder at different process parameter. It is clear that the bulk density values of powder decreased with decrease in sugar concentration. The effect of spray drying on bulk density of banana pseudostem juice powder were statistically examined and tabulated in Appendix I3 pseudostem juice powder were statistical analysis, it is evident that spray drying parameters and and I4. From the statistical analysis, it is evident that spray drying parameters and MD concentration affects the bulk density significantly (p<0.0001).

Moisture content (% w.b.)

2.500

15% MD

25% MD

20% MD

25% MD

15% MD

20% MD

25% MD

15% B

180°C

185°C

190°C

Treatments

20% Ð

3.000 3.500 4.000

15% Sugar

12% Sugar

4.500

5.000

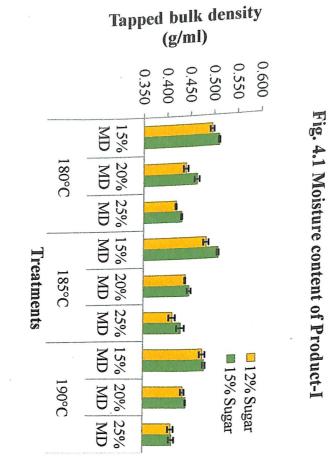
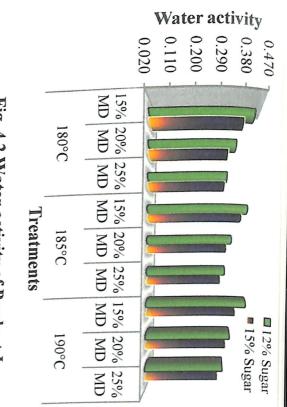


Fig. 4.3 Tapped bulk density of Product-I





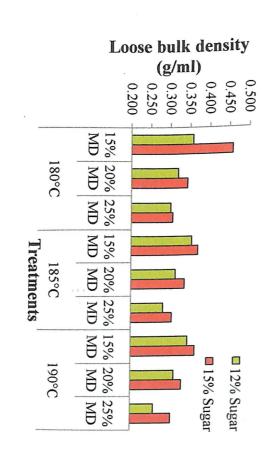


Fig. 4.4 Loose bulk density of Product-I

4.3.2.4 Colour characteristics

Colour is one of the important parameter as it determines consumer acceptance in food applications. It was observed that 'L^{*}, value indicating lightness varied from 77.50 to 92.30 and maximum 'L^{*}, value of 92.30 was recorded in treatment T6. The variation of 'L^{*}, values for different treatments are given in Table 4.2.

| Colour characteristics | Inlet temperature (°C) | T1 | T2 | Т3 | T4 | T5 | T6 |
|---------------------------|------------------------------|--------|--------|--------|--------|--------|-----------|
| | 180 | 78.850 | 80.000 | 83.000 | 86.140 | 88.060 | 92.030 |
| L^{\star} | 185 | 78.010 | 79.910 | 83.200 | 86.600 | 88.010 | 92.300 |
| 2 | 190 | 77.500 | 79.000 | 85.710 | 85.500 | 87.500 | 90.200 |
| a [*] | 180 | 1.520 | 0.610 | 0.500 | 0.430 | 0.400 | 0.270 |
| | 185 | 1.270 | 1.180 | 0.510 | 0.440 | 0.410 | 0.340 |
| | 190 | 2.240 | 0.530 | 0.510 | 0.470 | 0.420 | 0.380 |
| b [*] | 180 | 6.020 | 6.340 | 4.600 | 4.010 | 4.100 | 5.240 |
| | 185 | 5.210 | 6.000 | 5.000 | 5.800 | 6.700 | 5.770 |
| | 190 | 7.700 | 7.500 | 6.600 | 4.310 | 5.680 | 7.650 |

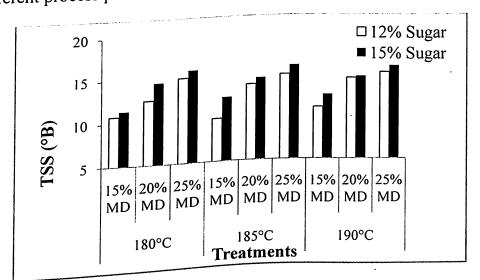
Table 4.2 Colour characteristics of Product-I

A significant increase in 'L^{*}' value was observed with increase in concentration of MD wall material. The variation in 'L^{*}' values from T1 to T6 might be due to the effect of MD concentration. The inherent whitish colour of MD contributes for the bright colour of final powder. Similar results were obtained during spray drying of tamarind pulp by Muzaffar and Kumar (2015). A significant decrease in a^{*} (0.270-2.240) and b^{*} (4.010-7.700) values were noted with increase in MD and decrease in inlet air temperature. The 'L^{*}', 'a^{*}' and 'b^{*}' values of spray dried pseudostem powder were analysed statistically and given in Appendix I5, I6 and I7 respectively.

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4.3.2.5 Total soluble solids (TSS)

Analysis of total soluble solids in reconstituted pseudostem juice powder (one gram powder in 10 ml distilled water) ranged from 10 to 16.10°B (Appendix A5). The result obtained was in agreement with observations of Nithya et al., (2014) in pseudostem juice powder. The experimental results assessed from current study revealed that increase in TSS value might be due to the increased MD concentration and sugar. High TSS value of 16.10°B was noted in treatment with 25% MD and 15% sugar. During this study, the effect of temperature was found to be insignificant. Fig. 4.5 represents the change in TSS with different process parameter.





4.3.2.6 pH and Titrable acidity

The pH of fresh pseudostem juice obtained from the present investigation was found to be 6.5 and for reconstituted powder it was 5.80 to 6.61. These values were similar to those determined by Nithya et al. (2014). The titrable acidity of pseudostem juice powder was determined between 0.115-0.140% in all treatments. Increase in pH and decrease in titrable acidity among treatments might be due to the addition of ginger extract (4%), sugar and wall material concentration. From Table 4.3, it is clear that pH value found to be less than the neutral value. So the powder can graded as low acid foods

increase in concentration of MD due to its lower dissolution capacity. However, powders with higher moisture content showed less time of wettability, due to its agglomeration tendency which helps in reconstitution process (Martinez et al., 2015).

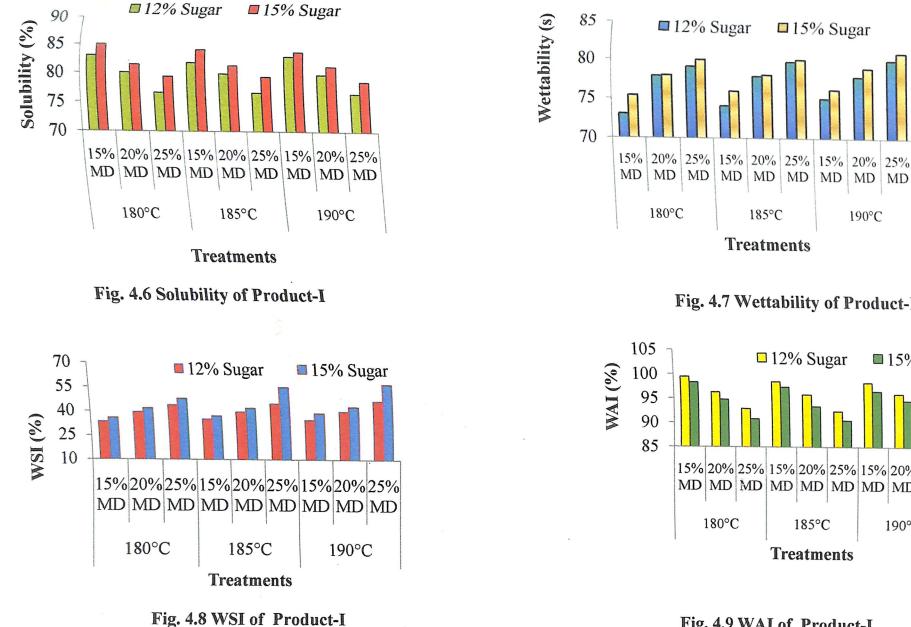
Highest wettability value (80.06 s) was found in T6 while minimum of 73.05 s for T1 (Appendix A7). The effect of wall material concentration and inlet air temperature on wettability of pseudostem powder was statistically analysed using analysis of variance (ANOVA) and is presented Appendix I9. Spray drying parameters and MD concentration showed significant effect (p<0.0001) on wettability of banana pseudostem juice powder.

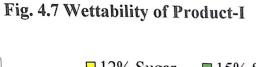
4.3.3.3 Water solubility index

The value of water solubility index (WSI) varied from 33.4 to 57.03% (Appendix A8) for the spray dried powder. From Fig 4.8, it is evident that the water solubility index increased with increase in inlet air temperature (i.e., 180°C to 190°C) and MD concentration. This might be due to low moisture content of powder formed at elevated temperature and high MD concentration which resulted in increased insoluble solids in powders (Jumah et al., 2000). Similar trend in water solubility index was observed in spray dried Garcinia combogia powder by Sunitha (2012).

4.3.3.4 Water absorption index

The water absorption index (WAI) of experimented powder samples ranged between 90.50 to 99.45% (Fig.4.9). WAI value decreased to 90.50% with increase in inlet air temperature (190°C) and MD concentration of 25% (Appendix A9). The obtained result agreed with the study of Phoungchandang and Sertwasana (2010) on ginger powder. Addition of MD during drying made a layer around the drops which affected the surface stickiness of particles and cohesion between them. Reduced cohesion between particles leads to less clustering resulted in low water absorption index (Grabowski et al., 2006 and Adhikari et al., 2003).





185°C

Treatments

□15% Sugar

20% 25%

190°C

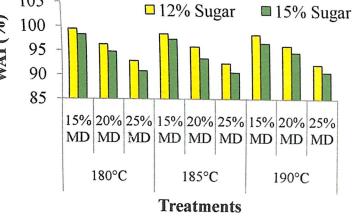


Fig. 4.9 WAI of Product-I

4.3.4 Flowability and Cohesiveness

4.3.4.1 Carr's index (CI) and Hausner ratio (HR)

Flowability and Cohesiveness property of any powder product can be well understand through Carr's index (CI) and Hausner ratio (HR). The results showed that Carr's index (CI) value ranges from 10.457 to 37.043% and the Hausner ratio (HR) of banana pseudostem juice powder varied from 1.117 to 1.588 (Table 4.4).

| Inlet temperature (°C) | T1 | T2 | Т3 | T4 | Τ5 | T6 |
|------------------------------|-----------------------------------|--|---|---|---|--|
| 180 | 27.995 | 10.457 | 27.500 | 26.024 | 28.400 | 29.134 |
| | 26.759 | 27.392 | 28.756 | 24.944 | 31.715 | 29.302 |
| 190 | 27.762 | 24.497 | 28.736 | 25.453 | 37.043 | 26.541 |
| 180 | 1.389 | 1.117 | 1.379 | 1.352 | 1.398 | 1.411 |
| - | 1.365 | 1.377 | 1.404 | 1.332 | 1.464 | 1.414 |
| | 1.384 | 1.324 | 1.403 | 1.342 | 1.588 | 1.361 |
| | temperature (°C) 180 185 | temperature (°C)T118027.99518526.75919027.7621801.3891851.365 | temperature (°C)T1T218027.99510.45718526.75927.39219027.76224.4971801.3891.1171851.3651.37712841.324 | temperature (°C)T1T2T318027.99510.45727.50018526.75927.39228.75619027.76224.49728.7361801.3891.1171.3791851.3651.3771.4041.2841.3241.403 | temperature (°C)T1T2T3T418027.99510.45727.50026.02418526.75927.39228.75624.94419027.76224.49728.73625.4531801.3891.1171.3791.3521851.3651.3771.4041.33212841.3241.4031.342 | temperature (°C)T1T2T3T4T518027.99510.45727.50026.02428.40018526.75927.39228.75624.94431.71519027.76224.49728.73625.45337.0431801.3891.1171.3791.3521.3981851.3651.3771.4041.3321.4641851.2841.3241.4031.3421.588 |

Table 4.4 Flow properties of Product-I

With respect to standard table values (Table3.6) of CI and HR, treatment T2 Spray dried at 180°C showed good flowability. Treatment T2 (190°C) and T4 (185°C) exhibited possible flowability values and other treatments showed poor flowability values. Flowability and cohesiveness of powders depends on size of the Particles and also cohesive and frictional forces within the particles. Particle size depends mainly on atomization airflow (Wang *et al.*, 2015). Apart from the particle size and internal forces flowability and cohesiveness was also affected by many size and internal forces flowability and cohesiveness was also affected by many other factors like moisture content, hardness, surface asperity, elasticity *etc* as reported by Chen *et al.* (2010). Therefore it was difficult to point out appropriate reason for the change in CI and HR values within treatments.

4.3.5 Product yield

Yield of banana pseudostem juice powder ranged from 28.601 to 56.738%. It was found that product yield having significant effect on wall material concentration and inlet air temperature. At low inlet air temperature and high wall material concentration product yield increased due to reduced powder stickiness and higher solid concentration (Arslan et al., 2015 and Fazaeli et al., 2012). During the study, product yield was taken as one of an important parameter for the product optimisation. The influence of spray drying parameters and wall material concentrations on yield of encapsulated banana pseudostem juice powder is presented in Table 4.5.

| | Inlet temperature (°C) | | Т2 | Т3 | T4 | Т5 | T6 |
|-----------|------------------------------|--------|--------|--------|-----------|--------|--------|
| | 180 | 29.164 | 32.034 | 36.834 | 42.022 | 43.451 | 56.738 |
| Product | 185 | 29.079 | 31.711 | 36.268 | 41.970 | 38.247 | 56.664 |
| yield (%) | 190 | 28.601 | 31.092 | 35.460 | 38.224 | 39.171 | 53.663 |

Table 4.5 Product yield of spray dried banana pseudostem juice powder

From the given table, it is evident that yield decreased with temperature and wall material concentration. Treatment T6 exhibited the higher yield at 180°C. The ANOVA associated with general factorial design was implemented (Appendix I10) for statistical analysis and verified the significance (p<0.0001).

4.3.6 Optimisation of spray dried parameters for banana pseudostem juice

powder

Based on the results obtained from physiochemical and reconstitution properties of spray dried pseudostem juice powder which was discussed in previous section, it was confirmed that treatments T6 (56% pseudostem juice +

25% MD+ 15% sugar) was superior in its powder qualities. The banana pseudostem juice powder produced under these treatments showed low moisture content and water activity also it exhibited appreciable pH, bulk density and TSS, better colour and reconstitution properties. The treatment T6 illustrated maximum product yield at different temperatures compared to the rest of other treatments. The optimised process parameters for the powder are presented in Table 4.6. The statistical analysis revealed that all the quality parameters were significant at 1% level. So T6 was selected as best treatment and the T6 sample produced from various temperature combinations (180, 185 and 190°C) were stored in ambient condition for further shelf life studies.

| Table 4.6 Optimised process | parameters | for | the | production | of | pseudostem |
|-----------------------------|------------|-----|-----|------------|----|------------|
| powder by spray drying | | | | | | |

| SI. No. | Process parameters | Optimised condition |
|---------|-----------------------------|---------------------|
| 1 | Speed of the atomizer (rpm) | 1800 |
| 1 | Inlet air temperature (°C) | 180 |
| 2 | Outlet temperature (°C) | 65-68 |
| 3 | | 15 |
| 4 | Feed pump speed (rpm) | |

4.3.7 Microbiological Analysis

The microbial analysis authenticates the shelf life of spray dried pseudostem juice powder. It was observed that, microbial colonies were absent in fresh pseudostem juice powder samples. So the spray dried powder was suggested to be safe for consumption based on microbial analysis. The absence of colony count might be due to low moisture content and water activity. The former was due to drying temperature which removed moisture while the later was due to both drying temperature and addition of sugar (Olivera et al., 2000 and Sariga, 2015). The details of microbial analysis of the powder were tabulated and are given Appendix G1.

4.3.8 Particle size

The morphology of the spray dried powder which was produced at optimised process conditions was examined using scanning electron microscope (SEM) and the observed images are presented in Plate 4.2. The particle size of spray dried powder sample was found to be 50 and 10 µm at 500 and 1000 resolutions respectively.

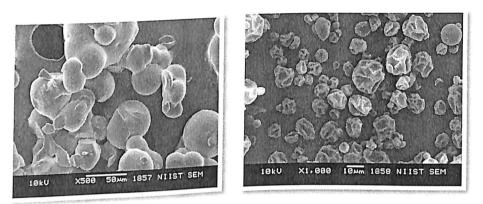


Plate 4.2 SEM micrographs of Product-I

The result obtained in present study was confirmatory with the observations of Krishnan et al. (2005). According to them the particle size of spray dried powder comes under the range of 5-50 μ m.

4.3.9 Proximate analysis

The pseudostem is naturally rich source of micro nutrients. Since the drying was carried out with MD as wall material which is a good source of carbohydrate. The experimentally optimised processed powders were examined for its carbohydrates, protein and minerals. The carbohydrate content of optimised powder was 76.31% and protein 3.03%. Pseudostem powder composed of several minerals like K, Mg, Ca. Mineral composition of wheat flour mixed pseudostem flour were analysed by Ho et al. 2012. Their study confessed the presence of higher Na, K, Ca, Mg and P. Mineral profile analysis of optimised samples were carried out through atomic absorption spectro photometry and is listed in the Table 4.7. Ash content of powder was also determined by muffle furnace method and presented in the Table 4.7. According to Patil et al. (2014) solids present in

guava powder increased with MD concentration which lowers the ash content. Treatment T6 exhibited a lowest ash content of 3.60% and is presented below.

| Composition | % |
|--------------|--|
| Carbohydrate | 76.31 |
| Protein | 3.03 |
| Total ash | 3.60 |
| Potassium | 0.48 |
| Calcium | 1.88 |
| Magnesium | 0.23 |
| | Carbohydrate Protein Total ash Potassium Calcium |

Table 4.7 Proximate composition of Product-I

4.3.10 Packaging of banana pseudostem juice powder

The optimised pseudostem powder samples were packed for shelf life studies in LDPE pouches covered with a laminated aluminum film and were sealed hermetically using hand sealing machine. Sealed packets were stored in ambient condition and tested for its quality attributes. Similar packaging practices were followed by Sunitha, (2012) and Sariga, (2015) for packaging spray dried *Garcinia* powder and vanilla extract powder respectively.

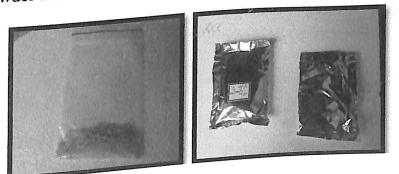


Plate 4.3 Packaged pseudostem juice powder samples in LDPE pouches and aluminium foil pouches

4.3.11 Storage studies

The packed and stored samples (Plate 4.3) at ambient condition were investigated for its quality attributes at two months interval as per analysis methods explained in chapter III.

4.3.11.1 Changes in moisture content and water activity of pseudostem powder during storage

It was observed that the moisture content and water activity of the powder changed significantly during storage (Fig. 4.10 and Fig. 4.11). During filling and sealing the powder is highly susceptible to absorb moisture. This might be due to exposure of product to the external environment. Permeability of packaging material was also an important factor for increased product moisture and water activity values during storage (Pua et al., 2007).

It was noted that the moisture value of T6 at 180, 185 and 190°C showed comparatively less variation during storage period Treatment T6 at 190°C maintained a lowest moisture value of 3.29% at the end of 6th month storage. Water activity at the 6th month of storage for T6 190°C was 0.326. The values of moisture content and water activity at 6th month were acceptable and offered appreciable stability to the product (Appendix B1 and B2). The effect of drying method and storage on moisture content and water activity of pseudostem juice powder were analysed statistically and confirmed the significance (p<0.0001). The ANOVA table related with moisture content and water activity are presented

4.3.11.2 Changes in Bulk density of pseudostem powder during storage Bulk density of the pseudostem powder increased linearly with storage. At the 6th month of storage the tapped bulk density exhibited maximum value 0.443 g/ml and loose bulk density was 0.356 g/ml (Appendix B3 and B4). This might be due to proportionality relation of bulk density with moisture content. It has been reported that bulk density have significant co-relation with the moisture content of the powder. Increase in moisture content tends to raise the bulking weight of the

product which apparently results with high bulk density. Powders having higher moisture content showed higher bulk density of 0.443 g/ml. Similar trend in bulk density was observed by Sunitha (2012) during the storage of *Garcinia cambogia* powder. The changes in tapped and loose bulk density values during the storage were poltted and are given in Fig. 4.12 and 4.13.

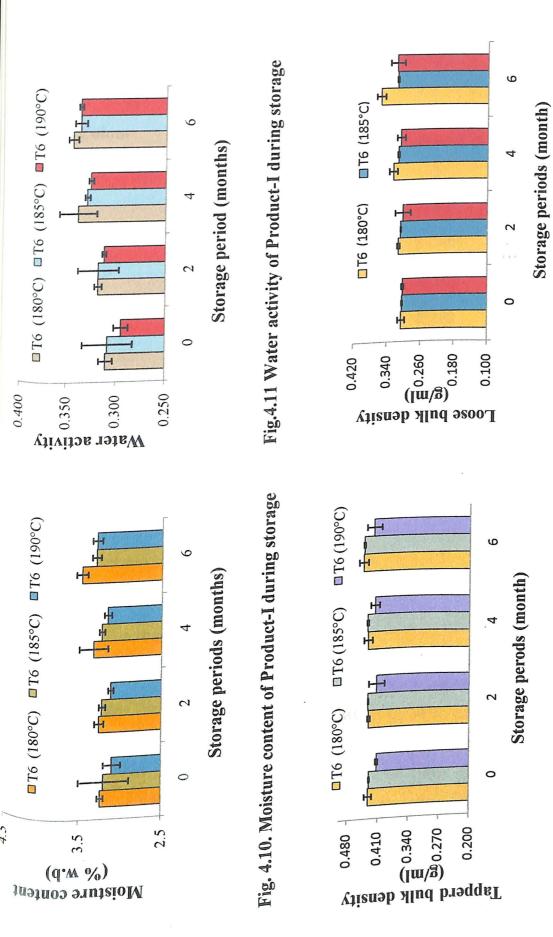
The effect of drying method and storage on bulk density of pseudostem juice powder were analysed statistically and confirmed the significance (p<0.0001). The ANOVA table related with tapped and loose bulk density is presented in appendix J3 and J4.

4.3.11.3 Changes in colour characteristics of pseudostem powder during storage

Lightness L^* value of the spray dried powder reduced during storage period. Chudy *et al.* (2015) stated that the enzymatic browning occurred due to Maillard reaction, lipid oxidation, deterioration of ascorbic acid *etc* contribute for reduction in lightness during storage. The lowest L^* value of 89.270 was observed in treatment T6 (190°C) at the end of 6th month storage.

During the study a^* (redness) and b^* (yellowness) values of powder found to be increased during storage and T6 (190°C) exhibited the highest a^* and b^* values (0.415 and 7.715). The variation in colour values is illustrated in Table 4.8.

The effect of drying method and storage on colour characteristics of banana pseudostem juice powder were analysed statistically and confirmed the significance of a* and b* (p<0.0001) while L* values showed non significance at 1% level. The ANOVA table related with colour characteristics of pseudostem 1% level is presented in appendix J5, J6 and J7.





| Colour characteristics | Treatments | Storage periods (month) | | | | | | |
|---------------------------|------------|-------------------------|--------|--------|--------|--|--|--|
| | - | 0 | 2 | 4 | 6 | | | |
| | T6 (180°C) | 92.030 | 92.000 | 92.000 | 91.870 | | | |
| T 4 | T6 (185°C) | 92.300 | 92.100 | 92.000 | 91.930 | | | |
| L* | Тб (190°С) | 90.200 | 90.100 | 89.380 | 89.270 | | | |
| | T6 (180°C) | 0.270 | 0.276 | 0.282 | 0.297 | | | |
| a* | T6 (185°C) | 0.340 | 0.350 | 0.356 | 0.404 | | | |
| | Т6 (190°С) | 0.380 | 0.350 | 0.357 | 0.415 | | | |
| | T6 (180°C) | 5.240 | 5.243 | 5.248 | 5.253 | | | |
| b* | T6 (185°C) | 5.770 | 5.830 | 5.836 | 5.850 | | | |
| | Т6 (190°С) | 7.650 | 7.661 | 7.663 | 7.715 | | | |

Table 4.8 Changes in colour characteristics of Product-I

4.3.11.4 Changes in pH and titrable acidity of banana pseudostem juice Powder during storage

The pH and titrable acidity of banana pseudostem juice powder varied with storage period. During storage pH of powder found to be decreased from 6.6-6.520 and titrable acidity increased from 0.115-0.138%. This difference in pH and titrable acidity values during storage might be due to formation of acids in powder through chemical conversion of sugars or by other chemical reactions that occurred during storage (Verma *et al.*, 2013). From the Table 4.9, it is clear that a significant increase in titrable acidity value was noted on 6th month of storage. It might be due to increased rate of chemical reaction during 6th month of storage. Sample dried at 190°C showed comparatively low chemical reactions during storage period resulting in slight variation in pH and titrable acidity.

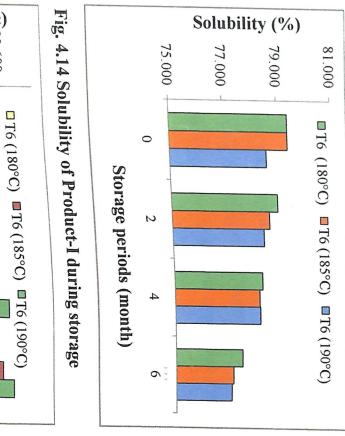
| | Treatments | St | orage period | | |
|-------------|------------|-------|--------------|-------|-------|
| | | 0 | 2 | 4 | 6 |
| | T6 (180°C) | 6.600 | 6.600 | 6.580 | 6.550 |
| | T6 (185°C) | 6.610 | 6.600 | 6.520 | 6.520 |
| рН | T6 (190°C) | 6.600 | 6.590 | 6.580 | 6.580 |
| | | 0.129 | 0.129 | 0.130 | 0.138 |
| Titrable | T6 (180°C) | 0.115 | 0.115 | 0.116 | 0.119 |
| acidity (%) | T6 (185°C) | • | 0.128 | 0.130 | 0.130 |
| actury (70) | T6 (190°C) | 0.128 | 0.120 | 0.150 | |

Table 4.9 Changes in pH and titrable acidity of Product-I

4.3.11.5 Changes in solubility and wettability of pseudostem juice powder during storage

Solubility of powder decreased with storage period, conversely wettability of the product increased (Appendix B5 and B6). This might be due to alteration in moisture content of powder. Samborska and Bienkowska, (2013) suggest that the changes in solubility of powders was accompanied with changes in proteins, moisture content and also the characteristics of drying aid and drying conditions. Change in wettability value also rest on the same reason as reported by Samborska and Bienkowska, (2013).

Samborska and Dichkornance Solubility of powder decreased from 79.40% to 77.13% (Fig. 4.14) and wettability increased from 80.00 to 80.63 s (Fig. 4.15) during storage. The statistical analysis of reconstitution properties showed non significant effect on storage. The ANOVA table related with reconstitution properties of banana storage. The ANOVA table related in appendix J8, and J9.



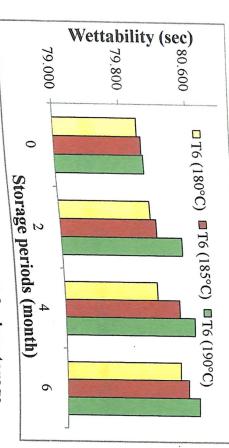


Fig. 4.15 Wettability of Product-I during storage

4.3.11.6 Microbial analysis

powder were tabulated and are given Appendix G4. content and water activity during storage. months. The presence The spray dried powder was suggested to be safe and stable during storage of six recommended by International commission for microbial specifications for foods. T6 (180°C) and in other samples the microbial colonies were absent. It was noted Bacterial colony of 1×10^1 cfu/g was noted at the end of sixth month storage for The microbial load of samples was well below permissible of colony count might be due to the increased moisture The details of microbial analysis of the examined using pour plate method.

4.3.12 Sensory analysis

Sensory analysis is an important parameter for consumer acceptability. The products were analysed by 12 semi – trained sensory panelists. The optimised treatments of banana pseudostem juice powder were reconstituted and kept for sensory evaluation. Powders were reconstituted with normal water in 1:10. The fresh banana pseudostem juice with ginger and sugar was taken as control. The results of the sensory evaluation of the samples are depicted in Fig. 4.16.

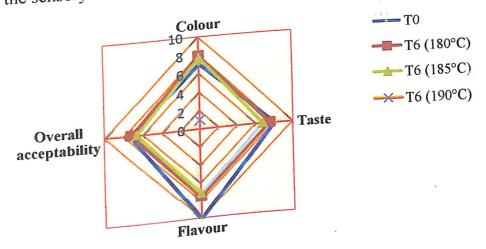


Fig. 4.16 Sensory score obtained for Product-I

Based on sensory analysis, treatment T6 (180°C) scored an overall acceptability of 7.22 which was the highest score among other samples except control. Control sample scored the best score of 7.38, which was on par with the treatment T6 (180°C). Apart from this, T6 (180°C) exhibited appreciable product quality and stability during six month storage. Thus T6 (a treatment combination of 56% pseudostem juice, 25% MD and 15% sugar at inlet air temperature of 180°C) was optimised as the best treatment.

4.4.1 Spray dried banana pseudo stem juice-horse gram extract blend

The aim of this work was to develop a nutraceutical powder from pre-(product-II) treated banana pseudostem juice and horse gram extract by utilizing the novel spray drying technology. The effect of spray drying conditions on powder quality

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was investigated during the study. Various independent and dependent variables related to this experiment were fixed by conducting preliminary experiments. Spray drying parameters such as feed pump and blower speed were kept constant throughout the study as mentioned in previous experiment (*i.e.*, 1800 and 15 rpm). Different temperature combinations such as 180, 185 and 190°C were chosen for product development. Wall material combinations (20 and 25%) and raw materials such as horse gram extract and pseudo stem juice varied from one trial to other (Table3.3).

Analysing final powder for quality verification was a key objective of this study and it includes analysis of:-

- Physico-chemical properties
- Reconstitution properties •
- Flow properties •
- Proximate composition •
- Particle size •

4.4.1.1 Physico-chemical properties

4.4.1.1.1 Moisture content

The moisture content of pseudostem juice powder ranged from 3.930-4.523% (w.b.) and is given in Appendix C1. The effect of process parameters like inlet air temperature and maltodextrin (MD) on the powder moisture content is plotted in Fig.4.17. From the figure, it is clear that the product moisture content followed a similar trend as reported in the earlier experiment. Increase in solid content in feed mixture at higher MD concentration and rapid evaporation at high temperature resulted in moisture reduction (Quek et al., 2007). Horuz et al. (2012) also reported similar fluctuation of product moisture in spray dried pomegranate juice powder, in which the moisture of pomegranate powder found to be increased from 3.44 to 9.13%. Comparing with the previous experiments, moisture content of current sample showed high moisture values. The addition of horse gram might be the reason for this variation.

During the study, higher moisture content was observed in T1 treatment (20% MD and 10% horse gram) spray dried at 180°C inlet air temperature and the lowest value was noted in T6 (3.93%) dried at 190°C. Effect of process conditions on the moisture content of powder was analysed statistically by one-way ANOVA for evaluating the significance of product moisture and its details are given in Appendix K1. Statistical results showed a significant effect of different process conditions on moisture content (p<0.0001).

4.4.1.1.2 Water activity

Water activity value indicates the microbial stability of food. The water activity assessed for banana pseudostem juice-horse gram blend powders were in the range of 0.314-0.407 (Appendix C2). Fennema, (1996) suggest that water activity values should be below 0.6 for stable foods. From the observed values, it was evident that the water activity values were in the safe range and microbiologically stable. Increased inlet air temperature reduced the moisture of product thus the available water for microbial growth gets to drop down resulted in low water activity. The concentration of MD and raw materials does not affect the water activity values significantly. However their combined outcome with temperature exhibited a significant effect on water activity values (Susantikarn and Donlao, 2016). Water activity of 0.407 was found in sample dried at 180°C having 20% MD concentration and 10% horse gram extract. The lowest value (0.314) was observed in powder produced at 190°C inlet air temperature with 25% MD (Fig. 4.18). Statistical results revealed a significant effect of different process parameters on water activity (p<0.0001) and is given in Appendix K2.

4.4.1.1.3 Bulk density

Bulk density values such as tapped bulk density (TBD) and loose bulk density values of spray dried pseudostem juice powders were calculated by standard procedures explained in earlier section. TBD values of powder ranged from 0.416-0.577 g/ml (Appendix C3) and LBD of powder ranged from 0.316-0.474 g/ml (Appendix C4). Obtained results were varied in such a manner that, at elevated inlet air temperature bulk density of the powder gets decreased. Fazaeli *et*

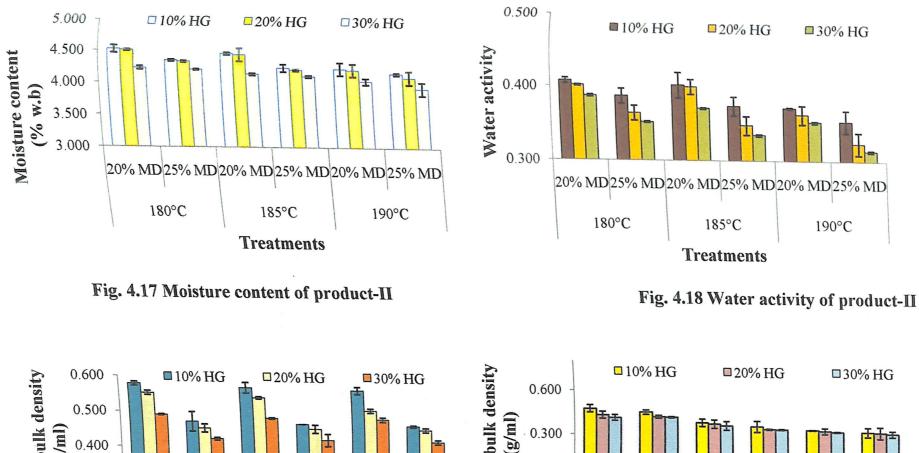
64

al. (2012) illustrates that decreased bulk density in powders at elevated temperatures was due to the poorer shrinkage of powder particles occurred with rapid evaporation.

The lower value of bulk density was obtained at high MD concentration owing to the fact that, the particle size of the powder increased when the feed concentration increased. Similar results were reported by Sarabandi et al. (2014). Their study revealed that the bulk density of spray dried grape juice powder decreased at higher feed concentration. In present study, higher bulk density value observed in treatments having 20% MD and lower values recorded in T6 treatment with 25% MD. Variation in TBD and LBD values are given Fig. 4.19 and Fig. 4.20. Details of the statistical study and ANOVA table were illustrated in Appendix K3 and K4. Statistical study on bulk density values revealed its significance at 1% level.

4.4.1.1.4 Colour characteristics

The colour characteristics such as L^* , a^* and b^* of banana pseudostem juicehorse gram extract blend powder were recorded as: L* (80.133-93.333), a* (0.296-2.216) and b* (5.000-12.800). From Table 4.10 it is clear that treatment T6 at 180°C had a higher L^{*} value. There was a decrease in L^{*} value, more reflected when inlet temperature increased. Maximum and minimum values of L^* were observed in T6 and T1 treatments respectively. The results revealed that L^* value was increased to maximum with higher ratio of MD concentration. Similar observation was reported in the spray dried barberry extract powder by Sharifi et al. (2015), in which powder showed highest L^* at 100% MD concentration.



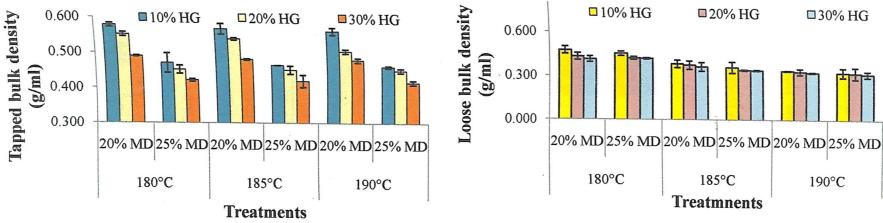


Fig. 4.19 Tapped bulk density of product-II

Fig. 4.20 Loose bulk density of product-II

Similar trend in a^{*} and b^{*} values were noted as mentioned in previous experiment. Higher a^{*}(2.216) and b^{*}(12.8) values were observed in T1 at 190°C. As mentioned earlier in first experiment, colour characteristics of powder were also influenced by inlet air temperatures. Reduction in L^{*} and increased a^{*} and b^{*} values were obtained at higher temperature. This result was confirmatory with Poungchandang and Sertwasana, (2010). Their study revealed that spray dried ginger powder exhibited best colour values (L^{*}- 78.06±0.47 and a^{*}/b^{*}-0.09) at 120°C among other samples dried at 135 and 150°C.



Plate 4.4 Spray dried banana pseudostem-horse gram extract blend powder

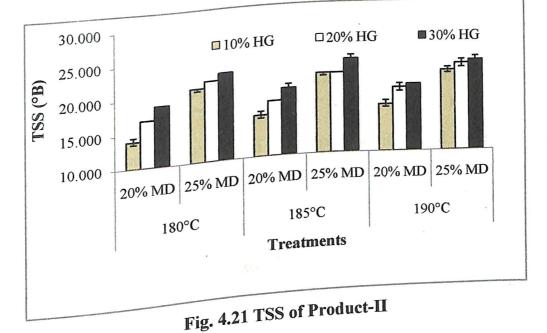
| Table 4.10 Co | Iour end | | | | | | |
|-----------------|----------------------|--------|--------|--------|------------|--------|-----------------|
| Colour | Inlet temperature | T1 | T2 | Т3 | T 4 | T5 | Т6 |
| characteristics | (°C) | 83.160 | 86.376 | 88.356 | 90.176 | 92.100 | 93.333 |
| | 180 | 80.370 | 83.366 | 88.193 | 90.003 | 91.466 | 9 <u>2</u> .526 |
| L | 185 | 80.370 | 85.346 | 86.450 | 88.594 | 90.766 | 92.366 |
| | 190 | 0.510 | 0.446 | 0.296 | 0.520 | 0.510 | 0.357 |
| | 180 | 1.830 | 0.782 | 0.403 | 0.817 | 0.477 | 0.403 |
| ล้ | 185 | | 1.293 | 0.446 | 1.400 | 0.782 | 0.403 |
| | 190 | 2.216 | 5.600 | 5.210 | 5.340 | 5.030 | 5.000 |
| | 180 | 5.600 | 6.680 | 5.600 | 6.800 | 6.500 | 6.340 |
| b [*] | 185 | 8.600 | 8.500 | 7.300 | 12.500 | 9.310 | 6.700 |
| L.Y | 190 | 12.800 | 8.300 | 7.2 | | | |

.10 Colour characteristics of Product-II

The colour characteristics of products showed non significance at 1% level. The corresponding ANOVA tables were given in Appendix K5, K6 and K7.

4.4.1.1.5 Total soluble solids (TSS)

Total soluble solids contained in banana pseudostem juice-horse gram extract blend powder were recorded by standard procedures as mentioned in previous chapter. The powder TSS found to be increased from T1 to T6 (Fig. 4.21) *i.e.*, $14 - 24^{\circ}B$ (Appendix C5). Increased solid concentration by addition of wall material and horse gram extract might be the reason for the increased TSS value (T6). Nithya *et al.* (2014) conducted studies on spray drying of pseudostem-sugar mix. During the study, it was observed that the value of TSS increased with concentration. The highest value of $15^{\circ}B$ was noted at higher feed concentration.



TSS of the current sample was comparatively higher than that of the previous sample as it contains horse gram extract. Evaluation of product TSS revealed that process temperatures have no influence on given results.

4.4.1.1.6 Titrable acidity and pH

The chemical properties like titrable acidity and pH of powder were obtained by standard lab procedures. These values were used to investigate the product's acidic characteristics. The values of titrable acidity and pH of pseudostem juice-horse gram extract blend powder varied from 0.118-0.153% and 5.28-6.910.

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| I able 4.11 Th | | | | | | | |
|-------------------------|------------------------------|-------|-------|-------|-------|-------|-------|
| | Inlet temperature (°C) | T1 | T2 | T3 | T4 | T5 | Т6 |
| | 180 | 6.880 | 6.600 | 6.910 | 6.530 | 6.410 | 6.430 |
| | 185 | 6.370 | 6.270 | 6.550 | 5.830 | 6.030 | 5.280 |
| рН | | 5.600 | 5.910 | 6.200 | 5.600 | 5.550 | 6.000 |
| | 190 | 0.118 | 0.128 | 0.125 | 0.143 | 0.136 | 0.148 |
| | 180 | 0.147 | 0.140 | 0.126 | 0.147 | 0.139 | 0.153 |
| Titrable acidity (%) | 185 | | 0.142 | 0.128 | 0.151 | 0.147 | 0.150 |
| actury (70) | 190 | 0.152 | 0 | | | | |

| Table 4.11 Titrable acidity | and pH | of | Product-I |
|-----------------------------|--------|----|-----------|
|-----------------------------|--------|----|-----------|

The results showed that as the inlet air temperature increased, the titrable acidity increased while the pH value got reduced (Table 4.11). As the moisture content of powder decreased, the feed becomes more concentrated resulting in lower pH and higher acidity values (Rodriguez et al., 2005). Highest titrable acidity of 0.153% and lowest pH of 5.28 observed in T6 treatment. Addition of ginger extract also influenced the titrable acidity and pH values as mentioned in earlier experiment and showed near values of pH and acidity.

4.4.1.2 Reconstitution properties

Reconstitution properties of powder having significant effect on powder quality. The powder reconstitution properties of pseudostem juice-horse gram extract blend powder were analysed and discussed in the following section.

4.4.1.2.1 Solubility

Solubility is one of the main quality aspects in powder products. In this investigation this parameter was found between 97.48 and 99.20% (Appendix C6). As mentioned previously in first experiment, solubility of spray dried banana pseudostem juice-horse gram extract blend powder also showed decrease in solubility at higher temperature and low MD concentration (Fig.4.22). Caparino *et al.* (2012) conducted studies on mango powders and its quality charactreistics. Their study revealed that maltodextrin acted as an encapsulating agent which increased the solubility at lower concentrations. During their study mango powder with 10% MD spray dried at 180°C exhibited a high solubility of 95.31±0.112%. The results obtained during the current study were confirmatory with the observations by Caparino *et al.* (2012).

Higher solubility value of 99.20% observed in treatment T1 at 180°C and lowest for T6 at 190°C. Statistical analysis showed that solubility values were significant at 1% level. The detailed ANOVA table for significance in solubility is presented in Appendix K8.

4.4.1.2.2 Wettability

The wettability values followed similar trend observed in previous powder sample. However the wettability values were high. During the experiments, the wettability values of powder found to be increased from 481.783 to 564.027 s (Appendix C7). Highest wettability value (564.027 s) was found in T6 while minimum of 481.783 s for T1 Martinez *et al.* (2015) analysed beetroot-orange juice powder and observed a higher wettability value of 192.7 ± 11.8 s when 7% MD was added and spray dried at 150° C. They stated that moisture content variation in powder and lower dissolution property of MD were the cause for increased wettability. Wettability values of spray dried banana pseudostem juicehorse gram extract blend powder was analysed statistically and confirmed its significance (p<0.0001) (Appendix K9).

4.4.1.2.3 Water solubility index and Water absorption index

The value of water solubility index (WSI) and water absorption index (WAI) of powder varied from 32.81-60.46% (Appendix C8) and 97.12-102.00% (Appendix C9). Both WSI and WAI values found to be directly proportional to inlet air temperature (Fig. 4.24 and 4.25). Increased inlet air temperature (190°C) led to increased WSI and WAI values (60.46% and 102.00%). According to Sunitha (2012) effect of MD towards WSI and WAI were due to moisture fluctuations and surface properties of powder. Higher WSI and WAI values were observed in treatment T6 at 190°C and T3 at 190°C respectively.

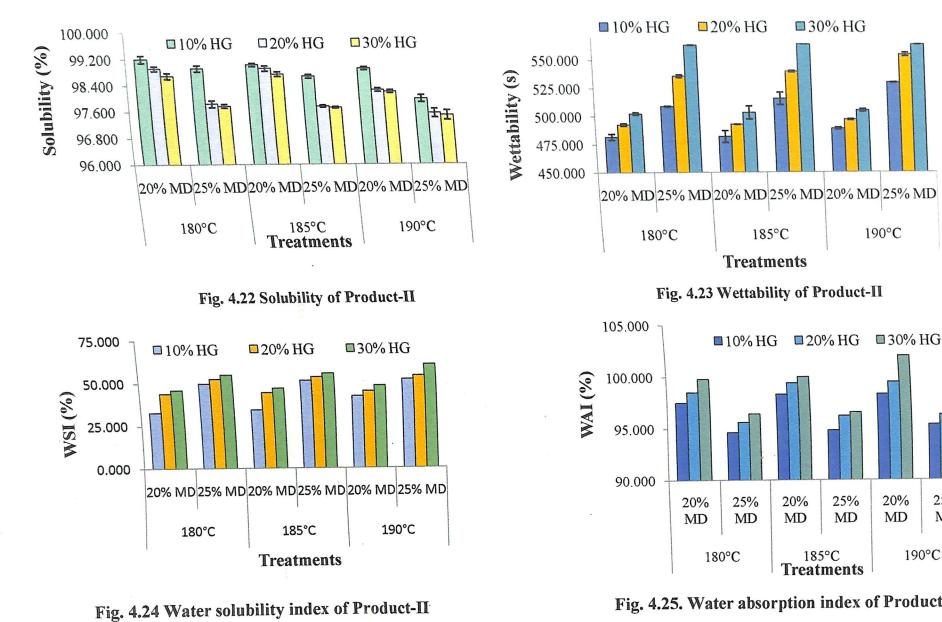


Fig. 4.25. Water absorption index of Product-II

25%

MD

72

4.4.1.3 Flow properties

4.4.1.3.1 Carr's index (CI) and Hausner ratio (HR)

Flow properties such as CI and HR were calculated from observed tapped bulk density and loose bulk density values. The results showed that CI of pseudostem juice powder varied from 0.316 to 39.750% and HR value ranged from 1.00 to 1.66 (Table 4.12). The values of spray dried banana pseudostem juice powder were compared with the standard table values (Table 3.6) and confirmed an excellent flowability of treatment T6 at 180°C.

| | Inlet temperature (°C) | T1 | T2 | Т3 | T4 | T5 | T6 |
|--------|------------------------------|--------|--------|--------|--------|--------|-----------|
| | 180 | 17.851 | 21.550 | 15.628 | 3.685 | 6.720 | 0.316 |
| | 185 | 32.235 | 30.383 | 24.166 | 22.623 | 24.118 | 18.615 |
| CI (%) | 185 | 39.750 | 34.083 | 31.642 | 29.166 | 28.444 | 24.038 |
| | 180 | 1.217 | 1.275 | 1.185 | 1.038 | 1.072 | 1.003 |
| | | 1.476 | 1.436 | 1.319 | 1.292 | 1.317 | 1.228 |
| HR | 185 190 | 1.660 | 1.517 | 1.463 | 1.412 | 1.398 | 1.316 |

Table 4.12 Flowability values of Product-II

As mentioned earlier, fluctuating trend in CI and HR values of remaining treatments were due to frictional and cohesive forces and also due to moisture content, particle size *etc*. Shishir *et al.* (2014) conducted studies on spray dried pink guava powder and flowability of powder noted as HR-1.18 and CI-15%. According to their study, MD and temperature were the key parameters that influenced the flow properties of powder. Increase in temperature and MD concentration increased the CI and HR values during their study.

4.4.1.4 Product yield

Yield of banana pseudostem juice-horse gram extract blend powder ranged from 47.47-78.21%. It was found that product yield had significant effect on wall material concentration and inlet air temperature and is given in Table 4.13. In present study, yield of powder reduced at higher inlet air temperature of 190°C due to stickiness and increased with MD concentration as reported by Papadakis et al. (2006). Fazeli et al. (2012) observed that powder yield of spray dried black mulberry powder varied from 45 to 82% at an inlet temperature range of 110-150°C and 8-16% wall material concentration.

| Table 4.13 Product yield of sp | oray dried | banana | pseudostem | juice- | horse |
|--------------------------------|------------|--------|------------|--------|-------|
| gram extract blend | | | | | |

| | Inlet temperature (°C) | T1 | Т2 | Т3 | · T4 | Т5 | Т6 |
|-----------|------------------------------|--------|--------|--------|-------------|--------|--------|
| | 180 | 52.370 | 54.460 | 69.960 | 56.460 | 64.810 | 73.080 |
| Product | 185 | 53.380 | 54.730 | 69.680 | 56.390 | 64.200 | 78.210 |
| yield (%) | 185 | 50.120 | | | | | 78.150 |

Treatment T6 at 185°C showed high yield of 78.21% and T4 spray dried at 190°C having low yield of 47.47%. Comparing with previous experiments, banana pseudostem juice-horse gram extract blend exhibited higher yield. This might be due to the higher feed concentration contributed by horse gram and MD. Statistical studies revealed that powder yield was non significant at 1% level

(Appendix K10).

4.4.1.5 Optimisation of spray dried parameters

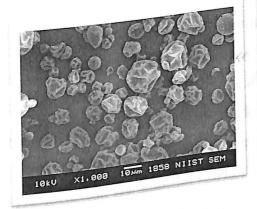
Statistical models were employed to determine numerical optimisation to fix the optimal process parameters for developing fortified pseudostem juice powders. In the present investigation, the independent variables were kept within the range and dependent variables were chosen as maximum and minimum. Inlet temperature of 180°C, outlet temperature of 65-68°C. feed pump and blower rpm of 15 and 1800 were optimised during the study. Optimised sample (T6) was kept for shelf life studies under ambient conditions. Treatment T6 developed at different inlet temperatures (180, 185 and 190°C) were selected to evaluate its quality during the storage period of six months.

4.4.1.6 Microbiological stability

The microbial analysis was carried out for the optimised powder samples. Shelf life and storage stability of the product were predicted based on its microbial load. The results showed that microbial colony was absent in fresh powder samples (Appendix G2). Arawwawala et al. (2011) analysed microbial growth in spray dried piper beetle extract and absence of Escherichia coli, Pseudomonas aeroginosa, Staphylococcus aureus and Salmonella spp. were confirmed.

4.4.1.7 SEM study

Particle size of product was analysed by SEM analysis as mentioned earlier. Particle size of powder varied from 10-50 μ m (Plate 4.5.) at 1000 and 500 resolutions. Sarkar et al. (2016) conducted studies on micro structural studies on spray dried emulsion of high oil content prepared from whey protein. The particle size of powders were analysed by scanning electron microscope and the particle size of powder found to be varied from 37.27- 42.88 μ m.



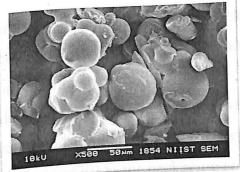


Plate 4.5. SEM micrographs of Product-II

4.4.1.8 Proximate analysis

Nutritional components like minerals. carbohydrate and protein present in the powder samples were analysed by standard procedures. The pseudostem juicehorse gram extract blend powder found to be highly rich in carbohydrate (85.01%), protein (3.8%) and minerals like K (0.69%), Ca (1.98%) and Mg (0.31%) (Table 4.14). Comparing with the previous sample (Experiment I), present sample showed more nutritional compounds. This might be due to the addition of horse gram extract which enriched the powder with high amount of nutrients. Increased concentration in powder reduced the ash content to 2.5%. However the ash content of Product-I was 3.60%.

| 4.14 FI0XIII | | % |
|------------------|--------------|-------|
| | COMPOSITION | 70 |
| SI. No | | 85.01 |
| 1. | Carbohydrate | 3.80 |
| 2. | Protein | 2.50 |
| 3. | Total ash | 0.69 |
| 4. | Potassium | 1.98 |
| 5. | Calcium | 0.31 |
| <i>5</i> . 6. | Magnesium | 0.51 |
| 0. | | |

Table 4.14 Proximate composition of Product-II

4.4.1.9 Packaging and shelf life study

To arrest quality deterioration, standardised powder samples were packed in LDPE pouches. The packed sample was covered with a laminated aluminium film and sealed hermetically using hand sealing machine. Aluminium foils having superior flavor retention capacity and check deterioration of product by providing barriers to light, air, water and most other gases and liquids. Similar packaging practices were followed by Sunitha (2012) and Sariga (2015) for packaging spray dried Garcinia powder and vanilla extract powder respectively. Sealed packets were stored in ambient condition and tested for its quality attributes. Changes in quality attributes during six month storage were analysed and discussed in following sections.

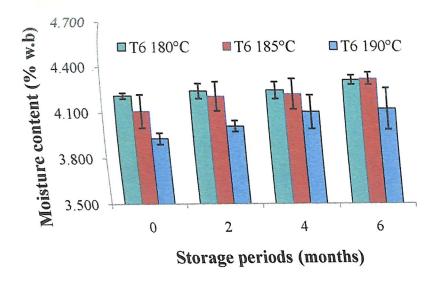
4.4.1.9.1 Changes in moisture content and water activity

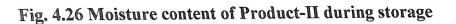
Moisture content and water activity of powder changed significantly during six months of storage period (Fig. 4.26 and 4.27). Moisture content of 4.110% of T6 (185°C) increased to 4.319% after six month (Appendix D, Table D1) and least moisture variation observed in T6 (190°C) after six month (*i.e.*, 3.930 to 4.122%). Water activity variation was comparatively high at T6 (180°C) *i.e.*, 0.355 and T6 at (190°C) showed less water activity value of 0.324 after six month of storage (Appendix D, Table D2). Samborska and Bienkowska, (2013) conducted storage studies on spray dried honey powder and reported that moisture content and water activity of honey powder found to be increased from 1.8-2.9% (w.b.) and 0.151-0.215 respectively during nine month storage period. The study concluded that, addition of MD and moisture absorption during storage might be the reason for moisture and water activity variation in powders. Variation of moisture content and water activity is given Fig. 4.26 and Fig. 4.27. Statistical studies revealed that moisture content and water activity values were significant and the related ANOVA table is given in Appendix L1 and L2.

4.4.1.9.2 Changes in bulk densities of powder

As reported in previous experiment, both tapped bulk density and loose bulk density of spray dried banana pseudostem juice-horse gram extract blend powder increased during storage (Fig. 4.28 and Fig. 4. 29). The tapped bulk density of 0.421 g/ml (T6-180°C) increased to 0.438 g/ml at the end of 6^{th} month (Appendix D3). Similar increase in loose bulk density was also observed and was 0.429 g/ml at the end of 6^{th} month (Appendix D, Table D4). Treatment T6 at 190°C showed lowest values of tapped and loose bulk density at the end of 6^{th} month storage (*i.e.*, 0.347 g/ml and 0.425 g/ml). Sunitha (2012) conducted studies on spray drying of *Garcinia combogia* powder and observed changes in bulk density values during six month storage. The study revealed that increase in bulk density from 0.428 to 0.540 g/cm³ might be due to the increased moisture absorption during storage. Statistical studies revealed that bulk density values absorption during storage. Statistical studies revealed that bulk density L3 and L4.

77





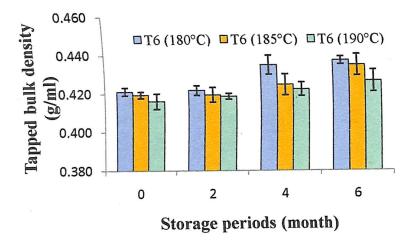
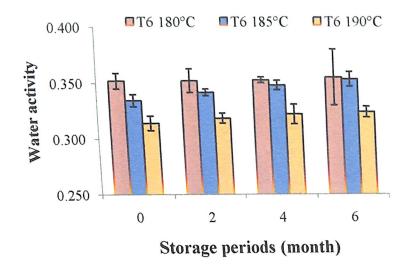
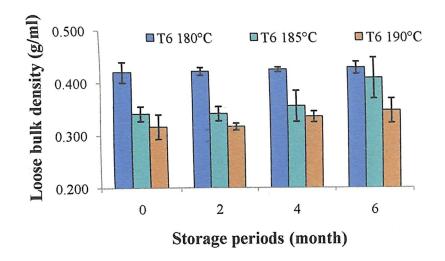
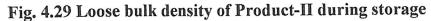


Fig. 4.28 Tapped bulk density of Product-II during storage









4.4.1.9.3 Changes in colour characteristics of powder

During storage colour values of powder changed slightly. It was noted that the L* value of powder decreased during storage and the change was more pronounced between 4^{th} and 6^{th} month (Table 4.15). According to Liu *et al.* (2010) decreased L* value might be the non enzymatic reaction occurred during storage and they observed a decrease in colour value in tomato powder $(37.60\pm0.25 \text{ to } 35.05\pm0.23)$ during five months storage. The colour values such as a* and b* of powder retained without appreciable fluctuations and are significant at 1% level.

٦

| Colour characteristics | Treatments | Storage periods (month) | | | |
|---------------------------|------------|-------------------------|--------|--------|--------|
| | | 0 | 2 | 4 | 6 |
| | | 93.333 | 93.333 | 93.310 | 93.010 |
| L* | T6 (185°C) | 94.366 | 94.364 | 94.361 | 94.267 |
| | T6 (190°C) | 94.366 | 94.360 | 94.342 | 94.200 |
| | | 0.403 | 0.403 | 0.403 | 0.404 |
| a* | T6 (180°C) | 0.357 | 0.357 | 0.358 | 0.358 |
| | T6 (185°C) | 0.403 | 0.403 | 0.403 | 0.404 |
| | T6 (190°C) | 5.000 | 5.000 | 5.001 | 5.010 |
| b* | T6 (180°C) | | 6.341 | 6.341 | 6.341 |
| | T6 (185°C) | 6.340 | | | |
| | T6 (190°C) | 6.700 | 6.701 | 6.701 | 6.705 |

Table 4.15 Colour characteristics of Product-II during storage

Statistical studies revealed that colour values were significant and the related ANOVA table is given in Appendix L5, L6 and L7.

4.4.1.9.4 Changes in pH and titrable acidity of powder

pH and titrable acidity of powder varied significantly during storage. Titrable acidity and pH of tomato powder were analysed by Liu *et al.* (2010) and reported that titrable acidity of tomato powder increased from 7.61 to 8.02% and pH varied from 4.17 to 3.87 during five month storage at 25°C. According to them, increased titrable acidity and pH might be due to the effect of storage conditions and the conversion of sugar to acids by maillard reaction. Similar trend in pH and titrable acidity was observed in current study and is given in Table 4.16

| | Treatments | Storage periods (month) | | | |
|-------------|------------|-------------------------|-------|-------|-------|
| | | 0 | 2 | 4 | 6 |
| | T6 (180°C) | 6.880 | 6.821 | 6.723 | 6.510 |
| рН | T6 (185°C) | 6.370 | 6.345 | 6.254 | 6.163 |
| | T6 (190°C) | 5.280 | 5.251 | 5.233 | 5.193 |
| | T6 (180°C) | 0.118 | 0.118 | 0.119 | 0.121 |
| Titrable | T6 (185°C) | 0.129 | 0.126 | 0.126 | 0.124 |
| acidity (%) | Тб (190°С) | 0.153 | 0.154 | 0.152 | 0.150 |

Table 4.16 Changes in pH and titrable acidity of Product-II during storage

4.4.1.9.5 Changes in solubility and wettability of powder

Solubility and wettability of pseudostem juice-horse gram extract blend powder was analysed during each month to confirm the stability. Experimental results showed a decreased solubility and increased wettability values during storage (Table 4.17). Solubility of T6 (185°C) retained higher solubility value at 6th month storage and T6 (180°C) showed higher wettability value at the end of 6th month. Ramachandran *et al.* (2014) observed a decrease in solubility value of papaya powder containing ($56.5\pm5\%$ to $50.5\pm5\%$) during 30 days storage and Sariga (2015) reported an increased wettability values *(i.e., 3.503-45.13 s)* in spray dried vanilla extract powder during six month storage. According to Ramachandran *et al.* (2014) and Sariga (2015), the solubility change and wettability variations might be due to the moisture absorption in powders during storage. The results obtained during present study were confirmatory with the former studies and followed similar trend in solubility and wettability values during storage. Statistical studies revealed that solubility and wettability values were significant and the related ANOVA table is given in Appendix L8 and L9.

| | Treatments | Storage periods (month) | | | |
|-----------------|------------|-------------------------|---------|---------|---------|
| | - | 0 | 2 | 4 | 6 |
| | T6 (180°C) | 97.770 | 97.770 | 97.721 | 97.540 |
| Solubility (%) | T6 (185°C) | 97.720 | 97.770 | 97.693 | 97.665 |
| | Т6 (190°С) | 97.480 | 97.453 | 97.450 | 97.361 |
| | T6 (180°C) | 563.300 | 563.345 | 563.440 | 563.501 |
| | T6 (185°C) | 564.027 | 564.054 | 563.074 | 563.088 |
| Wettability (s) | T6 (190°C) | 563.867 | 563.900 | 563.050 | 563.110 |

| able 4.17 Changes in solubility and wettability of Product-II during storage |
|--|
|--|

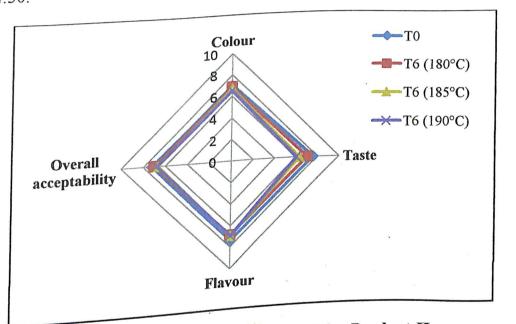
4.4.1.9.6 Microbial stability of powder during storage

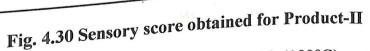
The microbial load of samples was examined using pour plate method and depicted in Appendix G3. Among stored samples T6 at 185°C showed comparatively higher colony count and at the end of sixth month storage and it was 3×10^{1} cfu/g. It was noted that colonies were well below permissible limit

 $(<10^4)$ (T6-190°C) recommended by International commission for microbial specifications for foods. The spray dried powder was suggested to be safe and stable during storage of six months. The presence of colony count might be due to increase in moisture content and water activity during storage.

4.4.1.10 Sensory analysis

Sensory analysis of products was done by 12- semi-trained sensory panelists. The optimised powder samples were reconstituted and kept for sensory evaluation. Powders were reconstituted with normal water in 1:10 ratio. The fresh pseudostem juice, freshly prepared horse gram extract with ginger and sugar was taken as control. The results of the sensory evaluation of the samples are given in Fig. 4.30.





Based on sensory evaluation treatment, T6 (180°C) scored an overall acceptability of 7.02 which was the highest secured among the other samples except control (Appendix D, Table D5). Control samples scored the best score of 7.24. Comparing with control treatment, T6 (180°C) maintained similar sensory quality. Apart from this T6 (180°C) exhibited appreciable product stability during six month storage. Thus T6 (43% banana pseudostem juice, 30% horse gram six month storage. Thus T6 (43% banana pseudostem juice, 30% horse gram six and 25% MD with 2% ginger extract) at inlet air temperature of 180°C was optimised as the best treatment.

4.5 Experiment III

Development and quality evaluation of milk fortified banana 4.5.1 pseudostem juice powder (product-III)

A process has been developed for the preparation of fortified banana pseudo stem juice powder using the spray drying technology. In order to improve the tastiness and nutritional value of powder, milk (15, 25, 30 and 50%) and horse gram extract (10, 20 and 30%) were incorporated in predefined concentration. Cardamom powder was added during drying to enhance the sensorial quality. Quality parameters of the powder which was obtained in different spray drying temperatures (185, 190 and 200°C) with selected feed pump and blower rpm of 15 and 1800 respectively were judged by standard methods and the results obtained are discussed in the following sections.

4.5.1.1 Quality evaluation of fortified pseudostem juice powder

4.5.1.1.1 Moisture content

Moisture of the pseudostem powder was found to be in the range of 3.64-4.5% w.b (Appendix E, Table E1). It was observed that an increase in total solid content in feed reduced the moisture of final powder. As the total solid content in feed increased, the available water for evaporation gets decreased which led to the reduction in moisture content. In present study feed with 30% horse gram extract and 50% milk exhibited the lowest moisture value of 3.64% (Fig.4.31).

Powder dried at different spray drying temperatures (i.e., 185 190 and 200°C) were analysed for moisture content. Efficient removal of water at high temperature yields a low moisture product. It can be elucidated that, elevated inlet air temperature offered an appreciable driving force for moisture deduction (Goula and Adamopoulos, 2010). A low moisture value of 3.64% was noted at 200°C and high moisture of 4.5% was present in samples dried at 185°C. However the product obtained at high temperatures were over dried. Statistical studies on moisture content of fortified pseudostem juice powder confirmed its significance at 1% level (p<0.0001) (Appendix M1).

4.5.1.1.2 Water activity

Water activity of the spray dried powder was analysed for predicting microbial intensification and shelf life stability. The water activity value observed for present study ranged from 0.221 to 0.376 (Appendix E2). Experimental results showed that the water activity and moisture content were related to each other. Water activity exhibited a sudden drop with an increase in inlet air temperature and decrease in moisture content (Fig. 4.32). These observations were comparable with the results illustrated by Schuck et al. (2005). Schuck et al. (2005) observed a lower water activity value of 0.16 in tomato powder spray dried at an inlet temperature of 210°C among the trials conducted at 140, 190 and 210°C. While, the higher water activity of 0.32 observed at 140°C.

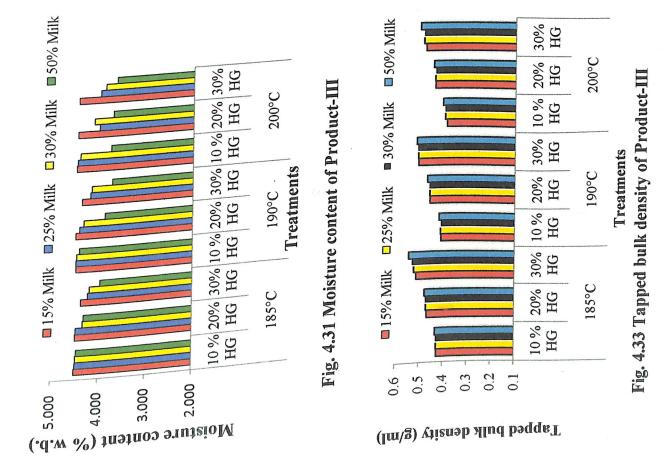
Water activity of fortified pseudostem juice powder showed a higher value of 0.376 at 185°C and minimum of 0.221 at 200°C. Water activity recorded for the powder was found to be in the permissible limit as suggested by Stapelfeldt et al. (1997). According to their study, water activity range for best quality whole milk powder was observed at 0.11-0.23. Comparing the water activity value of fortified pseudostem juice powder with whole milk powder, water activity values were safe for fortified pseudostem juice powder samples. The water activity of pseudostem juice powder was analysed statistically. It was observed that water activity was significant at 1% level (p < 0.0001) and the ANOVA table related with it is given in Appendix M2.

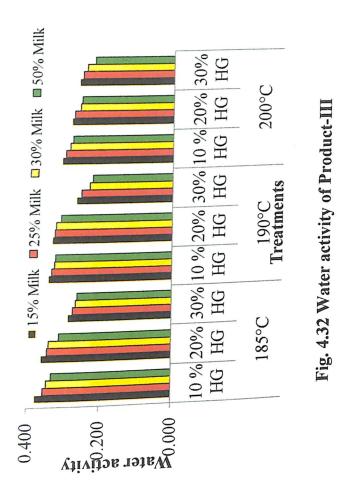
4.5.1.1.3 Bulk density

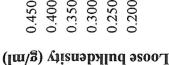
The loose and tapped bulk density of powdered samples under different experimental conditions was computed. The loose bulk density value ranged from 0.309-0.431 g/ml and tapped bulk density ranged from 0.387-0.543 g/ml (Appendix E3 and E4). Among different treatments, loose and tapped bulk density values were lowest for treatment T_1 (200°C) (*i.e.*, 0.309 g/ml and 0.387 g/ml) while, highest for treatment T_{12} (185°C) (*i.e.*, 0.431 and 0.543 g/ml). These experimental observations revealed that the bulk density values get decreased under elevated process temperatures. Research conducted on milk powder by

Nijdam and Langrish. (2006) provided key evidence for the former statement. According to their study. milk powders were produced at 120 and 200°C in which low bulk density values were observed at 200°C. Employing higher inlet air temperature resulted in enlarged particle formation due to rapid evaporation and layer development over feed droplets, thus leading to a low bulk density (Phisut, 2012, Jittanit *et al.*, 2011 and Reineccius, 2001).

From Fig.4.36, it is evident that an increase in bulk density values were observed with an increase in feed concentration. Chegini and Ghobadian (2008) and Kha *et al.* (2010) were stated that, increase in feed concentration leads to a decrease in particle volume and increase in bulk density. Samples with 30% horse gram extract and 50% milk showed the highest bulk density values of 0.431 and 0.543 g/ml at an inlet temperature of 185°C. Statistical analysis of product exhibited significance at 1% and the ANOVA table related with the bulk density values are presented in Appendix M3 and M4.







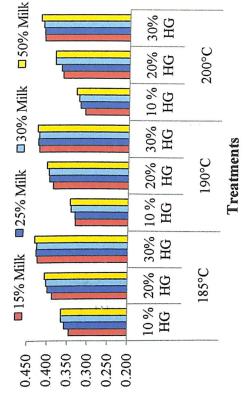


Fig. 4.34 Loose bulk density of Product-III

86

4.5.1.1.4 Colour characteristics

Colour characteristics of fortified pseudostem powder were examined using Hunter lab to verify the powder quality and consumer acceptability. Colour characteristics including L^* , a^* and b^* obtained from instrument were recorded as: L^* - 67 to 90.03, a^* - 4.94 to 10.63 and b^* - 15.19 to 24.71.



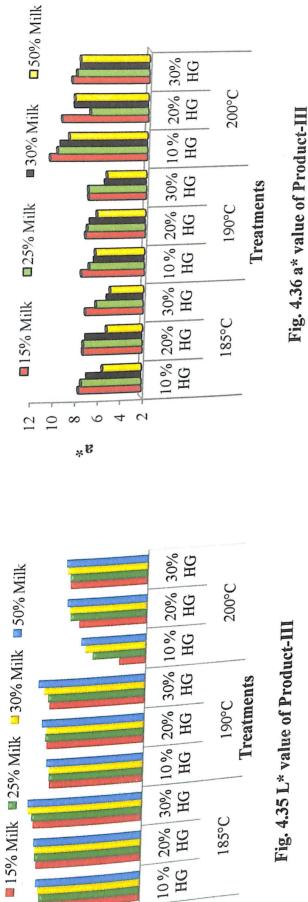
T12 (185°C)

T1 (200°C)

Plate 4.6 Fortified pseudostem juice powder at different inlet air temperature

The luminance (L^*) of powder was found to be fair and acceptable Influence of inlet temperature and raw material concentrations showed a significant effect on L^* value. Treatments conducted at low temperature showed appreciable increase in L^* values (Appendix E5). Highest L^* value of 90.03 was recorded in treatment T_{12} at 185°C. At elevated temperature the chance of browning gets increased due to product oxidation (Bento *et al.*, 2008 and Fonseca browning gets increased due to product oxidation (Fig. 4.35) from *et al.*, 2011). Thus at high temperature L^* values get decreased (Fig. 4.35) from 90.03 to 67. The a^{*} values of powder under different spray drying temperatures along with varying concentration of raw materials were observed (Appendix E7). The maximum a^{*} value of 10.63 was found in the treatment T1, whereas minimum value of 4.94 was recorded in T12. In this study, a^{*} values of fortified pseudostem juice powder were significantly affected by feed concentration as well as inlet air temperature (p<0.0001) (Appendix M5, M6 and M7). The variation in a^{*} value of fortified pseudostem juice powder at different process is depicted in Fig. 4.36 given below.

A significant variation in b^{*} value was noted at high inlet air temperature (Appendix E7). The average b^{*} value of fortified pseudostem juice powder ranged from 15.19 to 24.71 (Fig. 4.37). Highest value of 24.71 was recorded in T1 (200°C) treatment and lowest value of 15.19 for T12 (185°C). Occurance of non enzymatic browning as a result of maillard reaction might be the reason for increased b^{*} value (Bento *et al.*, 2008 and Fonseca *et al.*, 2011).





30

190°C Treatments

185°C

60

89

92

84

76

r*



20

*d

10



Treatments

200°C

190°C

185°C

 10 %
 20%
 30%
 10 %
 20%
 30%
 10 %
 20%
 30%
 10 %
 20%
 30%
 HG
 HG

89

4.5.1.1.5 Total soluble solids (TSS)

TSS of reconstituted fortified pseudostem juice powders (one gram powder in 10 ml distilled water) were tested by using a digital hand refractometer. TSS of the fortified pseudostem juice powder varied from 15-19.3°B (Appendix E8). For treatments with high feed concentration showed a slight variation in TSS. This might be due to the increased solid content in sample. Sample with 50% milk and 30% horse gram extract (T_{12} -190°C) showed high TSS of 19.3°B.

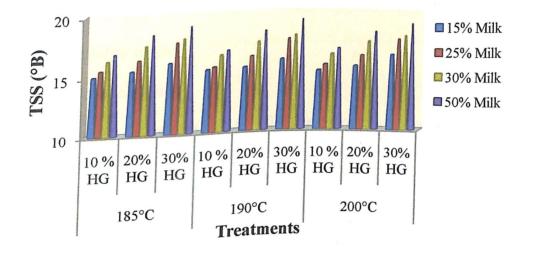


Fig. 4.38 Total soluble solids of Product-III

From Fig. 4.38, it is evident that spray drying temperature had no influence on the powder TSS. In the present study, TSS of fortified pseudostem juice powder was not significantly affected by concentration as well as by drying inlet air temperature.

4.5.1.1.6 Titrable acidity and pH

The titratable acidity of fortified pseudostem juice powder was found to be in range of 0.114-0.141% (Table 4.18). These results were analogous with those reported by Tracy et al. (1951). According to their study titrable acidity in whole milk powder was varied between 0.135 and 0.140%.

pH obtained for the reconstituted powders (one gram powder in 10 ml distilled water) were in range of 5.76-6.94 (Table 4.18). The results indicated that

with increase in milk concentration, pH value increased. High pH value of milk might be the reason for its increment in powder pH.

Fortified pseudostem powders developed from 50% milk and 30% horse gram extract combination exhibited high pH value of 6.94. The lowest pH value of 5.76 was observed in the treatment T1 (15% milk + 10% horse gram extract).

4.5.1.2 Reconstitution properties

- Solubility
- Wettability
- Water solubility index
- Water absorption index

Reconstitution properties of fortified pseudostem juice powders were tested for validating product quality and its consumer acceptability. Standard test procedures mentioned under chapter III were followed to determine reconstitution properties. The observations for each reconstitution properties listed above were examined for standardizing the best product. The range of values and reason behind its specific trend is explained in subsequent sections.

4.5.1.2.1 Solubility

Solubility is a key parameter to be evaluated for assuring the quality of final powder. The solubility of fortified pseudostem under different spray drying conditions was investigated. During the study, it was observed that the solubility varied with process conditions. The solubility value of fortified pseudostem juice powder was found to be in the range of 43.80 - 93% (Appendix E9). Among various treatments with high feed concentration (30% horse gram and 50% milk) scored less solubility value (Fig. 4.39). The solubility decreased with increase in feed concentration and temperature, this might be due to denaturation of protein at high temperature resulted in low solubility. Chegini and Taheri, (2013) observed that at high temperature, the solubility reduced in spray dried powder due to the formation of a rigid surface film above the powder particles. The formed film act

as barrier to water and therefore a larger fraction of powder remained as insoluble solids.

Fig. 4.39 shows the solubility variation of fortified pseudostem juice powder in different process conditions. From the graphical representation, it is evident that, a drastic reduction in solubility occurred from initial (T1) to final (T12) treatment. Higher solubility value of 93% was observed in T1 at lower temperature of 185°C and lower solubility value (43.8%) was found at 200°C for T12. Chudy et al. (2015) put forward the assumption that inappropriate production parameters may lead to reduction in solubility to below 50%. Statistical analysis showed its significance at 1% interval (p<0.0001) and the ANOVA related with it is given in Appendix M8.

4.5.1.2.2 Wettability

The wettability of fortified pseudostem juice powder under different spray drying inlet air temperatures and raw material concentrations was observed. From the results, wetting time of fortified pseudostem juice powder varied between 348 and 1290 s (Appendix E10). Kim et al. (2002) stated that good quality whole milk having a wetting time below 15 min (>900 s). From the experimental observations, fortified pseudostem juice powders of 185 and 190°C showed wetting time less than 900 s. The wetting time of fortified pseudostem juice powder values were in range at lower temperature and feed concentration. Effect of process parameters governing the wettability of fortified pseudostem juice powder was studied and depicted in Fig. 4.40.

The test results indicated a stated reduction in wettability with low inlet air temperature and raw material concentration. Moisture trend in powder occurred by different process parameters might be the reason for this result fluctuation. Apart from this, presence of free fat on droplet surface also contributed the wettability variation among trials (Chegini and Taheri, 2013 and Zbikowska and Zbikowski, 2006). Kim et al. (2002) propounds the view that deviation of wettability among treatments might be due to different particle size, surface area, density, porosity etc. Fair wettability value of 348 s and the unacceptable value of

1290 s were found in T1 (185°C) and T12 (200°C). Statistically the wettability values were significant on the process parameters and the ANOVA for the values were checked and tabulated in Appendix M9.

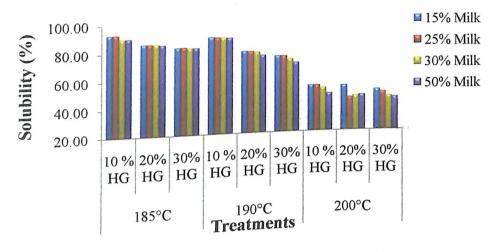


Fig. 4.39 Solubility of Product-III

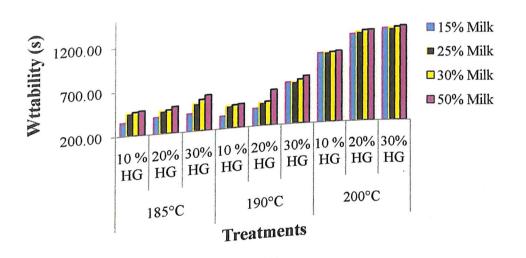


Fig. 4.40 Wettability of Product-II

4.5.1.2.3 Water solubility index (WSI)

Water solubility index of fortified pseudostem juice powder showed a slight variation among treatments. WSI values varied from 40.12% to 53.54% and are given in Table 4.19. Similar findings were observed by Jumah et al. according to them, as the inlet temperature increases the chance of formation of insoluble solids gets increased and resulted a higher WSI value. The result obtained for fortified pseudostem juice powder was comparable with the former study. At the higher temperature of 200°C maximum value of WSI (53.54%) was noted while a minimum of 40.12% was noted at 185°C.

4.5.1.2.4 Water absorption index (WAI)

WAI value of fortified pseudostem juice powder found to decrease from 93.35-88.24% (Table 4.19) with a decrease in inlet air temperature of 200°C. The obtained result agreed with the study of Phoungchandang *et al.* (2010) on ginger powder. At elevated temperature cohesion between particles gets reduced which led to less clustering and low water absorption index (Grabowski *et al.*, 2006 and Adhikari *et al.*, 2003). Low WAI value of 88.24 was recorded in T1 (200°C) treatment and the highest value of 93.35% was recorded in T12 (185°C).

| | Inlet air temperature (°C) | T1 | T2 | Т3 | T4 | T5 | Т6 | T7 | T 8 | Т9 | T10 | T11 | T12 |
|----------|----------------------------------|-------|-------|-------|-------|-------|-------|-----------|------------|-------|-------|-------|-------|
| Titrable | 185 | 0.140 | 0.132 | 0.113 | 0.138 | 0.140 | 0.124 | 0.133 | 0.131 | 0.141 | 0.128 | 0.140 | 0.140 |
| acidity | 190 | 0.134 | 0.133 | 0.129 | 0.135 | 0.121 | 0.128 | 0.131 | 0.121 | 0.132 | 0.130 | 0.143 | 0.130 |
| (%) | 200 | 0.134 | 0.121 | 0.132 | 0.114 | 0.113 | 0.114 | 0.118 | 0.131 | 0.135 | 0.129 | 0.131 | 0.119 |
| | 185 | 5.980 | 6.120 | 6.240 | 6.350 | 6.350 | 6.530 | 6.440 | 6.530 | 6.830 | 6.670 | 6.680 | 6.860 |
| pH | 190 | 5.760 | 6.140 | 6.220 | 6.320 | 6.330 | 6.550 | 6.440 | 6.550 | 6.860 | 6.670 | 6.680 | 6.940 |
| | 200 | 5.760 | 6.200 | 6.320 | 6.320 | 6.350 | 6.440 | 6.440 | 6.510 | 6.880 | 6.670 | 6.670 | 6.680 |

Table 4.18 pH and titrable acidity of Product-III

| | Inlet air temperature (°C) | T1 | T2 | T3 | T4 | T5 | Т6 | T7 | T8 | Т9 | T10 | T11 | T12 |
|-----|----------------------------------|--------|--------|--------|--------|--------|--------|-----------|--------|--------|--------|--------|------------------|
| WSI | 185 | 40.120 | 40.230 | 42.430 | 42.540 | 42.670 | 42.880 | 46.760 | 47.620 | 47.660 | 48.070 | 50.090 | 50.01/ |
| (%) | 190 | 40.180 | 40.250 | 42.530 | 42.540 | 42.690 | | | 47.770 | 47.670 | 48.500 | 50.090 | 50.84(51.43(|
| | 200 | 40.260 | 40.310 | 42.550 | 42.620 | 43.040 | 43.070 | 48.210 | 48.000 | 48.050 | 48.770 | 52.110 | 53.54(|
| WAI | 185 | 93.040 | 93.120 | 93.120 | 93.170 | 93.190 | 93.230 | 93.230 | 93.280 | 93.300 | 93.280 | 93.330 | 93.35(|
| (%) | 190 | 92.460 | 93.000 | 93.000 | 92.770 | 93.110 | 93.190 | 93.170 | 93.210 | 93.260 | 93.090 | 93.250 | 93.300 |
| | 200 | 88.240 | 88.240 | 90.000 | 89.250 | 90.030 | 89.950 | 89.460 | 90.090 | 89.950 | 90.020 | 90.110 | 90.05(|

Table 4.19 Water solubility index and water absorption index of Product-III

4.5.1.3 Flow properties

- Carr's index (CI) •
- Hausner ratio (HR)

Carr's index (CI) and Hausner ratio (HR) were the two key parameters that determining the flowability and cohesiveness characteristics of powder. The results showed that CI of fortified pseudostem juice powder varied from 1.15 to 1.26 and the value of HR ranges from 12.301 to 20.626 (Table 4. 20). Flowability and cohesiveness of powders depends on the size of the particles and also cohesive and frictional forces within the particles. Particle size depends mainly on atomization airflow (Wang et al., 2015). Besides the particle size, internal forces, flowability and cohesiveness is also affected by many other factors like moisture content, hardness, elasticity etc as reported by Chen et al. (2010). Therefore, it was difficult to point out the appropriate reason for the change in CI and HR values within treatments. With respect to standard table values of CI shown in Table 3.6 the obtained values of CI and HR values for spray dried pseudostem powders showed good and fair and poor flowability value for different treatments as given in Table 4.20.

4.5.1.4 Product yield

Product yield of fortified pseudostem juice found to be in the range of 10.63 to 79.25% (Table 4.20). It was found that product yield was affected significantly by wall material concentration and inlet air temperature. Product yield was taken as one of the important parameter for the product optimisation (Arslan et al., 2015 and Fazaeli et al., 2012). Dolinsky et al. (2000) hypothesized that higher inlet air temperature leads in a negative effect on product yield.

| t | Inlet air cemperature (°C) | T 1 | T2 | Т3 | T4 | T5 | Т6 | Т7 | Т8 | Т9 | T10 | T11 | T12 |
|---------|----------------------------------|------------|--------|--------|--------|--------|--------|--------|--------|--------|--------|--------|--------|
| | 185 | 1.230 | 1.200 | 1.200 | 1.200 | 1.180 | 1.220 | 1.180 | 1.180 | 1.250 | 1.180 | 1.170 | 1.260 |
| HR | 190 | 1.240 | 1.180 | 1.200 | 1.240 | 1.150 | 1.190 | 1.200 | 1.160 | 1.210 | 1.210 | 1.160 | 1.200 |
| | 200 | 1.250 | 1.190 | 1.160 | 1.230 | 1.240 | 1.180 | 1.230 | 1.140 | 1.181 | 1.220 | 1.150 | 1.190 |
| | 185 | 19.014 | 16.916 | 16.992 | 16.393 | 15.287 | 18.008 | 15.809 | 15.158 | 19.925 | 15.509 | 14.706 | 20.620 |
| CI (%) | 190 | 19.071 | 15.132 | 16.436 | 19.221 | 13.348 | 16.008 | 16.990 | 13.853 | 17.221 | 17.506 | 13.704 | 16.602 |
| | 200 | 20.155 | 16.284 | 13.836 | 18.734 | 19.256 | 15.226 | 18.546 | 12.301 | 15.133 | 18.069 | 13.288 | 15.968 |
| Product | 185 | 55.750 | 73.280 | 75.320 | 57.230 | 76.860 | 75.490 | 66.370 | 77.340 | 76.860 | 71.690 | 77.330 | 78.73(|
| yield | 190 | 55.480 | 73.280 | 75.280 | 60.720 | 76.360 | 77.230 | 69.080 | 74.230 | 78.380 | 71.590 | 72.820 | 79.25(|
| (%) | 200 | 10.630 | 19.890 | 33.490 | 18.150 | 23.850 | 46.620 | 19.870 | 25.760 | 46.820 | 21.300 | 29.750 | 47.050 |

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At elevated temperature powder melts and gets adhered to the drier wall there by resulting in a reduction in product yield. Product yield of 10.63% was found in T₁ treatment dried at 200°C and higher yield of 79.25% was achieved at 190°C (50% milk+30% horse gram extract+ 20% pseudostem juice). Statistical analysis showed a significant effect on yield at different process conditions (p<0.0001) and the ANOVA table with it is given in Appendix M10.

4.5.1.5 Microbiological stability

The microbial analysis was carried out for the standardised fortified pseudostem juice powder samples. Shelf life and storage stability of the product were predicted based on the microbial load. The results confirmed the absence of microbial colonies in spray dried powders (Appendix G3). Rajput *et al.* 2009 conducted microbal studies on 60 milk powder samples and quantified the toatal viable count as $3.1 \times 10^3 \pm 5 \times 10^2$ cfu/g.

4.5.1.6 Optimisation of process parameters

Optimisation of process variables especially inlet air temperature and raw material concentration were performed using the General composite design. Feed pump rpm and blower speed were already fixed based on preliminary trials. Statistical models were employed to determine numerical optimisation to fix the optimal process parameters for developing fortified banana pseudostem juice powders. In the present investigation, the independent variables were kept within the range and dependent variables were chosen as maximum and minimum. The experimental sample had the optimum process conditions of inlet air temperature (Table 4.20) and raw material concentration (30% horse gram extract and 50% milk+20% pseudostem juice).

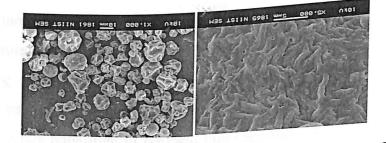
Optimised sample (T12) was kept for shelf life studies under ambient conditions. Treatment T12 developed at different inlet temperatures (185, 190 and 200°C) were taken to compare its storage period. Due to limited time period, product-III was analysed only three months during shelf life study.

| optimised Conditions | Process parameters | .oN .IS |
|----------------------|-----------------------------|---------|
| 0081 | Speed of the atomizer (rpm) | I |
| 58 I | Inlet air temperature (°C) | 7 |
| 74-92 | Outlet temperature (°C) | 4 |
| S I | Feed flow rate (rpm) | £ |

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4.5.1.7 Particle size

granules ranged in size from 2 to 18 µm ranged from <5 to 30 µm, while native tapioca starch had more homogeneous maltodextrin and reported that, modified tapioca starch had granules size spray-dried powers of modified tapioca starch, native tapioca starch, and 1000 resolutions respectively. Loksuwan, (2007) observed the particle size of earlier. Particle size of powder varied from 5-10 µm (Plate 4.7) at 5000 and Particle size of product was analysed by SEM analysis as mentioned



III-toubord to endergoroim MAS 7.4 otel9

sizylana olitorq laroniM 8.1.2.4

product and are listed below. done by absorption spectro-photometry and confirmed its presence in the final Mineral profile analysis for fortified banana pseudostem juice powder was

JOO

- Carbohydrate -85.31%
- Protein 3.80%
- Total ash- 4.10%
- Potassium 1.18%
- Calcium 2.88%
- Magnesium 0.31%

Feriotti and Igutti (2011) quantified pseudostem sap proximate composition *viz.* Total solid- 0.308%, Protein- 0.0141%, Lipid- 0.005%, Total sugar - 0.191%, Ash -0.104% and mineral composition of sap (*viz.* Sodium-88mg/l, Potassium - 874 mg/l, Calcium - 130 mg/l, Magnesium - 116 mg/l). Comparing the mineral analysis data obtained in this study for fortified banana pseudostem juice powder with the mineral content of fresh banana pseudostem sap, it is clear that the powder scored high in mineral content than fresh pulp. Thus for fortifying the pseudostem juice with horse gram extract and milk, the nutritious value of final powder could be significantly enhanced.

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4.5.1.9 Packaging of banana pseudostem juice powder

The optimised banana pseudostem juice powder samples were packed for shelf life studies in LDPE pouches and inserted in laminated aluminium pouches and was sealed hermetically using a hand sealing machine. Sealed packets were stored in ambient condition and tested for its quality attributes. Sharma *et al.*, (2003) studied storage studies of apple powder packed in aluminium pouches. From their six month storage study, the powder quality was found to remain unchanged.

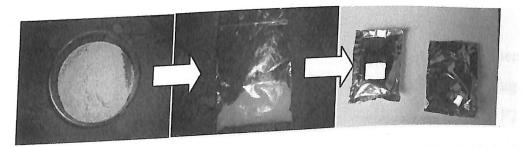


Plate 4.8 Packaged pseudostem juice powder samples in LDPE pouches and aluminium foil pouches

4.5.1.10 Storage stability

To consider the utility of spray-dried sample (fortified banana pseudo stem juice powder) for food industrial applications, it is important to assure its stability and quality during storage. In this context, the variation in different quality parameters during storage of three months was investigated and used as an indicator of product stability. Optimised samples of fortified pseudostem juice powder were stored under ambient conditions about three months. Shelf life study of experimental sample was done in every month to assess the fluctuations in powder characteristics. The data yielded during storage period were plotted in error graphs and its changes were discussed under this session.

4.5.1.10.1 Moisture content

The moisture content of the powder varied during storage period. Transfer of moisture to and from the product significantly influenced the stability of powder. A slight increase in moisture was noted during storage *i.e.*, 3.64-4.13% (Appendix F1). Cristina et al. (2008) also reported an increasing trend for moisture in whole milk powder.

Among three samples T12 at 185°C showed higher moisture absorption at 3rd month of storage. Moisture transfer might be due to the effect of storage conditions and permeability of packaging material. A significant change in moisture content was noted and was plotted, given below in Fig. 4.41. Statistical results revealed a significant effect of storage on moisture content (p<0.0001) and is given in Appendix N1.

4.5.1.10.2 Water activity

The change in water activity of packed fortified banana pseudostem samples was determined during three month storage study to verify the storage stability. Water activity of powder changed from 0.253 to 0.309 (Appendix F2) during storage. The shelf life study results showed that water activity of powder altered slightly during three month storage (Fig. 4.42).

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Pua *et al.* (2007) observed similar trend in water activity during the storage of jackfruit powders. Statistical analysis showed a significant effect on moisture content (p<0.0001) and ANOVA related with it is presented in Appendix N2.

4.5.1.10.3 Bulk density

Bulk density values showed a slight increase with storage life. Bulk density and moisture content showed an indirect relation. Increase in moisture during storage led to an increased bulk density (Fig. 4.43 and 4.44). Bulk density values such as tapped bulk density and loose bulk density of powder found to be increased during storage (Appendix F3 and F4). Powders with higher moisture value showed higher loose bulk density and tapped bulk density of values of 0.450 and 0.553 g/ml (T12-185°C) respectively in the 3^{rd} month. The increase in moisture content led to an increased bulk weight of product during storage. The increase in moisture might be due to the permeability of packaging material, variation in storage temperature *etc* (Pua *et al.*, 2007). Statistical study showed that bulk density values were significant (p> 0.0001) and the ANOVA related with it is given N3 and N4.

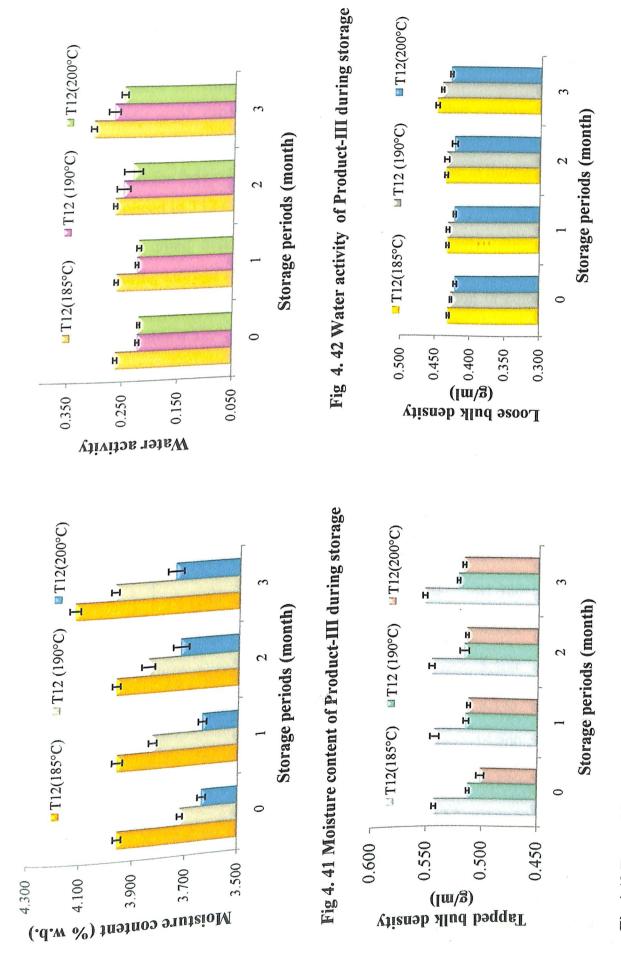




Fig 4. 44 Loose bulk density of Product-III during storage

4.5.1.10.4 Colour characteristics

Lightness L^* value of the spray dried powder reduced during storage period. Chudy *et al.* (2015) supported the prediction that enzymatic browning occurred due to Maillard reaction, lipid oxidiation, deterioration of ascorbic acid *etc* contributed for fade colour during storage. The decrease in lightness value was more pronounced in T12 (200°C) treatment at 2nd month (*i.e.*, 81.380).

It was noted that a^* (redness) and b^* (yellowness) values of powder increased during storage and T6 (200°C) exhibited the highest b^* and a^* values (8.147and 19.215) at the end of third month. The variation in colour values is illustrated in Table 4.22.

| Colour | Treatments | St | orage period | ls (month) | |
|-----------------|--|--------|--------------|------------|--------|
| characteristics | | 0 | 1 | 2 | 3 |
| | T6 (185°C) — | 90.030 | 90.300 | 90.000 | 90.000 |
| | T6 (190°C) | 88.000 | 88.000 | 87.810 | 87.871 |
| L* | T6 (200°C) | 81.490 | 81.410 | 81.380 | 81.380 |
| | T6 (185°C) | 4.940 | 4.930 | 4.935 | 4.915 |
| | | 5.500 | 5.500 | 5.500 | 5.504 |
| a* | T6 (190°C) | 8.140 | 8.140 | 8.146 | 8.147 |
| | T6 (200°C) | 15.190 | 15.200 | 15.200 | 15.213 |
| | T6 (185°C) | 18.820 | 18.830 | 18.830 | 18.850 |
| b* | T6 (^{190°C)} T6 (^{200°C)} | 19.210 | 19.210 | 19.251 | 19.251 |

| | Colour characteristics of Product-III during storage | |
|------------|--|--|
| Table 4.22 | Colour characteristic | |

The effect of drying method and storage on colour characteristics of fortified banana pseudostem juice powder were analysed statistically and

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confirmed the significance of L* and a* (p<0.0001) while b* showed non significance during storage. The ANOVA table related with colour characteristics of pseudostem juice powder is presented in appendix N5, N6 and N7.

4.5.1.10.5 pH and Titrable acidity

pH and titrable acidity values were analysed to investigate the extent of chemical reactions occurred during storage of fortified banana pseudo stem juice powder. The pH value of powders decreased after 3 months of storage and is given in Table 4.23. Verma et al. (2013) revealed that conversion of sugars into acids in powders led to increased acidic properties.

| | | Storage periods (month) | | | | | | | |
|----------------|-------------|-------------------------|-------|-------|-------|--|--|--|--|
| | Treatments | 0 | 1 | 2 | 3 | | | | |
| | T12 (185°C) | 6.860 | 6.860 | 6.841 | 6.840 | | | | |
| рН | T12 (190°С) | 6.940 | 6.940 | 6.940 | 6.939 | | | | |
| | T12 (200°C) | 6.680 | 6.680 | 6.680 | 6.680 | | | | |
| | T12 (185°C) | 0.140 | 0.140 | 0.140 | 0.136 | | | | |
| Titrable | T12 (190°C) | 0.130 | 0.130 | 0.130 | 0.128 | | | | |
| acidity (%) | T12 (200°C) | 0.119 | 0.119 | 0.119 | 0.118 | | | | |

Table 4.23 Change in pH and titrable acidity of product-III

4.5.1.10.6 Solubility and Wettability

Solubility and wettability of fortified banana pseudostem samples showed a significant effect on storage (Appendix F5 and F6). Supplee and Bellis et al. (1925) investigated the effect of storage on milk powders. Their study indicated a decrease in solubility during storage period (Fig. 4.45). It might be due to the increased moisture content of product. The observations on solubility of fortified banana pseudostem juice powder agreed with the results obtained in Product-I and II. Trend in wettability found to be contrary to the measured solubility. Wettability of powder increased with storage (Fig. 4.46), with similar reason as mentioned earlier for solubility (Sariga et al., 2015).

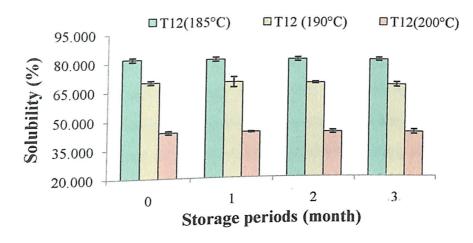


Fig. 4.45 Solubility of Product-III during storage

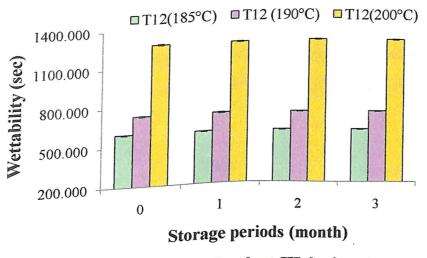


Fig. 4.46 Wettability of Product-III during storage

The effect of drying method and storage on solubility and wettability of fortified banana pseudostem juice powder were analysed statistically. Stastical studies reavealed that solubility and wettability of powders were non significant during storage (p<0.0001). The ANOVA table related with solubility and wettability characteristics of pseudostem juice powder is presented in appendix N8 and N9.

4.5.1.10.7 Microbial analysis

Stored samples were analysed for microbial load during storage period. Among stored samples T12 at 185°C showed comparatively higher colony count at the end of three month storage and it was 3×10^1 cfu/g. Sariga, (2015) conducted studies on spray dried vanilla extract and absence of microbial colony was reported in samples stored at ambient condition during six month periods. Details of microbial analysis are given in Appendix G4.

4.5.1.11 Sensory analysis

Sensory analysis is an important parameter for consumer acceptability. The products were analysed by 12 semi – trained sensory panelists. The optimised pseudostem juice powder was reconstituted with normal water in 1:10 ratio and sugar was added during sensory analysis. The fresh pseudostem juice with freshly prepared horse gram extract and pasteurized milk with sugar was taken as control. The results of the sensory evaluation of product are given Fig. 4.47.

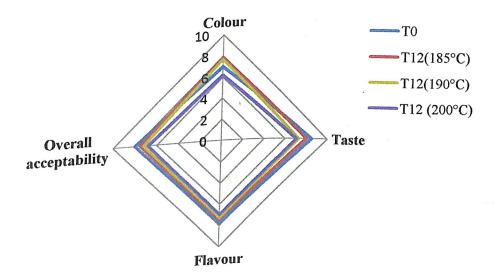


Fig. 4.47 Sensory score obtained for Product-III

Treatment T12 (185°C) scored highest value during sensory analysis. Control sample scored the best score of 8. Comparing with control treatment T12 (185°C) maintained similar sensory quality. Apart from this T12 (185°C) (185°C) maintained product stability during three months storage. Thus T12 (a treatment combination of banana pseudostem juice 20% and horse gram extract 30% with 50% milk at an inlet air temperature of 185°C) was optimised as the best treatment.

4.6 Cost analysis of pseudostem based powders

The computation of production cost for 1 kg and 50 g packet of spray dried pseudostem based powders were estimated by considering the variable cost and fixed cost of production and is given in Appendix N1. The cost of the production of 1 kg and 50 g banana pseudostem juice-sugar mix power was estimated to be Rs. 195/- and Rs. 9.75/-, respectively. The cost of powder produced from banana pseudostem juice and horse gram extract blend was calculated as Rs. 208/- for 1 kg pack and Rs. 10.40/- for its 50 g packs and for milk fortified powder, it was Rs. 243/- and Rs.12.15 /- respectively.

Summary and

Conclusion

CHAPTER V

SUMMARY AND CONCLUSION

India has a large area of banana cultivation and its fruit and other parts are utilized as a low-cost food source. Generally, a huge biomass is produced during the post-harvest banana cultivation. A major share of this biomass includes a fibrous soft stem termed as banana pseudostem. Its massive accumulation and disposal lead to unhygienic surroundings and environmental pollution. Safe disposal of banana residue has become an unsolved issue in banana cultivation. Scientific studies revealed that banana pseudostem is rich in minerals like K, Mg, Na etc. Minerals present in banana pseudostem have the ability to dissolve preformed kidney stones and also have anti-oxidant activities. In this context, utilisation and value addition of pseudostem will provide an additional income to banana growers. Though it has got high medicinal value, its usage is mostly constrained to non food applications like craft products. High perishability and its astringent tangy mouth feel might be the reason for its restricted application in food industry. Considering the above cited facts, a study was undertaken to obtain powdered product from pseudostem juice.

The intention of the study was to develop a process protocol for microencapsulated banana pseudostem juice powder, standardisation of the spray drying parameters for its development and quality analysis of fresh and stored product. Spray drying facilitates retention of product quality and preservation of heat sensitive components present in pseudostem juice. Microencapsulated banana pseudostem juice with suitable wall material could be suggested as a healthy ready to drink mix.

Three powder products were developed from banana pseudostem juice by spray drying technology. Banana pseudostem (Cv. Palayankodan) was collected from KCAET farm and pre-treated to arrest enzymatic activity. The pseudostem slices were dipped in 0.3% citric acid and blanched at 100°C for one minute to slices were dipped in 0.3% citric acid and catalase test were done for checking the inactivate enzymes. Peroxidase and catalase test were done for checking the effectiveness of blanching. The major constituents of feed combination include; banana pseudostem juice. sugar. horse gram extract and milk. Product-I comprised of pseudostem juice-sugar combination flavoured with ginger. Product-II consists of a blend of banana pseudostem juice and horse gram with ginger extract. However the third product, banana pseudostem juice was fortified with horse gram extract and milk flavoured cardamom.

Process parameters such as raw material concentration, wall material selection and drying conditions such as blower speed, feed speed and inlet air temperature combinations were fixed according to the preliminary trials. Blower speed of 1800 rpm and feed speed of 15 rpm were kept constant throughout the study. A process temperature of 180, 185 and 190°C were chosen for the development of product-I and II. However for product-III, the temperature selected was 185, 190 and 200°C. Maltodextrin and corn starch was selected as encapsulating agents for product-I and II, subsequently corn starch was omitted as it gave low yield during preliminary trails.

As part of quality evaluation of fresh product, physicochemical properties, reconstitution properties, and flow properties were analysed for the spray dried powder. Physicochemical properties of products such as moisture content, water activity, bulk density, TSS, titrable acidity, pH and colour characteristics were assessed by standard procedures. The reconstitution properties like solubility, wettability, water solubility index and water absorption index were done by standard laboratory methods and flow properties such as Carr's index and Hausner ratio were calculated using standard formulae.

The experimental results revealed that spray drying conditions and raw material concentrations significantly affected the powder characteristics. Moisture content and water activity of all the three powders decreased at higher inlet air temperature and feed concentration. For Product-I moisture values ranged between 2.99-4.6% (w.b) and for Product-II it was 3.93-4.52%. In both products-between 2.99-4.6% (w.b) and for Product-II it was 3.93-4.52%. In both products-between and II temperature as well as wall material concentration influenced the moisture I and II temperature as well as wall material showed a moisture range from 3.64 reduction. Product-III, milk fortified sample showed the same decreasing trend for to 4.5% (w.b). Water activity values also followed the same decreasing trend for

three products I. II and III and were found to be in the range of 0.295-0.430, 0.314 -0.407and 0.221-0.376. respectively.

Bulk density. an important parameter of powders found to decrease at elevated temperatures. The tapped and loose bulk densities of three products were assessed using standard laboratory procedures. Lower tapped bulk density of 0.410, 0.416 and 0.387g/ml were observed for the three products at low moisture conditions. Higher loose bulk density values noted at lower temperatures of 180 and 185°C and the highest loose bulk density value of 0.577 g/ml for product-II in was observed in Product-II at low temperature.

Total soluble solid (TSS) is an indicator of solids present in product and varied with raw material concentration. The TSS of pseudostem based products were 16, 24 and 19.3°B respectively, in which product-II exhibited the highest value (24°B) due to its high feed concentration contributed by maltodextrin (25%) and horse gram extract (30%).

Analysis of acidic properties revealed that pH and titrable acidity values varied from product to product. Product-I and II exhibited a low pH and higher acidity values compared to product-III, due to the addition of ginger extract. However, product-III showed a high pH value (5.76-6.94) and low titrable acidity (0.114-0.141%) due to the addition of milk.

Colour characteristics of powder having great importance in consumer acceptability and market value. During the study it varied with inlet temperatures and the same was reflected significantly in product-III. The lowest L* value of 67 and highest b* value of 24.71 were observed at a higher inlet temperature of and highest b* value of 24.71 were observed at a higher L* values within a range 200°C for product-III. Other two products showed higher L* values within a range of 77.50 to 94.52. The redness a* was not appreciable in products except product-III, which exhibited a higher value of 10.63 at 200°C.

Reconstitution properties such as wettability, water absorption index and water solubility index increased with increase in inlet air temperature. At the same time solubility showed a decreasing trend towards increasing temperature. Highest solubility was found in product-II (97.48-99.20%) and lowest was in product-I (76.55-85%). Product-III showed a moderate solubility of 43.80-93% and wettability of 348-1290 s.

Considering the flow properties such as Carr's index and Hausner ratio product-I showed good and fair flowability, Product-II exhibited excellent and fair and for product-III flow properties were rated good.

In terms of yield, all the three products were good and showed a product yield range of 28.60 to 56.70, 47.72 to 78.20, and 10.60 to 79.20% for product-I, II and III respectively. High TSS contributed to a higher yield in case of product-II. Higher temperature resulted in a lowest yield of 10.6% in product-III.

Mineral profile analysis ensures and quantifies the nutritional quality of the developed product. An increased nutritional profile was noted from product-I to III. Presence of potassium which is the key ingredient that reduces the kidney stone formation was existed in all the three products and was found to be maximum in product-III (1.88%). Presence of magnesium was confirmed in all the three products and was high in product-II and III (*i.e.*, 0.31%). Fortification of pseudostem juice resulted in a highly nutritious health drink powder. Addition of horse gram increased the carbohydrate content whereas milk enriched the powder with calcium and protein. Among the three products, product-III showed high calcium and protein content (Ca- 2.15% and protein- 3.03%).

Particle size of the standard samples were analysed and found to be in the range of 10-50 μ m for product I and II. and 5-10 μ m for product III.

Storage study of standardised samples (Product-I and II) was done for six and three month periods (Product-III). For product I and II storage studies were done for six months under ambient storage conditions. Product III stored only upto three months due to limited time. Shelf life studies of sample confirmed that no appreciable changes occurred in quality and stability of powder during storage.

Moisture absorption and chemical reactions were observed during storage. This led to variation in physicochemical properties and reconstitution properties. Moisture absorption increased the moisture content and water activity of powder, resulting in increased bulking weight of powder. Decreased pH and increased titrable acidity due to chemical reactions were also noted during storage. Degradation in colour characteristics was noted in all samples and it was more pronounced in product III. Reduced solubility and increased wettability values were the changes observed in reconstitution properties during storage period.

Microbial load in all the three products were found to be within the admissible limit and the products were safe for consumption.

Standardisation of best product from each combination was done by sensory analysis. Reconstituted powders along with their fresh feed samples were kept for sensory evaluation conducted with 12 semi trained panel members. Colour, taste, flavour and overall acceptability of reconstituted powder were judged and recorded in a 9- point Hedonic scale. From the sensory evaluation and quality analysis the best samples were selected from each combination. From the banana pseudostem juice - sugar combinations, trial T6 (15% sugar + 25% MD + 56% pseudostem juice) spray dried at 180°C scored high in quality evaluation, which was supported by statistical analysis and sensory analysis. In case of experiment-II. treatment T6 (25% maltodextrin + 30% horse gram extract + 43% pseudostem juice) at an inlet temperature 180°C showed he best result. However in third combination *i.e.*, pseudostem juice-horse gram extract blended with milk exhibited the best result at an inlet temperature of 185°C for the treatment T12 (50% milk + 30% horse gram extract + 20% pseudostem juice), based on the statistical and sensory analysis.

Cost analysis of the products were estimated by considering the variable and fixed cost of production. Cost of production for one kilogram and 50 g packets were estimated for all the three optimised product combinations. The production cost for one kilogram of product-I, II, and III was Rs.195/-, Rs.208/and Rs.243/- respectively. The study revealed that the production of banana pseudostem juice based powder has good nutritional value and is economically viable. The adoption of microencapsulation technology using spray drying to produce pseudostem juice powder will be a promising technology and it will help banana growers, and banana based food industries. Due to its high nutritional value and appreciable price, the pseudostem juice powder has good scope to fetch high market price and will be preferred by all health conscious consumers.

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CHAPTER VII

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<u>Appendices</u>

APPENDIX A

QUALITY PARAMETERS OF SPRAY DRIED PSEUDOSTEM JUICE POWDER (Product-I)

| | | Table A | A1 Moist | ure conte | nt | | |
|-------------------------------------|----------------------------------|-------------------------|-------------------------|-------------------------|-------------------------|-------------------------|-------------------------|
| | Inlet air temperature (°C) | T1 | T2 | Т3 | T 4 | Т5 | Т6 |
| Moisture content (% w.b.) | | 3.890 3.710 3.500 | 4.600 4.040 3.930 | 3.350 3.330 3.270 | 3.480 3.430 3.360 | 3.080 3.000 2.990 | 3.240 3.200 3.100 |
| | | Table | A2 Wate | r activity | | | |
| | Inlet air temperature (°C) | T1 | T2 | Т3 | T 4 | Т5 | T6 |
| Water activity | 180 185 190 | 0.430 0.403 0.400 | 0.387 0.380 0.360 | 0.355 0.340 0.338 | 0.325 0.322 0.324 | 0.320 0.315 0.312 | 0.313 0.300 0.295 |
| | Т | able A3 | Tapped b | oulk dens | ity | | |
| | Inlet air temperature (°C) | T1 | T2 | T3 | T4 | Т5 | Τ6 |
| Tapped bulk density (g/ml) | 180 185 190 | 0.495 0.483 0.477 | 0.510 0.509 0.480 | 0.440 0.439 0.435 | 0.464 0.448 0.441 | 0.419 0.412 0.410 | 0.431 0.430 0.412 |
| (g/m) | 7 | able A4 | Loose bu | ılk densit | y | | |
| | Inlet air temperature (°C) | T1 | Т2 | T3 | T4 | T5 | Т6 |
| Loose bulk density | 180 185 190 | 0.357 0.354 0.344 | 0.457 0.369 0.363 | 0.319 0.313 0.310 | 0.343 0.336 0.329 | 0.300 0.281 0.258 | 0.306 0.304 0.303 |
| (g/ml) | 190 | | | | | | |

| | Ta | able A5 To | tal solub | le solids (| TSS) | | |
|-------------|----------------------------------|----------------|------------|-------------|------------|--------|-----------|
| | Inlet air temperature (°C) | T1 | T2 | Т3 | T4 | Т5 | Т6 |
| | 180 | 10.800 | 11.400 | 12.540 | 14.600 | 15.000 | 15.890 |
| | 185 | 10.000 | 12.450 | 13.900 | 14.650 | 15.000 | 16.100 |
| TSS (°B) | 190 | 11.020 | 12.500 | 14.500 | 14.700 | 15.200 | 16.000 |
| | | Table | e A6 Solı | ıbility | .,, | | |
| | Inlet air temperature (°C) | e T1 | T2 | T3 | T4 | Т5 | Т6 |
| | 180 | 85.000 | 83.010 | 80.100 | 81.500 | 79.400 | 76.650 |
| Solubility | 185 | 84.120 | 81.802 | 79.900 | 81.300 | 79.400 | 76.650 |
| (%) | 190 | 83.700 | 82.900 | 79.800 | 81.200 | 78.600 | 76.530 |
| | | Table | A7 Wett | ability | | | |
| | Inlet air temperatur | re T1 | T2 | Т3 | T 4 | T5 | T6 |
| | (°C) | 73.050 | 75.430 | 77.900 | 78.000 | 79.100 | 80.000 |
| Wettability | 180 | 74.100 | | 77.900 | 78.060 | 79.760 | 80.040 |
| (s) | 185 190 | 75.100 | | 77.900 | 79.000 | 80.000 | 80.060 |
| | Table | e A8 Wate | r solubili | ty index | (WSI) | | |
| | Inlet air temperature | T1 | T2 | T3 | T4 | T5 | Т6 |
| | (°C) | 33.400 | 35.800 | 39.540 | 41.870 | 43.870 | 48.000 |
| | 180 | | 37.120 | 39.760 | 42.140 | 44.870 | 55.100 |
| WSI (%) | 185 190 | | 39.000 | 40.130 | 43.000 | 46.700 | 57.030 |
| | | A9 Water | absorpti | on index | (WAI) | | |
| t | Inlet air temperature | T1 | T2 | Т3 | T4 | Т5 | Т6 |
| | (°C) | 99.450 | 98.300 | 96.300 | 94.800 | 92.900 | 90.800 |
| | 180 | <i>y)</i> .138 | | 95.840 | 93.400 | 92.400 | 90.540 |
| WAI | 185 | 90.500 | | 96.000 | 94.620 | 92.100 | 90.500 |
| (%) | 190 | 90.500 | | | | | |

APPENDIX B

STORAGE STUDY OF SPRAY DRIED BANANA PSEUDOSTEM JUICE **POWDER (Product-I)**

| | Μ | loisture conte | nt (% w.b.) | | | | |
|--------------------------|--------------------------|----------------|-------------|-------|--|--|--|
| Treatments | Storage periods (months) | | | | | | |
| | 0 | 2 | 4 | 6 | | | |
| m((1909C) | 3.240 | 3.287 | 3.320 | 3.470 | | | |
| T6 (180°C) | 3.200 | 3.219 | 3.220 | 3.300 | | | |
| T6 (185°C) T6 (190°C) | 3.100 | 3.100 | 3.150 | 3.290 | | | |

Table B1 Changes in moisture content of Product-I during storage

| Table B2 Changes in water activity | of Product-I during storage |
|------------------------------------|-----------------------------|
| Table D2 Chang | |

| Water activity Storage periods (months) | | | | | |
|--|-------------------------------------|---|--|--|--|
| | | | | | |
| | 0.322 | 0.338 | 0.345 | | |
| - | 0.320 | 0.332 | 0.339 | | |
| - | 0.302 | 0.311 | 0.326 | | |
| 0.275 | | | | | |
| | 0 0.313 0.300 0.295 | Storage periods 0 2 0.313 0.322 0.300 0.320 0.202 0.302 | Storage periods (months) 0 2 4 0.313 0.322 0.338 0.300 0.320 0.332 | | |

Table B3 Changes in tapped bulk density of Product-I during storage

| | T | apped bulk de | nsity (g/ml) | • | | | |
|--------------|-------|--------------------------|--------------|-------|--|--|--|
| Treatments — | | Storage periods (months) | | | | | |
| Ireatmente | 0 | 2 | 4 | 6 | | | |
| | 0.431 | 0.431 | 0.432 | 0.443 | | | |
| T6 (180°C) | 0.430 | 0.432 | 0.433 | 0.442 | | | |
| T6 (185°C) | 0.412 | 0.413 | 0.417 | 0.420 | | | |
| T6 (200°C) | 0.412 | | | | | | |

| Loose bulk density (g/ml) Storage periods (months) | | | | | |
|---|----------------|---|--|--|--|
| | | | | | |
| 0.306 | 0.313 | 0.326 | 0.356 | | |
| 0.304 | 0.308 | 0.314 | 0.316 | | |
| 0.303 | 0.303 | 0.308 | 0.317 | | |
| | 0.306 0.304 | Storage periods 0 2 0.306 0.313 0.304 0.308 | Storage periods (months) 0 2 4 0.306 0.313 0.326 0.304 0.308 0.314 | | |

Table B4 Changes in loose bulk density of Product-I during storage

Table B5 Changes in solubility of Product-I during storage

| | Solubility (%) Storage periods (months) | | | | | |
|--------------|--|--------|--------|--------|--|--|
| Treatments – | | | | | | |
| | 0 | 2 | 4 | 6 | | |
| | 79.400 | 78.970 | 78.320 | 77.541 | | |
| T6 (180°C) | 79.400 | 78.650 | 78.206 | 77.198 | | |
| T6 (185°C) | 78.600 | 78.423 | 78.223 | 77.127 | | |
| T6 (190°C) | 78.000 | | | | | |

Table B6 Changes in wettability of Product-I during storage

| | Wettability (s) | | | | | |
|--------------|-----------------|-----------------|------------|--------|--|--|
| Treatments – | | Storage periods | s (months) | | | |
| Ireatments | 0 | 2 | 4 | 6 | | |
| | 80.000 | 80.090 | 80.120 | 80.390 | | |
| T6 (180°C) | 80.000 | 80.160 | 80.390 | 80.500 | | |
| т6 (185°С) | 80.040 | 80.460 | 80.570 | 80.630 | | |
| T6 (190°C) | 80.000 | | | | | |

APPENDIX C

QUALITY PARAMETERS OF SPRAY DRIED BANANA PSEUDO STEM JUICE-HORSE GRAM EXTRACT BLEND POWDER (Product-II)

| | Table | C1 Mois | ture cont | ent of Pro | oduct-II | | |
|-------------------------------------|----------------------------------|-------------------------|-------------------------|-------------------------|-------------------------|-------------------------|-------------------------|
| | Inlet air temperature | T1 | T2 | Т3 | T4 . | T5 | T 6 |
| Moisture content (%) | (°C) 180 185 190 | 4.523 4.460 4.231 | 4.510 4.442 4.211 | 4.230 4.140 4.040 | 4.350 4.240 4.160 | 4.333 4.210 4.100 | 4.210 4.110 3.930 |
| | Table | e C2 Wat | er activit | y of Prod | uct-II | | |
| | Inlet air temperature | T1 | T2 | Т3 | T4 | Т5 | Т6 |
| Water activity | (°C) 180 185 190 | 0.407 0.402 0.372 | 0.401 0.400 0.363 | 0.387 0.371 0.353 | 0.387 0.374 0.354 | 0.364 0.348 0.324 | 0.352 0.335 0.314 |
| | Table C3 | 3 Tapped | bulk der | isity of Pi | roduct-II | | |
| | Inlet air temperature (°C) | T1 | T2 | T3 | T4 | T5 | Т6 |
| Tapped bulk density (g/ml) | 180 185 190 | 0.577 0.567 0.561 | 0.551 0.539 0.504 | 0.490 0.480 0.478 | 0.470 0.464 0.460 | 0.451 0.451 0.450 | 0.421 0.419 0.416 |

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| | Inlet air temperature (°C) | T1 | T2 | Т3 | T4 | T5 | Т6 |
|-------------------|----------------------------------|-------|-------|-------|-------|-------|-------|
| Loose | 180 | 0.474 | 0.432 | 0.413 | 0.453 | 0.421 | 0.420 |
| bulk | 185 | 0.384 | 0.375 | 0.364 | 0.359 | 0.342 | 0.341 |
| density (g/ml) | 190 | 0.338 | 0.332 | 0.327 | 0.326 | 0.322 | 0.316 |

Table C4 Loose bulk density of Product-II

Table C5 Total soluble solid of Product-II

| <u> </u> | Inlet air temperature (°B) | T1 | T2 | T3 | T4 | Т5 | T6 |
|----------|----------------------------------|-----------|--------|--------|-----------|--------|-----------|
| | 180 | 14.000 | 17.000 | 19.500 | 21.000 | 22.000 | 23.000 |
| TSS | 185 | 16.150 | 18.200 | 20.000 | 22.000 | 22.000 | 24.000 |
| 100 | 190 | 17.300 | 19.000 | 20.000 | 22.000 | 23.000 | 23.500 |
| | | | | | | | |

Table C6 Solubility of Product-II

| | Inlet air temperature (°C) | T1 | T2 | T3 | · T4 | Т5 | Т6 |
|------------|----------------------------------|------------------|------------------|------------------|------------------|------------------|------------------|
| Solubility | 180 | 99.200 99.020 | 98.910 98.907 | 98.690 98.739 | 98.910 98.670 | 97.840 97.760 | 97.770 97.720 |
| (%) | 190 | 98.900 | 98.250 | 98.200 | 97.980 | 97.540 | 97.480 |

| | Inlet air temperature (°C) | T1 | T2 | Т3 | T 4 | Т5 | Т6 |
|-------------|----------------------------------|-----------|---------|---------|------------|---------|---------|
| | 180 | 481.783 | 492.600 | 502.367 | 508.667 | 535.833 | 563.300 |
| Wettability | 185 | 481.897 | 492.767 | 503.350 | 515.833 | 539.967 | 564.027 |
| (s) | 190 | 489.060 | 497.050 | 505.167 | 530.100 | 554.967 | 563.867 |
| | | | | , | • 1 | | |

Table C7 Wettability of Product-II

Table C8 Water solubility index of Product-II

| | Inlet air temperature (°C) | T1 | T2 | Т3 | T 4 | Т5 | T 6 |
|------------|----------------------------------|-----------|--------|--------|------------|--------|------------|
| Water | 180 | 32.810 | 43.833 | 46.007 | 49.640 | 52.467 | 54.853 |
| solubility | 185 | 34.267 | 44.500 | 47.047 | 51.427 | 53.403 | 55.466 |
| index | 190 | 42.427 | 45.037 | 48.377 | 51.933 | 53.910 | 60.467 |
| (%) | | | | | | | |

Table C9 Water absorption index of Product-II

| | Inlet air temperature (°C) | T1 | T2 | Т3 | . T 4 | Т5 | T6 |
|------------|----------------------------------|------------------|------------------|-------------------|------------------|------------------|------------------|
| Water | 180 | 97.500 | | 99.800 100.000 | 94.620 94.800 | 95.600 96.210 | 96.400 |
| absorption | 185 190 | 98.300 98.300 | 99.390 99.500 | 102.000 | 94.800 95.340 | 96.210 96.300 | 96.550 97.120 |
| index (%) | | | | | | | |

APPENDIX D

STORAGE STUDY OF SPRAY DRIED BANANA PSEUDO STEM JUICE-HORSE GRAM EXTRACT BLEND POWDER (Product-II)

| | Ν | loisture conte | nt (% w.b.) | |
|--------------------------|-------|----------------|--------------------|-------|
| Treatments | S | torage period | ls (months) | |
| | 0 | 2 | 4 | 6 |
| T((1000C)) | 4.210 | 4.240 | 4.244 [.] | 4.311 |
| T6 (180°C) | 4.110 | 4.209 | 4.220 | 4.319 |
| T6 (185°C) T6 (190°C) | 3.930 | 4.000 | 4.110 | 4.122 |

Table D1 Changes in moisture content of Product-II during storage

Table D2 Changes in water activity of Product-II during storage

| Treatments | | Moisture o | content (%) | |
|--|-------|--------------|--------------|-------|
| | | Storage peri | ods (months) | |
| | 0 | 2 | 4 | 6 |
| | 0.352 | 0.352 | 0.353 | 0.355 |
| T6 (180°C) | 0.335 | 0.342 | 0.348 | 0.354 |
| T6 (180°C) T6 (185°C) T6 (190°C) | 0.314 | 0.319 | 0.322 | 0.324 |

Table D3 Changes in tapped bulk density of Product-II during storage

| Treatments | | Tapped bulk | density (g/ml) | |
|--|-------|--------------|----------------|-------|
| | | Storage peri | ods (months) | |
| | 0 | 2 | 4 | 6 |
| | 0.421 | 0.422 | 0.435 | 0.438 |
| ть (180°С) | • | 0.421 | 0.425 | 0.436 |
| T6 (180°C) T6 (185°C) T6 (190°C) | 0.419 | 0.419 | 0.422 | 0.425 |
| 10(100%) | 0.416 | 01111 | | 0 |

Table D4 Changes in loose bulk density of Product-II during storage

| Treatments | | Loose buik C Storage peri | lensity (g/ml) ods (months) | |
|--------------------------|-------|------------------------------|--------------------------------|-------|
| | | 2 | 4 | 6 |
| | 0 | 0.422 | 0.424 | 0:429 |
| | 0.420 | 0.342 | 0.356 | 0.409 |
| T6 (180°C) | 0.341 | 0.317 | 0.335 | 0.347 |
| T6 (185°C) T6 (190°C) | 0.316 | 0.517 | | |

APPENDIX E

QUALITY PARAMETERS OF MILK FORTIFIED BANANA PSEUDOSTEM JUICE POWDER (Product-III)

| | Inlet air temperature (°C) | T 1 | T2 | T3 | Τ4 | Т5 | Т6 | T7 | Т8 | Т9 | T10 | T11 | T12 |
|-------------------|----------------------------------|--------------------|--------------------|-------------|----------------|--------------------|------------------|------------|--------------------|-----------|---------------------|---------------------|------------------------------|
| Moisture | 185 | 4.500 | 4.480 | 4.363 | 4.481 | 4.463 | 4.222 | 4.475 | 4.320 | 4.183 | 4.450 | 4.300 | 3,960 |
| content | 190 | 4.473 | 4.476 | 4.357 | 4.472 | 4.400 | 4.180 | 4.460 | 4.313 | 4.150 | 4.400 | 3.870 | 3.720 |
| (%) | 200 | 4.473 | 4.452 | 4.440 | 4.455 | 4.000 | 3.880 | 4.400 | 4.110 | 3.880 | 3.750 | 3.711 | 3.643 |
| | | | | Table | E2 XV-4 | | | | | | | | |
| | | | | Table | E2 Water | · activity o | of Product | -III | | | | | |
| | Inlet air temperature | T1 | T2 | | E2 Water T4 | | | | TQ | | T10 | | |
| | | T1 | T2 | Table T3 | E2 Water T4 | • activity o T5 | of Product T6 | -III T7 | Т8 | T9 | T10 | T11 | T12 |
| | temperature | T1 0.376 | T2 0.360 | | | | | | T8 0.340 | T9 | T10 0.332 | T11 0.312 | |
| Water activity | temperature (°C) | | | Т3 | T 4 | Т5 | Т6 | Τ7 | | • | | | T12 0.263 0.223 |

Table E1 Mosture content of Product-III

| | Inlet air temperature (°C) | T1 | T2 | T3 | T4 | T5 | Т6 | T 7 | Т8 | Т9 | T10 | T11 | T12 |
|-------------------------------------|----------------------------------|-------------------------|-------------------------|-------------------------|-------------------------|-------------------------|-------------------------|-------------------------|-------------------------|-------------------------|-------------------------|-------------------------|-------------------------|
| Tapped bulk density (g/ml) | 185 190 200 | 0.426 0.409 0.387 | 0.467 0.456 0.436 | 0.512 0.505 0.477 | 0.427 0.411 0.395 | 0.471 0.457 0.457 | 0.522 0.506 0.486 | 0.432 0.412 0.399 | 0.475 0.462 0.439 | 0.532 0.511 0.489 | 0.432 0.417 0.404 | 0.476 0.467 0.444 | 0.543 0.512 0.501 |

Table E3 Tapped bulk density of Product-III

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Table E4 Loose bulk density of Product-III

| | Inlet air temperature (°C) | T1 | T2 | T3 | T4 | Τ5 | Т6 | T7 | Τ8 | Т9 | T10 | T11 | T12 |
|------------------------------------|----------------------------------|-------------------------|-------------------------|-------------------------|-------------------------|-------------------------|-------------------------|-------------------------|-------------------------|-------------------------|-------------------------|-------------------------|-------------------------|
| Loose bulk density (g/ml) | 185 190 200 | 0.345 0.331 0.309 | 0.467 0.456 0.457 | 0.512 0.505 0.477 | 0.357 0.332 0.321 | 0.399 0.396 0.369 | 0.428 0.425 0.412 | 0.365 0.342 0.325 | 0.403 0.398 0.385 | 0.426 0.423 0.415 | 0.365 0.344 0.331 | 0.406 0.403 0.385 | 0.431 0.427 0.421 |

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| | Inlet air temperature | T1 | T2 | Т3 | T4 | Т5 | Т6 | Τ7 | Т8 | Т9 | T10 | | |
|-----------|-----------------------------------|----------------|----------|--------|---------------------|-----------------------------|-----------|--------------|-------------------------------|-------------------------------|--------------------------------|--------------------------------|--------------------------------|
| | (°C) | | | | | | | 1, | 10 | 19 | T10 | T11 | T12 |
| | 185 | 85.460 | 87.540 | 88.600 | 86.460 | 87.890 | 88.980 | 86.850 | 88.010 | 89.990 | 87.430 | 88.240 | 90.030 |
| 」* values | 190 | 84.540 | 85.460 | 84.990 | 84.650 | 85.870 | 85.870 | 84.760 | 85.680 | 86.560 | 85.440 | 86.900 | |
| | 200 | 67.000 | 77.800 | 80.350 | 74 000 | 05.070 | | | | 00.500 | 05.440 | 80.900 | 88.000 |
| | | | 77.000 | | 74.000 | 85.870 | 81.200 | 76.000 | 80.430 | 81.200 | 77.100 | 81.070 | 81.490 |
| | | | | | | | | | | | | | |
| | Inlet air | | | | Table E6 | b [*] values | of Produc | et-III | | | | | |
| | temperature | e T1 | T2 | Т3 | Table E6 | b [*] values T5 | of Produc | et-III T7 | T8 | T9 | T10 | T11 | T12 |
| | temperature (°C) | | | | T4 | | | | T8 | T9 | T10 | T11 | T12 |
| | temperature (°C) 185 | e T1 19.44(| | | T4 | | | Τ7 | | | | | |
| b* value: | temperature (°C) 185 190 | |) 19.370 | 18.280 | T4 19.430 | Т5 | T6 | T7 | T8 16.320 17.330 | T9 17.140 18.220 | T10 16.400 18.450 | T11 15.780 17.220 | T12 15.190 18.820 |

 Table E5 L^{*} values of Product-III

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| | Inlet air temperature (°C) | Tl | Т2 | Т3 | T4 | Τ5 | Т6 | Τ7 | Т8 | Т9 | T10 | T11 | T12 |
|-----------|----------------------------------|---------|--------|-------|-----------|-------|---------|-------|--------|-------|-------|-------|--------|
| | 185 | 7.510 | 7.220 | 7.090 | 7.300 | 7.200 | 6.160 | 6.830 | 7.010 | 5.240 | 5 420 | 5.170 | |
| a* values | 190 | 7 5 4 0 | 7 2 (0 | 7 000 | | | | 0.050 | 7.010 | 5.240 | 5.430 | 5.170 | 4.94() |
| a values | 190 | 7.540 | 7.260 | 7.090 | 6.860 | 7.140 | 7.130 | 6.470 | 6.950 | 5.710 | 6.350 | 6.300 | 5.500 |
| | 200 | 10.630 | 9.650 | 8.840 | 9.980 | 7.140 | 0 1 1 0 | 0 7(0 | 0 (1 0 | | | 0.500 | |
| | | | | | | 7.140 | 8.440 | 9.760 | 8.610 | 8.150 | 8.960 | 8.550 | 8.150 |

 Table E7 a^{*} values of Product-III

Table E8 Total soluble solids (TSS) in Product-III

| | Inlet air temperature (°C) | T 1 | T2 | T3 | T4 | Τ5 | Т6 | T7 | T8 | Т9 | T10 | T11 | T12 |
|----------|----------------------------------|------------|--------|--------|--------|--------|--------|-----------|--------|--------|------------------|------------------|------------------|
| | 185 | 15.000 | 15.400 | 16.000 | 15.500 | 16.300 | 17.700 | 16.300 | 17.500 | 18.000 | 16.070 | 10 400 | 10.000 |
| TSS (°B) | 190 | 15.300 | 15.400 | 16.000 | 15.500 | 16.300 | 17.700 | 16.500 | 17.500 | 18.000 | 16.870 16.860 | 18.400 18.400 | 19.000 19.300 |
| (-) | 200 | 15.000 | 15.400 | 16.400 | 15.500 | 16.300 | 17.700 | 16.400 | 18.000 | 18.000 | 16.860 | 18.300 | 19.300 |

.

| | Inlet air temperature (°C) | Tl | T2 | Т3 | T 4 | T5 | Т6 | T 7 | T 8 | Т9 | T10 | T11 | T12 |
|-------------------|----------------------------------|------------------|-----------------|------------------|------------------|------------------|------------------|------------------|------------------|------------------|------------------|------------------|------------------|
| | 185 | 93.000 | 85.860 | 82.730 | 93.000 | 85.740 | 82.720 | 90.120 | 85.700 | 82.300 | 90.010 | 85.000 | 82.060 |
| Solubility (%) | 190 200 | 89.500 53.000 | 78.940 44.00 | 74.961 49.400 | 89.000 53.000 | 78.510 44.000 | 74.670 47.540 | 88.770 51.200 | 78.010 45.000 | 72.610 44.560 | 88.520 47.000 | 75.750 45.600 | 70.000 43.800 |

 Table E9 Solubility of Product-III

 Table E10 Wettability of Product-III

| | Inlet air temperature (°C) | T 1 | T2 | Т3 | T4 | T5 | Т6 | Τ7 | Т8 | Т9 | T10 | T 11 | T12 |
|-------------|----------------------------------|------------|------|------|-----------|------|------|------|------|------|------|-------------|------|
| | 185 | 348 | 388 | 400 | 457 | 460 | 512 | 466 | 466 | 558 | 475 | 501 | 603 |
| Wettability | 190 | 344 | 400 | 675 | 457 | 462 | 677 | 463 | 462 | 703 | 470 | 603 | 741 |
| (s) | 200 | 995 | 1200 | 1260 | 1001 | 1223 | 1260 | 1001 | 1242 | 1278 | 1012 | 1246 | 1290 |

APPENDIX F

STORAGE STUDY OF MILK FORTIFIED BANANA PSEUDOSTEM JUICE POWDER (Product-III)

| | | Moisture c | ontent (%) | |
|-------------|-------|--------------|--------------|-------|
| Treatments | | Storage peri | ods (months) | |
| | 0 | 1 | 2 | 3 |
| T12 (185°C) | 3.960 | 3.960 | 3.967 | 4.127 |
| T12 (185°C) | 3.720 | 3.827 | 3.847 | 3.977 |
| T12 (190°C) | 3.643 | 3.643 | 3.723 | 3.747 |

Table F1 Changes in moisture content of Product-III during storage

Table F2 Changes in water activity of Product-III during storage

| | | Water | activity | |
|-------------|-------|--------------|--------------|-------|
| Treatments | | Storage peri | ods (months) | |
| | 0 | 1 | 2 | 3 |
| | 0.263 | 0.263 | 0.267 | 0.309 |
| T12 (185°C) | 0.223 | 0.226 | 0.252 | 0.271 |
| T12 (190°C) | 0.221 | 0.223 | 0.235 | 0.253 |
| T12 (200°C) | 0.221 | | | |

Table F3 Changes in tapped bulk density of Product-III during storage

| | | Tapped bulk | density (g/ml) | |
|----------------------------|-------|--------------|----------------|-------|
| T | | Storage peri | ods (months) | |
| Treatments | 0 | 1 | 2 | 3 |
| | 0.543 | 0.543 | 0.546 | 0.553 |
| T12 (185°C) | 0.512 | 0.515 | 0.517 | 0.523 |
| T12 (190°C) T12 (200°C) | 0.501 | 0.512 | 0.514 | 0.518 |

| | | Loose bulk o | lensity (g/ml) | |
|-------------|-------|--------------|----------------|-------|
| Treatments | | Storage peri | ods (months) | |
| | 0 | 1 | 2 | 3 |
| T12 (185°C) | 0.431 | 0.433 | 0.436 | 0.450 |
| T12 (185°C) | 0.427 | 0.432 | 0.435 | 0.443 |
| T12 (190°C) | 0.421 | 0.423 | 0.424 | 0.430 |

Table F4 Changes in loose bulk density of Product-III during storage

Table F5 Changes in Solubility of Product-III during storage

| | | Solubi | lity (%) | |
|----------------------------|--------|--------------|--------------|--------|
| Treatments | | Storage peri | ods (months) | |
|]] Cathlered | 0 | 1 | 2 | 3 |
| | 82.060 | 82.060 | 82.000 | 81.758 |
| T12 (185°C) | 70.000 | 70.000 | 69.463 | 68.417 |
| T12 (190°C) T12 (200°C) | 43.800 | 43.806 | 43.577 | 43.466 |

Table F6 Changes in wettability of Product-III during storage

| | | Wettab | oility (s) | |
|-------------|----------|--------------|--------------|----------|
| - (monts | | Storage peri | ods (months) | |
| Treatments | 0 | 1 | 2 | 3 |
| | 603.000 | 603.000 | 603.100 | 603.151 |
| T12 (185°C) | 741.000 | 741.000 | 741.053 | 741.110 |
| т12 (190°С) | 1290.000 | 1290.000 | 1300.000 | 1300.000 |
| T12 (200°C) | 1290.000 | | | |

APPENDIX G

MICROBIAL ANALYSIS OF SPRAY DRIED PSEUDOSTEM JUICE POWDERS

Table G1 Microbial load of Product-I

| | Mi | icrobial load (cf | u/g) |
|--|-----------------|-------------------|------|
| Treatments | 10 ¹ | 10^{2} | 104 |
| m((100°C) | 0 | 0 | 0 |
| 16(180C) | 0 | 0 | 0 |
| T6 (180°C) T6 (185°C) T6 (190°C) | ů | 0 | 0 |
| $T_{6}(190^{\circ}C)$ | 0 | | |

Table G2 Microbial load of Product-II

| | Μ | licrobial load (cfu | ı/g) |
|--|--------|---------------------|-----------------|
| Treatments | | 10 ² | 10 ⁴ |
| | 0 | 0 | 0 |
| T6 (180°C) | 0 | 0 | 0 |
| T6 (180°C) T6 (185°C) T6 (190°C) | ů 0 | 0 | 0 |
| T6(190°C) | 0 | | |

Table G3 Microbial load of Product-III

| | Microbial load (cfu | ı/g) |
|---|-----------------------|-----------------|
| Treatments 10 ¹ | 10^{2} | 10 ⁴ |
| 0 | 0 | 0 |
| T12 (185°C) 0 | 0 | . 0 |
| T12 (185°C) 0 T12 (190°C) 0 | 0 | 0 |
| (a, 0, 0, 0, 0, 0) | had of Product-I duri | ng storago |

12 (200 C) Table G4 Microbial load of Product-I during storage

| | Microbial | | Storage peri | ods (months) | |
|-------------------|----------------|---|--------------|--------------|---|
| Treatments | load (cfu/g) - | 0 | 2 | 4 | 6 |
| I I Catiller | 10/10 (02 0 | 0 | 0 | 0 | 1 |
| T((180°C)) | | 0 | 0 | 0 | 0 |
| T6 (180°C) | 10' | 0 | 0 | 0 | 0 |
| T6 (185°C) | | 0 | 0 | 0 | 0 |
| <u>T6 (190°C)</u> | | 0 | 0 | 0 | 0 |
| T6 (180°C) | 10^{2} | 0 | 0 | 0 . | 0 |
| T6 (185°C) | | 0 | 0 | 0 | 0 |
| T6 (190°C) | | 0 | 0 | 0 | 0 |
| T6 (180°C) | 10^{4} | 0 | 0 | 0 | 0 |
| T6 (185°C) | 10 | 0 | 0 | v | |
| TO(12) | | | | | |

T6 (190°C)

| Storage periods (months) | | | | IsidoroiM | 2 Treatments | |
|--------------------------|---|-----|---|-----------------|---------------------|--|
| 9 | t | 7 | 0 | — (g/utɔ) brol | | |
| 7 | 7 | 0 | 0 | | (J°081) 9T | |
| 3 | 0 | 0 | 0 | 101 | (J°281) 9T | |
| 5 | 0 | 0 | 0 | | (J.061) 9L | |
| I | 5 | 0 | 0 | | (J°081) 8T | |
| 7 | 0 | 0 | 0 | 10 ₅ | | |
| I | 0 | 0 | 0 | | (J°261) aT | |
| 0 | I | 0 | 0 | | (J.061) 91 | |
| 0 | 0 | 0 . | 0 | 10 ₄ | ()°081) aT | |
| 0 | 0 | 0 | 0 | 0.1 | (J°281) 9T | |
| | | | | | (J.061) 9L | |

Table G5 Microbial load of Product-II during storage

Table G6 Microbial load of Product-III during storage

| <u>v</u> | (sutnom) sboir | Storage per | | Microbial [] | 2119m100mT |
|----------|----------------|-------------|---|-----------------|-------------|
| 3 | 7 | T | 0 | – (g/ut2) brol | Treatments |
| 5 | 0 | 0 | 0 | | |
| 5 | 0 | 0 | 0 | · | T12(185°C) |
| 0 | 0 | 0 | 0 | 101 | LI2 (190°C) |
| I | 0 | 0 | 0 | | T12 (200°C) |
| I | 0 | 0 | 0 | | T12 (185°C) |
| 0 | 0 | 0 | 0 | ۱0 ₅ | LIS (100°C) |
| 0 | 0 . | 0 | 0 | | T12 (200°C) |
| 0 | 0 | 0 | 0 | | T12 (185°C) |
| 0 | 0 | 0 | 0 | 10 ₄ | 115(100C) |
| | | | | | T12 (200°C) |

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APPENDIX H

SENSORY EVALUATION OF BANANA PSEUDOSTEM BASED POWDERS

SENSORY SCORE CARD FOR PSEUDOSTEM BASED POWDERS

Date:

Name of judge:

You are requested to assess the product in terms of general acceptability on a 9 point hedonic scale

Score system:

Like extremely 9

Like very much 8

Like moderately 7

Like slightly 6

Neither like nor dislike 5

Dislike slightly 4

Dislike moderately 3

Dislike very much 2

Dislike extremely 1

| | Sample code | | | | | | | |
|--------------------------------|-------------|---|---|---|---|---|--|--|
| Characteristics | A | В | С | D | E | F | | |
| Colour & appearance | | | | | | | | |
| Flavor | | | | | | | | |
| Taste Overall acceptability | | | | | | | | |
| Overall acceptation | | | | | | | | |

Signature

Comments if any:

APPENDIX I

ANOVA FOR SPRAY DRIED BANANA PSEUDOSTEM JUICE POWDER (Product-I)

Factor –**A:** Inlet air temperature (°C)

Factor –B: Maltodextrin concentration (%)

Factor –C: Sugar concentration (%)

| Source | Sum of squares | df | Mean square | F value | p- value Prob > F | Std. Dev. | C.V. (%) |
|--------------------|----------------|----|----------------|------------|----------------------|--------------|-------------|
| | 8.03 | 5 | 1.61 | 42.35 | < 0.0001 | 0.19 | 5.57 |
| Model Factor –A | 0.40 | 2 | 0.20 | 5.22 | 0.0089 | | |
| Factor –B | 6.51 | 2 | 3.26 | 85.88 | < 0.0001 | | |
| Factor –C | 1.12 | 1 | 1.12 | 29.58 | < 0.0001 | | · |
| Pure error | 0.83 | 36 | 0.023 | | | | |
| Cor total | 9.85 | 53 | La 12 Wate | - ontivity | Signific | ant | |

Table 11 Moisture content (% w.b.)

Table I2 Water activity

| Source | Sum of squares | df | Mean square | F value | p- value Prob > F | Std. Dev. | C.V. (%) |
|---------------------|-------------------|----|----------------|------------|----------------------------|--------------|-------------|
| | 0.077 | 5 | 0.015 | 5.12 | 0.0008 | 0.055 | 15.73 |
| Model Factor – A | 3.299E-003 | 2 | 1.649E-003 | 0.55 | 0.5808 | | |
| Factor –B | 0.064 | 2 | 0.024 | 50.63 | 0.0002 | | |
| Factor –C | 9.699E-003 | 1 | 9.439E-003 | 3.23 | 0.0785 | | |
| | 0.14 | 36 | 2.585E-003 | | | | |
| Pure error | 0.22 | 53 | | | Signifi | cant | |
| Cor total | 0.2 | | | • | | | |

| Source | Sum of squares | df | Mean square | F value | p- value Prob > F | Std. Dev. | C.V. (%) |
|------------|-------------------|----|----------------|------------|----------------------------|--------------|-------------|
| Model | 0.056 | 5 | 0.011 | 3.65 | 0.0071 | 0.05 6 | 12.29 |
| Factor –A | 2.903E-003 | 2 | 1.451E-003 | 0.47 | 0.0282 | | |
| Factor –B | 0.051 | 2 | 0.026 | 8.32 | 0.0008 | | |
| Factor –C | 2.022E-003 | 1 | 2.022E-003 | 0.65 | 0.4227 | | |
| Pure error | 0.15 | 36 | 4.090E-003 | | | | |
| Cor total | 0.20 | 53 | | | Signifi | cant | |

Table 13 Tapped bulk density (g/ml)

Table I4 Loose bulk density (g/ml)

| Source | Sum of squares | df | Mean square | F value | p- value Prob > F | Std. Dev. | C.V. (%) |
|--------------------|-------------------|----|----------------|------------|----------------------------|--------------|-------------|
| | 0.059 | 5 | 0.012 | 5.48 | 0.0005 | 0.047 | 19.98 |
| Model Factor –A | 1.42E-003 | 2 | 7.146E-004 | 0.33 | 0.7203 | | |
| Factor –B | 0.048 | 2 | 0.024 | 11.18 | 0.0001 | | |
| Factor –C | 9.439E-003 | 2 | 9.439E-003 | 4.36 | 0.0420 | | |
| | 0.093 | 36 | 2.585E-003 | | | | |
| Pure error | 0.16 | 53 | | | Signifi | cant | |
| Cor total | - | | | | | | |

| Source | Sum of squares | df | Mean square | F value | p- value Prob > F | Std. Dev. | C.V. (%) |
|------------|----------------|----|----------------|---------|----------------------|--------------|-------------|
| Model | 1321.55 | 5 | 264.31 | 57.37 | < 0.0001 | 2.15 | 2.53 |
| Factor –A | 27.87 | 2 | 13.94 | 3.02 | 0.0579 | | |
| Factor –B | 17.60 | 2 | 587.17 | 127.44 | < 0.0001 | | |
| Factor –C | 119.35 | 1 | 119.35 | 25.90 | < 0.0001 | | |
| Pure error | 186.62 | 36 | 5.18 | | | | |
| Cor total | 1542.71 | 53 | | | Signific | cant | |

Table 15 L^{*} value

Table I6 a^{*} value

| Source | Sum of squares | df | Mean square 22.67 | F value 10.65 | p- value Prob > F <0.0001 | Std. Dev. 1.46 | C.V. (%) 115.38 |
|--------------------|----------------|--------|-------------------------|---------------------|---------------------------------|----------------------|--------------------|
| Model Factor –A | 113.33 7.89 | 5 2 | 3.94 | 1.85 | 0.1678 | 1.10 | 115.50 |
| Factor –B | 75.84 | 2 | 37.92 | 17.82 | <0.0001 | | |
| Factor –C | 29.60 | 1 | 29.60 | 13.91 | 0.0005 | | |
| | 8.82 | 36 | 0.24 | | | | |
| Pure error | -15 50 | 53 | | | Signif | icant | |
| Cor total | 215.50 | | | | | | |

| Source | Sum of squares | df | Mean square | F value | p- value Prob > F | Std. Dev. | C.V. (%) |
|------------|----------------|----|----------------|------------|----------------------------|--------------|-------------|
| Model | 38.95 | 5 | 7.79 | 5.96 | <0.000 1 | 1.14 | 19.50 |
| Factor –A | 21.34 | 2 | 10.67 | 8.16 | 0.1678 | | |
| Factor –B | 17.60 | 2 | 8.80 | 6.74 | <0.000 1 | | |
| Factor –C | 8.817E-003 | 1 | 8.8E-003 | 6.748E-003 | 0.0005 | | |
| Pure error | 22.41 | 36 | 0.62 | | | | |
| Cor total | 101.67 | 53 | | | Significan | t | |

Table 17 b^{*} value

Table I8 Solubility (%)

| Source | Sum of squares | df | Mean square | F value | p- value Prob > F | Std. Dev. | C.V. (%) |
|--------------------|----------------|----|----------------|---------|----------------------|--------------|-------------|
| | 372.84 | 5 | 74.57 | 12.78 | < 0.0001 | 2.42 | 2.98 |
| Model Factor –A | 18.35 | 2 | 9.17 | 1.57 | 0.2181 | | |
| Factor –B | 350.56 | 2 | 175.28 | 30.03 | <0.0001 | | |
| Factor –C | 3.93 | 1 | 3.93 | 0.67 | 0.4159 | | |
| | 152.43 | 36 | 4.23 | | | | |
| Pure error | 652.96 | 53 | | | Signific | ant | |

| Source | Sum of squares | df | Mean square | F value | p- value Prob > F | Std. Dev. | C.V. (%) |
|------------|----------------|----|----------------|---------|----------------------|--------------|-------------|
| Model | 182.89 | 5 | 36.56 | 13.79 | < 0.0001 | 1.63 | 2.10 |
| Factor –A | 15.60 | 2 | 7.80 | 2.94 | 0.625 | | |
| Factor –B | 149.06 | 2 | 74.53 | 28.09 | <0.0001 | | |
| Factor –C | 18.24 | 1 | 18.24 | 6.87 | 0.0117 | | |
| Pure error | 112.18 | 36 | 3.12 | | | | |
| Cor total | 310.22 | 53 | | | Signific | ant | |

Table 19 Wettability (s)

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Table I10 Product yield (%)

| | Sum of | df | Mean square | F value | p- value Prob > F | Std. Dev. | C.V. (%) |
|--------------------|------------------|----|----------------|---------|----------------------|--------------|-------------|
| Source | squares | 5 | 715.99 | 70.85 | <0.0001 | 3.18 | 8.18 |
| Model Factor –A | 3579.95 49.45 | 2 | 24.72 | 2.45 | 0.0973 | | |
| Factor –B | 2790.04 | 2 | 1395.02 | 138.04 | <0.0001 | | |
| Factor –C | 740.46 | 1 | 740.46 | 73.27 | < 0.0001 | | |
| Pure error | 35.59 | 36 | 0.99 | | | | |
| Cor total | 4065.03 | 53 | | | Signific | ant | |

APPENDIX J

ANOVA FOR SPRAY DRIED BANANA PSEUDOSTEM JUICE POWDER DURING STORAGE (Product-I)

Factor –A: Storage period (months)

Factor –B: Treatment temperature (°C)

Table J1 Moisture content of product during storage

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| Source | Sum of squares | df | Mean square | F value | p- value Prob > F | Std. Dev. | C.V. (%) |
|------------|----------------|----|----------------|---------|----------------------|--------------|-------------|
| Model | 0.32 | 5 | 0.063 | 5.71 | 0.0008 | 0.11 | 3.25 |
| Factor –A | 0.17 | 3 | 0.056 | 5.02 | 0.0061 | | |
| Factor –B | 0.15 | 2 | 0.075 | 6.74 | 0.0038 | | |
| Pure error | 0.31 | 24 | 0.013 | | | | |
| Cor total | 0.65 | 35 | | | Signific | cant | |

Table J2 Water activity of product during storage

| Source | Sum of squares | df | Mean square | F value | p- value Prob > F | Std. Dev. | C.V. (%) |
|------------|----------------|----|----------------|------------|----------------------|--------------|-------------|
| | 9.369E-003 | 5 | 1.874E-003 | 15.41 | < 0.0001 | 0.11 | 3.46 |
| Model | | 3 | 2.007E-003 | 16.51 | < 0.0001 | | |
| Factor –A | 6.020E-003 | - | · (75E 003 | 13.78 | < 0.0001 | | |
| Factor –B | 3.349E-003 | 2 | 1.675E-003 | 15170 | | | |
| | 3.276E-003 | 24 | 1.365E-004 | | | | |
| Pure error | | | | | Significa | ant | |
| Cor total | 0.013 | 35 | | | | | |

| Source | Sum of squares | df | Mean square | F value | p- value Prob > F | Std. Dev. | C.V. (%) |
|------------|-------------------|----|----------------|------------|-------------------------|-------------|-------------|
| Model | 3.521E-003 | 5 | 7.042E-004 | 10.08 | <0.0001 | 8.358E-003 | 1.95 |
| Factor –A | 6.371E-003 | 3 | 2.124E-004 | 3.04 | 0.0442 | | |
| Factor –B | 2.884E-003 | 2 | 1.442E-004 | 20.64 | <0.0001 | | |
| Pure error | 2.030E-003 | 24 | 8.459E-005 | | | | |
| Cor total | 5.616E-003 | 35 | | | | Significant | |

Table J3 Tapped bulk density of product during storage

Table J4 Loose bulk density of product during storage

| Source | Sum of squares | df | Mean square | F value | p- value Prob > F | Std. Dev. | C.V. (%) |
|------------|-------------------|----|----------------|------------|-------------------------|-----------|-------------|
| Model | 0.013 | 5 | 2.647E-003 | 6.87 | 0.0002 | 0.020 | 6.56 |
| Factor –A | 9.237E-003 | 3 | 3.079E-003 | 7.99 | 0.0005 | | |
| Factor –B | 3.998E-003 | 2 | 1.999E-003 | 5.19 | 0.0116 | | |
| Pure error | 4.932E-003 | 24 | 2.055E-004 | | | | |
| Cor total | 0.025 | 35 | | | Sigr | nificant | |

| Source | Sum of squares | df | Mean square | F value | p- value Prob > F | Std. Dev. | C.V. (%) |
|------------|----------------|----|----------------|------------|----------------------|------------|-------------|
| Model | 1.68 | 5 | 0.34 | 1.99 | 0.1090 | 0.41 | 0.45 |
| Factor –A | 0.017 | 3 | 5.641E-003 | 0.033 | 0.9916 | | |
| Factor –B | 1.66 | 2 | 0.83 | 4.92 | 0.0142 | | |
| Pure error | 5.06 | 24 | 1.077E-003 | | | | |
| Cor total | 6.74 | 35 | | | Not s | ignificant | |

Table J5 L* value of product during storage

Table J6 a* value of product during storage

| Source | Sum of squares | df | Mean square | F value | p- value Prob > F | Std. Dev. | C.V. (%) |
|--------------------|-------------------|----|----------------|------------|----------------------|------------|-------------|
| | 1.605E-003 | 5 | 3.210E-004 | 17.98 | < 0.0001 | 4.226E-003 | 1.36 |
| Model Factor –A | 1.313E-004 | 3 | 4.375E-005 | 2.45 | 0.0828 | | |
| Factor –B | 1.474E-003 | 2 | 7.370E-004 | 41.27 | <0.0001 | | |
| | 3.720E-004 | 24 | 1.550E-005 | | | | |
| Pure error | 2.141E-003 | 35 | | | Sig | nificant | |
| Cor total | 2.19115 | | | | | | |

Table J7 b* value of product during storage

| | Sum of | df | Mean square | F value | p- value Prob > F | Std. Dev. | C.V. (%) |
|-------------------------|--------------------|---------|----------------|------------|----------------------|-----------|-------------|
| Source | squares | 5 | 5.779E-003 | 11.52 | < 0.0001 | 0.022 | 0.42 |
| Model | 0.029 | 3 | 3.439E-005 | 0.069 | 0.9762 | | |
| Factor –A | 1.032E-004 | 2 | 0.014 | 28.70 | <0.0001 | | |
| Factor –B | 0.029 5.060.015 | 2 24 | 6.120E-004 | | Sig | nificant | |
| Pure error Cor total | 5.080.013 0.044 | 35 | | | 016 | | |

| Source | Sum of squares | df | Mean square | F value | p- value Prob > F | Std. Dev. | C.V. (%) |
|------------|-------------------|----|----------------|------------|----------------------|-----------|-------------|
| Model | 902.32 | 5 | 180.46 | 1.32 | 0.2842 | 11.71 | 15.76 |
| Factor –A | 644.94 | 3 | 214.98 | 1.57 | 0.2179 | | |
| Factor –B | 257.39 | 2 | 128.69 | 0.94 | 0.4026 | | |
| Pure error | 3287.74 | 24 | 136.99 | | | | |
| Cor total | 5018.69 | 35 | | | Not si | gnificant | |

Table J8 Solubility of product during storage

Table J9 Wettability of product during storage

| Source | Sum of squares | df | Mean square | F value | p- value Prob > F | Std. Dev. | C.V. (%) |
|-------------------------|----------------|----------|----------------|------------|----------------------|-----------|-------------|
| Model | 14.84 | 5 | 2.97 | 66.10 | < 0.0001 | 0.21 | 0.26 |
| Factor –A | 12.98 | 3 | 4.33 | 96.38 | <0.0001 | | |
| Factor –B | 1.86 | 2 | 0.93 | 20.68 | <0.0001 | | |
| Pure error Cor total | 0.38 16.19 | 24 35 | 0.016 | | Not si | gnificant | |

APPENDIX K

ANOVA FOR SPRAY DRIED BANANA PSEUDOSTEM JUICE-HORSE GRAM EXTRACT BLEND POWDER (Product-II)

Factor –**A:** Inlet air temperature (°C)

Factor –B: Maltodextrin (%)

Factor –C: Horse gram extract (%)

. . .

| Source | Sum of squares | df | Mean square | F value | Pro | value ob > F | Std. Dev. | C.V. (%) | - |
|--------------------|------------------|-----|----------------|------------|-----|---------------------|--------------|-------------|-------------|
| Model | 1.13 | 3 | 0.38 | 5.56 | 0. | 0023 | 0.26 | 6.18 | |
| Factor –A | 0.52 | 1 | 0.52 | 7.60 | 0. | 0081 | | | |
| Factor –B | 0.50 | 1 | 0.50 | 7.37 | 0. | 0091 | | | |
| Factor –C | 0.12 | 1 | 0.12 | 1.72 | 0. | 1963 | | | |
| Pure error | 2.17 | 36 | 0.060 | | | | | | |
| | 4.53 | 53 | | | | Significa | int | | _ |
| Cor total | 1.00 | Tab | le K2 Wate | r activity | 7 | | | | |
| | Sum of | df | Mean square | F val | ue | p- valu Prob > 2 | | td. Dev. | C.V. (%) |
| Source | squares 0.035 | 6 | 5.857E-00 | 3 197. | 43 | < 0.000 | 5.4 | 47E-003 | 1.49 |
| Model Factor –A | 0.035 | 1 | 0.012 | 391.9 | 96 | < 0.000 | l | | |
| Factor –B | 0.015 | 1 | 0.015 | 502.2 | 26 | < 0.0001 | [| | |
| Factor –C | 8.070E-003 | 1 | 8.070E-00 | 3 272.0 | 03 | < 0.0001 | | | |
| AB | 1.469E-005 | 1 | 1.469E-00 | 5 0.50 |) | 0.4850 | | | |
| AC | 8.167E-006 | 1 | 8.167E-00 | 6 0.28 | 3 | 0.6023 | | | |
| ВС | 5.214E-004 | 1 | 5.214E-004 | 4 17.5 | 7 | 0.0001 | | | |
| - | 2.860E-004 | 36 | 7.944E-000 | 6 | | с. | • ~ | | |
| Pure error | 0.027 | 53 | | | | Sig | nifican | t | |
| Cor total | 0.037 | | | | | | | | |

| Source | Sum of squares | df | Mean square | F value | p- value Prob > F | Std. Dev. | C.V. (%) |
|------------|----------------|----------|--------------------------|---------|----------------------|------------|-------------|
| Model | 0.14 | 6 | 0.023 | 477.04 | <0.0001 | 6.918E-003 | 1.42 |
| Factor –A | 1.845E-003 | 1 | 1.845E-003 | 38.54 | <0.0001 | | |
| Factor –B | 0.092 | 1 | 0.092 | 1920.74 | <0.0001 | | |
| Factor –C | 0.039 | 1 | 0.039 | 806.79 | <0.0001 | | |
| AB | 9.724E-004 | 1 | | | <0.0001 | | |
| | 9.724E-004 | 1 | 9.724E-004 | 20.32 | | | |
| AC | 7.315E-005 | | 7.315E-005 | 1.53 | 0.2225 | | |
| BC | 7.510- | 1 | | | <0.0001 | | |
| BC | 3.556E-003 | 26 | 3.556E-003 4.021E-006 | 74.30 | <0.0001 | | |
| Pure error | 1.448E-004 | 36 53 | 4.0212 000 | | Signifi | cant | |
| Cor total | 0.14 | | | | ~.5 | | |

Table K3 Tapped bulk density (g/ml)

| Source | Sum of squares | df | Mean square | F value | p- value Prob > F | Std. Dev. | C.V. (%) |
|--|--|-----------------------------|--|--|--|--------------|-------------|
| Model | 0.14 | 6 | 0.023 | 54.52 | < 0.0001 | 0.020 | 5.40 |
| Factor-A | 0.13 | 1 | 0.13 | 311.1 2 | < 0.0001 | | |
| Factor-B Factor-C AB AC BC Pure | 4.178E-003 9.818E-004 1.190E-003 9.375E-006 2.668E-004 2.349E-003 | 1 1 1 | 4.178E-003 9.818E-004 1.190E-003 9.375E-006 2.668E-004 6.526E-005 | 10.08 2.37 2.87 0.023 0.64 | 0.0026 0.1306 0.0968 0.8811 0.4265 | | |
| Error Cor Total | 0.16 | 53 | L of Drod | unt II | signi | ficant | |
| 10101 | Table | K5 L* v | alue of Prod | uct-11 | | | |
| Source | Sum of squares | df | Mean square | F value | p- value Prob > F | Std. Dev. | C.V. (%) |
| Model | 112.58 | 6 | 18.76 0.55 | 0.96 0.028 | 0.4606 0.8671 | 4.41 | 5.01 |
| Factor-A Factor-B C AB AC BC Pure Error | 0.55 11.36 14.03 0.71 67.18 18.74 656.62 | 1 1 1 1 1 36 | 0.33 11.36 14.03 0.71 67.18 18.74 18.24 | 0.028 0.58 0.72 0.037 3.45 0.96 | 0.8071 0.4491 0.4004 0.8492 0.0696 0.3317 Not signif | īcant | |
| Cor Total | 1028.46 | 53 | | | | | |

Table K4 Loose bulk density (g/ml)

| Source | Sum of squares | df | Mean square | F value | p- value Prob > F | Std. Dev. | C.V. (%) |
|--------------|-------------------|----|----------------|---------|----------------------------|--------------|-------------|
| Model | 2.61 | 6 | 0.44 | 1.54 | 0.1876 | 0.53 | 71.38 |
| Factor-A | 0.089 | 1 | 0.089 | 0.32 | 0.5772 | | |
| Factor-B | 0.33 | 1 | 0.33 | 1.17 | 0.2854 | | |
| Factor-C | 0.10 | 1 | 0.10 | 0.35 | 0.5544 | | |
| AB | 0.078 | 1 | 0.078 | 0.28 | 0.6020 | | |
| AD AC | 1.93 | 1 | 1.93 | 6.80 | 0.0122 | | |
| AC BC | 0.085 | 1 | 0.085 | 0.30 | 0.5871 | | |
| Pure | 9.88 | 36 | 0.27 | | | | |
| Error | | | | | | | |
| Cor Total | 15.93 | 53 | | | Not signi | ficant | |

Table K6 a* values

.

| Table | K7 | b* | va] | lue |
|-------|----|----|-----|-----|
|-------|----|----|-----|-----|

| Source | Sum of squares | df | Mean square | F value | p- value Prob > F | Std. Dev. | C.V. (%) |
|---|--|---------------------------------------|---|--|--|--------------|-------------|
| Model Factor-A Factor-B Factor-C AB AC BC Pure | 25.00 7.51 4.72 1.18 6.32 0.015 5.25 186.02 | 6 1 1 1 1 1 1 36 | 4.17 7.51 4.72 1.18 6.32 0.015 5.25 5.17 | 0.78 1.40 0.88 0.22 1.18 2.709E- 003 0.98 | 0.5911 0.2423 0.3524 0.6403 0.2829 0.9587 0.3270 | 2.31 | 34.11 |
| Error Cor Total | 276.61 | 53 | | | Not signi | ficant | |

| Source | Sum of squares | df | Mean square | F value | p- value Prob > F | Std. Dev. | C.V. (%) |
|------------|----------------|----|----------------|------------|----------------------|-----------|-------------|
| Model | 14.79 | 6 | 2.47 | 51.43 | <0.0001 | 0.22 | 0.22 |
| Factor-A | 2.03 | 1 | 2.03 | 42.38 | <0.0001 | | |
| Factor-B | 8.10 | 1 | 8.10 | 168.9 2 | <0.0001 | | |
| Factor-C | 4.29 | 1 | 4.29 | 89.46 | <0.0001 | | |
| AB | 9.404E-004 | 1 | 9.404E-004 | 0.020 | 0.8892 | | |
| AC | 0.10 | 1 | 0.10 | 2.10 | 0.1538 | | |
| BC | 0.27 | 1 | 0.27 | 5.71 | 0.0209 | | |
| Pure Error | 0.49 | 36 | 0.014 | | | | |
| Cor Total | 17.05 | 53 | | | Signifi | cant | |

Table K8 Solubility

| Source | Sum of squares | df | Mean square | F value | p- value Prob > F | Std. Dev. | C.V. (%) |
|------------------|-------------------|---------------|-----------------|---------|----------------------|--------------|-------------|
| Model | 42930.46 | 6 | 7155.08 | 877.52 | <0.0001 | 2.86 | 0.55 |
| Factor-A | 792.68 | 1 | 792.68 | 97.22 | < 0.0001 | | |
| Factor-B | 30634.57 | 1 | 30634.57 | 3757.13 | <0.0001 | | |
| Factor-C | 9532.45 | 1 | 9532.45 | 1169.09 | <0.0001 | | |
| AB | 165.02 | 1 | 165.02 | 20.24 | <0.0001 | | |
| AC | 238.10 | 1 | 238.10 | 20.20 | <0.0001 | | |
| BC Pure Error | 1567.65 22.93 | 1 36 53 | 1567.65 0,64 | 192.26 | <0.0001 Significa | nt | |
| Cor Total | 43313.69 | | | | | | |

Table K9 Wettability

| Source | Sum of squares | df | Mean square | F value | p- value Prob > F | Std. Dev. | C.V. (%) |
|--|------------------------------------|------------------|------------------------------------|------------------------------|--------------------------------------|-----------|-------------|
| Model | 528.98 | 6 | 88.16 | 1.08 | 0.3861 | 9.02 | 15.0 4 |
| Factor-A Factor-B Factor-C AB | 17.57 31.93 109.13 102.56 | 1 1 1 1 | 17.57 31.93 109.13 102.56 | 0.22 0.39 1.34 1.26 | 0.6444 0.5341 0.2527 0.2673 | | |
| AC | 254.31 | 1 | 254.31 | 3.13 | 0.0836 | | |
| BC Pure Error Cor Total | 13.47 2981.66 4353.78 | 1 36 53 | 13.47 82.82 | 0.17 0.93 | 0.6860 Not ig | nificant | |

Table K10 Product yield

APPENDIX L

ANOVA FOR SPRAY DRIED BANANA PSEUDOSTEM JUICE-HORSE GRAM EXTRACT BLEND POWDER DURING STORAGE (Product-II)

Factor –A: Storage period (months)

Factor –**B**: Treatment temperature (°C)

Table L1 Moisture content of product during storage

| | Sum of | df | Mean square | F value | p- value Prob > F | Std. Dev. | C.V. (%) |
|------------|-----------------|----|----------------|-------------|----------------------|--------------|-------------|
| Source | squares 0.43 | 5 | 0.087 | 15.61 | < 0.0001 | 0.075 | 1.79 |
| Model | | 3 | 0.044 | 7.95 | 0.0005 | | |
| Factor –A | 0.13 | _ | 0.15 | 27.10 | < 0.0001 | | |
| Factor –B | 0.30 | 2 | | <u>,,,,</u> | | | |
| | 0.15 | 24 | 6.066E- 003 | | | | |
| Pure error | 0.15 | 35 | | | Signific | cant | _ |
| Cor total | | | | | | | |

Cor total

| Source | Sum of squares | df | Mean square | F value | p- value Prob > F | Std. Dev. | C.V. (%) |
|------------|-------------------|----|----------------|------------|----------------------|----------------|-------------|
| Model | 7.747E-003 | 5 | 1.549E-003 | 19.99 | < 0.0001 | 8.803E- 003 | 2.59 |
| Factor – A | 5.821E-004 | 3 | 1.940E-004 | 2.50 | 0.0782 | | |
| Factor –B | 7.165E-003 | 2 | 3.583E-003 | 46.23 | < 0.0001 | | |
| Pure error | 2.107E-003 | 24 | 8.778E-005 | | | | |
| Cor total | 0.010 | 35 | | | Signif | ficant | |

Table L2 Water activity of product during storage

Table L3 Tapped bulk density of product during storage

| Source | Sum of squares | df | Mean square | F value | p- value Prob > F | Std. Dev. | C.V. (%) |
|------------|-------------------|----|----------------|------------|----------------------|-----------|-------------|
| Model | 0.064 | 5 | 0.013 | 28.74 | < 0.0001 | 0.021 | 5.67 |
| Factor –A | 7.390E-003 | 3 | 2.463E-003 | 5.56 | 0.0037 | | |
| Factor –B | 0.056 | 2 | 0.028 | 63.52 | < 0.0001 | | |
| Pure error | 9.254E-003 | 24 | 4.434E-004 | | | • | |
| Cor total | 0.077 | 35 | 3.856E-004 | | Signifi | | |

| Source | Sum of squares | df | Mean square | F value | p- value Prob > F | Std. Dev. | C.V. (%) |
|------------|----------------|----|----------------|------------|----------------------|-----------|-------------|
| Model | 0.064 | 5 | 0.013 | 28.74 | < 0.0001 | 0.021 | 5.67 |
| Factor –A | 7.390E-003 | 3 | 2.463E-003 | 5.56 | 0.0037 | | |
| Factor –B | 0.056 | 2 | 0.028 | 63.52 | < 0.0001 | | |
| Pure error | 9.254E-003 | 24 | 3.856E-004 | | 5 1 2 | | |
| Cor total | 0.077 | 35 | | | Signif | icant | |

Table L4 Loose bulk density of product during storage

Table L5 L* value of product during storage

| Source | Sum of squares | df | Mean square | F value | p- value Prob > F | Std. Dev. | C.V. (%) |
|------------|----------------|----|----------------|------------|----------------------|---------------------------------------|-------------|
| | | 5 | 1.22 | 5.93 | 0.0006 | 0.45 | 0.48 |
| Model | 6.10 | 3 | 0.004 | 0.46 | 0 7120 | | |
| Factor –A | 0.28 | J | 0.094 | 0.46 | 0.7130 | | |
| Factor –B | 5.82 | 2 | 2.91 | 14.14 | < 0.0001 | | |
| Dung arror | 5.02 | 24 | 0.21 | | | | |
| Pure error | 5.02 | 35 | | | Signif | ficant | |
| Cor total | 12.27 | | | | | · · · · · · · · · · · · · · · · · · · | |

Table L6 a* value of product during storage

| | Sum of | df | Mean square | F value | p- value Prob > F | Std. Dev. | C.V. (%) |
|------------|------------|----|----------------|---------|----------------------|------------|-------------|
| Source | squares | 5 | 3.364E-003 | 1823.74 | < 0.0001 | 1.358E-003 | 0.35 |
| Model | 0.017 | 3 | 4.472E-006 | 2.42 | 0.0851 | | |
| Factor – A | 1.342E-005 | 2 | 8.403E-003 | 4555.72 | < 0.0001 | | |
| Factor –B | 0.017 | 24 | 1.583E-006 | | | | |
| Pure error | 3.800E-005 | 35 | 1.3850 000 | | Signi | ficant | |
| Cor total | 0.017 | | | | | | |

| Source | Sum of squares | df | Mean square | F value | p- value Prob > F | Std. Dev. | C.V. (%) |
|------------|----------------|----|----------------|---------|----------------------|-----------|-------------|
| Model | 11.63 | 5 | 2.33 | 74.30 | < 0.0001 | 0.18 | 2.89 |
| Factor –A | 0.36 | 3 | 0.12 | 3.87 | 0.0189 | | |
| Factor –B | 11.26 | 2 | 5.63 | 179.95 | < 0.0001 | | |
| Pure error | 0.78 | 24 | 0.033 | | · • • | | |
| Cor total | 12.57 | 35 | | | Signifi | cant | |

Table L7 b* value of product during storage

 Table L8 Solubility of product during storage

| Source | Sum of | df | Mean square | F value | p- value Prob > F | Std. Dev. | C.V. (%) |
|------------|-----------------|---------|----------------|------------|----------------------|---------------------------------------|-------------|
| Model | squares 0.86 | 5 | 0.17 | 23.47 | < 0.0001 | 0.085 | 0.08 7 |
| Factor –A | 0.18 | 3 | 0.061 | 8.34 | 0.0003 | | |
| Factor –B | 0.67 | 2 24 | 0.34 | 46.17 | < 0.0001 | | |
| Pure error | 0.16 | 35 | 6.502E-003 | | Signi | ficant | |
| Cor total | 1.07 | | | | | · · · · · · · · · · · · · · · · · · · | |

| Source | Sum of squares | df | Mean square | F value | p- value Prob > F | Std. Dev. | C.V. (%) |
|------------|----------------|----|----------------|------------|----------------------|-----------|-------------|
| Model | 3.57 | 5 | 0.71 | 12.61 | < 0.0001 | 0.24 | 0.04 2 |
| Factor –A | 3.52 | 3 | 1.17 | 20.72 | < 0.0001 | | |
| Factor –B | 0.051 | 2 | 0.025 | 0.45 | 0.6438 | | |
| Pure error | 0.48 | 24 | 0.057 | | Signi | ficant | |
| Cor total | 5.26 | 35 | 0.020 | | | | |

Table L9 Wettability of product during storage

APPENDIX M

ANOVA FOR MILK FORTIFIED BANANA PSEUDOSTEM JUICE POWDER DURING STORAGE (Product-III)

Factor –A: Storage period (months)

Factor –B: Milk (%)

Factor –C: Horse gram extract (%)

| Source | Sum of squares | df | Mean square | F value | p- value Prob > F | Std. Dev. | C.V. (%) |
|----------------|----------------|-----|----------------|---------|----------------------|-----------|-------------|
| Model | 24.96 | 9 | 2.77 | 96.87 | < 0.0001 | 0.17 | 6.00 |
| Factor –A | 0.74 | 1 | 0.74 | 25.81 | < 0.0001 | | |
| Factor –B | 2.00 | 1 | 2.00 | 69.97 | < 0.0001 | | |
| Factor –C | 20.33 | 1 | 20.33 | 710.24 | < 0.0001 | | |
| AB | 0.035 | 1 | 0.035 | 1.22 | 0.2727 | | |
| AC | 0.043 | 1 | 0.043 | 1.48 | 0.2259 | | • |
| BC | 0.67 | 1 | 0.67 | 23.48 | < 0.0001 | | |
| | 5.316E-003 | 1 | 5.316E-003 | 0.19 | 0.6675 | | |
| A ² | 0.43 | 1 | 0.43 | 15.08 | 0.0002 | | |
| B ² | 0.29 | 1 | 0.29 | 10.25 | 0.0018 | | |
| C^2 | 0.27 | 72 | 8.851E-003 | | | | |
| Pure error | | 107 | | | Signifi | cant | |
| Cor total | 27.76 | | | | | | |

Table M1 Moisture content

| Source | Sum of squares | df | Mean square | F value | p- value Prob > F | Std. Dev. | C.V. (%) |
|----------------|----------------|-----|----------------|---------|----------------------|-----------|-------------|
| Model | 0.18 | 9 | 0.020 | 121.15 | < 0.0001 | 0.013 | 4.51 |
| Factor –A | 5.580E-004 | 1 | 5.580E-004 | 3.34 | 0.0709 | | |
| Factor –B | 6.685E-003 | 1 | 6.685E-003 | 39.96 | < 0.0001 | | |
| Factor –C | 0.16 | 1 | 0.16 | 971.58 | < 0.0001 | | |
| AB | 2.318E-004 | 1 | 2.318E-004 | 1.39 | 0.2420 | | |
| AC | 8.143E-005 | 1 | 8.143E-005 | 0.49 | 0.4870 | | |
| BC | 2.716E-003 | 1 | 2.716E-003 | 16.24 | 0.0001 | | |
| A ² | 8.792E-005 | 1 | 8.792E-005 | 0.53 | 0.4702 | | |
| B ² | 3.359E-005 | 1 | 3.359E-005 | 0.20 | 0.6551 | | |
| C^2 | 7.782E-003 | - | 7.782E-003 | 46.52 | < 0.0001 | | |
| Pure error | 6.900E-003 | 72 | 1.673E-004 | | Signif | icant | |
| Cor total | 0.20 | 107 | 9.583E-005 | | | | |

Table M2 Water activity

| Source | Sum of squares | df | Mean square | F value | p- value Std. Dev. Prob > F | C.V (%) |
|--|---|----------------------------|--|--|--|------------|
| Model Factor –A Factor –B Factor –C AB AC | 0.19 5.827E-004 0.014 0.15 1.763E-004 8.976E-004 | 9 1 1 1 1 1 | 0.021 5.827E-004 0.014 0.15 1.763E-004 8.976E-004 | 27.54 0.75 17.88 195.41 0.23 1.16 | < 0.0001 0.028 0.3872 < 0.0001 < 0.0001 0.6339 0.2836 | 5.80 |
| BC | 2.237E-003 | 1 | 2.237E-003 | 2.90 | 0.0919 | |
| A^2 B^2 | 1.239E-004 0.017 | 1 | 1.239E-004 0.017 | 0.16 21.92 | 0.6896 < 0.0001 | |
| C ² Pure error | 3.082E-004 7.328E-003 | 1 72 | 3.082E-004 1.018E-004 | 0.40 | 0.5291 Significant | |
| Cor total | 0.27 | 107 | | <u></u> | · · · · · · · · · · · · · · · · · · · | |

Table M3 Tapped bulk density (g/ml)

| Source | Sum of squares | df | Mean square | F value | p- value Prob > F | Std. Dev. | C.V. (%) |
|------------------------------|----------------|-----|----------------|---------|----------------------|-----------|-------------|
| Model | 0.013 | 9 | 1.483E-003 | 6.65 | < 0.0001 | 0.015 | 4.37 |
| Factor –A | 4.004E-006 | 1 | 4.004E-006 | 0.018 | 0.8937 | | |
| Factor –B | 2.142E-003 | 1 | 2.142E-003 | 9.61 | 0.0025 | | |
| Factor –C | 5.273E-005 | 1 | 5.273E-005 | 0.24 | 0.6279 | | |
| AB | 3.577E-006 | 1 | 3.577E-006 | 0.016 | 0.8995 | | |
| AC | 4.353E-004 | 1 | 4.353E-004 | 1.95 | 0.1656 | | |
| BC | 2.777E-005 | 1 | 2.777E-005 | 0.12 | 0.725 | | |
| | 3.934E-004 | 1 | 3.934E-004 | 1.76 | 0.1872 | | |
| A ² | 3.875E-003 | 1 | 3.875E-003 | 17.37 | < 0.0001 | | |
| B ² | 5.099E-003 | 1 | 5.099E-003 | 22.86 | < 0.0001 | | |
| C ² Pure error | 7.187E-003 | 72 | 9.982E-005 | • | | | |
| Cor total | 0.035 | 107 | | | Signific | cant | |

Table M4 Loose bulk density (g/ml)

| Source | Sum of squares | df | Mean square | F value | p- value Prob > F | Std. Dev. | C.V. (%) |
|----------------|----------------|-----|----------------|----------------|----------------------|-----------|-------------|
| Model | 1623.47 | 9 | 180.48 | 144.29 | < 0.0001 | 1.12 | 1.33 |
| Factor –A | 145.18 | 1 | 143.09 | 114.40 | < 0.0001 | | |
| Factor –B | 101.99 | 1 | 101.22 | 80.92 | < 0.0001 | | |
| Factor –C | 1247.14 | 1 | 1247.14 | 997.02 | < 0.0001 | | |
| AB | 0.036 | 1 | 1247.14 | 0.029 | 0.8648 | | |
| AC | 37.89 | 1 | 37.89 | 30.29 | < 0.0001 | | |
| BC | 23.99 | 1 | 23.99 | 19.18 | < 0.0001 | | |
| A 2 | 0.43 | 1 | 0.43 | 0.35 | 0.5568 | | |
| A^2 | 0.46 | 1 | 0.46 | 0.36 | 0.5478 | | |
| B^2 C^2 | 6.588E-003 | 1 | 6.588E-003 | 5.267E- 003 | 0.9423 | | |
| Pure error | 33.49 | 72 | 0.47 | | | | |
| Cor total | 1746.95 | 107 | | | Signific | cant | |

Table M5 L* value

| Source | Sum of squares | df | Mean square | F value | p- value Prob > F | Std. Dev. | C.V. (%) |
|---------------------|----------------|-----|----------------|---------|----------------------|--------------|-------------|
| Model | 3.76 | 9 | 0.42 | 4.20 | 0.0001 | 0.32 | 12.67 |
| Factor –A | 0.095 | 1 | 0.095 | 0.96 | 0.3295 | | |
| Factor –B | 0.34 | 1 | 0.34 | 3.40 | , 0.0682 | | |
| Factor –C | 0.69 | 1 | 0.69 | 6.93 | 0.0098 | | |
| AB | 0.42 | 1 | 0.42 | 4.27 | 0.0413 | | |
| AC | 0.96 | 1 | 0.96 | 9.70 | 0.0024 | | |
| BC | 0.28 | 1 | 0.28 | 2.85 | 0.0947 | | |
| A ² | 0.21 | 1 | 0.21 | 2.10 | 0.1500 | | |
| А В ² | 0.029 | 1 | 0.029 | 0.29 | 0.5889 | | |
| C^2 | 0.026 | 1 | 0.026 | 0.27 | 0.6069 | | |
| Pure error | 7.17 | 72 | 0.100 | | 0 | | |
| Cor total | 13.48 | 107 | | | Signifi | cant | · |

Table M6 a* value

| Source | Sum of squares | df | Mean square | F value | p- value Prob > F | Std. Dev. | C.V. (%) |
|----------------|----------------|-----|----------------|---------------------------------------|----------------------|--------------|-------------|
| Model | 74.03 | 9 | 8.23 | 12.10 | < 0.0001 | 0.82 | 7.66 |
| Factor –A | 56.12 | 1 | 56.12 | 82.58 | < 0.0001 | | |
| Factor –B | 0.20 | 1 | 0.20 | 0.29 | •• 0.5918 | | |
| Factor –C | 0.86 | 1 | 0.86 | 1.27 | 0.2632 | | |
| AB | 1.49 | 1 | 1.49 | 2.19 | 0.1417 | | |
| AC | 3.90 | 1 | 3.90 | 5.73 | 0.0185 | | |
| BC | 2.30 | 1 | 2.30 | 3.38 | 0.0690 | | |
| A ² | 2.58 | 1 | 2.58 | 3.79 | 0.0544 | | |
| B^2 | 2.11 | 1 | 2.11 | 3.10 | 0.0813 | | |
| C^2 | 1.92 | 1 | 1.92 | 2.83 | 0.0958 | | |
| Pure error | 36.99 | 72 | 0.51 | | <u> </u> | | |
| Cor total | 140.63 | 107 | | · · · · · · · · · · · · · · · · · · · | Signifi | | |

Table M7 b* value

| Source | Sum of squares | df | Mean square | F value | p- value Prob > F | Std. Dev. | C.V. (%) |
|------------------------------|----------------|-----|----------------|---------|----------------------|--------------|-------------|
| Model | 574.03 | 9 | 63.78 | 59.91 | < 0.0001 | 1.03 | 1.08 |
| Factor –A | 30.97 | 1 | 30.97 | 29.09 | < 0.0001 | | |
| Factor –B | 62.28 | 1 | 62.28 | 58.49 | · < 0.0001 | | |
| Factor –C | 448.01 | 1 | 448.01 | 420.80 | < 0.0001 | | |
| AB | 0.062 | 1 | 0.062 | 0.058 | 0.8102 | | |
| AC | 0.37 | 1 | 0.37 | 0.35 | 0.5580 | | |
| BC | 54.50 | 1 | 54.50 | 51.19 | < 0.0001 | | |
| | 0.26 | 1 | 0.26 | 0.24 | 0.6244 | | |
| A ² | 3.30 | 1 | 3.30 | 3.10 | 0.0816 | | |
| B ² | 2.40 | 1 | 2.40 | 2.25 | 0.1367 | | |
| C ² Pure error | 18.57 | 72 | 0.26 | | | | |
| Cor total | 678.37 | 107 | | | Signifi | cant | |

Table M8 Solubility

| Source | Sum of squares | df | Mean square | F value | p- value Prob > F | Std. Dev. | C.V. (%) |
|----------------|----------------|---------|-------------|---------|----------------------|--------------|-------------|
| Model | 4.431E+005 | 9 | 49232.48 | 29.97 | < 0.0001 | 40.53 | 8.50 |
| Factor –A | 19.91 | 1 | 19.91 | 0.012 | 0.9126 | | |
| Factor –B | 20213.76 | 1 | 20213.76 | 12.31 | 0.0007 | | |
| Factor –C | 2.433E+005 | 1 | 2.433E+005 | 148.10 | < 0.0001 | | |
| AB | 460.12 | 1 | 460.12 | 0.28 | 0.5978 | | |
| AC | 56578.57 | 1 | 56578.57 | 34.45 | < 0.0001 | | |
| BC | 43704.94 | 1 | 43704.94 | 26.61 | < 0.0001 | | |
| A ² | 1530.65 | 1 | 1530.65 | 0.93 | 0.3368 | | |
| B^2 | 4923.91 | 1 | 4923.91 | 3.00 | 0.0865 | | |
| C^2 | 71232.24 | 1 | 71232.24 | 43.37 | < 0.0001 | | |
| Pure error | 64615.46 | 72 | 3705.90 | | | | |
| Cor total | 6.041E+005 | 10 7 | | | Signific | ant | |

Table M9 Wettability

| Source | Sum of squares | df | Mean square | F value | p- value Prob > F | Std. Dev. | C.V. (%) |
|------------------------------|----------------|---------|-------------|---------|----------------------|--------------|-------------|
| Model | 4312.35 | 9 | 479.15 | 94.98 | < 0.0001 | 2.25 | 3.95 |
| Factor –A | 119.75 | 1 | 119.75 | 23.74 | < 0.0001 | | |
| Factor –B | 307.28 | 1 | 307.28 | 60.91 | < 0.0001 | | |
| Factor –C | 3592.78 | 1 | 3592.78 | 712.17 | < 0.0001 | | |
| AB | 4.41 | 1 | 4.41 | 0.87 | 0.3523 | | |
| AC | 9.03 | 1 | 9.03 | 1.79 | 0.1841 | | |
| ВС | 104.47 | 1 | 104.47 | 20.71 | < 0.0001 | | |
| A ² | 33.58 | 1 | 33.58 | 6.66 | 0.0114 | | |
| B^2 | 86.15 | 1 | 86.15 | 17.08 | < 0.0001 | | • |
| | 121.58 | 1 | 121.58 | 24.10 | < 0.0001 | | |
| C ² Pure error | 63.38 | 72 | 0.88 | | | | |
| Cor total | 4806.75 | 10 7 | | | Significant | | |

Table M10 Product yield

APPENDIX N

ANOVA FOR MILK FORTIFIED BANANA PSEUDOSTEM JUICE POWDER DURING STORAGE (Product-III)

. . .

Factor –A: Storage period (months)

Factor –**B:** Treatment temperature (°C)

| T UDIC - | | df | Mean | F value | p- value | Std. | C.V. |
|------------|----------------|----|------------|----------------|-----------|-------|------|
| Source | Sum of squares | ui | square | | Prob > F | Dev. | (%) |
| Model | 0.76 | 5 | 0.15 | 112.60 | < 0.0001 | 0.037 | 0.96 |
| Factor –A | 0.16 | 3 | 0.053 | 39.54 | < 0.0001 | | |
| Factor –B | 0.60 | 2 | 0.30 | 222.19 | < 0.0001 | | |
| Pure error | 9.719E-003 | 24 | 1.350E-003 | | | | |
| Cor total | 0.80 | 24 | 4.049E-004 | | Significa | ant | |

Table N1 Moisture content of product during storage

| tivity of product | during | storage |
|-------------------|--------|---------|
|-------------------|--------|---------|

| Table N2 Wa | ater activity of | f pro | duct during s | torage | | | |
|-------------|------------------|-------|----------------|------------|----------------------|----------------|-------------|
| Source | Sum of | df | Mean square | F value | p- value Prob > F | Std. Dev. | C.V. (%) |
| 002- | squares | | | , | | | |
| Model | 0.022 | 5 | 4.391E-003 | 56.28 | < 0.0001 | 8.833E- 003 | 3.53 |
| Factor – A | 0.010 | 3 | 3.423E-003 | 43.87 | < 0.0001 | | |
| | 0.010 | | 5.843E-003 | 74.89 | < 0.0001 | | |
| Factor –B | 0.012 | 2 | 5.843E-005 | / 1.0 / | 0.0001 | | |
| Pure error | 1.418E-003 | | 5.908E-005 | | | | |
| | 1.4100 | 24 | | | Significant | | |
| Cor total | 0.024 | | | | | | |

| Source | Sum of squares | df | Mean square | F value | p- value Prob > F | Std. Dev. | C.V. (%) |
|------------|----------------|----|----------------|---------|----------------------|------------|-------------|
| Model | 9.180E-003 | 5 | 1.836E-003 | 193.18 | < 0.0001 | 3.083E-003 | 0.59 |
| Factor –A | 6.990E-004 | 3 | 2.330E-004 | 24.52 | < 0.0001 | | |
| Factor –B | 8.481E-003 | 2 | 4.240E-003 | 446.18 | < 0.0001 | | |
| Pure error | 1.573E-004 | 24 | 6.556E-006 | | | | |
| Cor total | | 2. | | | Signi | ficant | |
| Cortota | 9.465E-003 | | | | | | |

Table N3 Tapped bulk density of product during storage

Table N4 Loose bulk density of product during storage

| Table N4 L0 | Sum of | df | Mean square | F value | p- value Prob > F | Std. Dev. | C.V. (%) |
|-------------|------------|----|----------------|---------|----------------------|------------|-------------|
| | squares | | | | | | |
| Model | 2.098E-003 | 5 | 4.196E-004 | 44.18 | < 0.0001 | 3.082E-003 | 0.71 |
| Factor –A | 1.027E-003 | 3 | 3.423E-004 | 36.04 | < 0.0001 | | |
| Factor –B | 1.071E-003 | | 5.355E-004 | 56.38 | < 0.0001 | | |
| Pure error | 1.633E-004 | | 6.806E-006 | | | | |
| | 1.0- | 24 | | | Signi | ficant | |
| Cor total | 2.383E-003 | | | | | | |

| Source | Sum of squares | df | Mean square | F value | p- value Prob > F | Std. Dev. | C.V. (%) |
|------------|-------------------|----|----------------|---------|----------------------|--------------|-------------|
| Model | 2013.67 | 5 | 402.73 | 0.77 | 0.5757 | 22.80 | 105.79 |
| Factor –A | 1546.90 | 3 | 515.63 | 0.99 | 0.4100 | | |
| Factor –B | 466.77 | 2 | 233.38 | 0.45 | 0.6426 | | |
| Pure error | 12485.53 | 24 | 520.23 | | | | |
| | | | | | Not sig | nificant | |
| Cor total | 17611.29 | | | | | | |

Table N7 b* value of product during storage

Table N8 Solubility of product during storage

| Source | Sum of squares | df | Mean square | F value | p- value Prob > F | Std. Dev. | C.V. (%) |
|------------|-------------------|----|----------------|---------|----------------------|--------------|-------------|
| Model | 9163.07 | 5 | 1832.61 | 1536.12 | < 0.0001 | 1.09 | 1.68 |
| Factor –A | 3.31 | 3 | 1.10 | 0.92 | 0.4410 | | |
| Factor –B | 9159.76 | 2 | 4579.88 | 3838.91 | < 0.0001 | | |
| Pure error | 33.64 | 24 | 1.40 | | Not sign | nificant | |
| Cor total | 9198.86 | | | | | | |

| Source | Sum of squares | df | Mean square | F value | p- value Prob > F | Std. Dev. | C.V. (%) |
|------------|----------------|----|--------------------|------------|----------------------|--------------|-------------|
| Model | 3.219E+006 | 5 | 6.438E+005 | 81000.43 | < 0.0001 | 2.82 | 0.32 |
| Factor –A | 104.20 | 3 | 34.73 | 4.37 | 0.0115 | | |
| Factor –B | 3.219E+006 | 2 | 1.609E+006 7.95 | 2.025E+005 | < 0.0001 | | |
| Pure error | 42.55 | 24 | ,,,,, | N | ot significan | ıt | |
| Cor total | 3.219E+006 | | 1.77 | | | | |

Table N9 Wettability of product during storage

APPENDIX O

COST ESTIMATION OF SPRAY DRIED BANANA PSEUDOSTEM JUICE BASED POWDERS

| | Fixed cost | |
|------------------------------|------------|----------------|
| Machinery cost | | |
| | = | Rs 15,00,000/- |
| Cost of spray dryer | | Rs 68,681/- |
| Cost of steam blancher | = | Rs 25,000/- |
| Cost of slicer | = | Rs 10,000/- |
| Cost of juicer cum filter | = | Rs 10,000/- |
| Cost of electric balance | = | Rs 1500/- |
| Cost of hand sealing machine | = | Rs 1,615,181/- |
| Total cost of machineries | | |

Building cost

Furniture cost

Water charge per annum

Installation charge

Assumptions

Working hours per shift

No. of shifts per day

Total capacity of unit/day

Life span of unit (L)

Annual working hours (H)

Interest on initial cost (i) Repair and maintenance Fixed cost

Where, C

Hence, Fixed cost

Total fixed cost

- = Rs 72.00.000/-
- = Rs 1,00,000/-
- = Rs 3,60,000/-
- = Rs 50,000/-
- = 8 h
- = .2, .
- = 250 kg/day
- = 15 Years
- = 300 days (per day 16 h)
- = 4800 h
- = 11% annually
- = 2% of initial cost

$$= \frac{i(i+1)^n}{(i+1)^n+1} \times C$$

= Total cost of (machineries

+building + furniture +

installation)

= 1615181+7200000+100000

+50000

- = Rs 8965181/-
- $= \frac{0.11(0.11+1)^{15}}{(0.11+1)^{15}+1} \times (8965181)$
- = Rs 814,935/-
- Total fixed cost of
 (machinery + building cost
 + furniture + installation+
 water charge)

| | = | 814935+360000 |
|---------------|---|------------------------|
| | = | Rs 1174935/- |
| Fixed cost /h | = | $\frac{1174935}{4800}$ |
| | = | Rs 245/- |

Variable cost

| a) Repair and maintenance charge of (equipments | = | 2% of initial cost | |
|---|---|--------------------|--|
| + furnitures + building) | | | |

| = | $\frac{2}{100 \times 4800} \times 8915181$ |
|---|--|
| = | Rs 37.14/h |

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. . .

| b) Cost of energy | | |
|--|---|----------------|
| i) Energy requirement (motor and heating coil) | = | 131.57 KWh/16h |
| i) Energy required for steam blancherii) Energy required for steam blancher | = | 30 KWh/16 h |
| ii) Energy required for slicer and juicer cum filter iii) Energy required for slicer and juicer cum filter | = | 11.2 KWh/16h |
| iii) Energy requirement for electric balance andiv) Energy requirement for electric balance and | = | 7.2 KWh/16h |
| iv) Energy requirement for the | | |

sealer

| Total energy requirement | = | i + ii + iii + iv + v 182.21 KWh/16 h |
|--|---|--|
| Electric charges Electric consumption charges | = | Rs 7.5 /KWh Power × duration × cost for 1 unit |
| | = | 182.21×16×7.5 |
| | = | Rs 21865.200/h |
| c) Labour charges Three women labour @ Rs 300/shift | = | Rs. 900/- Rs 1000/- |
| Two men labour @ Rs 500/shift Labour charge /day | = | Rs 3800/- |

d) Cost of raw material required per day

| a tity (kg) | Unit rate (per kg) | Total amount (Rs) |
|-------------|-------------------------|---|
| | 3 | 3000 |
| - | 40 | 10000 |
| _ | 35 | 5250 |
| - | 80 | 320 |
| | aterials | Rs 18570.00/- |
| | 1000 250 150 4 | 1000 3 250 40 150 35 80 |

| Product-II | Quantity (kg) | Unit rate (per kg) | Total amount (Rs) |
|----------------------------|---------------|--------------------|-------------------|
| Raw materials | | 3 | 2460 |
| Banana | 820 | 40 | 10000 |
| pseudostem Maltodextrin | 250 | 40 | |
| | 150 | 60 | 9000 |
| Horse gram | 4 | 80 | 320 |
| Ginger | materials | | Rs 21780.00 /- |
| Total cost of raw | Illateration | | |

| Product-III Raw materials | Quantity (kg) | Unit rate (per kg) | Total amount (Rs) |
|-----------------------------------|-------------------|--------------------|--------------------------|
| | 200 | 3 | 1200 |
| Banana | 200 | | |
| pseudostem Milk | 500 | 40 | 20000 |
| | 150 | 60 | 9000 |
| Horse gram Cardamom | 0.5 | 600 | 300 |
| Total cost of raw | materials | | Rs 30500/- |
| l otal cost of fatt | | | |
| e) Packaging cos | st | = | 4 kg |
| · ···· ···· | mired/day | _ | , ng |
| Total quantity rec | | = | Rs. 150 |
| | Unit rate | _ | |
| | Total amount | = | Rs 600/- |
| | | | |
| | | = | a + b + c + d + e |
| Total variable cos | st | | |
| | | = | Rs 44872.340/- |
| Product-I | | = | Rs 48082.340/- |
| Product-II | | = ' | Rs 56802.340/- |
| Product-III | | 1 | |
| Product-III Total cost of proc | luction of 250 kg | powder (m) = | Total fixed cost + Total |
| | | | variable cost |
| | | = | Rs 48712.340/- |
| Product-I | | = | Rs 52002.340/- |
| Product-II | | = | Rs 60722.34/- |
| Product-III | | | |
| 11000 | | | |

| Cost of production of 1 Kg of powder/year | = | <u>m</u> 250 |
|---|-----|--------------------------|
| Product-I | = | Rs 195/kg |
| Product-II | = | Rs 208/kg |
| Product-III | = | Rs 243/kg |
| Cost of 50 g packet | | |
| Product-I | = | Rs. 9.75/- |
| | = | Rs 10.40/- |
| Product-II | = | Rs. 12.15/- |
| Product-III | | |
| Expected selling cost | | |
| Product-I | = | Rs 400/kg |
| | = | Rs 450/kg |
| Product-II | = | Rs 550/kg |
| Product-III | | |
| Benefit cost ratio | | |
| Product-I | = | $\frac{400}{195} = 2.05$ |
| Product-II | = . | $\frac{450}{208} = 2.16$ |
| Product-III | = | $\frac{550}{243} = 2.26$ |

Therefore the total production cost of 1kg of micro encapsulated banana pseudostem based powders (Product-I, II and III) was estimated as Rs. 195/-, Rs 208/- and Rs 243 respectively and the benefit cost ratio of the production was found to be 2.05:1, 2.16:1 and 2.26:1 respectively.

• • **Abstract**

DEVELOPMENT AND QUALITY EVALUATION OF MICROENCAPSULATED BANANA PSEUDOSTEM JUICE POWDER

by

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SARANYA S (2014-18-110)

ABSTRACT OF THE THESIS

Submitted in partial fulfillment of the requirement for the degree of MASTER OF TECHNOLOGY

IN

AGRICULTURAL ENGINEERING

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Abstract

Banana pseudostem, often discarded after the harvest of bunch is very good for health. Its disposal in the field lead to unhygienic surroundings and environmental pollution. Juice from banana stem is a well-known remedy for urinary disorders. But the major problem associated with the pseudostem juice is its perishability and immediate browning reactions which lead to reduction of its acceptability by consumers. Considering these facts, a study was undertaken to obtain powdered products from pseudostem juice. The intention of the study was to develop a process protocol for microencapsulated banana pseudostem juice powder, standardisation of the spray drying parameters, and quality analysis of developed product. Three powder based products were developed from banana pseudostem juice by spray drying technology. Product-I comprised of pseudostem juice-sugar combination with ginger as flavourant. Product-II consists of a blend of banana pseudostem and horse gram with ginger extract. However, the third product from banana pseudostem juice was fortified with milk, horse gram extract and cardamom flavour. The process parameters were optimised as inlet temperature of 180°C and outlet temperature of 65-68°C for product-I & II, whereas inlet air temperature of 185°C and outlet temperature of 74-92°C were chosen for Product-III. The feed pump rpm of 15 and main blower rpm of 1800 were kept constant for developing all three products. The physicochemical characteristics, reconstitution and flow properties were determined. Standardised products were stored in aluminium pouches and quality parameters of product-I and II were analysed up to six months at an interval of two months and Productand the months for verifying its stability during storage. Based on quality analysis and sensory evaluation, best samples were selected from on your selected from product-I, II and III *i.e.*, T6-180°C (15% sugar + 25% maltodextrin + 56% pseudostem juice), T6-180°C (25% maltodextrin + 30% horse gram extract + 43% pseudostem juice), and T12-185°C (50% milk + 30% horse gram extract + 20% pseudostem juice), respectively. Cost analysis of the products was done and cost pseudostem store kilo gram was set production of one kilo gram was estimated as Rs.195/-, Rs.208/- and Rs.243/-of production III respectively. for product I, II and III, respectively.