

**INTERACTION OF PHOSPHORUS AND SULPHUR IN BLACK  
COTTON SOILS OF PALAKKAD (AEU: 23) UNDER  
GROUNDNUT (*Arachis hypogaea* L.) CULTIVATION**

**SHAHANA, C. K.  
(2017-11-040)**

**THESIS**

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**SOIL SCIENCE AND AGRICULTURAL CHEMISTRY**

**COLLEGE OF HORTICULTURE**

**VELLANIKKARA, THRISSUR**

**KERALA, INDIA**

**2020**

## DECLARATION

I, hereby declare that the thesis entitled “**Interaction of phosphorus and sulphur in black cotton soils of Palakkad (AEU: 23) under groundnut (*Arachis hypogaea* L.) cultivation**” is a bonafide record of research work done by me during the course of research and the thesis has not previously formed the basis for the award to me of any degree, diploma, associateship, fellowship or other similar title, of any other university or society.

Vellanikkara

Date: 18/01/2020



**SHAHANA, C. K.**

**2017-11-040**

## CERTIFICATE

Certified that the thesis entitled “**Interaction of phosphorus and sulphur in black cotton soils of Palakkad (AEU: 23) under groundnut (*Arachis hypogaea* L.) cultivation**” is a record of research work done independently by **Ms. Shahana, C.K.** under my guidance and supervision and that it has not previously formed the basis for the award of any degree, diploma, fellowship or associateship to her.



**Dr. Beena V. I.**

(Major Advisor)

Assistant Professor (SSAC)

and Head in charge (Radio Tracer  
Laboratory)

College of Horticulture, Vellanikkara.

Vellanikkara

Date: 18/01/2020

## CERTIFICATE

We, the undersigned members of the advisory committee of **Ms. Shahana, C. K.** (2017-11-040), a candidate for the degree of **Master of Science in Agriculture**, with major field in Soil Science and Agricultural Chemistry, agree that the thesis entitled “**Interaction of phosphorus and sulphur in black cotton soils of Palakkad (AEU: 23) under groundnut (*Arachis hypogaea* L.) cultivation**” may be submitted by **Ms. Shahana, C. K.** in partial fulfilment of the requirement for the degree.

**Dr. Beena, V. I.**  
Assistant Professor (SSAC)  
and Head in charge (Radio Tracer Laboratory)  
College of Horticulture, Vellanikkara

**Dr. Jayasree Sankar S.**  
Professor & Head  
Department of Soil Science &  
Agricultural Chemistry  
College of Horticulture, Vellanikkara

**Dr. P. Sureshkumar**  
Professor & Head (Retd.) Radiological  
Safety Officer  
Radiotracer Laboratory  
College of Horticulture, Vellanikkara

**Dr. C. Beena**  
Professor, AICRP on Medicinal and  
Aromatic Plants  
College of Horticulture, Vellanikkara



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## ABBREVIATIONS

B	Boron
Ca	Calcium
CD	Critical difference
CEC	Cation exchange capacity
cm	Centimeter
Cu	Copper
DAP	Days after planting
dS	Deci seimens
EC	Electrical conductivity
Fe	Iron
FYM	Farm yard manure
Ha	Hectare
K	Potassium
kg	Kilogram
LAI	Leaf area index
M	Meter
Mg	Magnesium
mg	Milligram
Mn	Manganese
MOP	Muriate of potash
N	Nitrogen
OC	Organic carbon
P	Phosphorus

S Sulphur

Zn Zinc

# Introduction

## 1. INTRODUCTION

In Kerala, black cotton soils are seen in Chittur taluk of Palakkad district occupying an area of 2000 ha (Padmaja *et al.*, 1994). These soils are dark, low in organic matter, calcareous, neutral to alkaline (pH 7.0 to 8.5), high in clay content and CEC. The texture of soil ranges from clay loam to clay. The level of total nitrogen in the soil is satisfactory but, only very small fraction of phosphorus is in available form (less than 1 per cent) due to the process of fixation under high pH and clay content.

Even though these soils are fertile, the nutrient imbalances and poor physical conditions may adversely affect the yield of the crop (Krishnakumar, 1978; Padmaja *et al.*, 1994). Balanced supply of nutrients is one of the most important factors determining crop yield. Sometimes the applied nutrients may not be available for plant use, as their availability depends on interaction between the applied nutrients. When the supply of one of the nutrients affects absorption and utilization of the other nutrient element, the elements are said to be in interaction. Interaction between nutrients in soil affects the overall performance of crop. Nutrient interaction may be negative or positive. When nutrients are in combination and results in a greater response than individual response, the interaction is positive (synergism). When combination results in a lesser response, then interaction is negative (antagonism).

The black cotton soils of Chittur area is deficient in available P and S. The availability of P to plants for uptake and utilization is impaired in alkaline soils due to the formation of poorly soluble calcium phosphate.

The groundnut (*Arachis hypogaea* L.) belonging to the family *leguminosae*, originated in South America (southern Bolivia/north west Argentina region) and cultivation started as early as 1000 B.C. Groundnut is an important oilseed crop, about two third of world's groundnut production is used for oil production.

Palakkad is the only district in Kerala where groundnut cultivation is practiced in a large and commercial scale. Thus the area and production are also high in Palakkad district. Chittur taluk contributes a major share to this.

The nut (kernel) of groundnut is a rich source of edible oil, containing 36 to 54% oil and 25 to 32% protein. Even though groundnut can grow in soils of marginal fertility, proper fertilizer application will help to achieve full yield potential of crop. Groundnut has the capacity to utilize soil nutrients that are relatively unavailable to other crops, and therefore can make good use of residual fertility (Cox *et al.*, 1982).

Phosphorus is the second limiting nutrient for crop production (Mallikarjuna *et al.*, 2003). In majority of Indian soils, available P ranges from low to medium. It stimulates setting of pods, decreases the number of unfilled pods (pops) and hastens the maturity of the crop. P enhances nitrogen use efficiency by plants. It is essential for energy storage and transfer and hence called “energy currency” of the living system.

Sulphur is one of the most limiting nutrient for groundnut production, as it has a role in oil content, protein content and quality of kernels. Sulphur has vital role in the metabolism of groundnut plant. It is important for the synthesis of proteins. It helps in biological oxidation-reduction processes. Sulphur deficiency leads to stunted growth and chlorosis and delay maturity in groundnut crop.

In this context present study entitled “Interaction of phosphorus and sulphur in black cotton soils of Palakkad (AEU: 23) under groundnut (*Arachis hypogaea* L.) cultivation” was carried out with following objectives:

1. To find out the interactions of sulphur and phosphorus in black cotton soils of Palakkad.
2. To assess the treatment level of sulphur and phosphorus for maximizing groundnut yield.



# Review of Literature

## 2. REVIEW OF LITERATURE

The results of various experiments conducted in India and elsewhere on nutrient uptake, growth, yield and quality parameters of groundnut and some related oilseed crops under the influence of different nutrients and their interaction effects are reviewed in this chapter.

### 2.1 EFFECT OF PRIMARY NUTRIENTS ON GROWTH PARAMETERS

#### *2.1.1 Effect of nitrogen on growth parameters*

Nitrogen (N) has an important role in the metabolism of plants and is an important structural constituent of the plant cell (Mahapatra *et al.*, 1985). Groundnut is a leguminous crop, with a biological nitrogen fixation capacity of 200-260 kg N ha<sup>-1</sup> with the help of root nodules, which reduces the demand for applied N. Even though groundnut can fix atmospheric N, it shows good response to the application of nitrogenous fertilizers (York and Colwell, 1951 and Williams, 1979).

Application of N in early stages has beneficial effect on growth parameters of groundnut (Reddy and Rao, 1965). There was a significant increase in the number of leaves, branches and height of groundnut plant as a result of application of nitrogen (Punnoose, 1968). Singh and Ahuja (1985) observed increase in growth of groundnut due to the application of N at 25 kg ha<sup>-1</sup>.

#### *2.1.2 Effect of phosphorus on growth parameters*

Phosphorus (P) is the second major essential nutrient element for crop growth. The most important effect of P is in the plant root system. Phosphorus requirement in nodulating legumes is higher compared to non-nodulating crops as it has an integral role in nodule formation and also in the fixation of atmospheric nitrogen (Brady and Weil, 2002). Phosphorus is essential for the storage and transfer of energy. Phosphate group is the major constituent of adenosine di and

tri phosphate (ADP and ATP) known as energy currency of plants (Tisdale *et al.*, 1993). P requirement of plants is met by the uptake of phosphate anions from soil solution. P is important for the formation and growth of roots and N fixation (Lakshamma and Raj, 1997).

Phosphorus application at 132 kg ha<sup>-1</sup> increased haulm yield in groundnut (York and Colwell, 1951). Punnoose (1968) studied the effect of P on growth, yield and quality of groundnut and observed that application of P increased the number of leaves, branches, height of the plant and weight of nodules per plant. Patel *et al.* (1994) reported that higher dose of phosphatic fertilizer application increased the number of root nodules. The number and weight of root nodules, activity of nitrogenase enzyme, leghaemoglobin content, leaf area and dry matter production increased significantly by enhancing the P<sub>2</sub>O<sub>5</sub> content from 0 to 30, 60 and 90 kg ha<sup>-1</sup>.

Shelke and Khuspe (1980) observed highest dry matter production and P uptake by groundnut cv. Latur No.33 as a result of the application of 17.5 kg P<sub>2</sub>O<sub>5</sub> ha<sup>-1</sup>. Basha and Rao (1980) reported decrease in number of leaves and length of shoot in groundnut due to the deficiency of P. Higher levels of P increased growth of root and shoot significantly (Patel *et al.*, 1994). Change in the rate of P application from 30 to 90 kg ha<sup>-1</sup> found to enhance the growth of plants (Singh and Ahuja, 1985).

Sebale and Khuspe (1986) observed higher plant height, number of leaves, branches and dry weight per plant due to the application of P at the rate of 60 kg ha<sup>-1</sup>. P application increased the plant height and dry matter yield in groundnut crop (Juan *et al.* 1986). Kamara *et al.* (2011) reported an increase in biomass of groundnut after the application of phosphorus fertilizer and attributed it to the availability of soluble phosphate that enhanced extensive root development.

### **2.1.3 Effect of potassium on growth parameters**

Rao (1979) reported an increased dry matter production with the application of higher levels of K in groundnut cv. TMV-2. Mathew *et al.* (1983) observed that growth parameters such as plant height, number of branches, number of leaves per plant and Leaf Area Index (LAI) were increased with potassic fertilizer application.

## **2.2 EFFECT OF PRIMARY NUTRIENTS ON YIELD AND YIELD ATTRIBUTES**

### **2.2.1 Effect of nitrogen on yield and yield attributes**

Plants require N in larger amounts compared to other elements. As groundnut belongs to *leguminosae* family, it can fix 40-80 kg N ha<sup>-1</sup> yr<sup>-1</sup> (Islam and Noor, 1982). Dart *et al.* (1983) reported that about 86-92 per cent of N uptake by groundnut occurred through biological nitrogen fixation (BNF) which is equivalent to 125-178 kg N ha<sup>-1</sup>.

Jadhar and Narkhende (1980) concluded that N played a significant role on the number of pods as well as number of filled pods per plant. According to Saradhi *et al.* (1990) higher doses of nitrogen led to production of more number of flowers and pegs in groundnut crop. Hasan (2018) concluded that there was a significant increase in yield of groundnut due to the application of nitrogen. Increase in pod yield and its subsequent reduction was observed due to the application of nitrogen @ 10-30 kg ha<sup>-1</sup> (Pant and Katiyar, 1996; Patel *et al.*, 1994) whereas, application of 40-60 kg ha<sup>-1</sup> nitrogen increased number of pods per plant (Reddy *et al.*, 1984).

Reddy and Rao (1965) reported significant reduction in yield of groundnut by the application of nitrogen @ 40 kg ha<sup>-1</sup>. Nijhawan and Maini (1966) observed increase in yield of groundnut crop even at application of small doses of nitrogen. Study conducted by Puntamkar and Bathkal (1967) indicated that application of nitrogen at the rate of 20 kg ha<sup>-1</sup> significantly increased the

number of pods per plant and its weight in groundnut. Saini and Tripathi (1973) also investigated the effect of nitrogen on groundnut and concluded that application of  $15 \text{ kg ha}^{-1}$  nitrogen showed highest pod yield and oil content.

### ***2.2.2 Effect of phosphorus on yield and yield attributes***

Legume plants require higher amount of phosphorus compared to non-legume crops because of its role in the formation of nodules and atmospheric nitrogen fixation (Brady and Weil, 2002). Balasubramanian and Palaniappan (1991) revealed that higher the amount of phosphorus, higher will be the quantity of nitrogen fixed.

Total uptake of nitrogen and its proportion in kernel were highly influenced by the level of phosphorus and interaction between phosphorus and potassium showed significant influence on kernel yield (Balasubramanian and Palaniappan, 1991). Bala *et al.* (2011) opined that increased pod and seed index and shelling per cent of groundnut were due to early and greater availability of nitrogen and phosphorus to plants which favorably influenced the development and size of kernels. According to Samtana *et al.* (1994) there was a significant improvement in yield attributes of groundnut by the application of P. This improvement was due to the production and proliferation of new roots which led to their improved functional activity.

Ae *et al.* (1996) opined that in acid soils, groundnut showed superior ability to take up phosphorus from a soil with low P fertility status compared with sorghum and soybean. They also concluded that root cell walls of groundnut are characterized by higher P-solubilizing activity compared to those of soybean or sorghum. Response of crop to phosphatic fertilizer application is influenced by initial available P content in the soil. Agasimani and Hosmani (1989) revealed that response of applied P could be obtained when the available soil phosphorus content is less than  $35 \text{ kg ha}^{-1}$ .

Rao *et al.* (1984) reported that application of P above  $60 \text{ kg ha}^{-1}$  had no significant effect on number of pods and it depended on the fertility status of soil.

Hasan (2018) reported that there was a significant increase in yield of bambara groundnut due to the application of phosphorus along with nitrogen. Chauhan *et al.* (1987) opined that there was increase in shelling per cent as a result of application of moderate to high level of P. Banerjee *et al.* (1967) viewed an increase in yield of groundnut by the application of  $P_2O_5$  up to the level of 67.2 kg ha<sup>-1</sup>. Puri (1969) observed a significant response of groundnut crop to the application of superphosphate. According to Choudhary (1979), the pod yield of irrigated groundnut variety, TMV-2 was higher when applied with 60 kg  $P_2O_5$  ha<sup>-1</sup> than with 30 kg  $P_2O_5$  ha<sup>-1</sup>. Nakagawa *et al.* (1981) reported that application of 40 kg  $P_2O_5$  ha<sup>-1</sup> led to increase in pod yield from 1.42 to 2.5 t ha<sup>-1</sup> and seed yield from 0.91 to 1.58 t ha<sup>-1</sup>. The P application increased seed size and 100 pod weight.

### ***2.2.3 Effect of potassium on yield and yield attributes***

Potassium nutrition showed favorable effect on photosynthesis as well as in translocation of food reserves from leaves to developing pods (Koch and Mengal, 1977). Groundnut is a heavy feeder of potassium. Adequate supply of this nutrient must be given to obtain a better yield (Geethalakshmi *et al.*, 1993). York and Colwell (1951) observed that groundnut grew well even in potassium deficient soils where other crops could not grow.

Yakadri and Sathyanarayana (1992) investigated the effect of application of  $K_2O$ , and found that 40-60 kg ha<sup>-1</sup> was the optimum dose of K for groundnut. Whereas, Nair *et al.* (1981) revealed that application of potassic fertilizer @ 80 kg ha<sup>-1</sup> increased the number of pods per plant. According to Ramanathan *et al.* (1982) application K fertilizer at 50 kg ha<sup>-1</sup> resulted in maximum number of pods per plant and highest test weight of seed.

According to Hadwani and Gundalia (2005), K fertilizer application increased the yield of pod and haulm. Study conducted by Loganathan and Krisnamoorthy (1980) concluded that there was an increase in yield and yield contributing characters as a result of increased K application level. If the K level

in the pod zone is high, it is undesirable as it resulted in pod rot and interfered with uptake of Ca by pegs and pods, which in turn led to a higher per cent of pods formation and Ca deficiency in the seeds (Hallock and Garren, 1968; Csinos and Gaines, 1986).

Higher level of K increased the number of pods per plant and test weight of seeds in groundnut variety, TMV-2 (Rao, 1979). Loganathan and Krishnamoorthy (1980) emphasized that optimum dose of potassium for irrigated groundnut crop was  $52 \text{ kg ha}^{-1}$  and for rainfed crop was  $26 \text{ kg ha}^{-1}$ . It was identified by Chavan and Kalra (1983) that dry pod yield, shelling per cent, 1000 grain weight and oil content of groundnut cv. TG- 1 were higher when applied with  $50 \text{ kg ha}^{-1}$  of K than with  $25 \text{ kg ha}^{-1}$ .

### 2.3 EFFECT OF COMBINED APPLICATION OF PRIMARY NUTRIENTS ON GROUNDNUT

Venkateswaralu and Nath (1989) showed the importance of balanced fertilizer schedule and its influence on groundnut. Das (1982) reported that growth components were increased by the application of NPK. Combined application of NPK at the rate of 20: 40: 40  $\text{kg ha}^{-1}$  gave highest yield (Pradhan and Das, 1989). Yadav (1990) also reported that the application of NPK at 20: 60: 40  $\text{kg ha}^{-1}$  resulted in highest yield in groundnut crop. Whereas Balasubramaniam and Palaniappan (1991) opined that the application rate of 150 kg N and 50  $\text{kg K}_2\text{O ha}^{-1}$  resulted in higher yield.

According to Patel *et al.* (1994) application of nitrogen @  $25 \text{ kg ha}^{-1}$  along with  $50 \text{ kg P}_2\text{O}_5 \text{ ha}^{-1}$  increased the pod and haulm yield of groundnut cv. GAUG-1. Kachot *et al.* (1984) recorded higher number of pegs per plant, number of pods per plant, pod weight and test weight per plant when groundnut crop was supplied with  $12.5 \text{ kg N ha}^{-1}$  and  $50 \text{ kg P}_2\text{O}_5 \text{ ha}^{-1}$ . Rana *et al.* (1984) observed that higher pod yield of  $23.19 \text{ q ha}^{-1}$  was obtained by the application of  $20 \text{ kg N ha}^{-1}$ ,  $60 \text{ kg P}_2\text{O}_5 \text{ ha}^{-1}$  and  $40 \text{ kg K}_2\text{O ha}^{-1}$ .

Application of NPK at the rate of 50: 100: 50 kg NPK ha<sup>-1</sup> significantly increased the number of branches per plant (Dholaria *et al.* 1972).

Long term application of manures and fertilizers significantly influenced the yield and productivity of groundnut crop grown in alfisols of Chittoor taluk in Andhra Pradesh (Parvathi *et al.*, 2015). Ghadekar *et al.* (1993) reported that pod yield was highest at the fertilizer application rate of 40 kg ha<sup>-1</sup> N, 80 kg ha<sup>-1</sup> P<sub>2</sub>O<sub>5</sub> and 30 kg ha<sup>-1</sup> K<sub>2</sub>O. The application of NPK @ the rate of 25 kg ha<sup>-1</sup> N + 75 kg ha<sup>-1</sup> P<sub>2</sub>O<sub>5</sub> + 37.5 kg ha<sup>-1</sup> K<sub>2</sub>O gave mean pod yield of 3.55, 4.10 and 4.99 t ha<sup>-1</sup> respectively (Thimmegowda, 1993).

A study conducted by Sireesha *et al.* (2017) found that higher yield from groundnut crop was obtained when supplied with 50 per cent of recommended dose along with 4 t ha<sup>-1</sup> FYM. Dahatonde (1982) reported that combined application of organic manure and inorganic fertilizers recorded favorable effects on various growth parameters and yield attributing characters of groundnut. They revealed that application of 25 kg N and 50 kg P<sub>2</sub>O<sub>5</sub> ha<sup>-1</sup> along with 5 t ha<sup>-1</sup> FYM recorded highest plant height, spread, no.of branches per plant and total dry matter per plant at harvest and yield attributes *viz.*, filled pod per plant, dry pod per plant and pod and haulm yield of summer groundnut.

Kuchanwar *et al.* (1997) opined that highest nitrogen and phosphorus uptake was observed with combination of 25:50 kg N and P<sub>2</sub>O<sub>5</sub> ha<sup>-1</sup> respectively. According to Dubey (1997) application of single super phosphate (SSP) at 60 kg P<sub>2</sub>O<sub>5</sub> ha<sup>-1</sup> gave highest, but was on par with 30 kg P<sub>2</sub>O<sub>5</sub> ha<sup>-1</sup> as SSP along with *Pseudomonas striata* with regard to N, P and K uptake (straw + grain) in black or medium clay soil (Vertisol). Shipkule *et al.* (2008) observed that application of 80:60:20 kg ha<sup>-1</sup> N, P<sub>2</sub>O<sub>5</sub> and K<sub>2</sub>O respectively gave maximum nutrient content as well as highest uptake of nitrogen, phosphorus and potassium by kernel and haulm of groundnut. According to Sanchez and Owen (1978), application of 150 kg P<sub>2</sub>O<sub>5</sub> ha<sup>-1</sup> along with nitrogen and potassium increased pod yield from 0.75 to 2.07 t ha<sup>-1</sup>.



Babu *et al.* (2007) observed that highly fertilized plots had higher uptake of NPK by the groundnut crop. But this higher uptake did not contribute to higher yield but enhanced vegetative growth. Dholaria *et al.* (1972) reported that pod weight, number of pods and number of branches per plant increased under higher fertilizer application rate (50: 100: 50 kg NPK ha<sup>-1</sup>).

## 2.4 EFFECT OF SECONDARY NUTRIENTS ON GROWTH AND YIELD ATTRIBUTES

### 2.4.1 Effect of Ca on growth and yield parameters

Groundnut plants require calcium (Ca) from the beginning of pegging stage, fruit formation, until the maturity of pods (Walker, 1975). Ca deficiency led to high per cent of aborted seeds (empty pods), improperly filled pods and caused aborted or shrivelled fruit, including darkened plumules and production of pods without seed (Singh and Oswalt, 1995).

Mandal *et al.* (2005) reported that application of gypsum in summer and rainy season groundnut in sandy loam soils of West Bengal @ the rate of 400 kg ha<sup>-1</sup> showed highest plant height (65.1cm). Calcium plays an important role in proper development of pod and production of high quality seeds (Cox *et al.*, 1982). Calcium deficiency led to lowering of yield, darkening of plumule in the seed, empty pods and sometimes plants remained green and continued to produce flowers and pegs without pods, that might be infertile (Sumner and Farina, 1986).

Application of soluble source of Ca helped to avoid Ca deficiency at early flowering stage. The surrounding soil of developing pods require high Ca level, as the calcium absorbed by root did not translocate into pods and the required calcium for pod development was directly absorbed from the soil solution (Skelton and Shear, 1971).

According to Sumner and Farina (1986) and Kvien *et al.* (1988) soil Ca level in the range of 600-800 mg kg<sup>-1</sup> produced good quality groundnut kernel.

Kvien *et al.* (1988) reported that, morphological characters of pod such as surface area, volume, number of days for maturation of and shell thickness significantly influenced Ca uptake by the pods. Ca requirement showed variation with the size of seeds. Small seeded cultivars required less quantity of calcium compared to larger seeded types, because of its larger surface to volume ratio. Soil characters such as soil moisture content, soluble and exchangeable calcium and type of mineral present in the soil affected the uptake of Ca by groundnut (Keisling *et al.*, 1983).

#### ***2.4.2 Effect of magnesium on growth and yield parameters***

Dowood (1982) opined that application of three levels of Mg *i.e.* 0, 120 and 240 kg MgSO<sub>4</sub>.7H<sub>2</sub>O ha<sup>-1</sup> significantly increased phosphorus uptake by the plant. Al-lami (1999) reported that increase in addition of MgSO<sub>4</sub>.7 H<sub>2</sub>O from 0 to 80 kg ha<sup>-1</sup> resulted in significant increase in available phosphorus content in soil from 0.23 to 0.25 c mol kg<sup>-1</sup>.

#### ***2.4.3 Effect of sulphur on growth and yield parameters***

Rao *et al.* (2013) found that application of S at 45 kg ha<sup>-1</sup> as gypsum recorded highest plant height (71.45 cm) in sandy loam soils of Andhra Pradesh. Application of S influenced growth, yield attributing characters, yield and oil content regardless of the sources and levels of S. Addition of S at 45 kg ha<sup>-1</sup> recorded highest plant height, number of filled pods per plant, 100 pod weight, 100 kernel weight, pod yield, haulm yield and oil content of kernels in groundnut (Rao *et al.*, 2013). The S uptake by pods increased significantly with increasing levels of S and maximum uptake (10.89 kg ha<sup>-1</sup>) was noticed with application of 60 kg S ha<sup>-1</sup>.

Application of sulphur @ 60 kg ha<sup>-1</sup> in groundnut gave higher number of total pods plant<sup>-1</sup> (37.80), 100 pods weight (96.82 g), 100 seeds weight (46.25 g), shelling per cent (85.29%), pod yield (3.13 t ha<sup>-1</sup>), seed yield (2.67 t ha<sup>-1</sup>), stover yield (6.84 t ha<sup>-1</sup>), and harvest index (31.37 %) when compared with other treatments such as 0, 15, 30 and 45 kg ha<sup>-1</sup> of S (Nurezannat *et al.*, 2019). Singh

and Chaudhari (2008) observed that in calcareous soil, plants grown by addition of S had increased plant height, number of flowers, number and weight of nodules, higher dry matter, seed, haulm (leaves and stems), and oil content compared to those without S. Babu *et al.* (2007) reported that application of S through gypsum @ 40 kg ha<sup>-1</sup> recorded highest pod and haulm yield. Giri *et al.* (2014) observed that number of pods per plant was highest in sandy loam soils when S was applied at 15 kg ha<sup>-1</sup> (25.52). Umadevi *et al.* (1999) reported that pod yield enhanced by increasing S levels from 15 to 30 and 75 kg ha<sup>-1</sup> in red loamy sandy soils of Ananthapur, Andhra Pradesh. Highest pod yield was recorded by addition of S at 75 kg ha<sup>-1</sup>.

## 2.5 EFFECT OF MICRONUTRIENTS ON GROWTH AND YIELD ATTRIBUTES

### 2.5.1 Effect of micronutrients on growth and yield parameters

Soil pH, cation and anion exchange capacity and nutrient interactions are the factors affecting availability of micronutrients in soils. The application of B promoted absorption of N by groundnut and increased plant height, dry weight and the total number of pods per plant. Study conducted by Bharthi *et al.* (2010) revealed that application of micronutrients (Fe, Zn, B) along with recommended dose of fertilizers resulted in improvement of growth characters and chlorophyll content in groundnut. They observed that application of 20 kg ha<sup>-1</sup> ZnSO<sub>4</sub> and 5 kg borax gave highest pod yield and dry matter content.

Mahamoud *et al.* (2006) observed that foliar application of boron at 25-50 ppm increased plant height, leaf area, total dry matter, number of pods and seed yield. Study conducted by Ravichandra *et al.* (2015) indicated that foliar spray of boron along with rhizobium in flowering and pod formation stages had positive impact on growth and yield of groundnut as it increased plant height, number of branches, number of pods per plant, plant dry weight, 100 pod weight, pod yield and seed index. Excess foliar application of boron led to decrease in the above mentioned growth and yield parameters.

Kamalakaran and Ravichandran (2013) recorded highest plant height at all critical stages of groundnut as a result of application of 100 per cent NPK, 25 kg ha<sup>-1</sup> of ZnSO<sub>4</sub>, 10 kg ha<sup>-1</sup> of boron and 12.5 t ha<sup>-1</sup> FYM. Subrahmaniyan *et al.* (2001) suggested that combined application of borax at 5 kg ha<sup>-1</sup>, ZnSO<sub>4</sub> at 5 kg ha<sup>-1</sup> and ferrous sulphate at 10 kg ha<sup>-1</sup> recorded maximum number of pods per plant. Reddy *et al.* (2011) emphasized that soil application of micronutrients *viz.*, ZnSO<sub>4</sub> at 10 kg ha<sup>-1</sup>, borax at 5 kg ha<sup>-1</sup> and copper sulphate at 5 kg ha<sup>-1</sup> resulted in increase in number of pods per plant.

Mahajan *et al.* (1994) reported that boron at 0.5 kg ha<sup>-1</sup> applied as boronated super phosphate or borax increased dry pod yield (3200 kg ha<sup>-1</sup>) followed by spray of 0.5 ppm boron in 2 sprays in such a way that first at 30 days after germination and second at the time of flowering gave higher yield than control. According to Singh *et al.* (2009) soil application of boron at 1 kg ha<sup>-1</sup> as, solubor, agricol and borosol increased pod yield by 18, 23 and 12 per cent respectively, compared to the spray of 5 per cent as boric acid and 9 per cent as borax.

## 2.6 EFFECT OF NUTRIENTS ON PLANT NUTRIENT CONTENT AND UPTAKE

### 2.6.1 Effect of nitrogen on nutrient content and uptake

Nitrogen uptake is more intensive in flowering as well as pod formation stages. During reproductive stage, there is continuous mobilization of nitrogen from leaves to developing fruit, and this resulted in appearance of N deficiency symptoms on leaves (Kvien *et al.*, 1988). Chahal *et al.* (1983) reported that nitrogen content in shoot was high at early and mid-flowering stages. Reddy *et al.* (1984) studied uptake of nitrogen in groundnut and it was maximum by the application of 10 kg N as basal and 20 kg N at 30 DAP. Boote *et al.* (1985) observed that during seed filling stage, N content in leaf decreased from 4.01 to 2.85 per cent, in stem from 1.65 to 1.13 per cent and in root N from 2.19 to 1.50 per cent. Reddy and Murthy (1985) observed that N content was highest in kernel

and lowest in the shell. N content decreased along with crop growth.

### **2.6.2 Effect of phosphorus on nutrient content and uptake**

Basha and Rao (1980) reported that deficiency of P decreased N,P,K and Ca contents in 30 days old groundnut plants. According to Shelke and Khuspe (1980) P uptake and dry matter production by groundnut cv. Latur No. 33 was highest with the application of P at 17.5 kg ha<sup>-1</sup> than with 0 or 35 kg ha<sup>-1</sup>. Nakagawa *et al.* (1981) revealed that highest rate of P application significantly increased P content in seeds. Patel *et al.* (1994) recorded the effect of application of 100 ppm P<sub>2</sub>O<sub>5</sub> on nutrient uptake and growth of groundnut in calcareous soil. High phosphorus level increased growth of shoot and uptake of P by root and shoot. Higher level of P was also effective in increasing the uptake of nitrogen by the plant. Chahal *et al.* (1983) observed that application of P increased the uptake of N and P and dry matter yield.

Application of P<sub>2</sub>O<sub>5</sub> @ 60 kg ha<sup>-1</sup> increased uptake and content of N and P in seed (Islam and Noor, 1982). Chavan and Kalra (1983) reported that P increased the N content and NPK uptake in plants. Bell (1985) reported that the tissue P content during vegetative growth was 0.3 per cent of dry matter and declined during reproductive stage as 0.27 per cent at 60 Days After Emergence to 0.12 per cent at 100 days.

### **2.6.3 Effect of potassium on nutrient content and uptake**

Rao (1979) revealed that uptake of N and P increased with application of K @ 0, 40, 80 kg ha<sup>-1</sup> in groundnut variety, TMV-2 under irrigated condition. According to Reddy *et al.* (1983) uptake of K in groundnut was maximum when it was applied as basal dose at the rate of 40 kg ha<sup>-1</sup>. Survase *et al.* (1986) concluded that average plant N, P and K contents at flowering stage were 2.7, 0.21, and 2.28 per cent respectively.

#### **2.6.4 Effect of secondary nutrients on nutrient content and uptake**

Giri *et al.* (2014) reported that uptake of nutrients such as N, P, K and S by kernel, shell and haulm of groundnut and also total uptake of nutrients by groundnut were significantly influenced by levels of sulphur. Umadevi *et al.* (1999) reported that S application @ 75 kg ha<sup>-1</sup> recorded highest nitrogen (100.7 kg ha<sup>-1</sup>), phosphorus (10.40 kg ha<sup>-1</sup>), potassium (40.4 kg ha<sup>-1</sup>), sulphur (12.21 kg ha<sup>-1</sup>), calcium (34.6 kg ha<sup>-1</sup>) and magnesium (15.59 kg ha<sup>-1</sup>) uptake by groundnut. Singh *et al.* (2009) viewed that S uptake by groundnut pods increased significantly with increased levels of S up to 60 kg ha<sup>-1</sup> (10.89 kg ha<sup>-1</sup>). Patel *et al.* (2009) observed that successive increase in sulphur application rate up to 40 kg ha<sup>-1</sup> improved NPS uptake by groundnut. The maximum uptake of nutrients was observed at S application of 15 kg ha<sup>-1</sup> and minimum uptake in no S treatment.

Rao and Shaktawat (2002) reported that gypsum application at the rate of 250 kg ha<sup>-1</sup> (half at sowing + half at 35 DAS) in groundnut crop significantly increased nitrogen, phosphorus, potassium, calcium, sulphur and magnesium uptake by 13.2, 11.0, 10.6, 11.1, 10.4 and 8.9 per cent respectively over control.

Study conducted by Veerabhadrapa and Yeledhalli (2005) revealed that foliar application (N, P, K, Ca and S - commercial formulation of urea, SSP and MOP at 1 per cent level each) 60 DAS along with the application of 100 per cent RDF recorded higher levels of nitrogen (252 kg ha<sup>-1</sup>), phosphorus (28.9 kg ha<sup>-1</sup>), potassium (204 kg ha<sup>-1</sup>), calcium (74.8 kg ha<sup>-1</sup>) and sulphur (31.4 kg ha<sup>-1</sup>) uptake by groundnut.

#### **2.6.5 Effect of micronutrients on nutrient content and uptake**

Mahajan *et al.* (1994) concluded that soil application of B at the rate of 0.5 kg ha<sup>-1</sup> through boronated superphosphate recorded higher nitrogen (127.4 kg ha<sup>-1</sup>) and phosphorus (11.7 kg ha<sup>-1</sup>) uptake in clayey soils. Study conducted by Kamalakannan and Ravichandran (2013) indicated that application of 100 per cent NPK, borox at 10 kg ha<sup>-1</sup>, zinc sulphate at 25 kg ha<sup>-1</sup> and FYM at 12.5 t ha<sup>-1</sup>

showed highest NPK uptake in all the growth stages of groundnut crop. Nadaf and Chidanandappa (2015) reported that borax at 5 kg ha<sup>-1</sup> and zinc sulphate at 20 kg ha<sup>-1</sup> recorded highest uptake of nitrogen (95.72 kg ha<sup>-1</sup>), phosphorus (23.50 kg ha<sup>-1</sup>), potassium (92.68 kg ha<sup>-1</sup>), calcium (38.34 kg ha<sup>-1</sup>), magnesium (20.87 kg ha<sup>-1</sup>) and sulphur (28.16 kg ha<sup>-1</sup>).

## 2.7 EFFECT OF PRIMARY NUTRIENTS ON QUALITY PARAMETERS

### 2.7.1 Oil content

#### 2.7.1.1 *Effect of nitrogen*

Most of the experiments on the influence of nitrogen on oil content of oil seed crops emphasized that there was a reduction as a result of application of nitrogen in the oil content.

Maini and Bhandar (1965) investigated and concluded that oil content of seeds was adversely affected by the application of nitrogen in oil seed crops. Punnoose (1968) noticed significant reduction in oil content due to application of nitrogen fertilizers on groundnut in a trial conducted at College of Agriculture, Vellayani.

According to Salini and Tripathi (1973) nitrogen application @ 15 kg ha<sup>-1</sup> produced better oil content and it was decreased with the increase in the nitrogen dose.

Singh and Ahuja (1985) opined that growth, nutrient uptake, pod yield and oil content increased significantly with seed inoculation of nitrogen @ 25 kg ha<sup>-1</sup>.

#### 2.7.1.2 *Effect of phosphorus*

Cheema *et al.*, (2001) reported that application of phosphorus resulted in higher protein content in oilseed crops. The oil concentration increased from 43.2 to 47.3 g per 100 g of seeds due to the application of phosphorus (Lickfett, 1999), whereas Brennan and Bolland (2007) opined that application of phosphorus had no effect on the concentration of oil in oilseeds.

### **2.7.1.3 Effect of potassium**

Farhad *et al.* (2010) reported that combined application of potassium at the rate of 40 kg ha<sup>-1</sup> and sulphur at the rate of 20 kg ha<sup>-1</sup> resulted in the production of high amount of oil.

### **2.7.2 Protein content**

#### **2.7.2.1 Effect of nitrogen**

Nijhawan (1962) reported beneficial effect of nitrogen application in increasing the protein content.

#### **2.7.2.2 Effect of phosphorus**

Punnoose (1968) observed that graded doses of phosphorus resulted in the enhancement of protein content in groundnut kernels. Study conducted by Basha and Rao (1980) indicated that P deficiency led reduction in protein content of groundnut. Nair and Sadanandan (1981) reported that protein content showed increase with increased phosphatic fertilizer application at the rate of 50-100 kg ha<sup>-1</sup>.

#### **2.7.2.3 Effect of potassium**

According to Nair and Sadanandan (1981) the protein content decreased with increased dose of potassium from 25 to 75 kg ha<sup>-1</sup>.

## **2.8 EFFECT OF COMBINED APPLICATION OF PRIMARY NUTRIENTS ON QUALITY PARAMETERS**

Bandopadhyay *et al.* (2003) reported increased oil production with increased N and P fertilizer application.



## 2.9. EFFECT OF SECONDARY NUTRIENTS ON QUALITY PARAMETERS

### 2.9.1 Oil content

Tripathi and Hazra (2003) found that there was significant improvement in the oil per cent of groundnut pods along with the application of sulphur containing fertilizers. Bandopadhyay *et al.* (2003) also reported increased oil production with increased S fertilizer application. Gypsum or SSP application followed by elemental S increased the oil content in groundnut seeds (Dutta and Patra, 2005).

### 2.9.2 Protein content

According to Tripathi and Hazra (2003) application of S fertilizers improved the protein content in groundnut pods. Sulphur application showed significant increase in kernel protein content (Kalaiyarasan *et al.*, 2003).

## 2.10 EFFECT OF MICRONUTRIENTS ON QUALITY PARAMETERS OF GROUNDNUT

### 2.10.1 Effect of micronutrients on oil and protein content

Foliar application of  $\text{FeSO}_4$  increased oil content in groundnut kernels (Akhtar *et al.*, 2018).

## 2.11 INTERACTION OF NUTRIENTS

### 2.11.1 Interaction of P and S

Teotia *et al.*, (2000) observed that increase in the levels of P and S significantly increased the grain and straw yield in rice. According to Aulakh *et al.* (1990) there was increase in yield of vegetative tissues and grains as a result of application of S and P individually but decreased when they were applied as different combinations. Sulphur application led to increase in sulphur content whereas decrease in phosphorus content in grains as well as in straw. They also

observed that increased total P content with the application of P and decreased with the application of sulphur. By the application of phosphorus, protein content decreased and it increased by sulphur fertilization in grains of moong.

Das (2017) conducted an experiment in green gram with four levels of sulphur (0, 20, 40 and 60 kg ha<sup>-1</sup>) and two levels of phosphorus (30 and 60 kg ha<sup>-1</sup>). He concluded that interaction of P and S at higher doses had a negative impact on the yield of crop.

### **2.11.2 Interaction of P with other nutrients**

Phosphorus deficiency is a major yield limiting factor for crop production in acid as well as alkaline soils (Fageria, 1983). Assessment of interaction of phosphorus with other nutrients is critical to keep up a balanced nutrient supply for enhancing crop growth and yield. Phosphorus showed a positive interaction with nitrogen and also in plant development (Sumner and Farina, 1986). Sumner and Farina (1986) also concluded that increased growth required more quantity of both N and P and the mutually synergistic effects resulted in growth stimulation and enhanced uptake of nitrogen and phosphorus.

If large quantity of P is supplied, P: Fe and P: Zn ratio in plant tissues increased and led to deficiency of these nutrients (Loneragan *et al.*, 1979; Loneragan *et al.*, 1982).

High available P resulted in the deficiencies of Zn and Mn in potato (Adriano *et al.*, 1971) and maize (Adriano and Murphy, 1970). There was formation of chemical bond by P with Zn, at high levels of P and P bounds large quantity of Zn resulted in P induced Zn deficiency, which led to reduction in shoot growth. Friesen *et al.* (1980) observed increase in total uptake of Zn with P addition which led to increased root growth, however extreme levels of P caused Zn deficiency. According to Saeed and Fox (1979) there was an increase in Zn sorption in Hawaiian soil due to the application of P. Sorption of P on the surface of Fe and Al oxides led to increase in negative charges on them and resulted in

increased sorption of Zn. Gupta and Raj (1983) viewed positive interaction between Zn and K on yield of wheat.

Heavy application of P also led to the deficiency of Fe. Interaction of P with Fe produced Fe-phosphates which resulted in Fe chlorosis in plants (Ayed, 1970). Inhibition of Fe absorption by roots occurred due to higher level of P. It also inhibited Fe transport from roots to shoots and inactivates Fe content in plants (Elliott and Laeuchli, 1985; Moraghan and Mascagni, 1991).

Interaction of P with Ca was complex; it showed both synergistic and antagonistic effects. The synergistic effect was due to simultaneous uptake of Ca and P. Antagonistic effect was due to precipitation of P into less soluble calcium phosphate (Jakobsen, 1979). Fageria (1983) reported decreased uptake of P and Ca with increased concentration of potassium in rice. Lundergardh (1934) observed a higher absorption of P and Ca at lower concentration of potassium. Acidifying effect of S application played an important role in mobilization of P, Fe, Zn and Mn in calcareous soils (Soliman *et al.*, 1992).

### **2.11.3 Interaction of S with other nutrients**

Abdin *et al.* (2003) reported that sulphur is known to interact with almost all essential macronutrients, secondary and micronutrients. These interactions can either enhance or reduce growth and yield of crops by influencing the nutrient uptake and utilization. Soliman *et al.*, (1992) reported that S application increased Mn content in corn plants grown in calcareous soils.

# **Materials and Methods**

### 3. MATERIALS AND METHODS

The present study entitled “Interaction of phosphorus and sulphur in black cotton soils of Palakkad (AEU: 23) under groundnut (*Arachis hypogaea* L.) cultivation” was carried out in College of Horticulture, Vellanikkara during 2017-19. The study consists of a field experiment with groundnut in black cotton soils of Chittur Taluk in Palakkad district followed by laboratory analysis of soil and plant samples taken from the experimental field. The materials used and methods followed in the study are described in this chapter.

#### 3.1 COLLECTION OF SOIL SAMPLES AND ANALYSIS

Soil samples were collected from different locations of Chittur Taluk and analyzed the status of available phosphorus and sulphur. Four to five samples were collected from each location at a depth of 0-15 cm. Collected samples were air dried, processed and sieved through 2mm sieve. These samples were analyzed for available P and S. The details of available P and S status of soil samples collected from different locations of Chittur are given below.

**Table 1: Available status of P and S in soil samples collected from different locations of Chittur**

Sl. no.	Place	Available P (kg ha <sup>-1</sup> )	Available S (mg kg <sup>-1</sup> )
1	Kambalathara	11.65	4.90
2	Kannimari	13.51	4.68
3	Kulappurakkad	7.33	6.03
4	Erimedu	7.04	6.32
5	Meenakshipuram	9.09	5.94
6	Plachimada	16.30	6.33
7	Mullamthodu	12.30	4.29
8	Velloor	8.94	5.14
9	Nellimedu-1	8.13	4.50
10	Nellimedu-2	9.01	3.19





**Plate 1: Initial soil sample collection from the experimental field**



**Plate 2: View of the experimental field after ploughing**

The experiment was conducted in a field deficient in both P and S. The methods of analysis followed for estimating the physico – chemical properties of soil are given below.

**Table 2: Methods followed for the analysis of soil samples**

Parameter	Method	Reference
Bulk density	Keen – Raczkowski cup	Piper ( 1966)
Texture	International pipette method	
pH and electrical conductivity	1: 2.5 soil water suspension- pH meter and conductivity meter	Jackson (1973)
Organic carbon	Walkley and Black method	Walkley and Black (1934)
Available nitrogen	Alkaline permanganate method	Subbiah and Asija ( 1956)
Available phosphorus	Olsen extraction (0.5 M NaHCO <sub>3</sub> at pH 8.5) and estimation by spectrophotometer	Watanabe and Olsen (1965)
Available potassium	Neutral normal ammonium acetate extraction and estimation by flame photometer	Jackson ( 1973)
Available calcium and magnesium	Neutral normal ammonium acetate extraction and estimation by AAS	
Available sulphur	CaCl <sub>2</sub> extraction and estimation by Spectrophotometer	
Available boron	Hot water extraction and estimation by spectrophotometer	
Available micronutrients (Fe, Mn, Zn, Cu)	DTPA extraction and estimation by ICP-OES	
		Lindsay and Norvell ( 1978)



**Table 3: Physico – chemical properties of soil in the field before experiment**

Parameter	Value
Bulk density ( $\text{Mg m}^{-3}$ )	1.35
Texture	Sandy clay loam
Coarse sand (%)	31.80
Fine sand (%)	27.30
Silt (%)	18.65
Clay (%)	22.25
pH	7.89
Electrical conductivity ( $\text{dS m}^{-1}$ )	0.245
Organic carbon (%)	1.24
Available nitrogen ( $\text{kg ha}^{-1}$ )	286.50
Available phosphorus ( $\text{kg ha}^{-1}$ )	9.10
Available potassium ( $\text{kg ha}^{-1}$ )	234.90
Available calcium ( $\text{mg kg}^{-1}$ )	1654.97
Available magnesium ( $\text{mg kg}^{-1}$ )	497
Available sulphur ( $\text{mg kg}^{-1}$ )	3.19
Available B ( $\text{mg kg}^{-1}$ )	2.99
Available Fe ( $\text{mg kg}^{-1}$ )	18.34
Available Mn ( $\text{mg kg}^{-1}$ )	7.78
Available Zn ( $\text{mg kg}^{-1}$ )	1.09
Available Cu ( $\text{mg kg}^{-1}$ )	3.10

## 3.2 GENERAL DETAILS OF FIELD EXPERIMENT

### 3.2.1 Experimental site

The experiment was conducted in black soils at Chittur, Palakkad. Geographically it is situated at eastern side of Palakkad district at  $10^{\circ} 38' 3.88''$  N latitude and  $76^{\circ} 44' 53.90''$  E longitude and at an elevation of 129 m from mean sea level.



### **3.2.2 Climate and weather**

The climate was humid tropical during the experiment. Temperature range was 30-33°C.

### **3.2.3 Cropping season**

Experiment was conducted during May - August 2018.

### **3.2.4 Cropping history of field**

Maize and cowpea were the main crops cultivated in the field till 2016 and then field was left fallow for one year.

### **3.2.5 Crop variety**

Groundnut variety used for the study is K-6 (Kadiri-6). This variety was released from Agricultural Research station, Kadiri, Andhra Pradesh, which is semi spreading in nature. It is a short duration variety suitable for both kharif and rabi seasons. The variety is resistant to leaf spot disease.

## **3.3 EXPERIMENTAL DETAILS**

Soil samples were collected from different locations of Chittur and experiment was conducted where deficiencies of both P and B are noticed. The experimental details are given below.

Crop : Groundnut

Variety : K-6

Season : May - August

Design : RBD (factorial)

Treatments :  $4^2+1$

Replications : 3

Spacing : 15cm x 15cm.

Plot size : 4.05 x 2.5 m<sup>2</sup>

POP recommendations of groundnut- 10:75:75 N:P:K kg ha<sup>-1</sup>

Seeds were collected from Agricultural Research Station, Kadiri, Andhra Pradesh.

### **3.3.1 Treatment details**

Different doses of P and S and their combinations were used as treatments. Treatment combinations are made with four levels of P and four levels of S. Soil test based recommendations is taken as control.

#### **Factor A**

##### **Levels of phosphorus (P) – 4**

P<sub>0</sub> -Control

P<sub>1</sub> -60 kg P<sub>2</sub>O<sub>5</sub> ha<sup>-1</sup>

P<sub>2</sub> -75 kg P<sub>2</sub>O<sub>5</sub> ha<sup>-1</sup>

P<sub>3</sub> -90 kg P<sub>2</sub>O<sub>5</sub> ha<sup>-1</sup>

#### **Factor B**

##### **Levels of sulphur (S) – 4**

S<sub>0</sub> - Control

S<sub>1</sub> – 10 kg ha<sup>-1</sup>

S<sub>2</sub> – 20 kg ha<sup>-1</sup>

S<sub>3</sub> – 30 kg ha<sup>-1</sup>



**Plate 3: Groundnut crop at flowering stage**

## Treatment combinations

T<sub>1</sub> : Soil test based recommendations

Treatment	Notation	Treatment	Notation
T <sub>2</sub>	P <sub>0</sub> S <sub>0</sub>	T <sub>10</sub>	P <sub>2</sub> S <sub>0</sub>
T <sub>3</sub>	P <sub>0</sub> S <sub>1</sub>	T <sub>11</sub>	P <sub>2</sub> S <sub>1</sub>
T <sub>4</sub>	P <sub>0</sub> S <sub>2</sub>	T <sub>12</sub>	P <sub>2</sub> S <sub>2</sub>
T <sub>5</sub>	P <sub>0</sub> S <sub>3</sub>	T <sub>13</sub>	P <sub>2</sub> S <sub>3</sub>
T <sub>6</sub>	P <sub>1</sub> S <sub>0</sub>	T <sub>14</sub>	P <sub>3</sub> S <sub>0</sub>
T <sub>7</sub>	P <sub>1</sub> S <sub>1</sub>	T <sub>15</sub>	P <sub>3</sub> S <sub>1</sub>
T <sub>8</sub>	P <sub>1</sub> S <sub>2</sub>	T <sub>16</sub>	P <sub>3</sub> S <sub>2</sub>
T <sub>9</sub>	P <sub>1</sub> S <sub>3</sub>	T <sub>17</sub>	P <sub>3</sub> S <sub>3</sub>

Nitrogen and potassium levels are kept same based on POP recommendations of Kerala Agricultural University for all the treatments except for the first treatment where soil test based recommendations were followed.

### 3.4.1 Land preparation

Land was made into fine tilth by ploughing thoroughly using tractor. Gross area of the selected field was 18 cents in which net area used for the experiment was 13 cents. The experimental field was divided into three blocks and it was further divided into 17 treatment plots. Drainage furrow of 1m width was taken in between the blocks.

### 3.4.2 Application of manures and fertilizers

All the fertilizers were applied as basal dose at the time of sowing. Equal quantity of farmyard manure was applied in all the plots. Based on POP recommendations of KAU, nitrogen was applied in equal quantities in all experimental plots except in T<sub>1</sub> where all the fertilizer nutrients are given according

4 m

T5
T7
T6
T12
T15
T16
T14
T17
T13
T4
T11
T3
T2
T9
T1
T10
T8

0.5-1m

4 m

T14
T5
T12
T16
T6
T9
T11
T1
T15
T17
T2
T10
T7
T13
T3
T8
T4

4 m

T7
T8
T1
T9
T4
T10
T5
T17
T16
T3
T15
T14
T13
T6
T11
T12
T2





**Plate 4: Field view at pegging stage of groundnut crop**



**Plate 5: Pod formation stage of groundnut crop**

to soil test based recommendations. Potassium dihydrogen phosphate ( $\text{KH}_2\text{PO}_4$ ) and elemental sulphur (ES) were used as the source of P and S respectively. Different levels of  $\text{KH}_2\text{PO}_4$  and elemental sulphur and their combinations were supplied based on the treatment requirements. MOP was used to supplement potassium requirement of the crop.

**Table 4: Rate of application of fertilizers in the experimental field**

Treatments	Urea ( $\text{kg ha}^{-1}$ )	MKP* ( $\text{kg ha}^{-1}$ )	MOP ( $\text{kg ha}^{-1}$ )	Elemental sulphur ( $\text{kg ha}^{-1}$ )
T <sub>1</sub>	18.26	152.88	46.04	0
T <sub>2</sub>	21.74	0	150	0
T <sub>3</sub>	21.74	0	150	11.11
T <sub>4</sub>	21.74	0	150	22.22
T <sub>5</sub>	21.74	0	150	33.33
T <sub>6</sub>	21.74	115.38	71.54	0
T <sub>7</sub>	21.74	115.38	71.54	11.11
T <sub>8</sub>	21.74	115.38	71.54	22.22
T <sub>9</sub>	21.74	115.38	71.54	33.33
T <sub>10</sub>	21.74	144.23	51.92	0
T <sub>11</sub>	21.74	144.23	51.92	11.11
T <sub>12</sub>	21.74	144.23	51.92	22.22
T <sub>13</sub>	21.74	144.23	51.92	33.33
T <sub>14</sub>	21.74	173.08	32.30	0
T <sub>15</sub>	21.74	173.08	32.30	11.11
T <sub>16</sub>	21.74	173.08	32.30	22.22
T <sub>17</sub>	21.74	173.08	32.30	33.33

\*MKP- Monopotassium phosphate

### **3.4.3 Irrigation**

Irrigation was given through furrows made in between the blocks. Furrows were irrigated at an interval of 7 days.

### **3.4.4 Weed management**

Hand weeding was done at an interval of 15 days. At the time of flowering, along with weeding, earthing up was also done to improve anchorage of tiny roots. After 45 days of sowing field was kept undisturbed.

### **3.4.5 Plant protection**

Pest and disease incidence was very less in the field. Peacock menace was controlled by tying bird repellent ribbons across the field.

### **3.4.6 Harvesting**

Harvesting was started when groundnut leaves started yellowing and began to dry up. Plants were ready for harvest at 90 days after sowing. The plants were uprooted and pods were separated manually.

## **3.5 OBSERVATIONS**

### **3.5.1 Biometric observations**

Five plants were selected randomly from each experimental plot and tagged for taking biometric observations. The biometric observations recorded from these plants during the period of experiment were at four different growth stages namely flowering, pegging, pod formation and harvesting. The mean values were calculated for all the observations made. The biometric observations recorded are given below.

#### **Plant height**

Height of plant was measured from labelled plants at flowering, pegging, pod formation and harvesting stages and mean values were computed.





**Plate 6: Harvesting of groundnut crop**



### **Number of leaves**

Counted the number of leaves at flowering, pegging, pod formation and harvesting stages and mean values were calculated.

### **Number of pods/plant**

No. of pods/plant were counted at harvesting stage and computed the mean values.

### **Yield**

The pods and haulm were harvested, weighed separately and mean values were recorded.

## **3.5.2 Soil analysis**

Soil samples were collected from a depth of 0-15 cm for analysis. The collected samples were analyzed and estimated pH, EC, OC, major nutrients (N, P, K), secondary nutrients (Ca, Mg, S) and micronutrients (Fe, Mn, Zn, Cu, B) both before and after the crop. Physical properties of soil *viz.* soil texture, bulk density and soil moisture were also analyzed. The methods employed for soil analysis are given in table no. 2.

## **3.5.3 Plant analysis**

### **Collection of samples**

Plant samples were collected at flowering, pegging, pod formation and harvesting stages. In order to remove dirt and soil, plant samples were first washed with tap water. These were then washed with single and double distilled water, and kept for shade drying for a period of one week. The shade dried samples were kept in an oven @ 60 °C and dry weight was recorded. The samples were powdered, labelled and stored in polythene bags. The content of major nutrients (N, P, and K), secondary nutrients (Ca, Mg, and S) and micronutrients (Fe, Mn, Zn, Cu, and B) were analyzed. The methods followed to determine the nutrients in samples are given in the table below.

**Table 5: Methods of plant analysis**

Parameter	Method	Reference
Nitrogen	Micro kjeldahl distillation	Jackson (1973)
Phosphorus	Vanado – molybdo – phosphoric (Bartons reagent) yellow color method	Jackson (1973)
Potassium	Nitric acid digestion and estimation by flame photometer	
Calcium and magnesium	Nitric acid digestion and estimation by ICP- OES	Piper (1966)
Sulphur	Nitric acid digestion and estimation by turbidimetry	Black (1965)
Boron	Nitric acid digestion and estimation by ICP- OES	Page <i>et al.</i> , (1982)
Micro nutrients (Fe, Mn, Zn, Cu)	Nitric acid digestion and estimation by ICP- OES	Piper (1966)

### 3.5.4 Uptake of nutrients

Uptake of major, secondary, and micro nutrients were calculated by using the formula,

$$\text{Uptake of nutrient (kg ha}^{-1}\text{)} = \frac{\text{Nutrient concentration (\%)} \times \text{biomass (kg ha}^{-1}\text{)}}{100}$$

100





**Plate 7: Harvested groundnut pods**

## **3.6 Quality parameters**

### **3.6.1 Protein content of kernels**

Per cent of nitrogen was estimated by micro-kjeldahl method. Crude protein content was calculated by multiplying nitrogen content of kernels with the constant 6.25.

### **3.6.2 Oil content**

Oil extraction was done in soxhlet apparatus using petroleum benzene as solvent. The weight difference of round bottom flask gave the amount of oil extracted (Pearson, 1981).

## **3.7 Statistical analysis**

Statistical analysis of experimental data was done by Fisher's method of analysis of variance as outlined by Gomez and Gomez (1984).

# Results



## 4. RESULTS

The results of the experiment entitled “Interaction of phosphorus and sulphur in black cotton soils of Palakkad (AEU: 23) under groundnut (*Arachis hypogaea* L.) cultivation” are presented in this chapter.

### 4.1 GROWTH AND YIELD ATTRIBUTES OF GROUNDNUT

The effect of different levels of P and S application on growth parameters of groundnut such as plant height, number of leaves and yield parameters such as number of pods at flowering, pegging, pod formation and harvesting stages are given.

#### 4.1.1 Growth parameters

##### 4.1.1.1 Plant height

Plant heights at flowering, pegging, pod development and harvest stages are shown in table 6, 7, 8, 9 respectively. The treatment, P<sub>3</sub>S<sub>3</sub> (P,90 kg ha<sup>-1</sup> and S,30 kg ha<sup>-1</sup>) showed highest plant height at flowering (22.13 cm) stage. . At pegging stage (35.00 cm) and harvesting stage (41.47 cm) the treatment T<sub>16</sub>, P<sub>3</sub>S<sub>2</sub> (P, 90 kg ha<sup>-1</sup> and S, 20 kg ha<sup>-1</sup>) showed highest plant height. Whereas, during pod formation stage, both the treatments T<sub>16</sub> and T<sub>17</sub> recorded highest (36.67 cm) plant height. Plant heights at different stages were found to be significantly influenced due to main effect and interaction effect. Plant height was enhanced by increased doses of P as well as S. F statistic for treatments Vs control was calculated and was found to be significantly different at flowering, pegging and harvesting stages and was on par at pod formation stage.

**Table 6: Effect of application of P and S on plant height at flowering stage (cm)****T<sub>1</sub>: 20.00**

	<b>S<sub>0</sub></b>	<b>S<sub>1</sub></b>	<b>S<sub>2</sub></b>	<b>S<sub>3</sub></b>	<b>Mean</b>
<b>P<sub>0</sub></b>	18.03	18.07	18.17	18.13	<b>18.10</b>
<b>P<sub>1</sub></b>	18.63	18.43	18.47	18.63	<b>18.54</b>
<b>P<sub>2</sub></b>	18.83	18.93	19.83	19.27	<b>19.22</b>
<b>P<sub>3</sub></b>	19.73	21.43	21.63	22.13	<b>21.23</b>
<b>Mean</b>	<b>18.81</b>	<b>19.22</b>	<b>19.53</b>	<b>19.54</b>	
<b>CD (0.05) P; 0.074</b>		<b>CD (0.05) S; 0.074</b>		<b>CD (0.05) PxS; 0.147</b>	
<b>SE (m) P; 0.025</b>		<b>SE (m) S; 0.025</b>		<b>SE (m) PxS; 0.051</b>	

**Table 7: Effect of application of P and S on plant height at pegging stage (cm)****T<sub>1</sub>: 32.00**

	<b>S<sub>0</sub></b>	<b>S<sub>1</sub></b>	<b>S<sub>2</sub></b>	<b>S<sub>3</sub></b>	<b>Mean</b>
<b>P<sub>0</sub></b>	26.33	26.30	26.53	26.27	<b>26.36</b>
<b>P<sub>1</sub></b>	28.60	28.43	28.30	28.27	<b>28.40</b>
<b>P<sub>2</sub></b>	32.27	32.90	32.63	31.97	<b>32.44</b>
<b>P<sub>3</sub></b>	34.47	34.50	35.00	34.27	<b>34.56</b>
<b>Mean</b>	<b>30.42</b>	<b>30.53</b>	<b>30.62</b>	<b>30.19</b>	
<b>CD (0.05) P; 0.343</b>		<b>CD (0.05) S; NS</b>		<b>CD (0.05) PxS; NS</b>	
<b>SE (m) P; 0.118</b>		<b>SE (m) S; 0.118</b>		<b>SE (m) PxS; 0.237</b>	



**Table 8: Effect of application of P and S on plant height at pod formation stage (cm)****T<sub>1</sub>: 36.00**

	<b>S<sub>0</sub></b>	<b>S<sub>1</sub></b>	<b>S<sub>2</sub></b>	<b>S<sub>3</sub></b>	<b>Mean</b>
<b>P<sub>0</sub></b>	31.30	31.07	31.40	31.80	<b>31.39</b>
<b>P<sub>1</sub></b>	33.60	33.63	33.30	33.50	<b>33.51</b>
<b>P<sub>2</sub></b>	35.60	35.63	35.30	35.60	<b>35.53</b>
<b>P<sub>3</sub></b>	36.27	36.30	36.67	36.67	<b>36.48</b>
<b>Mean</b>	<b>34.19</b>	<b>34.16</b>	<b>34.17</b>	<b>34.39</b>	
<b>CD (0.05) P; 0.311</b>		<b>CD (0.05) S; NS</b>		<b>CD (0.05) PxS; NS</b>	
<b>SE (m) P; 0.107</b>		<b>SE (m) S; 0.107</b>		<b>SE (m) PxS; 0.214</b>	

**Table 9: Effect of application of P and S on plant height at harvest stage (cm)****T<sub>1</sub>: 37.60**

	<b>S<sub>0</sub></b>	<b>S<sub>1</sub></b>	<b>S<sub>2</sub></b>	<b>S<sub>3</sub></b>	<b>Mean</b>
<b>P<sub>0</sub></b>	34.67	34.30	34.67	34.67	<b>34.58</b>
<b>P<sub>1</sub></b>	36.33	36.37	36.33	36.47	<b>36.38</b>
<b>P<sub>2</sub></b>	38.40	38.70	38.07	39.00	<b>38.54</b>
<b>P<sub>3</sub></b>	41.33	41.47	41.17	41.30	<b>41.32</b>
<b>Mean</b>	<b>37.68</b>	<b>37.71</b>	<b>37.56</b>	<b>37.86</b>	
<b>CD (0.05) P; 0.283</b>		<b>CD (0.05) S; NS</b>		<b>CD (0.05) PxS; NS</b>	
<b>SE (m) P; 0.097</b>		<b>SE (m) S; 0.097</b>		<b>SE (m) PxS; 0.195</b>	

**4.1.1.2 Number of leaves**

Number of leaves at flowering, pegging, pod development and harvest stages are shown in the tables 10, 11, 12, 13 respectively. Both the main effect and interaction effect of treatments significantly influenced the number of leaves at pegging, pod formation and harvesting stages, whereas at flowering stage the

interaction effect was found to be non significant. In flowering stage, the number of leaves in treatments P<sub>2</sub>S<sub>3</sub> (P, 75 kg ha<sup>-1</sup> and S, 30 kg ha<sup>-1</sup>) and P<sub>3</sub>S<sub>3</sub> (P, 90 kg ha<sup>-1</sup> and S, 30 kg ha<sup>-1</sup>) were found to be on par. Treatment P<sub>3</sub>S<sub>3</sub> (P, 90 kg ha<sup>-1</sup> and S, 30 kg ha<sup>-1</sup>) showed highest number of leaves at pegging, (53.81), pod development (60.06) and harvesting (55.06) stages. F statistic for treatments Vs control was calculated and was found to be significantly different at flowering, pegging, pod formation and harvesting stages.

**Table 10: Effect of application of P and S on number of leaves at flowering stage**

**T<sub>1</sub>: 41.98**

	S <sub>0</sub>	S <sub>1</sub>	S <sub>2</sub>	S <sub>3</sub>	Mean
P <sub>0</sub>	37.30	42.87	43.67	47.40	<b>42.81</b>
P <sub>1</sub>	37.80	44.30	46.70	47.67	<b>44.12</b>
P <sub>2</sub>	39.20	45.07	45.30	48.80	<b>44.59</b>
P <sub>3</sub>	42.70	45.33	45.30	49.33	<b>45.67</b>
Mean	<b>39.25</b>	<b>44.39</b>	<b>45.24</b>	<b>48.30</b>	
<b>CD (0.05) P; 1.5</b>		<b>CD (0.05) S; 1.5</b>		<b>CD (0.05) PxS; NS</b>	
<b>SE (m) P; 0.517</b>		<b>SE (m) S; 0.517</b>		<b>SE (m) PxS; 1.303</b>	

**Table 11: Effect of application of P and S on number of leaves at pegging stage**

**T<sub>1</sub>: 45.55**

	S <sub>0</sub>	S <sub>1</sub>	S <sub>2</sub>	S <sub>3</sub>	Mean
P <sub>0</sub>	41.93	46.63	47.30	51.33	<b>46.80</b>
P <sub>1</sub>	41.67	48.60	50.93	52.44	<b>48.41</b>
P <sub>2</sub>	43.13	49.19	49.51	52.87	<b>48.67</b>
P <sub>3</sub>	46.37	49.23	49.60	53.81	<b>49.75</b>
Mean	<b>43.27</b>	<b>48.41</b>	<b>49.34</b>	<b>52.61</b>	
<b>CD (0.05) P; 0.573</b>		<b>CD (0.05) S; 0.573</b>		<b>CD (0.05) PxS; 1.146</b>	
<b>SE (m) P; 0.197</b>		<b>SE (m) S; 0.197</b>		<b>SE (m) PxS; 0.395</b>	

**Table 12: Effect of application of P and S on number of leaves at pod formation stage****T<sub>1</sub>: 50.55**

	S <sub>0</sub>	S <sub>1</sub>	S <sub>2</sub>	S <sub>3</sub>	Mean
P <sub>0</sub>	46.94	51.64	52.30	56.34	<b>51.80</b>
P <sub>1</sub>	46.67	53.60	55.94	57.44	<b>53.41</b>
P <sub>2</sub>	48.13	54.19	54.52	57.87	<b>53.68</b>
P <sub>3</sub>	51.37	54.23	54.61	60.06	<b>55.07</b>
<b>Mean</b>	<b>48.28</b>	<b>53.42</b>	<b>54.34</b>	<b>57.93</b>	
<b>CD (0.05) P; 0.489</b>		<b>CD (0.05) S; 0.489</b>		<b>CD (0.05) PxS; 0.978</b>	
<b>SE (m) P; 0.168</b>		<b>SE (m) S; 0.168</b>		<b>SE (m) PxS; 0.337</b>	

**Table 13: Effect of application of P and S on number of leaves at harvest stage****T<sub>1</sub>: 45.56**

	S <sub>0</sub>	S <sub>1</sub>	S <sub>2</sub>	S <sub>3</sub>	Mean
P <sub>0</sub>	41.94	46.64	47.30	51.34	<b>46.80</b>
P <sub>1</sub>	41.67	48.60	50.94	52.44	<b>48.41</b>
P <sub>2</sub>	43.13	49.19	49.52	52.87	<b>48.68</b>
P <sub>3</sub>	46.37	49.23	49.61	55.06	<b>50.07</b>
<b>Mean</b>	<b>43.28</b>	<b>48.42</b>	<b>49.34</b>	<b>52.93</b>	
<b>CD (0.05) P; 0.489</b>		<b>CD (0.05) S; 0.489</b>		<b>CD (0.05) PxS; 0.977</b>	
<b>SE (m) P; 0.168</b>		<b>SE (m) S; 0.168</b>		<b>SE (m) PxS; 0.337</b>	

#### 4.1.2 Yield parameters

##### 4.1.2.1 Number of pods per plant

Number of pods per plant is significantly influenced by different levels of P and S. Increase in pod number per plant can be observed with increased dose of P and S application (table 14). The treatment, P<sub>3</sub>S<sub>3</sub> (P, 90 kg ha<sup>-1</sup> and S, 30 kg ha<sup>-1</sup>)

produced the highest number of pods per plant. Number of pods increased with increased dose of P and S application. F statistic for treatments Vs control was calculated and was found to be significantly different.

#### **4.1.2.2 Yield**

The data given in table 15 indicates that yield was significantly influenced by application of different doses of P and S fertilizers. The highest yield (3.68 t ha<sup>-1</sup>) was recorded in P<sub>3</sub> S<sub>3</sub> (P, 90 kg ha<sup>-1</sup> and S, 30 kg ha<sup>-1</sup>). The lowest yield (3.07 t ha<sup>-1</sup>) was in P<sub>0</sub> S<sub>0</sub> (P, 0 kg ha<sup>-1</sup> and S, 0 kg ha<sup>-1</sup>). Yield was increased with increased dose of P and S fertilizer. F statistic for treatments Vs control was calculated and was found to be significantly different.

## **4.2 PHYSICO – CHEMICAL CHARACTERISTICS OF SOIL**

### **4.2.1 Soil pH**

The effect of application of different levels of P and S on soil pH is given in the table 16. Data showed that none of main effects and interaction effect had significant influence on the pH. There was only a slight change in pH after the harvest. pH varied from 7.73 to 7.85. F statistic for treatments Vs control was calculated and was found to be on par.

### **4.2.2 Electrical conductivity**

The effect of application of different levels of P and S on soil EC is given in the table 17. Data showed that both main effects and interaction effect has no significant influence on EC. F statistic for treatments Vs control was calculated and was found to be on par.

### **4.2.3 Organic carbon**

The data on effect of application of different levels of P and S on soil OC is given in the table 18. Data showed that both main effects and interaction effect has no significant influence on OC content. F statistic for treatments Vs control was calculated and was found to be on par.

**Table 14: Effect of application of P and S on number of pods per plant**

**T<sub>1</sub>: 15.33**

	<b>S<sub>0</sub></b>	<b>S<sub>1</sub></b>	<b>S<sub>2</sub></b>	<b>S<sub>3</sub></b>	<b>Mean</b>
<b>P<sub>0</sub></b>	9.33	10.20	10.28	10.37	<b>10.05</b>
<b>P<sub>1</sub></b>	12.00	12.67	12.70	13.10	<b>12.62</b>
<b>P<sub>2</sub></b>	14.33	14.83	14.87	15.13	<b>14.79</b>
<b>P<sub>3</sub></b>	15.41	16.67	17.00	18.17	<b>16.81</b>
<b>Mean</b>	<b>12.77</b>	<b>13.59</b>	<b>13.71</b>	<b>14.19</b>	
<b>CD (0.05) P; 0.745</b>		<b>CD (0.05) S; 0.745</b>		<b>CD (0.05) PxS; NS</b>	
<b>SE (m) P; 0.257</b>		<b>SE (m) S; 0.257</b>		<b>SE (m) PxS; 0.513</b>	

**Table 15: Effect of application of P and S on groundnut yield (t ha<sup>-1</sup>)**

**T<sub>1</sub>: 3.37**

	<b>S<sub>0</sub></b>	<b>S<sub>1</sub></b>	<b>S<sub>2</sub></b>	<b>S<sub>3</sub></b>	<b>Mean</b>
<b>P<sub>0</sub></b>	3.07	3.12	3.19	3.23	<b>3.15</b>
<b>P<sub>1</sub></b>	3.16	3.21	3.25	3.29	<b>3.23</b>
<b>P<sub>2</sub></b>	3.24	3.29	3.31	3.42	<b>3.31</b>
<b>P<sub>3</sub></b>	3.35	3.46	3.59	3.68	<b>3.52</b>
<b>Mean</b>	<b>3.21</b>	<b>3.27</b>	<b>3.34</b>	<b>3.40</b>	
<b>CD (0.05) P; 0.054</b>		<b>CD (0.05) S; 0.054</b>		<b>CD (0.05) PxS; 0.108</b>	
<b>SE (m) P; 0.001</b>		<b>SE (m) S; 0.001</b>		<b>SE (m) PxS; 0.002</b>	

**Table 16: Effect of application of P and S on pH of soil**

**T<sub>1</sub>: 7.73**

	<b>S<sub>0</sub></b>	<b>S<sub>1</sub></b>	<b>S<sub>2</sub></b>	<b>S<sub>3</sub></b>	<b>Mean</b>
<b>P<sub>0</sub></b>	7.79	7.78	7.77	7.74	<b>7.77</b>
<b>P<sub>1</sub></b>	7.85	7.84	7.83	7.82	<b>7.91</b>
<b>P<sub>2</sub></b>	7.83	7.82	7.82	7.81	<b>7.82</b>
<b>P<sub>3</sub></b>	7.80	7.79	7.79	7.73	<b>7.78</b>
<b>Mean</b>	<b>7.82</b>	<b>7.81</b>	<b>7.80</b>	<b>7.77</b>	
<b>CD (0.05) P; NS</b>		<b>CD (0.05) S; NS</b>		<b>CD (0.05) PxS; NS</b>	
<b>SE (m) P; 0.044</b>		<b>SE (m) S; 0.044</b>		<b>SE (m) PxS; 0.087</b>	

**Table 17: Effect of application of P and S on EC of soil (dS m<sup>-1</sup>)**

**T<sub>1</sub>: 0.25**

	<b>S<sub>0</sub></b>	<b>S<sub>1</sub></b>	<b>S<sub>2</sub></b>	<b>S<sub>3</sub></b>	<b>Mean</b>
<b>P<sub>0</sub></b>	0.28	0.28	0.28	0.29	<b>0.28</b>
<b>P<sub>1</sub></b>	0.29	0.27	0.27	0.25	<b>0.27</b>
<b>P<sub>2</sub></b>	0.28	0.28	0.28	0.28	<b>0.28</b>
<b>P<sub>3</sub></b>	0.28	0.29	0.28	0.28	<b>0.28</b>
<b>Mean</b>	<b>0.28</b>	<b>0.28</b>	<b>0.28</b>	<b>0.27</b>	
<b>CD (0.05) P; NS</b>		<b>CD (0.05) S; NS</b>		<b>CD (0.05) PxS; NS</b>	
<b>SE (m) P; 0.011</b>		<b>SE (m) S; 0.011</b>		<b>SE (m) PxS; 0.022</b>	

**Table 18: Effect of application of P and S on organic carbon in soil (%)**

**T<sub>1</sub>: 1.03**

	<b>S<sub>0</sub></b>	<b>S<sub>1</sub></b>	<b>S<sub>2</sub></b>	<b>S<sub>3</sub></b>	<b>Mean</b>
<b>P<sub>0</sub></b>	1.10	1.06	1.08	1.02	<b>1.07</b>
<b>P<sub>1</sub></b>	1.07	1.08	0.99	1.01	<b>1.04</b>
<b>P<sub>2</sub></b>	1.02	1.08	1.02	1.01	<b>1.03</b>
<b>P<sub>3</sub></b>	1.06	1.02	1.11	1.02	<b>1.05</b>
<b>Mean</b>	<b>1.06</b>	<b>1.06</b>	<b>1.05</b>	<b>1.01</b>	
<b>CD (0.05) P; NS</b>	<b>CD (0.05) S; NS</b>		<b>CD (0.05) PxS; NS</b>		
<b>SE (m) P; 0.015</b>	<b>SE (m) S; 0.015</b>		<b>SE (m) PxS; 0.030</b>		

#### 4.3 NUTRIENT STATUS OF SOIL

##### 4.3.1 Available nitrogen

The effects of application of different levels of P and S on available N are given in the table 19. Data showed that main effect, and interaction effect were significantly influenced the available nitrogen in the soil. N increased with increased dose of P and S application. The treatment, T<sub>17</sub>, P<sub>3</sub>S<sub>3</sub> showed highest soil available N (293.22 kg ha<sup>-1</sup>). F statistic for treatments Vs control was calculated and was found to be significantly different.

##### 4.3.2 Available phosphorus

The influence of application of different levels of P and S on available P is given in the table 20. Data showed that main effect of P and S and interaction effects were found to be significant. In comparison with initial P status, available P increased in all treatments due to fertilizer application. Treatment, P<sub>3</sub> S<sub>3</sub> (P, 90 kg ha<sup>-1</sup> and S, 30 kg ha<sup>-1</sup>) showed highest available P (29.11 kg ha<sup>-1</sup>). An increase in available P with increased dose of S can be observed among treatments.

**Table 19: Effect of application of P and S on available nitrogen in soil (kg ha<sup>-1</sup>)****T<sub>1</sub>: 284.91**

	<b>S<sub>0</sub></b>	<b>S<sub>1</sub></b>	<b>S<sub>2</sub></b>	<b>S<sub>3</sub></b>	<b>Mean</b>
<b>P<sub>0</sub></b>	284.84	285.54	286.78	287.57	<b>286.18</b>
<b>P<sub>1</sub></b>	286.63	287.18	286.99	287.68	<b>287.12</b>
<b>P<sub>2</sub></b>	286.97	288.43	288.34	288.77	<b>288.13</b>
<b>P<sub>3</sub></b>	287.53	288.81	289.35	293.22	<b>289.73</b>
<b>Mean</b>	<b>286.49</b>	<b>287.49</b>	<b>287.87</b>	<b>289.31</b>	
<b>CD (0.05) P; 0.267</b>		<b>CD (0.05) S; 0.267</b>		<b>CD (0.05) PxS; 0.533</b>	
<b>SE (m) P; 0.092</b>		<b>SE (m) S; 0.092</b>		<b>SE (m) PxS; 0.184</b>	

**Table 20: Effect of application of P and S on available phosphorus in soil (kg ha<sup>1</sup>)****T<sub>1</sub>: 13.33**

	<b>S<sub>0</sub></b>	<b>S<sub>1</sub></b>	<b>S<sub>2</sub></b>	<b>S<sub>3</sub></b>	<b>Mean</b>
<b>P<sub>0</sub></b>	6.73	10.93	11.01	11.76	<b>10.11</b>
<b>P<sub>1</sub></b>	12.21	12.43	13.63	17.88	<b>14.04</b>
<b>P<sub>2</sub></b>	20.13	20.28	20.55	24.27	<b>21.31</b>
<b>P<sub>3</sub></b>	25.03	25.76	28.59	29.11	<b>27.12</b>
<b>Mean</b>	<b>16.03</b>	<b>17.35</b>	<b>18.45</b>	<b>20.75</b>	
<b>CD (0.05) P; 0.093</b>		<b>CD (0.05) S; 0.093</b>		<b>CD (0.05) PxS; 0.185</b>	
<b>SE (m) P; 0.032</b>		<b>SE (m) S; 0.032</b>		<b>SE (m) PxS; 0.064</b>	



### 4.3.3 Available potassium

The influence of application of different levels of P and S on available K is given in the table 21. The main effect of P and S and interaction effects were found to be significant. Treatment,  $P_1S_0$  (P,  $60 \text{ kg ha}^{-1}$  and S,  $0 \text{ kg ha}^{-1}$ ) showed highest K content ( $298.72 \text{ kg ha}^{-1}$ ). F statistic for treatments Vs control was calculated and was found to be significantly different.

### 4.3.4 Available calcium

The influence of application of different levels of P and S on available Ca is given in the table 22. With increased dose of P application available Ca content in soil decreased. Highest available calcium was in treatment,  $P_1S_0$  ( $1700.87 \text{ mg kg}^{-1}$ ). F statistic for treatments Vs control was calculated and was found to be significantly different.

### 4.3.5 Available magnesium

The influence of application of different levels of P and S on available Mg is given in the table 23. Data showed that available Mg content in soil significantly varied among different treatments due to main effects and interaction effects of P and S. Treatment,  $P_3S_3$  had highest available Mg ( $796.40 \text{ mg kg}^{-1}$ ). Mg showed positive interaction with P and S application. F statistic for treatments Vs control was calculated and was found to be significantly different.

### 4.3.6 Available sulphur

The influence of application of different levels of P and S on available S is given in the table 24. There was a noticeable change in available S due to different levels of fertilizer application. Available S content increased in every treatments as compared to initial sample due to S fertilizer application. Highest available S was noticed in treatment,  $P_2S_3$  ( $10.95 \text{ mg kg}^{-1}$ ). Available S was influenced by both main effects and interaction effect of P and S. F statistic for treatments Vs control was calculated and was found to be significantly different.

**Table 21: Effect of application of P and S on available potassium in soil (kg ha<sup>-1</sup>)****T<sub>1</sub>: 232.80**

	<b>S<sub>0</sub></b>	<b>S<sub>1</sub></b>	<b>S<sub>2</sub></b>	<b>S<sub>3</sub></b>	<b>Mean</b>
<b>P<sub>0</sub></b>	284.21	297.53	296.27	279.12	<b>289.28</b>
<b>P<sub>1</sub></b>	298.72	283.47	283.76	283.58	<b>287.38</b>
<b>P<sub>2</sub></b>	247.61	245.46	244.90	242.84	<b>245.20</b>
<b>P<sub>3</sub></b>	210.76	213.80	211.98	212.59	<b>212.28</b>
<b>Mean</b>	<b>260.32</b>	<b>260.07</b>	<b>259.23</b>	<b>254.53</b>	
<b>CD (0.05) P; 4.188</b>		<b>CD (0.05) S; 4.188</b>		<b>CD (0.05) PxS; 8.376</b>	
<b>SE (m) P; 1.443</b>		<b>SE (m) S; 1.443</b>		<b>SE (m) PxS; 2.886</b>	

**Table 22: Effect of application of P and S on available calcium in soil (mg kg<sup>-1</sup>)****T<sub>1</sub>: 1613.23**

	<b>S<sub>0</sub></b>	<b>S<sub>1</sub></b>	<b>S<sub>2</sub></b>	<b>S<sub>3</sub></b>	<b>Mean</b>
<b>P<sub>0</sub></b>	1,666.30	1,618.93	1,664.37	1,694.17	<b>1,660.94</b>
<b>P<sub>1</sub></b>	1,700.87	1,661.23	1,611.20	1,612.20	<b>1,646.38</b>
<b>P<sub>2</sub></b>	1,605.17	1,601.70	1,608.20	1,578.17	<b>1,598.31</b>
<b>P<sub>3</sub></b>	1,567.90	1,589.40	1,577.63	1,598.53	<b>1,583.37</b>
<b>Mean</b>	<b>1,635.06</b>	<b>1,617.82</b>	<b>1,615.35</b>	<b>1,620.77</b>	
<b>CD (0.05) P; 0.77</b>		<b>CD (0.05) S; 0.77</b>		<b>CD (0.05) PxS; 1.54</b>	
<b>SE (m) P; 0.265</b>		<b>SE (m) S; 0.265</b>		<b>SE (m) PxS; 0.531</b>	

**Table 23: Effect of application of P and S on available magnesium in soil (mg kg<sup>-1</sup>)**

**T<sub>1</sub>: 735.43**

	<b>S<sub>0</sub></b>	<b>S<sub>1</sub></b>	<b>S<sub>2</sub></b>	<b>S<sub>3</sub></b>	<b>Mean</b>
<b>P<sub>0</sub></b>	516.50	407.30	465.57	546.50	<b>483.97</b>
<b>P<sub>1</sub></b>	573.53	574.50	609.47	626.43	<b>595.98</b>
<b>P<sub>2</sub></b>	632.07	660.67	639.60	746.33	<b>669.67</b>
<b>P<sub>3</sub></b>	666.33	685.22	707.77	796.40	<b>713.93</b>
<b>Mean</b>	<b>597.11</b>	<b>581.92</b>	<b>605.60</b>	<b>678.92</b>	
<b>CD (0.05) P; 4.029</b>		<b>CD (0.05) S; 4.029</b>		<b>CD (0.05) PxS; 8.059</b>	
<b>SE (m) P; 1.388</b>		<b>SE (m) S; 1.388</b>		<b>SE (m) PxS; 2.777</b>	

**Table 24: Effect of application of P and S on available sulphur in soil (mg kg<sup>-1</sup>)**

**T<sub>1</sub>: 6.06**

	<b>S<sub>0</sub></b>	<b>S<sub>1</sub></b>	<b>S<sub>2</sub></b>	<b>S<sub>3</sub></b>	<b>Mean</b>
<b>P<sub>0</sub></b>	3.51	6.37	9.99	10.35	<b>7.55</b>
<b>P<sub>1</sub></b>	3.53	6.70	9.58	10.55	<b>7.59</b>
<b>P<sub>2</sub></b>	3.63	6.78	9.22	10.95	<b>7.65</b>
<b>P<sub>3</sub></b>	3.40	6.29	9.86	10.56	<b>7.53</b>
<b>Mean</b>	<b>3.52</b>	<b>6.53</b>	<b>9.66</b>	<b>10.60</b>	
<b>CD (0.05) P; NS</b>		<b>CD (0.05) S; 0.285</b>		<b>CD (0.05) PxS; NS</b>	
<b>SE (m) P; 0.098</b>		<b>SE (m) S; 0.098</b>		<b>SE (m) PxS; 0.197</b>	

#### **4.3.7 Available iron**

The influence of application of different levels of P and S on available Fe is given in the table 25. Highest available Fe was in treatment, P<sub>0</sub>S<sub>3</sub> (21.63 mg kg<sup>-1</sup>). Available Fe was low in soil treated with high dose of P. The available Fe content in soil varied significantly due to the main effect of different doses of P and S and interaction effect. Fe content decreased with increased dose of P application and increased with increased dose of S application. F statistic for treatments Vs control was calculated and was found to be significantly different.

#### **4.3.8 Available manganese**

The influence of application of different levels of P and S on available Mn is given in the table 26. Data showed that main effects of P and S and interaction effects affect the available Mn in soil. Treatment, P<sub>0</sub>S<sub>3</sub> showed highest available Mn (7.84 mg kg<sup>-1</sup>). Available Mn decreased with increased dose of P application. Mn content decreased with increased dose of P application and increased with increased dose of S application. F statistic for treatments Vs control was calculated and was found to be significantly different.

#### **4.3.9 Available zinc**

The effect of application of different levels of P and S on available Zn is given in the table 27. Data showed that main effects of P and S and interaction effects affect the available Zn in the soil. Highest Zn was observed in treatment, P<sub>0</sub> S<sub>3</sub> (3.04 mg kg<sup>-1</sup>). Zn content was decreased with increased dose of P application. F statistic for treatments Vs control was calculated and was found to be significantly different.

#### **4.3.10 Available copper**

The influence of application of different levels of P and S on available Cu is given in the table 28. Both main effects and interaction effect were found to have no significant effect on available Cu in soil. F statistic for treatments Vs control was calculated and was found to be on par.

**Table 25: Effect of application of P and S on available iron in soil (mg kg<sup>-1</sup>)**

**T<sub>1</sub>: 17.00**

	<b>S<sub>0</sub></b>	<b>S<sub>1</sub></b>	<b>S<sub>2</sub></b>	<b>S<sub>3</sub></b>	<b>Mean</b>
<b>P<sub>0</sub></b>	19.56	19.57	21.27	21.63	<b>20.51</b>
<b>P<sub>1</sub></b>	18.46	18.58	19.17	19.33	<b>18.88</b>
<b>P<sub>2</sub></b>	17.26	17.34	17.45	17.88	<b>17.48</b>
<b>P<sub>3</sub></b>	11.87	12.58	15.34	17.05	<b>14.21</b>
<b>Mean</b>	<b>16.79</b>	<b>17.02</b>	<b>18.31</b>	<b>18.97</b>	
<b>CD (0.05) P; 0.107</b>		<b>CD (0.05) S; 0.107</b>		<b>CD (0.05) PxS; 0.214</b>	
<b>SE (m) P; 0.037</b>		<b>SE (m) S; 0.037</b>		<b>SE (m) PxS; 0.074</b>	

**Table 26: Effect of application of P and S on available manganese in soil (mg kg<sup>-1</sup>)**

**T<sub>1</sub>: 7.580**

	<b>S<sub>0</sub></b>	<b>S<sub>1</sub></b>	<b>S<sub>2</sub></b>	<b>S<sub>3</sub></b>	<b>Mean</b>
<b>P<sub>0</sub></b>	7.78	7.79	7.80	7.84	<b>7.80</b>
<b>P<sub>1</sub></b>	7.62	7.66	7.67	7.69	<b>7.66</b>
<b>P<sub>2</sub></b>	7.52	7.56	7.57	7.60	<b>7.56</b>
<b>P<sub>3</sub></b>	7.25	7.29	7.31	7.34	<b>7.30</b>
<b>Mean</b>	<b>7.54</b>	<b>7.58</b>	<b>7.59</b>	<b>7.62</b>	
<b>CD (0.05) P; 0.049</b>		<b>CD (0.05) S; NS</b>		<b>CD (0.05) PxS; NS</b>	
<b>SE (m) P; 0.017</b>		<b>SE (m) S; 0.017</b>		<b>SE (m) PxS; 0.034</b>	

**Table 27: Effect of application of P and S on available zinc in soil (mg kg<sup>-1</sup>)**

**T<sub>1</sub>: 1.22**

	<b>S<sub>0</sub></b>	<b>S<sub>1</sub></b>	<b>S<sub>2</sub></b>	<b>S<sub>3</sub></b>	<b>Mean</b>
<b>P<sub>0</sub></b>	1.18	1.35	2.12	3.04	<b>1.92</b>
<b>P<sub>1</sub></b>	1.01	1.18	2.18	2.24	<b>1.65</b>
<b>P<sub>2</sub></b>	0.88	1.03	1.20	2.23	<b>1.34</b>
<b>P<sub>3</sub></b>	0.78	1.14	1.23	1.24	<b>1.10</b>
<b>Mean</b>	<b>0.96</b>	<b>1.18</b>	<b>1.68</b>	<b>2.19</b>	
<b>CD (0.05) P; 0.003</b>		<b>CD (0.05) S; 0.003</b>		<b>CD (0.05) PxS; 0.007</b>	
<b>SE (m) P; 0.011</b>		<b>SE (m) S; 0.011</b>		<b>SE (m) PxS; 0.022</b>	

**Table 28: Effect of application of P and S on available copper in soil (mg kg<sup>-1</sup>)**

**T<sub>1</sub>: 2.03**

	<b>S<sub>0</sub></b>	<b>S<sub>1</sub></b>	<b>S<sub>2</sub></b>	<b>S<sub>3</sub></b>	<b>Mean</b>
<b>P<sub>0</sub></b>	2.04	2.08	2.07	2.04	<b>2.06</b>
<b>P<sub>1</sub></b>	2.02	2.01	2.02	2.03	<b>2.02</b>
<b>P<sub>2</sub></b>	2.02	2.07	2.04	2.06	<b>2.05</b>
<b>P<sub>3</sub></b>	2.02	2.04	2.01	2.05	<b>2.03</b>
<b>Mean</b>	<b>2.02</b>	<b>2.05</b>	<b>2.04</b>	<b>2.04</b>	
<b>CD (0.05) P; NS</b>		<b>CD (0.05) S; NS</b>		<b>CD (0.05) PxS; NS</b>	
<b>SE (m) P; 0.011</b>		<b>SE (m) S; 0.011</b>		<b>SE (m) PxS; 0.022</b>	

### 4.3.11 Water soluble boron

Data showed (table 29) that water soluble B in soil significantly varied due to application of different levels of P and S in the soil. B in soil decreased with application of increased dose of P. The treatment, P<sub>0</sub>S<sub>3</sub> (P, 0 kg ha<sup>-1</sup> and S, 30 kg ha<sup>-1</sup>) showed highest B (4.04 mg kg<sup>-1</sup>) among treatments. B content decreased with increased dose of P application and increased with increased dose of S application. F statistic for treatments Vs control was calculated and was found to be significantly different.

**Table 29: Effect of application of P and S on water soluble boron (mg kg<sup>-1</sup>)**

**T<sub>1</sub>: 1.32**

	<b>S<sub>0</sub></b>	<b>S<sub>1</sub></b>	<b>S<sub>2</sub></b>	<b>S<sub>3</sub></b>	<b>Mean</b>
<b>P<sub>0</sub></b>	1.14	1.45	2.50	4.04	<b>2.28</b>
<b>P<sub>1</sub></b>	1.18	1.45	1.54	1.73	<b>1.48</b>
<b>P<sub>2</sub></b>	0.77	0.86	1.74	2.32	<b>1.42</b>
<b>P<sub>3</sub></b>	0.73	0.77	0.86	1.23	<b>0.90</b>
<b>Mean</b>	<b>0.96</b>	<b>1.13</b>	<b>1.66</b>	<b>2.33</b>	
<b>CD (0.05) P; 0.006</b>		<b>CD (0.05) S; 0.006</b>		<b>CD (0.05) PxS; 0.012</b>	
<b>SE (m) P; 0.002</b>		<b>SE (m) S; 0.002</b>		<b>SE (m) PxS; 0.004</b>	

## 4.4 ANALYSIS OF PLANT SAMPLES

### 4.4.1 Nitrogen content in groundnut plant

Nitrogen content of plant samples at flowering, pegging, pod development and harvest stages are given in the tables 30, 31, 32 and 33 respectively. Nitrogen content of plant during pegging and pod formation stages was significantly influenced due to main effect and interaction effect of treatments, whereas, the interaction effect was found to be non-significant at flowering and harvesting stages. Plants showed highest nitrogen content at flowering stage, there after nitrogen content decreased with plant growth. Nitrogen content in plant increased with application of increased dose of phosphorus as well as S. Highest nitrogen (2.39 per

cent) was in the treatment, P<sub>3</sub>S<sub>3</sub> (P, 90 kg ha<sup>-1</sup> and S, 30 kg ha<sup>-1</sup>) at flowering, pegging (2.207 per cent), pod formation (2.117 per cent) and harvesting (1.78 per cent) stages. F statistic for treatments Vs control was calculated and found to be significantly different at flowering, pegging, pod formation and harvesting stages.

**Table 30: Effect of application of P and S on nitrogen content in plant samples at flowering stage (%)**

**T<sub>1</sub>: 2.12**

	S <sub>0</sub>	S <sub>1</sub>	S <sub>2</sub>	S <sub>3</sub>	Mean
P <sub>0</sub>	2.18	2.22	2.25	2.29	<b>2.24</b>
P <sub>1</sub>	2.21	2.25	2.27	2.30	<b>2.26</b>
P <sub>2</sub>	2.22	2.25	2.30	2.34	<b>2.28</b>
P <sub>3</sub>	2.25	2.30	2.31	2.39	<b>2.31</b>
Mean	<b>2.21</b>	<b>2.26</b>	<b>2.28</b>	<b>2.33</b>	
<b>CD (0.05) P; 0.015</b>		<b>CD (0.05) S; 0.015</b>		<b>CD (0.05) PxS; NS</b>	
<b>SE (m) P; 0.005</b>		<b>SE (m) S; 0.005</b>		<b>SE (m) PxS; 0.010</b>	

**Table 31: Effect of application of P and S on nitrogen content in plant samples at pegging stage (%)**

**T<sub>1</sub>: 1.79**

	S <sub>0</sub>	S <sub>1</sub>	S <sub>2</sub>	S <sub>3</sub>	Mean
P <sub>0</sub>	1.813	1.840	1.893	1.923	<b>1.868</b>
P <sub>1</sub>	1.830	1.877	1.850	1.960	<b>1.879</b>
P <sub>2</sub>	1.897	1.903	1.953	2.050	<b>1.951</b>
P <sub>3</sub>	1.927	1.927	2.053	2.207	<b>2.028</b>
Mean	<b>1.867</b>	<b>1.887</b>	<b>1.938</b>	<b>2.035</b>	
<b>CD (0.05) P; 0.04</b>		<b>CD (0.05) S; 0.04</b>		<b>CD (0.05) PxS; 0.079</b>	
<b>SE (m) P; 0.014</b>		<b>SE (m) S; 0.014</b>		<b>SE (m) PxS; 0.027</b>	



**Table 32: Effect of application of P and S on nitrogen content in plant samples at pod setting stage (%)**

**T<sub>1</sub>: 1.69**

	<b>S<sub>0</sub></b>	<b>S<sub>1</sub></b>	<b>S<sub>2</sub></b>	<b>S<sub>3</sub></b>	<b>Mean</b>
<b>P<sub>0</sub></b>	1.717	1.747	1.770	1.810	<b>1.761</b>
<b>P<sub>1</sub></b>	1.703	1.767	1.773	1.827	<b>1.768</b>
<b>P<sub>2</sub></b>	1.770	1.760	1.793	1.860	<b>1.796</b>
<b>P<sub>3</sub></b>	1.817	1.843	1.833	2.117	<b>1.903</b>
<b>Mean</b>	<b>1.752</b>	<b>1.779</b>	<b>1.793</b>	<b>1.903</b>	
<b>CD (0.05) P; 0.015</b>		<b>CD (0.05) S; 0.015</b>		<b>CD (0.05) PxS; 0.031</b>	
<b>SE (m) P; 0.005</b>		<b>SE (m) S; 0.005</b>		<b>SE (m) PxS; 0.011</b>	

**Table 33: Effect of application of P and S on nitrogen content in plant samples at harvest stage (%)**

**T<sub>1</sub>: 1.60**

	<b>S<sub>0</sub></b>	<b>S<sub>1</sub></b>	<b>S<sub>2</sub></b>	<b>S<sub>3</sub></b>	<b>Mean</b>
<b>P<sub>0</sub></b>	1.52	1.53	1.56	1.56	<b>1.54</b>
<b>P<sub>1</sub></b>	1.56	1.57	1.59	1.59	<b>1.58</b>
<b>P<sub>2</sub></b>	1.64	1.67	1.68	1.71	<b>1.67</b>
<b>P<sub>3</sub></b>	1.72	1.74	1.76	1.78	<b>1.75</b>
<b>Mean</b>	<b>1.61</b>	<b>1.63</b>	<b>1.65</b>	<b>1.66</b>	
<b>CD (0.05) P; 0.014</b>		<b>CD (0.05) S; 0.014</b>		<b>CD (0.05) PxS; NS</b>	
<b>SE (m) P; 0.005</b>		<b>SE (m) S; 0.005</b>		<b>SE (m) PxS; 0.010</b>	

#### **4.4.2 Phosphorus content in plant**

Phosphorus content in plant sample at flowering, pegging, pod development and harvest stages are given in the tables 34, 35, 36 and 37 respectively. P content in

plant was increased with increased dose of P and S fertilizer application. P content in all the stages significantly varied due to main effect and interaction effect. The treatment, P<sub>3</sub> S<sub>3</sub> (P, 90 kg ha<sup>-1</sup> and S, 30 kg ha<sup>-1</sup>) showed highest P content (0.37 per cent at flowering stage, 0.34 per cent at pegging stage, 0.32 per cent at pod formation stage and 0.27 per cent at harvesting stage). F statistic for treatments Vs control was calculated and were found to be significantly different in all stages.

**Table 34: Effect of application of P and S on phosphorus content in plant samples at flowering stage (%)**

**T<sub>1</sub>: 0.31**

	S <sub>0</sub>	S <sub>1</sub>	S <sub>2</sub>	S <sub>3</sub>	Mean
P <sub>0</sub>	0.23	0.27	0.31	0.31	<b>0.28</b>
P <sub>1</sub>	0.25	0.29	0.32	0.34	<b>0.30</b>
P <sub>2</sub>	0.27	0.32	0.34	0.36	<b>0.32</b>
P <sub>3</sub>	0.31	0.35	0.35	0.37	<b>0.34</b>
<b>Mean</b>	<b>0.27</b>	<b>0.31</b>	<b>0.33</b>	<b>0.34</b>	
<b>CD (0.05) P; 0.013</b>		<b>CD (0.05) S; 0.013</b>		<b>CD (0.05) PxS; 0.025</b>	
<b>SE (m) P; 0.011</b>		<b>SE (m) S; 0.011</b>		<b>SE (m) PxS; 0.023</b>	

**Table 35: Effect of application of P and S on phosphorus content in plant samples at pegging stage (%)**

**T<sub>1</sub>: 0.30**

	S <sub>0</sub>	S <sub>1</sub>	S <sub>2</sub>	S <sub>3</sub>	Mean
P <sub>0</sub>	0.21	0.21	0.26	0.28	<b>0.24</b>
P <sub>1</sub>	0.22	0.22	0.28	0.31	<b>0.26</b>
P <sub>2</sub>	0.23	0.25	0.30	0.32	<b>0.28</b>
P <sub>3</sub>	0.25	0.27	0.31	0.34	<b>0.29</b>
<b>Mean</b>	<b>0.23</b>	<b>0.24</b>	<b>0.29</b>	<b>0.31</b>	
<b>CD (0.05) P; 0.002</b>		<b>CD (0.05) S; 0.002</b>		<b>CD (0.05) PxS; 0.004</b>	
<b>SE (m) P; 0.022</b>		<b>SE (m) S; 0.022</b>		<b>SE (m) PxS; 0.044</b>	

**Table 36: Effect of application of P and S on phosphorus content in plant samples at pod formation stage (%)**

**T<sub>1</sub>: 0.26**

	<b>S<sub>0</sub></b>	<b>S<sub>1</sub></b>	<b>S<sub>2</sub></b>	<b>S<sub>3</sub></b>	<b>Mean</b>
<b>P<sub>0</sub></b>	0.18	0.18	0.22	0.23	<b>0.20</b>
<b>P<sub>1</sub></b>	0.18	0.21	0.24	0.25	<b>0.22</b>
<b>P<sub>2</sub></b>	0.20	0.23	0.25	0.27	<b>0.24</b>
<b>P<sub>3</sub></b>	0.22	0.25	0.27	0.32	<b>0.26</b>
<b>Mean</b>	<b>0.20</b>	<b>0.22</b>	<b>0.25</b>	<b>0.27</b>	
<b>CD (0.05) P; 0.005</b>		<b>CD (0.05) S; 0.005</b>		<b>CD (0.05) PxS; 0.010</b>	
<b>SE (m) P; 0.002</b>		<b>SE (m) S; 0.002</b>		<b>SE (m) PxS; 0.005</b>	

**Table 37: Effect of application of P and S on phosphorus content in plant samples at harvest stage (%)**

**T<sub>1</sub>: 0.18**

	<b>S<sub>0</sub></b>	<b>S<sub>1</sub></b>	<b>S<sub>2</sub></b>	<b>S<sub>3</sub></b>	<b>Mean</b>
<b>P<sub>0</sub></b>	0.12	0.15	0.18	0.22	<b>0.17</b>
<b>P<sub>1</sub></b>	0.14	0.17	0.21	0.22	<b>0.19</b>
<b>P<sub>2</sub></b>	0.16	0.20	0.21	0.25	<b>0.21</b>
<b>P<sub>3</sub></b>	0.18	0.22	0.24	0.27	<b>0.23</b>
<b>Mean</b>	<b>0.15</b>	<b>0.19</b>	<b>0.21</b>	<b>0.24</b>	
<b>CD (0.05) P; 0.006</b>		<b>CD (0.05) S; 0.006</b>		<b>CD (0.05) PxS; 0.013</b>	
<b>SE (m) P; 0.002</b>		<b>SE (m) S; 0.002</b>		<b>SE (m) PxS; 0.004</b>	

#### **4.4.3 Potassium content in plant**

Potassium content in plant samples was significantly influenced by application of P and S (table no.s 38, 39, 40 and 41). The treatment, P<sub>0</sub>S<sub>3</sub> (P, 0 kg ha<sup>-1</sup> and S, 30 kg ha<sup>-1</sup>) had highest K content at flowering (2.23 per cent), pegging

(2.12 per cent), pod formation (2.01 per cent) and harvesting (1.96 per cent) stages. K content in plants decreased with increased dose of P and increased with increased dose of S fertilizers. F statistic for treatments Vs control was calculated and was found to be significantly different.

**Table 38: Effect of application of P and S on potassium content in plant at flowering stage (%)**

**T<sub>1</sub>: 1.96**

	S <sub>0</sub>	S <sub>1</sub>	S <sub>2</sub>	S <sub>3</sub>	Mean
P <sub>0</sub>	1.39	1.66	1.83	2.23	<b>1.78</b>
P <sub>1</sub>	1.32	1.59	1.80	2.06	<b>1.69</b>
P <sub>2</sub>	1.23	1.59	1.69	1.93	<b>1.61</b>
P <sub>3</sub>	1.23	1.45	1.69	1.91	<b>1.57</b>
<b>Mean</b>	<b>1.29</b>	<b>1.57</b>	<b>1.76</b>	<b>2.03</b>	
<b>CD (0.05) P; 0.028</b>		<b>CD (0.05) S; 0.028</b>		<b>CD (0.05) PxS; 0.057</b>	
<b>SE (m) P; 0.010</b>		<b>SE (m) S; 0.010</b>		<b>SE (m) PxS; 0.020</b>	

**Table 39: Effect of application of P and S on potassium content in plant at pegging stage (%)**

**T<sub>1</sub>: 1.95**

	S <sub>0</sub>	S <sub>1</sub>	S <sub>2</sub>	S <sub>3</sub>	Mean
P <sub>0</sub>	1.35	1.64	1.81	2.12	<b>1.73</b>
P <sub>1</sub>	1.27	1.51	1.72	2.01	<b>1.63</b>
P <sub>2</sub>	1.21	1.48	1.59	1.96	<b>1.56</b>
P <sub>3</sub>	1.21	1.42	1.63	1.84	<b>1.52</b>
<b>Mean</b>	<b>1.26</b>	<b>1.51</b>	<b>1.69</b>	<b>1.98</b>	
<b>CD (0.05) P; 0.026</b>		<b>CD (0.05) S; 0.026</b>		<b>CD (0.05) PxS; 0.052</b>	
<b>SE (m) P; 0.009</b>		<b>SE (m) S; 0.009</b>		<b>SE (m) PxS; 0.018</b>	

**Table 40: Effect of application of P and S on potassium content in plant at pod setting stage (%)**

**T<sub>1</sub>: 1.92**

	<b>S<sub>0</sub></b>	<b>S<sub>1</sub></b>	<b>S<sub>2</sub></b>	<b>S<sub>3</sub></b>	<b>Mean</b>
<b>P<sub>0</sub></b>	1.33	1.60	1.74	2.01	<b>1.67</b>
<b>P<sub>1</sub></b>	1.23	1.42	1.69	1.95	<b>1.57</b>
<b>P<sub>2</sub></b>	1.13	1.42	1.53	1.86	<b>1.49</b>
<b>P<sub>3</sub></b>	1.12	1.37	1.56	1.75	<b>1.45</b>
<b>Mean</b>	<b>1.21</b>	<b>1.45</b>	<b>1.63</b>	<b>1.89</b>	
<b>CD (0.05) P; 0.021</b>		<b>CD (0.05) S; 0.021</b>		<b>CD (0.05) PxS; 0.042</b>	
<b>SE (m) P; 0.007</b>		<b>SE (m) S; 0.007</b>		<b>SE (m) PxS; 0.014</b>	

**Table 41: Effect of application of P and S on potassium content in plant at harvest stage (%)**

**T<sub>1</sub>: 1.89**

	<b>S<sub>0</sub></b>	<b>S<sub>1</sub></b>	<b>S<sub>2</sub></b>	<b>S<sub>3</sub></b>	<b>Mean</b>
<b>P<sub>0</sub></b>	1.30	1.57	1.72	1.96	<b>1.64</b>
<b>P<sub>1</sub></b>	1.22	1.38	1.62	1.90	<b>1.53</b>
<b>P<sub>2</sub></b>	1.07	1.37	1.48	1.84	<b>1.44</b>
<b>P<sub>3</sub></b>	1.09	1.33	1.47	1.70	<b>1.40</b>
<b>Mean</b>	<b>1.17</b>	<b>1.42</b>	<b>1.57</b>	<b>1.85</b>	
<b>CD (0.05) P; 0.011</b>		<b>CD (0.05) S; 0.011</b>		<b>CD (0.05) PxS; 0.023</b>	
<b>SE (m) P; 0.004</b>		<b>SE (m) S; 0.004</b>		<b>SE (m) PxS; 0.008</b>	

#### 4.4.4 Calcium content in plant

Calcium content in plant samples at all the four stages were found to be significantly varied due to application of different levels of fertilizers (tables 42, 43,44 and 45). Ca content was highest in treatment, P<sub>1</sub>S<sub>0</sub> (P, 60 kg ha<sup>-1</sup> and S, 0 kg ha<sup>-1</sup> at flowering (1.99 per cent) stage. At pegging stage, treatment P<sub>0</sub> S<sub>0</sub> showed high Ca content (1.617 per cent). At pod formation (1.21 per cent) and harvest (1.03 per cent) stages P<sub>0</sub>S<sub>3</sub> showed highest Ca content. F statistic for treatments Vs control was calculated and was found to be significantly different.

**Table 42: Effect of application of P and S on calcium content in plant at flowering stage (%)**

**T<sub>1</sub>: 1.68**

	<b>S<sub>0</sub></b>	<b>S<sub>1</sub></b>	<b>S<sub>2</sub></b>	<b>S<sub>3</sub></b>	<b>Mean</b>
<b>P<sub>0</sub></b>	1.967	1.717	1.887	1.673	<b>1.811</b>
<b>P<sub>1</sub></b>	1.990	1.847	1.637	1.597	<b>1.768</b>
<b>P<sub>2</sub></b>	1.877	1.737	1.567	1.557	<b>1.684</b>
<b>P<sub>3</sub></b>	1.847	1.710	1.560	1.517	<b>1.658</b>
<b>Mean</b>	<b>1.920</b>	<b>1.753</b>	<b>1.663</b>	<b>1.586</b>	
<b>CD (0.05) P; 0.012</b>		<b>CD (0.05) S; 0.012</b>		<b>CD (0.05) PxS; 0.023</b>	
<b>SE (m) P; 0.004</b>		<b>SE (m) S; 0.004</b>		<b>SE (m) PxS; 0.008</b>	

**Table 43: Effect of application of P and S on calcium content in plant at pegging stage (%)**

**T<sub>1</sub>: 1.450**

	<b>S<sub>0</sub></b>	<b>S<sub>1</sub></b>	<b>S<sub>2</sub></b>	<b>S<sub>3</sub></b>	<b>Mean</b>
<b>P<sub>0</sub></b>	1.617	1.593	1.477	1.503	<b>1.548</b>
<b>P<sub>1</sub></b>	1.543	1.583	1.480	1.437	<b>1.511</b>
<b>P<sub>2</sub></b>	1.553	1.527	1.447	1.417	<b>1.486</b>
<b>P<sub>3</sub></b>	1.500	1.520	1.383	1.237	<b>1.410</b>
<b>Mean</b>	<b>1.553</b>	<b>1.556</b>	<b>1.447</b>	<b>1.398</b>	
<b>CD (0.05) P; 0.010</b>		<b>CD (0.05) S; 0.010</b>		<b>CD (0.05) PxS; 0.019</b>	
<b>SE (m) P; 0.003</b>		<b>SE (m) S; 0.003</b>		<b>SE (m) PxS; 0.007</b>	

**Table 44: Effect of application of P and S on calcium content in plant at pod formation stage (%)**

**T<sub>1</sub>: 0.78**

	<b>S<sub>0</sub></b>	<b>S<sub>1</sub></b>	<b>S<sub>2</sub></b>	<b>S<sub>3</sub></b>	<b>Mean</b>
<b>P<sub>0</sub></b>	0.85	1.05	1.15	1.21	<b>1.07</b>
<b>P<sub>1</sub></b>	0.82	0.91	1.09	1.18	<b>1.00</b>
<b>P<sub>2</sub></b>	0.78	0.86	1.03	1.13	<b>0.95</b>
<b>P<sub>3</sub></b>	0.68	0.77	0.91	1.07	<b>0.86</b>
<b>Mean</b>	<b>0.78</b>	<b>0.90</b>	<b>1.05</b>	<b>1.15</b>	
<b>CD (0.05) P; 0.011</b>		<b>CD (0.05) S; 0.011</b>		<b>CD (0.05) PxS; 0.021</b>	
<b>SE (m) P; 0.004</b>		<b>SE (m) S; 0.004</b>		<b>SE (m) PxS; 0.007</b>	

**Table 45: Effect of application of P and S on calcium content in plant at harvest stage (%)**

**T<sub>1</sub>: 0.51**

	<b>S<sub>0</sub></b>	<b>S<sub>1</sub></b>	<b>S<sub>2</sub></b>	<b>S<sub>3</sub></b>	<b>Mean</b>
<b>P<sub>0</sub></b>	0.78	0.84	0.94	1.03	<b>0.90</b>
<b>P<sub>1</sub></b>	0.75	0.80	0.89	1.02	<b>0.87</b>
<b>P<sub>2</sub></b>	0.68	0.75	0.82	0.87	<b>0.78</b>
<b>P<sub>3</sub></b>	0.62	0.70	0.72	0.82	<b>0.72</b>
<b>Mean</b>	<b>0.71</b>	<b>0.77</b>	<b>0.84</b>	<b>0.94</b>	
<b>CD (0.05) P; 0.017</b>	<b>CD (0.05) S; 0.017</b>		<b>CD (0.05) PxS; 0.034</b>		
<b>SE (m) P; 0.006</b>	<b>SE (m) S; 0.006</b>		<b>SE (m) PxS; 0.012</b>		

#### **4.4.5 Magnesium content in plant**

Magnesium content in plant samples was significantly influenced by application of P and S ( tables 46, 47, 48 and 49). The treatment, P<sub>3</sub>S<sub>3</sub> (P, 90 kg ha<sup>-1</sup> and S, 30 kg ha<sup>-1</sup>) had highest Mg content at flowering (2.98 per cent), pegging (1.98 per cent), pod formation (1.08 per cent) and harvesting (1.04 per cent) stages. Mg content in plant samples increased with increased dose of both P and S fertilizer application. F statistic for treatments Vs control was calculated and was found to be significantly different.



**Table 46: Effect of application of P and S on magnesium content in plant at flowering stage (%)**

**T<sub>1</sub>: 2.51**

	<b>S<sub>0</sub></b>	<b>S<sub>1</sub></b>	<b>S<sub>2</sub></b>	<b>S<sub>3</sub></b>	<b>Mean</b>
<b>P<sub>0</sub></b>	2.47	2.62	2.72	2.80	<b>2.65</b>
<b>P<sub>1</sub></b>	2.53	2.69	2.70	2.85	<b>2.69</b>
<b>P<sub>2</sub></b>	2.59	2.74	2.83	2.93	<b>2.77</b>
<b>P<sub>3</sub></b>	2.64	2.89	2.67	2.98	<b>2.82</b>
<b>Mean</b>	<b>2.56</b>	<b>2.18</b>	<b>2.73</b>	<b>2.89</b>	
<b>CD (0.05) P; 0.008</b>		<b>CD (0.05) S; 0.008</b>		<b>CD (0.05) PxS; 0.016</b>	
<b>SE (m) P; 0.003</b>		<b>SE (m) S; 0.003</b>		<b>SE (m) PxS; 0.006</b>	

**Table 47: Effect of application of P and S on magnesium content in plant at pegging stage (%)**

**T<sub>1</sub>: 1.67**

	<b>S<sub>0</sub></b>	<b>S<sub>1</sub></b>	<b>S<sub>2</sub></b>	<b>S<sub>3</sub></b>	<b>Mean</b>
<b>P<sub>0</sub></b>	1.67	1.70	1.75	1.78	<b>1.72</b>
<b>P<sub>1</sub></b>	1.68	1.74	1.82	1.88	<b>1.78</b>
<b>P<sub>2</sub></b>	1.77	1.81	1.88	1.92	<b>1.85</b>
<b>P<sub>3</sub></b>	1.82	1.87	1.92	1.98	<b>1.90</b>
<b>Mean</b>	<b>1.74</b>	<b>1.78</b>	<b>1.84</b>	<b>1.89</b>	
<b>CD (0.05) P; 0.010</b>		<b>CD (0.05) S; 0.010</b>		<b>CD (0.05) PxS; 0.020</b>	
<b>SE (m) P; 0.003</b>		<b>SE (m) S; 0.003</b>		<b>SE (m) PxS; 0.007</b>	

**Table 48: Effect of application of P and S on magnesium content in plant at pod formation stage (%)**

**T<sub>1</sub>: 0.96**

	<b>S<sub>0</sub></b>	<b>S<sub>1</sub></b>	<b>S<sub>2</sub></b>	<b>S<sub>3</sub></b>	<b>Mean</b>
<b>P<sub>0</sub></b>	0.72	0.72	0.82	0.87	<b>0.78</b>
<b>P<sub>1</sub></b>	0.69	0.78	0.88	0.91	<b>0.82</b>
<b>P<sub>2</sub></b>	0.78	0.85	0.94	1.05	<b>0.90</b>
<b>P<sub>3</sub></b>	0.82	0.94	1.03	1.08	<b>0.97</b>
<b>Mean</b>	<b>0.75</b>	<b>0.82</b>	<b>0.92</b>	<b>0.98</b>	
<b>CD (0.05) P; 0.009</b>		<b>CD (0.05) S; 0.009</b>		<b>CD (0.05) PxS; 0.018</b>	
<b>SE (m) P; 0.003</b>		<b>SE (m) S; 0.003</b>		<b>SE (m) PxS; 0.006</b>	

**Table 49: Effect of application of P and S on magnesium content in plant at harvest stage (%)**

**T<sub>1</sub>: 0.74**

	<b>S<sub>0</sub></b>	<b>S<sub>1</sub></b>	<b>S<sub>2</sub></b>	<b>S<sub>3</sub></b>	<b>Mean</b>
<b>P<sub>0</sub></b>	0.67	0.71	0.80	0.85	<b>0.76</b>
<b>P<sub>1</sub></b>	0.67	0.78	0.86	0.89	<b>0.80</b>
<b>P<sub>2</sub></b>	0.77	0.83	0.94	1.02	<b>0.89</b>
<b>P<sub>3</sub></b>	0.80	0.91	0.98	1.04	<b>0.94</b>
<b>Mean</b>	<b>0.73</b>	<b>0.81</b>	<b>0.90</b>	<b>0.95</b>	
<b>CD (0.05) P; 0.008</b>		<b>CD (0.05) S; 0.008</b>		<b>CD (0.05) PxS; 0.015</b>	
<b>SE (m) P; 0.003</b>		<b>SE (m) S; 0.003</b>		<b>SE (m) PxS; 0.005</b>	

#### 4.4.6 Sulphur content in plant

Sulphur content of plant at different stages varied significantly due to main effect and interaction effect (tables 50, 51, 52 and 53). S content at all the stages were influenced by treatments. The treatment, P<sub>3</sub>S<sub>3</sub> produced high S content. Treatment, P<sub>3</sub>S<sub>3</sub> (P, 90 kg ha<sup>-1</sup> and S, 30 kg ha<sup>-1</sup>) showed highest S content at flowering (0.64 per cent), pegging (0.51 per cent), pod formation (0.34 per cent) and harvest stages (0.123 per cent). F statistic for treatments Vs control was calculated and was found to be significantly different.

**Table 50: Effect of application of P and S on sulphur content in plant at flowering stage (%)**

**T<sub>1</sub>: 0.22**

	<b>S<sub>0</sub></b>	<b>S<sub>1</sub></b>	<b>S<sub>2</sub></b>	<b>S<sub>3</sub></b>	<b>Mean</b>
<b>P<sub>0</sub></b>	0.11	0.21	0.24	0.29	<b>0.21</b>
<b>P<sub>1</sub></b>	0.13	0.22	0.27	0.33	<b>0.24</b>
<b>P<sub>2</sub></b>	0.18	0.22	0.25	0.34	<b>0.25</b>
<b>P<sub>3</sub></b>	0.18	0.23	0.32	0.64	<b>0.34</b>
<b>Mean</b>	<b>0.15</b>	<b>0.22</b>	<b>0.27</b>	<b>0.40</b>	
<b>CD (0.05) P; 0.005</b>		<b>CD (0.05) S; 0.005</b>		<b>CD (0.05) PxS; 0.010</b>	
<b>SE (m) P; 0.002</b>		<b>SE (m) S; 0.002</b>		<b>SE (m) PxS; 0.003</b>	

**Table 51: Effect of application of P and S on sulphur content in plant at pegging stage (%)**

**T<sub>1</sub>: 0.18**

	<b>S<sub>0</sub></b>	<b>S<sub>1</sub></b>	<b>S<sub>2</sub></b>	<b>S<sub>3</sub></b>	<b>Mean</b>
<b>P<sub>0</sub></b>	0.08	0.14	0.17	0.18	<b>0.14</b>
<b>P<sub>1</sub></b>	0.09	0.14	0.16	0.24	<b>0.16</b>
<b>P<sub>2</sub></b>	0.11	0.15	0.17	0.25	<b>0.17</b>
<b>P<sub>3</sub></b>	0.12	0.17	0.24	0.51	<b>0.26</b>
<b>Mean</b>	<b>0.10</b>	<b>0.15</b>	<b>0.19</b>	<b>0.30</b>	
<b>CD (0.05) P; 0.006</b>		<b>CD (0.05) S; 0.006</b>		<b>CD (0.05) PxS; 0.012</b>	
<b>SE (m) P; 0.002</b>		<b>SE (m) S; 0.002</b>		<b>SE (m) PxS; 0.004</b>	

**Table 52: Effect of application of P and S on sulphur content in plant at pod formation stage (%)**

**T<sub>1</sub>: 0.16**

	<b>S<sub>0</sub></b>	<b>S<sub>1</sub></b>	<b>S<sub>2</sub></b>	<b>S<sub>3</sub></b>	<b>Mean</b>
<b>P<sub>0</sub></b>	0.06	0.07	0.10	0.10	<b>0.08</b>
<b>P<sub>1</sub></b>	0.07	0.08	0.11	0.12	<b>0.09</b>
<b>P<sub>2</sub></b>	0.08	0.09	0.12	0.16	<b>0.11</b>
<b>P<sub>3</sub></b>	0.09	0.14	0.16	0.34	<b>0.18</b>
<b>Mean</b>	<b>0.08</b>	<b>0.09</b>	<b>0.12</b>	<b>0.18</b>	
<b>CD (0.05) P; 0.004</b>		<b>CD (0.05) S; 0.004</b>		<b>CD (0.05) PxS; 0.008</b>	
<b>SE (m) P; 0.001</b>		<b>SE (m) S; 0.001</b>		<b>SE (m) PxS; 0.002</b>	

**Table 53: Effect of application of P and S on sulphur content in plant at harvest stage (%)**

**T<sub>1</sub>: 0.030**

	<b>S<sub>0</sub></b>	<b>S<sub>1</sub></b>	<b>S<sub>2</sub></b>	<b>S<sub>3</sub></b>	<b>Mean</b>
<b>P<sub>0</sub></b>	0.019	0.019	0.028	0.092	<b>0.039</b>
<b>P<sub>1</sub></b>	0.015	0.019	0.019	0.025	<b>0.019</b>
<b>P<sub>2</sub></b>	0.022	0.028	0.031	0.058	<b>0.035</b>
<b>P<sub>3</sub></b>	0.043	0.052	0.095	0.123	<b>0.079</b>
<b>Mean</b>	<b>0.025</b>	<b>0.029</b>	<b>0.043</b>	<b>0.075</b>	
<b>CD (0.05) P; 0.001</b>		<b>CD (0.05) S; 0.001</b>		<b>CD (0.05) PxS; 0.002</b>	
<b>SE (m) P; 0.011</b>		<b>SE (m) S; 0.011</b>		<b>SE (m) PxS; 0.023</b>	

#### **4.4.7 Iron content in plant**

Iron content in plant at all stages was significantly affected by main effect and interaction effect (tables 54, 55, 56 and 57). P<sub>0</sub>S<sub>3</sub> (P, 0 kg ha<sup>-1</sup> and S, 30 kg ha<sup>-1</sup>) showed highest Fe content at flowering stage (799.00 mg kg<sup>-1</sup>), pegging (707.23 mg kg<sup>-1</sup>), pod formation (687.83 mg kg<sup>-1</sup>) and harvesting stages (663.20 mg kg<sup>-1</sup>). Fe content in plant samples decreased with increased dose of P fertilizer application and increased with increased dose of S application. F statistic for treatments Vs control was calculated and was found to be significantly different.

**Table 54: Effect of application of P and S on iron content in plant at flowering stage (mg kg<sup>-1</sup>)**

**T<sub>1</sub>: 488.50**

	<b>S<sub>0</sub></b>	<b>S<sub>1</sub></b>	<b>S<sub>2</sub></b>	<b>S<sub>3</sub></b>	<b>Mean</b>
<b>P<sub>0</sub></b>	504.33	621.00	664.33	799.00	<b>647.17</b>
<b>P<sub>1</sub></b>	395.50	618.50	661.83	741.50	<b>604.33</b>
<b>P<sub>2</sub></b>	366.50	564.00	639.50	720.83	<b>572.71</b>
<b>P<sub>3</sub></b>	303.00	524.50	637.00	706.95	<b>542.86</b>
<b>Mean</b>	<b>392.33</b>	<b>582.00</b>	<b>650.67</b>	<b>742.07</b>	
<b>CD (0.05) P; 0.972</b>		<b>CD (0.05) S; 0.972</b>		<b>CD (0.05) PxS; 1.944</b>	
<b>SE (m) P; 0.335</b>		<b>SE (m) S; 0.335</b>		<b>SE (m) PxS; 0.670</b>	

**Table 55: Effect of application of P and S on iron content in plant at pegging stage (mg kg<sup>-1</sup>)**

**T<sub>1</sub>: 472.10**

	<b>S<sub>0</sub></b>	<b>S<sub>1</sub></b>	<b>S<sub>2</sub></b>	<b>S<sub>3</sub></b>	<b>Mean</b>
<b>P<sub>0</sub></b>	498.63	604.43	611.43	707.23	<b>605.43</b>
<b>P<sub>1</sub></b>	324.43	593.83	623.70	703.30	<b>561.32</b>
<b>P<sub>2</sub></b>	362.83	523.10	617.60	701.43	<b>551.24</b>
<b>P<sub>3</sub></b>	302.37	511.87	604.83	697.43	<b>529.13</b>
<b>Mean</b>	<b>372.07</b>	<b>558.31</b>	<b>614.39</b>	<b>702.35</b>	
<b>CD (0.05) P; 0.204</b>		<b>CD (0.05) S; 0.204</b>		<b>CD (0.05) PxS; 0.407</b>	
<b>SE (m) P; 0.070</b>		<b>SE (m) S; 0.070</b>		<b>SE (m) PxS; 0.014</b>	

**Table 56: Effect of application of P and S on iron content in plant at pod formation stage (mg kg<sup>-1</sup>)**

**T<sub>1</sub>: 454.50**

	<b>S<sub>0</sub></b>	<b>S<sub>1</sub></b>	<b>S<sub>2</sub></b>	<b>S<sub>3</sub></b>	<b>Mean</b>
<b>P<sub>0</sub></b>	472.13	582.77	590.63	687.83	<b>583.34</b>
<b>P<sub>1</sub></b>	321.67	570.57	587.53	672.10	<b>537.97</b>
<b>P<sub>2</sub></b>	319.27	519.03	553.53	650.87	<b>510.68</b>
<b>P<sub>3</sub></b>	300.73	504.43	543.87	622.03	<b>492.77</b>
<b>Mean</b>	<b>353.45</b>	<b>544.20</b>	<b>568.89</b>	<b>658.21</b>	
<b>CD (0.05) P; 0.188</b>		<b>CD (0.05) S; 0.188</b>		<b>CD (0.05) PxS; 0.376</b>	
<b>SE (m) P; 0.065</b>		<b>SE (m) S; 0.065</b>		<b>SE (m) PxS; 0.130</b>	

**Table 57: Effect of application of P and S on iron content in plant at harvest stage (mg kg<sup>-1</sup>)**

**T<sub>1</sub>: 421.70**

	<b>S<sub>0</sub></b>	<b>S<sub>1</sub></b>	<b>S<sub>2</sub></b>	<b>S<sub>3</sub></b>	<b>Mean</b>
<b>P<sub>0</sub></b>	439.50	554.63	567.10	663.20	<b>556.11</b>
<b>P<sub>1</sub></b>	317.60	543.20	553.30	657.43	<b>517.88</b>
<b>P<sub>2</sub></b>	312.20	509.00	541.10	638.20	<b>500.13</b>
<b>P<sub>3</sub></b>	297.63	497.20	522.13	608.03	<b>481.25</b>
<b>Mean</b>	<b>341.73</b>	<b>526.01</b>	<b>545.91</b>	<b>641.72</b>	
<b>CD (0.05) P; 0.048</b>		<b>CD (0.05) S; 0.048</b>		<b>CD (0.05) PxS; 0.097</b>	
<b>SE (m) P; 0.017</b>		<b>SE (m) S; 0.017</b>		<b>SE (m) PxS; 0.033</b>	

#### 4.4.8 Manganese content in plant

Manganese content in plant samples at flowering, pegging, pod development and harvest stages are given in the tables 58, 59, 60 and 61 respectively. Mn content was high in treatment having low P and high S. The treatment, P<sub>0</sub>S<sub>3</sub> (P, 0 kg ha<sup>-1</sup> and S, 30 kg ha<sup>-1</sup>) showed highest Mn content at flowering (87.50 mg kg<sup>-1</sup>), pegging (85.30 mg kg<sup>-1</sup>), pod setting (83.10 mg kg<sup>-1</sup>) and harvest (80.90 mg kg<sup>-1</sup>) stages. Mn content was found to be significantly different in all stages. F statistic for treatments Vs control was calculated and was found to be significantly different.

**Table 58: Effect of application of P and S on manganese content in plant at flowering stage (mg kg<sup>-1</sup>)**

**T<sub>1</sub>: 73.50**

	<b>S<sub>0</sub></b>	<b>S<sub>1</sub></b>	<b>S<sub>2</sub></b>	<b>S<sub>3</sub></b>	<b>Mean</b>
<b>P<sub>0</sub></b>	40.83	53.00	69.50	87.50	<b>62.71</b>
<b>P<sub>1</sub></b>	40.83	50.00	68.00	85.50	<b>61.08</b>
<b>P<sub>2</sub></b>	38.70	46.00	65.00	80.17	<b>57.47</b>
<b>P<sub>3</sub></b>	36.10	45.00	53.00	71.42	<b>51.38</b>
<b>Mean</b>	<b>39.12</b>	<b>48.50</b>	<b>63.88</b>	<b>81.15</b>	
<b>CD (0.05) P; 0.191</b>	<b>CD (0.05) S; 0.191</b>			<b>CD (0.05) PxS; 0.381</b>	
<b>SE (m) P; 0.066</b>	<b>SE (m) S; 0.066</b>			<b>SE (m) PxS; 0.131</b>	



**Table 59: Effect of application of P and S on manganese content in plant at pegging stage (mg kg<sup>-1</sup>)**

**T<sub>1</sub>: 70.10**

	<b>S<sub>0</sub></b>	<b>S<sub>1</sub></b>	<b>S<sub>2</sub></b>	<b>S<sub>3</sub></b>	<b>Mean</b>
<b>P<sub>0</sub></b>	38.90	51.53	67.10	85.30	<b>60.71</b>
<b>P<sub>1</sub></b>	39.20	48.30	66.03	83.60	<b>59.28</b>
<b>P<sub>2</sub></b>	35.10	43.70	63.10	78.30	<b>55.05</b>
<b>P<sub>3</sub></b>	35.50	43.20	49.83	68.60	<b>49.28</b>
<b>Mean</b>	<b>37.18</b>	<b>46.68</b>	<b>61.52</b>	<b>78.95</b>	
<b>CD (0.05) P; 0.123</b>		<b>CD (0.05) S; 0.123</b>		<b>CD (0.05) PxS; 0.246</b>	
<b>SE (m) P; 0.042</b>		<b>SE (m) S; 0.042</b>		<b>SE (m) PxS; 0.085</b>	

**Table 60: Effect of application of P and S on manganese content in plant at pod setting stage (mg kg<sup>-1</sup>)**

**T<sub>1</sub>: 67.70**

	<b>S<sub>0</sub></b>	<b>S<sub>1</sub></b>	<b>S<sub>2</sub></b>	<b>S<sub>3</sub></b>	<b>Mean</b>
<b>P<sub>0</sub></b>	37.43	48.80	65.70	83.10	<b>58.76</b>
<b>P<sub>1</sub></b>	38.03	46.70	63.53	80.90	<b>57.29</b>
<b>P<sub>2</sub></b>	33.07	40.20	60.73	75.10	<b>52.28</b>
<b>P<sub>3</sub></b>	33.80	40.70	47.70	65.73	<b>46.98</b>
<b>Mean</b>	<b>35.58</b>	<b>44.10</b>	<b>59.42</b>	<b>76.21</b>	
<b>CD (0.05) P; 0.246</b>		<b>CD (0.05) S; 0.246</b>		<b>CD (0.05) PxS; 0.491</b>	
<b>SE (m) P; 0.085</b>		<b>SE (m) S; 0.085</b>		<b>SE (m) PxS; 0.169</b>	

**Table 61: Effect of application of P and S on manganese content in plant at harvest stage (mg kg<sup>-1</sup>)**

**T<sub>1</sub>: 65.20**

	<b>S<sub>0</sub></b>	<b>S<sub>1</sub></b>	<b>S<sub>2</sub></b>	<b>S<sub>3</sub></b>	<b>Mean</b>
<b>P<sub>0</sub></b>	36.13	46.30	63.20	80.90	<b>56.63</b>
<b>P<sub>1</sub></b>	35.90	44.43	62.30	78.40	<b>55.26</b>
<b>P<sub>2</sub></b>	30.90	38.43	57.60	70.30	<b>49.31</b>
<b>P<sub>3</sub></b>	32.30	38.23	45.30	61.53	<b>44.34</b>
<b>Mean</b>	<b>33.81</b>	<b>41.85</b>	<b>57.10</b>	<b>72.78</b>	
<b>CD (0.05) P; 0.279</b>		<b>CD (0.05) S; 0.279</b>		<b>CD (0.05) PxS; 0.559</b>	
<b>SE (m) P; 0.096</b>		<b>SE (m) S; 0.096</b>		<b>SE (m) PxS; 0.193</b>	

#### **4.4.9 Zinc content in plant**

Zinc content in plant samples at flowering, pegging, pod development and harvesting stages were significantly influenced by main effect and interaction effect. Whereas, at pegging stage, interaction effect was found to be non-significant. Zn content was highest in treatment having low P. Treatment P<sub>0</sub>S<sub>3</sub> (P, 0 kg ha<sup>-1</sup> and S, 30 kg ha<sup>-1</sup>) showed highest Zn content in all stages. F statistic for treatments Vs control was calculated and was found to be significantly different.

**Table 62: Effect of application of P and S on zinc content in plant at flowering stage (mg kg<sup>-1</sup>)**

**T<sub>1</sub>: 48.78**

	<b>S<sub>0</sub></b>	<b>S<sub>1</sub></b>	<b>S<sub>2</sub></b>	<b>S<sub>3</sub></b>	<b>Mean</b>
<b>P<sub>0</sub></b>	50.10	52.30	50.80	53.43	<b>51.66</b>
<b>P<sub>1</sub></b>	49.10	49.50	51.10	52.70	<b>50.60</b>
<b>P<sub>2</sub></b>	47.70	47.20	50.10	51.63	<b>49.16</b>
<b>P<sub>3</sub></b>	45.60	46.10	49.23	50.20	<b>47.78</b>
<b>Mean</b>	<b>48.13</b>	<b>48.78</b>	<b>50.31</b>	<b>51.99</b>	
<b>CD (0.05) P; 0.216</b>		<b>CD (0.05) S; 0.216</b>		<b>CD (0.05) PxS; 0.432</b>	
<b>SE (m) P; 0.075</b>		<b>SE (m) S; 0.075</b>		<b>SE (m) PxS; 0.149</b>	

**Table 63: Effect of application of P and S on zinc content in plant at pegging stage (mg kg<sup>-1</sup>)**

**T<sub>1</sub>: 46.70**

	<b>S<sub>0</sub></b>	<b>S<sub>1</sub></b>	<b>S<sub>2</sub></b>	<b>S<sub>3</sub></b>	<b>Mean</b>
<b>P<sub>0</sub></b>	47.10	48.70	51.50	52.53	<b>49.96</b>
<b>P<sub>1</sub></b>	47.30	46.63	50.40	51.70	<b>49.01</b>
<b>P<sub>2</sub></b>	44.63	45.50	49.10	50.20	<b>47.36</b>
<b>P<sub>3</sub></b>	43.20	44.63	47.20	48.80	<b>45.96</b>
<b>Mean</b>	<b>45.56</b>	<b>46.37</b>	<b>49.55</b>	<b>50.81</b>	
<b>CD (0.05) P; 0.249</b>		<b>CD (0.05) S; 0.249</b>		<b>CD (0.05) PxS; NS</b>	
<b>SE (m) P; 0.086</b>		<b>SE (m) S; 0.086</b>		<b>SE (m) PxS; 0.171</b>	

**Table 64: Effect of application of P and S on zinc content in plant at pod setting stage ( $\text{mg kg}^{-1}$ )**

**T<sub>1</sub>: 45.30**

	<b>S<sub>0</sub></b>	<b>S<sub>1</sub></b>	<b>S<sub>2</sub></b>	<b>S<sub>3</sub></b>	<b>Mean</b>
<b>P<sub>0</sub></b>	47.40	45.80	48.60	51.43	<b>48.31</b>
<b>P<sub>1</sub></b>	45.10	46.07	47.20	48.10	<b>46.62</b>
<b>P<sub>2</sub></b>	44.70	43.50	46.10	47.20	<b>45.38</b>
<b>P<sub>3</sub></b>	43.43	42.40	45.30	46.40	<b>44.38</b>
<b>Mean</b>	<b>45.16</b>	<b>44.44</b>	<b>46.80</b>	<b>48.28</b>	
<b>CD (0.05) P; 0.164</b>		<b>CD (0.05) S; 0.164</b>		<b>CD (0.05) PxS; 0.327</b>	
<b>SE (m) P; 0.056</b>		<b>SE (m) S; 0.056</b>		<b>SE (m) PxS; 0.113</b>	

**Table 65: Effect of application of P and S on zinc content in plant at harvest stage ( $\text{mg kg}^{-1}$ )**

**T<sub>1</sub>: 44.10**

	<b>S<sub>0</sub></b>	<b>S<sub>1</sub></b>	<b>S<sub>2</sub></b>	<b>S<sub>3</sub></b>	<b>Mean</b>
<b>P<sub>0</sub></b>	44.40	47.30	46.20	49.03	<b>46.73</b>
<b>P<sub>1</sub></b>	43.10	42.70	45.40	46.50	<b>44.43</b>
<b>P<sub>2</sub></b>	41.60	42.60	44.73	45.30	<b>43.56</b>
<b>P<sub>3</sub></b>	41.70	39.53	43.10	44.30	<b>42.16</b>
<b>Mean</b>	<b>42.70</b>	<b>43.03</b>	<b>44.86</b>	<b>46.28</b>	
<b>CD (0.05) P; 0.209</b>		<b>CD (0.05) S; 0.209</b>		<b>CD (0.05) PxS; 0.419</b>	
<b>SE (m) P; 0.072</b>		<b>SE (m) S; 0.072</b>		<b>SE (m) PxS; 0.144</b>	

#### 4.4.10 Copper content in plant

Copper content in plant samples at flowering, pegging, pod development and harvest stages are given in the tables 66, 67, 68 and 69 respectively. Cu content at flowering, pegging and pod formation stages was significant due to main effect of P and S and interaction effect. During flowering stage, Cu content in plants was highest in treatment P<sub>0</sub> S<sub>0</sub> (P, 0 kg ha<sup>-1</sup> and S, 0 kg ha<sup>-1</sup>). During pegging and pod formation stages Cu content was highest in treatments P<sub>0</sub> S<sub>3</sub> (P, 0 kg ha<sup>-1</sup> and S, 30 kg ha<sup>-1</sup>). At harvest stage Cu content was non-significant due to main effect of S and interaction effect. F statistic for treatments Vs control was calculated and was found to be significant at flowering, pegging and pod formation stages, whereas, at harvesting stage it was found to be non-significant.

**Table 66: Effect of application of P and S on copper content in plant at flowering stage (mg kg<sup>-1</sup>)**

T<sub>1</sub>: 17.20

	S <sub>0</sub>	S <sub>1</sub>	S <sub>2</sub>	S <sub>3</sub>	Mean
P <sub>0</sub>	19.63	18.70	19.20	18.80	<b>19.08</b>
P <sub>1</sub>	17.30	16.80	16.90	17.80	<b>17.20</b>
P <sub>2</sub>	15.63	14.70	14.90	15.20	<b>15.11</b>
P <sub>3</sub>	14.10	14.50	14.73	15.10	<b>14.61</b>
Mean	<b>16.67</b>	<b>16.18</b>	<b>16.43</b>	<b>16.73</b>	
CD (0.05) P; 0.209		CD (0.05) S; 0.209		CD (0.05) PxS; 0.419	
SE (m) P; 0.072		SE (m) S; 0.072		SE (m) PxS; 0.144	

**Table 67: Effect of application of P and S on copper content in plant at pegging stage (mg kg<sup>-1</sup>)**

**T<sub>1</sub>: 16.70**

	<b>S<sub>0</sub></b>	<b>S<sub>1</sub></b>	<b>S<sub>2</sub></b>	<b>S<sub>3</sub></b>	<b>Mean</b>
<b>P<sub>0</sub></b>	16.53	17.30	18.10	18.30	<b>17.56</b>
<b>P<sub>1</sub></b>	16.80	15.60	15.40	16.20	<b>16.00</b>
<b>P<sub>2</sub></b>	15.43	14.30	14.10	15.00	<b>14.71</b>
<b>P<sub>3</sub></b>	12.90	13.50	13.40	14.07	<b>13.47</b>
<b>Mean</b>	<b>15.42</b>	<b>15.18</b>	<b>15.25</b>	<b>15.89</b>	
<b>CD (0.05) P; 0.175</b>		<b>CD (0.05) S; 0.175</b>		<b>CD (0.05) PxS; 0.351</b>	
<b>SE (m) P; 0.060</b>		<b>SE (m) S; 0.060</b>		<b>SE (m) PxS; 0.121</b>	

**Table 68: Effect of application of P and S on copper content at in plant pod setting stage (mg kg<sup>-1</sup>)**

**T<sub>1</sub>: 14.30**

	<b>S<sub>0</sub></b>	<b>S<sub>1</sub></b>	<b>S<sub>2</sub></b>	<b>S<sub>3</sub></b>	<b>Mean</b>
<b>P<sub>0</sub></b>	14.43	15.30	15.10	15.40	<b>15.06</b>
<b>P<sub>1</sub></b>	14.20	14.40	14.10	14.07	<b>14.19</b>
<b>P<sub>2</sub></b>	13.70	13.50	13.03	13.30	<b>13.38</b>
<b>P<sub>3</sub></b>	12.50	12.20	12.10	12.13	<b>12.23</b>
<b>Mean</b>	<b>13.71</b>	<b>13.85</b>	<b>13.58</b>	<b>13.73</b>	
<b>CD (0.05) P; 0.124</b>		<b>CD (0.05) S; 0.124</b>		<b>CD (0.05) PxS; 0.249</b>	
<b>SE (m) P; 0.043</b>		<b>SE (m) S; 0.043</b>		<b>SE (m) PxS; 0.086</b>	

**Table 69: Effect of application of P and S on copper content in plant at harvest stage (mg kg<sup>-1</sup>)**

**T<sub>1</sub>: 11.73**

	<b>S<sub>0</sub></b>	<b>S<sub>1</sub></b>	<b>S<sub>2</sub></b>	<b>S<sub>3</sub></b>	<b>Mean</b>
<b>P<sub>0</sub></b>	11.93	11.90	11.80	11.90	<b>11.88</b>
<b>P<sub>1</sub></b>	11.50	11.70	11.70	11.60	<b>11.63</b>
<b>P<sub>2</sub></b>	11.63	11.50	11.60	11.20	<b>11.48</b>
<b>P<sub>3</sub></b>	11.20	11.40	11.73	11.10	<b>11.36</b>
<b>Mean</b>	<b>11.57</b>	<b>11.63</b>	<b>11.71</b>	<b>11.45</b>	
<b>CD (0.05) P; NS</b>		<b>CD (0.05) S; NS</b>		<b>CD (0.05) PxS; NS</b>	
<b>SE (m) P; 0.072</b>		<b>SE (m) S; 0.072</b>		<b>SE (m) PxS; 0.144</b>	

#### **4.4.11 Boron content in plant**

Boron content at flowering, pegging, pod development and harvest stages are given in the tables 70, 71, 72 and 73 respectively. The B content was significantly influenced by different levels of P application. The treatment with higher dose of P significantly reduced the B content. Application of S was not found to cause significant difference between treatments. F statistic for treatments Vs control was calculated and was found to be significantly different.

**Table 70: Effect of application of P and S on boron content in plant at flowering stage ( $\text{mg kg}^{-1}$ )**

**T<sub>1</sub>: 16.30**

	<b>S<sub>0</sub></b>	<b>S<sub>1</sub></b>	<b>S<sub>2</sub></b>	<b>S<sub>3</sub></b>	<b>Mean</b>
<b>P<sub>0</sub></b>	18.13	18.17	18.00	18.07	<b>18.09</b>
<b>P<sub>1</sub></b>	17.27	17.20	17.43	17.33	<b>17.31</b>
<b>P<sub>2</sub></b>	16.43	16.30	16.10	16.13	<b>16.24</b>
<b>P<sub>3</sub></b>	15.33	15.30	15.10	15.23	<b>15.24</b>
<b>Mean</b>	<b>16.79</b>	<b>16.74</b>	<b>16.66</b>	<b>16.69</b>	
<b>CD (0.05) P; 0.277</b>		<b>CD (0.05) S; NS</b>		<b>CD (0.05) PxS; NS</b>	
<b>SE (m) P; 0.095</b>		<b>SE (m) S; 0.095</b>		<b>SE (m) PxS; 0.191</b>	

**Table 71: Effect of application of P and S on boron content in plant at pegging stage ( $\text{mg kg}^{-1}$ )**

**T<sub>1</sub>: 15.02**

	<b>S<sub>0</sub></b>	<b>S<sub>1</sub></b>	<b>S<sub>2</sub></b>	<b>S<sub>3</sub></b>	<b>Mean</b>
<b>P<sub>0</sub></b>	16.20	16.27	16.23	16.23	<b>16.23</b>
<b>P<sub>1</sub></b>	15.47	15.40	15.30	15.43	<b>15.40</b>
<b>P<sub>2</sub></b>	14.67	14.51	14.61	14.63	<b>14.60</b>
<b>P<sub>3</sub></b>	13.47	13.70	13.26	13.42	<b>13.46</b>
<b>Mean</b>	<b>14.95</b>	<b>14.97</b>	<b>14.85</b>	<b>14.93</b>	
<b>CD (0.05) P; 0.439</b>		<b>CD (0.05) S; NS</b>		<b>CD (0.05) PxS; NS</b>	
<b>SE (m) P; 0.151</b>		<b>SE (m) S; 0.151</b>		<b>SE (m) PxS; 0.303</b>	



**Table 72: Effect of application of P and S on boron content at in plant pod setting stage (mg kg<sup>-1</sup>)**

**T<sub>1</sub>: 13.04**

	S <sub>0</sub>	S <sub>1</sub>	S <sub>2</sub>	S <sub>3</sub>	Mean
P <sub>0</sub>	14.35	14.24	14.13	14.15	14.22
P <sub>1</sub>	13.14	13.37	13.29	13.25	13.26
P <sub>2</sub>	13.10	13.08	13.05	13.07	13.08
P <sub>3</sub>	12.30	12.24	12.18	12.16	12.22
Mean	13.22	13.23	13.16	13.16	
CD (0.05) P; 0.232		CD (0.05) S; NS		CD (0.05) PxS; NS	
SE (m) P; 0.08		SE (m) S; 0.08		SE (m) PxS; 0.16	

**Table 73: Effect of P and B application on boron content in plant at harvest stage (mg kg<sup>-1</sup>)**

**T<sub>1</sub>: 11.70**

	S <sub>0</sub>	S <sub>1</sub>	S <sub>2</sub>	S <sub>3</sub>	Mean
P <sub>0</sub>	12.27	12.34	12.36	12.27	12.31
P <sub>1</sub>	11.26	11.34	11.21	11.23	11.26
P <sub>2</sub>	11.04	11.02	11.03	11.04	11.03
P <sub>3</sub>	10.42	10.21	10.40	10.42	10.36
Mean	11.25	11.23	11.25	11.24	
CD (0.05) P; 0.149		CD (0.05) S; NS		CD (0.05) PxS; NS	
SE (m) P; 0.052		SE (m) S; 0.052		SE (m) PxS; 0.103	

## 4.5 NUTRIENT CONTENT IN POD SAMPLES

### 4.5.1 Nitrogen content in kernels

Nitrogen content in kernels varied significantly due to main effect and interaction effect. N content was increased with increased dose of P and S application (table 74). Highest N content was observed in  $P_3S_3$  (P, 90 kg ha<sup>-1</sup> and S, 30 kg ha<sup>-1</sup>) which had 3.83 per cent N. F statistic for treatments Vs control was calculated and was found to be significantly different.

### 4.5.2 Phosphorus content in kernels

Data (table 75) on P content in kernels showed that treatments varied significantly due to main effect and interaction effect. Treatment,  $P_3S_3$  (P, 90 kg ha<sup>-1</sup> and S, 30 kg ha<sup>-1</sup>) showed highest P content (0.414 per cent). P content in kernel increased with increased dose of P and S application. F statistic for treatments Vs control was calculated and was found to be significantly different.

### 4.5.3 Potassium content in kernels

Effect of application of P and S on K content in kernels is shown in the table 76. The K content varied significantly due to main effect whereas interaction effect was found to be non-significant. Treatment,  $P_0S_3$  (P, 0 kg ha<sup>-1</sup> and S, 30 kg ha<sup>-1</sup>) showed highest K content (1.631 per cent). F statistic for treatments Vs control was calculated and was found to be significantly different.

**Table 74: Effect of application of P and S on nitrogen content in kernel (%)**

**T<sub>1</sub>: 3.24**

	<b>S<sub>0</sub></b>	<b>S<sub>1</sub></b>	<b>S<sub>2</sub></b>	<b>S<sub>3</sub></b>	<b>Mean</b>
<b>P<sub>0</sub></b>	3.15	3.25	3.49	3.60	<b>3.37</b>
<b>P<sub>1</sub></b>	3.28	3.44	3.49	3.69	<b>3.48</b>
<b>P<sub>2</sub></b>	3.37	3.48	3.65	3.66	<b>3.54</b>
<b>P<sub>3</sub></b>	3.47	3.57	3.68	3.83	<b>3.64</b>
<b>Mean</b>	<b>3.32</b>	<b>3.44</b>	<b>3.58</b>	<b>3.69</b>	
<b>CD (0.05) P; 0.054</b>		<b>CD (0.05) S; 0.054</b>		<b>CD (0.05) PxS; NS</b>	
<b>SE (m) P; 0.019</b>		<b>SE (m) S; 0.019</b>		<b>SE (m) PxS; 0.037</b>	

**Table 75: Effect of application of P and S on phosphorus content in kernel (%)**

**T<sub>1</sub>: 0.35**

	<b>S<sub>0</sub></b>	<b>S<sub>1</sub></b>	<b>S<sub>2</sub></b>	<b>S<sub>3</sub></b>	<b>Mean</b>
<b>P<sub>0</sub></b>	0.332	0.344	0.353	0.363	<b>0.348</b>
<b>P<sub>1</sub></b>	0.342	0.346	0.352	0.382	<b>0.356</b>
<b>P<sub>2</sub></b>	0.351	0.355	0.377	0.393	<b>0.369</b>
<b>P<sub>3</sub></b>	0.351	0.366	0.392	0.414	<b>0.381</b>
<b>Mean</b>	<b>0.344</b>	<b>0.353</b>	<b>0.368</b>	<b>0.388</b>	
<b>CD (0.05) P; 0.003</b>		<b>CD (0.05) S; 0.003</b>		<b>CD (0.05) PxS; 0.005</b>	
<b>SE (m) P; 0.001</b>		<b>SE (m) S; 0.001</b>		<b>SE (m) PxS; 0.002</b>	

**Table 76: Effect of application of P and S on potassium content in kernel (%)****T<sub>1</sub>: 1.43**

	<b>S<sub>0</sub></b>	<b>S<sub>1</sub></b>	<b>S<sub>2</sub></b>	<b>S<sub>3</sub></b>	<b>Mean</b>
<b>P<sub>0</sub></b>	1.338	1.463	1.543	1.631	<b>1.494</b>
<b>P<sub>1</sub></b>	1.283	1.393	1.430	1.531	<b>1.409</b>
<b>P<sub>2</sub></b>	1.243	1.283	1.320	1.413	<b>1.315</b>
<b>P<sub>3</sub></b>	1.193	1.223	1.253	1.377	<b>1.262</b>
<b>Mean</b>	<b>1.265</b>	<b>1.341</b>	<b>1.387</b>	<b>1.488</b>	
<b>CD (0.05) P; 0.034</b>		<b>CD (0.05) S; 0.034</b>		<b>CD (0.05) PxS; NS</b>	
<b>SE (m) P; 0.012</b>		<b>SE (m) S 0.012</b>		<b>SE (m) PxS; 0.023</b>	

**4.5.4 Calcium content in kernels**

Effect of application of P and S on Ca content in pod is shown in the table 77. Calcium content varied significantly due to the application of P and S fertilizers. Ca showed a synergistic interaction with P. Ca content increased with increased dose of P application. Highest Ca content was noted in P<sub>3</sub>S<sub>1</sub> (P, 90 kg ha<sup>-1</sup> and S, 10 kg ha<sup>-1</sup>), where the Ca content was 0.245 per cent. F statistic for treatments Vs control was calculated and was found to be significantly different.

**4.5.5 Magnesium content in kernels**

Effect of application of P and S on Mg content in kernel is shown in the table 78. Mg content varied significantly due to the application of P and S fertilizers. Highest Mg content was observed in P<sub>2</sub>S<sub>3</sub> (P, 75 kg ha<sup>-1</sup> and S, 30 kg ha<sup>-1</sup>) (0.351 per cent). F statistic for treatments Vs control was calculated and was found to be significantly different.

**4.5.6 Sulphur content in kernels**

Sulphur content varied significantly due to the application of P and S fertilizers. The S content showed a synergistic interaction with P upto medium dose of P (75 kg ha<sup>-1</sup>). S content increased with increased dose of P application (table 79).

Highest S content was noted in P<sub>2</sub>S<sub>3</sub> (P, 75 kg ha<sup>-1</sup> and S, 30 kg ha<sup>-1</sup>) and it was recorded as 0.224 per cent S. F statistic for treatments Vs control was calculated and was found to be significantly different.

**Table 77: Effect of application of P and S on calcium content in kernel (%)**

**T<sub>1</sub>: 0.132**

	S <sub>0</sub>	S <sub>1</sub>	S <sub>2</sub>	S <sub>3</sub>	Mean
P <sub>0</sub>	0.113	0.138	0.148	0.134	<b>0.133</b>
P <sub>1</sub>	0.147	0.155	0.155	0.156	<b>0.153</b>
P <sub>2</sub>	0.152	0.166	0.186	0.191	<b>0.174</b>
P <sub>3</sub>	0.197	0.254	0.219	0.214	<b>0.221</b>
Mean	<b>0.152</b>	<b>0.178</b>	<b>0.177</b>	<b>0.174</b>	
CD (0.05) P; 0.013		CD (0.05) S; 0.013		CD (0.05) PxS; 0.026	
SE (m) P; 0.005		SE (m) S; 0.005		SE (m) PxS; 0.009	

**Table 78: Effect of application of P and S on magnesium content in kernel (%)**

**T<sub>1</sub>: 0.25**

	S <sub>0</sub>	S <sub>1</sub>	S <sub>2</sub>	S <sub>3</sub>	Mean
P <sub>0</sub>	0.231	0.291	0.321	0.341	<b>0.296</b>
P <sub>1</sub>	0.231	0.291	0.313	0.340	<b>0.294</b>
P <sub>2</sub>	0.241	0.261	0.281	0.351	<b>0.283</b>
P <sub>3</sub>	0.241	0.241	0.250	0.331	<b>0.266</b>
Mean	<b>0.236</b>	<b>0.271</b>	<b>0.291</b>	<b>0.341</b>	
CD (0.05) P; 0.002		CD (0.05) S; 0.002		CD (0.05) PxS; 0.003	
SE (m) P; 0.001		SE (m) S; 0.001		SE (m) PxS; 0.001	

**Table 79: Effect of application of P and S on sulphur content in kernel (%)**

**T<sub>1</sub>: 0.152**

	S <sub>0</sub>	S <sub>1</sub>	S <sub>2</sub>	S <sub>3</sub>	Mean
P <sub>0</sub>	0.149	0.174	0.195	0.222	<b>0.186</b>
P <sub>1</sub>	0.152	0.175	0.194	0.218	<b>0.185</b>
P <sub>2</sub>	0.167	0.182	0.193	0.224	<b>0.192</b>
P <sub>3</sub>	0.152	0.163	0.188	0.219	<b>0.181</b>
Mean	<b>0.155</b>	<b>0.174</b>	<b>0.193</b>	<b>0.222</b>	
CD (0.05) P; 0.001		CD (0.05) S; 0.001		CD (0.05) P x S; 0.002	
SE (m) P; 0.011		SE (m) S; 0.011		SE (m) P x S; 0.022	

#### 4.5.7 Iron content in kernels

Effect of application of P and S on Fe content in kernel is shown in the table 80. Fe content was influenced by both main effect and interaction effect. Highest Fe content was noted in P<sub>0</sub>S<sub>3</sub> (P, 90 kg ha<sup>-1</sup> and S, 30 kg ha<sup>-1</sup>). F statistic for treatments Vs control was calculated and was found to be significantly different.

#### 4.5.8 Manganese content in kernels

Manganese content varied significantly due to main effect and interaction effect. Mn content decreased with increased dose of P application (table 81). The highest Mn content was observed in P<sub>0</sub>S<sub>3</sub> (P, 90 kg ha<sup>-1</sup> and S, 30 kg ha<sup>-1</sup>). F statistic for treatments Vs control was calculated and was found to be significantly different.

#### 4.5.9 Zinc content in kernels

Data (table 82) on Zn content showed that Zn was influenced by P and S application. Zn content decreased with increased dose of P application. Highest Zn was noted in P<sub>0</sub> S<sub>3</sub> (P, 0 kg ha<sup>-1</sup> and S, 0 kg ha<sup>-1</sup>). F statistic for treatments Vs control was calculated and was found to be significantly different.

#### 4.5.10 Copper content in kernels

Copper content was found to be significant due to main effect and interaction effect (table 83). Highest Cu content was observed in P<sub>0</sub>S<sub>3</sub> (P, 0 kg ha<sup>-1</sup> and S, 30 kg ha<sup>-1</sup>). F statistic for treatments Vs control was calculated and was found to be significantly different.

#### 4.5.11 Boron content in kernels

Data (table 84) on B content showed that B was significantly influenced by P and S application. B content decreased with increased dose of P application. Highest B was noted in P<sub>0</sub>S<sub>2</sub> (P, 0 kg ha<sup>-1</sup> and S, 20 kg ha<sup>-1</sup>). It was on par with treatment, P<sub>0</sub> S<sub>0</sub> (P, 0 kg ha<sup>-1</sup> and S, 0 kg ha<sup>-1</sup>). F statistic for treatments Vs control was calculated and was found to be significantly different.

**Table 80: Effect of application of P and S on iron content in kernel (mg kg<sup>-1</sup>)**

**T<sub>1</sub>: 123.00**

	S <sub>0</sub>	S <sub>1</sub>	S <sub>2</sub>	S <sub>3</sub>	Mean
P <sub>0</sub>	206.00	274.00	343.00	399.33	305.58
P <sub>1</sub>	182.00	215.00	281.00	354.33	258.08
P <sub>2</sub>	167.00	186.00	245.33	313.00	227.83
P <sub>3</sub>	117.33	156.00	197.00	276.00	186.58
Mean	168.08	207.75	266.58	335.67	
CD (0.05) P; 0.248		CD (0.05) S; 0.248		CD (0.05) PxS; 0.496	
SE (m) P; 0.085		SE (m) S; 0.085		SE (m) PxS; 0.171	

**Table 81: Effect of application of P and S on manganese content in kernel (mg kg<sup>-1</sup>)**

**T<sub>1</sub>: 20.64**

	<b>S<sub>0</sub></b>	<b>S<sub>1</sub></b>	<b>S<sub>2</sub></b>	<b>S<sub>3</sub></b>	<b>Mean</b>
<b>P<sub>0</sub></b>	19.96	21.95	24.30	26.75	<b>23.24</b>
<b>P<sub>1</sub></b>	18.00	20.00	22.50	24.00	<b>21.13</b>
<b>P<sub>2</sub></b>	17.03	17.95	18.75	21.35	<b>18.77</b>
<b>P<sub>3</sub></b>	15.00	16.25	17.55	19.07	<b>16.97</b>
<b>Mean</b>	<b>17.50</b>	<b>19.04</b>	<b>20.78</b>	<b>22.79</b>	
<b>CD (0.05) P; 0.027</b>		<b>CD (0.05) S; 0.027</b>		<b>CD (0.05) PxS; 0.053</b>	
<b>SE (m) P; 0.009</b>		<b>SE (m) S; 0.009</b>		<b>SE (m) PxS; 0.018</b>	

**Table 82: Effect of application of P and S on zinc content in kernel (mg kg<sup>-1</sup>)**

**T<sub>1</sub>: 52.21**

	<b>S<sub>0</sub></b>	<b>S<sub>1</sub></b>	<b>S<sub>2</sub></b>	<b>S<sub>3</sub></b>	<b>Mean</b>
<b>P<sub>0</sub></b>	53.00	55.00	57.65	60.83	<b>56.62</b>
<b>P<sub>1</sub></b>	53.25	54.25	53.75	56.00	<b>54.31</b>
<b>P<sub>2</sub></b>	47.25	48.00	49.07	51.25	<b>48.89</b>
<b>P<sub>3</sub></b>	45.50	46.75	47.25	46.10	<b>46.40</b>
<b>Mean</b>	<b>49.75</b>	<b>51.00</b>	<b>51.93</b>	<b>53.55</b>	
<b>CD (0.05) P; 0.130</b>		<b>CD (0.05) S; 0.130</b>		<b>CD (0.05) PxS; 0.260</b>	
<b>SE (m) P; 0.045</b>		<b>SE (m) S; 0.045</b>		<b>SE (m) PxS; 0.090</b>	



**Table 83: Effect of application of P and S on copper content in kernel (mg kg<sup>-1</sup>)**

**T<sub>1</sub>: 13.75**

	<b>S<sub>0</sub></b>	<b>S<sub>1</sub></b>	<b>S<sub>2</sub></b>	<b>S<sub>3</sub></b>	<b>Mean</b>
<b>P<sub>0</sub></b>	13.57	13.00	13.75	14.00	<b>13.58</b>
<b>P<sub>1</sub></b>	12.75	12.27	12.50	12.00	<b>12.38</b>
<b>P<sub>2</sub></b>	10.75	11.25	10.50	11.50	<b>11.00</b>
<b>P<sub>3</sub></b>	10.13	10.25	9.50	10.07	<b>9.99</b>
<b>Mean</b>	<b>11.80</b>	<b>11.69</b>	<b>11.56</b>	<b>11.89</b>	
<b>CD (0.05) P; 0.117</b>		<b>CD (0.05) S; 0.117</b>		<b>CD (0.05) PxS; 0.233</b>	
<b>SE (m) P; 0.040</b>		<b>SE (m) S; 0.040</b>		<b>SE (m) PxS; 0.080</b>	

**Table 84: Effect of application of P and S on boron content in kernel (mg kg<sup>-1</sup>)**

**T<sub>1</sub>: 17.50**

	<b>S<sub>0</sub></b>	<b>S<sub>1</sub></b>	<b>S<sub>2</sub></b>	<b>S<sub>3</sub></b>	<b>Mean</b>
<b>P<sub>0</sub></b>	20.23	20.07	20.24	20.16	<b>20.18</b>
<b>P<sub>1</sub></b>	19.03	17.50	18.25	19.50	<b>18.57</b>
<b>P<sub>2</sub></b>	16.13	17.00	16.25	17.25	<b>16.66</b>
<b>P<sub>3</sub></b>	15.07	15.25	14.97	14.99	<b>15.07</b>
<b>Mean</b>	<b>17.62</b>	<b>17.45</b>	<b>17.43</b>	<b>17.98</b>	
<b>CD (0.05) P; 0.060</b>		<b>CD (0.05) S; 0.060</b>		<b>CD (0.05) PxS; 0.121</b>	
<b>SE (m) P; 0.021</b>		<b>SE (m) S; 0.021</b>		<b>SE (m) PxS; 0.042</b>	

#### 4.5.12 Nitrogen content in shell

Nitrogen content in shell varied significantly due to main effect and interaction effect. N content was increased with increased dose of P and S application (table 85). Highest N content was observed in  $P_3S_3$  (P, 90 kg ha<sup>-1</sup> and S, 30 kg ha<sup>-1</sup>) which had 1.193 per cent. F statistic for treatments Vs control was calculated and was found to be significantly different.

#### 4.5.13 Phosphorus content in shell

Data (table 86) on P content in shell showed that treatments varied significantly due to main effect and interaction effect. Treatment,  $P_3S_3$  (P, 90 kg ha<sup>-1</sup> and S, 30 kg ha<sup>-1</sup>) showed highest P content (0.223%). P content in shell increased with increased dose of P and S application. F statistic for treatments Vs control was calculated and was found to be significantly different.

#### 4.5.14 Potassium content in shell

Effect of application of P and S on K content in shell is shown in the table 87. The K content varied significantly due to main effect whereas interaction effect was found to be non-significant. Treatment,  $P_0S_3$  (P, 0 kg ha<sup>-1</sup> and S, 30 kg ha<sup>-1</sup>) showed highest K content (0.769%). F statistic for treatments Vs control was calculated and was found to be significantly different.

#### 4.5.15 Calcium content in shell

Effect of application of P and S on Ca content in shell is shown in the table 88. Calcium content varied significantly due to the application of P and S fertilizers. Ca showed a synergistic interaction with P. Ca content increased with increased dose of P application. Highest Ca content was noted in  $P_3S_0$  (P, 90 kg ha<sup>-1</sup> and S, 0 kg ha<sup>-1</sup>). F statistic for treatments Vs control was calculated and was found to be significantly different.

#### **4.5.16 Magnesium content in shell**

Effect of application of P and S on Mg content in shell is shown in the table 89. Mg content varied significantly due to the application of P and S fertilizers. Highest Mg content was observed in  $P_0S_3$  (P, 0 kg ha<sup>-1</sup> and S, 30 kg ha<sup>-1</sup>). F statistic for treatments Vs control was calculated and was found to be significantly different.

#### **4.5.17 Sulphur content in shell**

Sulphur content varied significantly due to the application of P and S fertilizers. The S showed a synergistic interaction with P upto medium dose of P (P, 75 kg ha<sup>-1</sup>). S content increased with increased dose of P application (table 90). Highest S content was noted in  $P_3S_3$  (P, 90 kg ha<sup>-1</sup> and S, 30 kg ha<sup>-1</sup>). F statistic for treatments Vs control was calculated and was found to be significantly different.

#### **4.5.18 Iron content in shell**

Effect of application of P and S on Fe content in shell is shown in the table 91. Fe content was influenced by both main effect and interaction effect. Highest Fe content was noted in  $P_0S_3$  (P, 0 kg ha<sup>-1</sup> and S, 30 kg ha<sup>-1</sup>). F statistic for treatments Vs control was calculated and was found to be significantly different.

#### **4.5.19 Manganese content in shell**

Manganese content varied significantly due to main effect and interaction effect. Mn content decreased with increased dose of P application (table 92). The highest Mn content was observed in  $P_0S_3$  (P, 0 kg ha<sup>-1</sup> and S, 30 kg ha<sup>-1</sup>). F statistic for treatments Vs control was calculated and was found to be significantly different.

#### **4.5.20 Zinc content in shell**

Data (table 93) showed that Zn content was influenced by P and S application. Zn content decreased with increased dose of P application. Highest Zn was noted in  $P_0S_3$  (P, 0 kg ha<sup>-1</sup> and S, 30 kg ha<sup>-1</sup>). F statistic for treatments Vs control was calculated and was found to be significantly different.

#### 4.5.21 Copper content in shell

Copper content was found to be significant due to main effect and interaction effect (table 94). Highest Cu content was observed in P<sub>0</sub>S<sub>2</sub> (P, 0 kg ha<sup>-1</sup> and S, 20 kg ha<sup>-1</sup>). F statistic for treatments Vs control was calculated and was found to be significantly different.

#### 4.5.22 Boron content in shell

Data (table 95) on B content showed that B was influenced by P and S application. B content decreased with increased dose of P application. Highest B (15.35 mg kg<sup>-1</sup>) was noted in P<sub>0</sub>S<sub>3</sub> (P, 0 kg ha<sup>-1</sup> and S, 30 kg ha<sup>-1</sup>). F statistic for treatments Vs control was calculated and was found to be significantly different.

**Table 85: Effect of application of P and S on nitrogen content in shell (%)**

T<sub>1</sub>: 1.020

	S <sub>0</sub>	S <sub>1</sub>	S <sub>2</sub>	S <sub>3</sub>	Mean
P <sub>0</sub>	0.973	0.983	1.043	1.073	1.018
P <sub>1</sub>	0.983	1.073	1.083	1.083	1.056
P <sub>2</sub>	1.057	1.103	1.123	1.163	1.112
P <sub>3</sub>	1.093	1.123	1.173	1.193	1.146
Mean	1.027	1.071	1.106	1.128	
CD (0.05) P; 0.043	CD (0.05) S; 0.043			CD (0.05) PxS; NS	
SE (m) P; 0.015	SE (m) S; 0.015			SE (m) PxS; 0.029	

**Table 86: Effect of application of P and S on phosphorus content in shell (%)**

T<sub>1</sub>: 0.150

	S <sub>0</sub>	S <sub>1</sub>	S <sub>2</sub>	S <sub>3</sub>	Mean
P <sub>0</sub>	0.077	0.093	0.103	0.123	0.099
P <sub>1</sub>	0.093	0.123	0.143	0.163	0.131
P <sub>2</sub>	0.073	0.124	0.163	0.173	0.134
P <sub>3</sub>	0.133	0.153	0.183	0.223	0.173
Mean	0.094	0.124	0.148	0.171	
CD (0.05) P; 0.005	CD (0.05) S; 0.005			CD (0.05) PxS; 0.01	
SE (m) P; 0.002	SE (m) S; 0.002			SE (m) PxS; 0.004	

**Table 87: Effect of application of P and S on potassium content in shell (%)**

**T<sub>1</sub>: 0.548**

	<b>S<sub>0</sub></b>	<b>S<sub>1</sub></b>	<b>S<sub>2</sub></b>	<b>S<sub>3</sub></b>	<b>Mean</b>
<b>P<sub>0</sub></b>	0.453	0.479	0.517	0.769	<b>0.555</b>
<b>P<sub>1</sub></b>	0.417	0.469	0.497	0.617	<b>0.500</b>
<b>P<sub>2</sub></b>	0.417	0.465	0.490	0.565	<b>0.484</b>
<b>P<sub>3</sub></b>	0.407	0.459	0.486	0.531	<b>0.471</b>
<b>Mean</b>	<b>0.424</b>	<b>0.468</b>	<b>0.497</b>	<b>0.621</b>	
<b>CD (0.05) P; 0.006</b>		<b>CD (0.05) S; 0.006</b>		<b>CD (0.05) PxS; 0.012</b>	
<b>SE (m) P; 0.002</b>		<b>SE (m) S; 0.002</b>		<b>SE (m) PxS; 0.004</b>	

**Table 88: Effect of application of P and S on calcium content in shell (%)**

**T<sub>1</sub>: 1.91**

	<b>S<sub>0</sub></b>	<b>S<sub>1</sub></b>	<b>S<sub>2</sub></b>	<b>S<sub>3</sub></b>	<b>Mean</b>
<b>P<sub>0</sub></b>	1.52	1.78	1.78	1.97	<b>1.76</b>
<b>P<sub>1</sub></b>	1.93	2.12	2.02	2.17	<b>2.06</b>
<b>P<sub>2</sub></b>	2.10	2.02	2.09	2.21	<b>2.11</b>
<b>P<sub>3</sub></b>	2.50	2.39	2.43	2.22	<b>2.39</b>
<b>Mean</b>	<b>2.01</b>	<b>2.08</b>	<b>2.08</b>	<b>2.14</b>	
<b>CD (0.05) P; 0.012</b>		<b>CD (0.05) S; 0.012</b>		<b>CD (0.05) PxS; 0.024</b>	
<b>SE (m) P; 0.004</b>		<b>SE (m) S; 0.004</b>		<b>SE (m) PxS; 0.008</b>	

**Table 89: Effect of application of P and S on magnesium content in shell (%)**

**T<sub>1</sub>: 1.304**

	<b>S<sub>0</sub></b>	<b>S<sub>1</sub></b>	<b>S<sub>2</sub></b>	<b>S<sub>3</sub></b>	<b>Mean</b>
<b>P<sub>0</sub></b>	1.297	1.413	1.583	1.733	<b>1.507</b>
<b>P<sub>1</sub></b>	1.123	1.263	1.473	1.643	<b>1.376</b>
<b>P<sub>2</sub></b>	1.023	1.163	1.384	1.590	<b>1.290</b>
<b>P<sub>3</sub></b>	0.907	1.010	1.245	1.486	<b>1.162</b>
<b>Mean</b>	<b>1.088</b>	<b>1.213</b>	<b>1.421</b>	<b>1.613</b>	
<b>CD (0.05) P; 0.012</b>		<b>CD (0.05) S; 0.012</b>		<b>CD (0.05) PxS; 0.023</b>	
<b>SE (m) P; 0.004</b>		<b>SE (m) S; 0.004</b>		<b>SE (m) PxS; 0.008</b>	

**Table 90: Effect of application of P and S on sulphur content in shell (%)**

**T<sub>1</sub>: 0.05**

	<b>S<sub>0</sub></b>	<b>S<sub>1</sub></b>	<b>S<sub>2</sub></b>	<b>S<sub>3</sub></b>	<b>Mean</b>
<b>P<sub>0</sub></b>	0.045	0.121	0.162	0.193	<b>0.130</b>
<b>P<sub>1</sub></b>	0.034	0.140	0.135	0.156	<b>0.116</b>
<b>P<sub>2</sub></b>	0.102	0.109	0.172	0.163	<b>0.136</b>
<b>P<sub>3</sub></b>	0.102	0.108	0.172	0.200	<b>0.145</b>
<b>Mean</b>	<b>0.071</b>	<b>0.119</b>	<b>0.160</b>	<b>0.178</b>	
<b>CD (0.05) P; 0.019</b>		<b>CD (0.05) S; 0.019</b>		<b>CD (0.05) PxS; 0.037</b>	
<b>SE (m) P; 0.008</b>		<b>SE (m) S; 0.008</b>		<b>SE (m) PxS; 0.016</b>	

**Table 91: Effect of application of P and S on iron content in shell (mg kg<sup>-1</sup>)**

**T<sub>1</sub>: 124.00**

	<b>S<sub>0</sub></b>	<b>S<sub>1</sub></b>	<b>S<sub>2</sub></b>	<b>S<sub>3</sub></b>	<b>Mean</b>
<b>P<sub>0</sub></b>	176.00	227.00	253.00	274.00	<b>232.50</b>
<b>P<sub>1</sub></b>	155.00	204.00	237.00	258.00	<b>213.50</b>
<b>P<sub>2</sub></b>	137.00	184.00	221.00	234.33	<b>194.08</b>
<b>P<sub>3</sub></b>	109.33	143.00	182.00	217.00	<b>162.83</b>
<b>Mean</b>	<b>144.33</b>	<b>189.50</b>	<b>223.25</b>	<b>245.83</b>	
<b>CD (0.05) P; 0.174</b>		<b>CD (0.05) S; 0.174</b>		<b>CD (0.05) PxS; 0.348</b>	
<b>SE (m) P; 0.060</b>		<b>SE (m) S; 0.060</b>		<b>SE (m) PxS; 0.120</b>	

**Table 92: Effect of application of P and S on manganese content in shell (mg kg<sup>-1</sup>)**

**T<sub>1</sub>: 20.12**

	<b>S<sub>0</sub></b>	<b>S<sub>1</sub></b>	<b>S<sub>2</sub></b>	<b>S<sub>3</sub></b>	<b>Mean</b>
<b>P<sub>0</sub></b>	19.23	20.33	22.75	24.00	<b>19.23</b>
<b>P<sub>1</sub></b>	17.44	18.64	20.45	22.35	<b>17.44</b>
<b>P<sub>2</sub></b>	15.01	16.39	18.65	20.58	<b>15.01</b>
<b>P<sub>3</sub></b>	11.23	15.97	16.28	19.34	<b>11.23</b>
<b>Mean</b>	<b>15.73</b>	<b>17.83</b>	<b>19.53</b>	<b>21.57</b>	<b>15.73</b>
<b>CD (0.05) P; 0.021</b>		<b>CD (0.05) S; 0.021</b>		<b>CD (0.05) PxS; 0.042</b>	
<b>SE (m) P; 0.007</b>		<b>SE (m) S; 0.007</b>		<b>SE (m) PxS; 0.014</b>	





**Table 93: Effect of application of P and S on zinc content in shell (mg kg<sup>-1</sup>)**

**T<sub>1</sub>: 41.30**

	S <sub>0</sub>	S <sub>1</sub>	S <sub>2</sub>	S <sub>3</sub>	Mean
P <sub>0</sub>	38.20	41.60	42.70	45.33	<b>41.96</b>
P <sub>1</sub>	35.40	40.17	41.30	43.20	<b>40.02</b>
P <sub>2</sub>	32.53	38.20	39.20	41.70	<b>37.91</b>
P <sub>3</sub>	29.30	34.30	37.40	39.60	<b>35.15</b>
Mean	<b>33.86</b>	<b>38.57</b>	<b>40.15</b>	<b>42.46</b>	
CD (0.05) P; 0.125		CD (0.05) S; 0.125		CD (0.05) PxS; 0.250	
SE (m) P; 0.043		SE (m) S; 0.043		SE (m) PxS; 0.086	

**Table 94: Effect of application of P and S on copper content in shell (mg kg<sup>-1</sup>)**

**T<sub>1</sub>: 11.20**

	S <sub>0</sub>	S <sub>1</sub>	S <sub>2</sub>	S <sub>3</sub>	Mean
P <sub>0</sub>	11.73	11.50	12.00	11.40	<b>11.66</b>
P <sub>1</sub>	11.20	11.10	10.80	10.90	<b>11.00</b>
P <sub>2</sub>	10.80	10.70	10.90	10.60	<b>10.75</b>
P <sub>3</sub>	9.80	9.17	9.10	9.30	<b>9.34</b>
Mean	<b>10.88</b>	<b>10.62</b>	<b>10.70</b>	<b>10.55</b>	
CD (0.05) P; 0.062		CD (0.05) S; 0.062		CD (0.05) PxS; 0.123	
SE (m) P; 0.021		SE (m) S; 0.021		SE (m) PxS; 0.042	

**Table 95: Effect of application of P and S on boron content in shell (mg kg<sup>-1</sup>)**

**T<sub>1</sub>: 10.50**

	S <sub>0</sub>	S <sub>1</sub>	S <sub>2</sub>	S <sub>3</sub>	Mean
P <sub>0</sub>	15.29	15.10	15.10	15.35	<b>15.21</b>
P <sub>1</sub>	14.17	14.25	13.95	13.50	<b>13.97</b>
P <sub>2</sub>	12.15	12.30	11.85	11.50	<b>11.95</b>
P <sub>3</sub>	9.75	9.20	9.25	8.85	<b>9.27</b>
Mean	<b>12.84</b>	<b>12.71</b>	<b>12.54</b>	<b>12.30</b>	
<b>CD (0.05) P; 0.054</b>		<b>CD (0.05) S; 0.054</b>		<b>CD (0.05) PxS; 0.108</b>	
<b>SE (m) P; 0.019</b>		<b>SE (m) S; 0.019</b>		<b>SE (m) PxS; 0.037</b>	

#### 4.6 UPTAKE OF NUTRIENTS BY PLANT

##### 4.6.1 Nitrogen uptake by plant

Uptake of N by plant varied significantly due to different levels of fertilizer application (table no.s 96, 97, 98 and 99). Significantly the highest N uptake by plant was noticed in treatment, P<sub>3</sub>S<sub>3</sub> (P, 90 kg ha<sup>-1</sup> and S, 30 kg ha<sup>-1</sup>) at all stages. F statistic for treatments Vs control was calculated and was found to be significantly different.

**Table 96: Effect of application of P and S on nitrogen uptake by plant at flowering stage (kg ha<sup>-1</sup>)**

**T<sub>1</sub>: 12.11**

	S <sub>0</sub>	S <sub>1</sub>	S <sub>2</sub>	S <sub>3</sub>	Mean
P <sub>0</sub>	6.91	12.72	14.70	18.45	<b>13.19</b>
P <sub>1</sub>	7.95	12.34	18.33	21.25	<b>14.97</b>
P <sub>2</sub>	8.54	13.37	20.04	22.96	<b>16.23</b>
P <sub>3</sub>	9.76	14.96	23.67	25.05	<b>18.36</b>
Mean	<b>8.29</b>	<b>13.35</b>	<b>19.19</b>	<b>21.93</b>	
<b>CD (0.05) P; 0.244</b>		<b>CD (0.05) S; 0.244</b>		<b>CD (0.05) PxS; 0.488</b>	
<b>SE (m) P; 0.084</b>		<b>SE (m) S; 0.084</b>		<b>SE (m) PxS; 0.168</b>	

**Table 97: Effect of application of P and S on nitrogen uptake by plant at pegging stage (kg ha<sup>-1</sup>)**

**T<sub>1</sub>: 34.22**

	<b>S<sub>0</sub></b>	<b>S<sub>1</sub></b>	<b>S<sub>2</sub></b>	<b>S<sub>3</sub></b>	<b>Mean</b>
<b>P<sub>0</sub></b>	27.12	31.81	34.40	37.42	<b>32.69</b>
<b>P<sub>1</sub></b>	30.38	35.05	36.65	39.76	<b>35.46</b>
<b>P<sub>2</sub></b>	33.25	38.82	39.58	45.97	<b>39.40</b>
<b>P<sub>3</sub></b>	38.15	40.70	48.45	52.29	<b>44.90</b>
<b>Mean</b>	<b>32.23</b>	<b>36.60</b>	<b>39.77</b>	<b>43.86</b>	
<b>CD (0.05) P; 0.842</b>		<b>CD (0.05) S; 0.842</b>		<b>CD (0.05) PxS; 1.685</b>	
<b>SE (m) P; 0.29</b>		<b>SE (m) S; 0.29</b>		<b>SE (m) PxS; 0.581</b>	

**Table 98: Effect of application of P and S on nitrogen uptake by plant at pod setting stage (kg ha<sup>-1</sup>)**

**T<sub>1</sub>: 41.34**

	<b>S<sub>0</sub></b>	<b>S<sub>1</sub></b>	<b>S<sub>2</sub></b>	<b>S<sub>3</sub></b>	<b>Mean</b>
<b>P<sub>0</sub></b>	38.85	40.33	45.43	51.36	<b>43.99</b>
<b>P<sub>1</sub></b>	41.97	45.20	50.55	56.98	<b>48.68</b>
<b>P<sub>2</sub></b>	45.54	48.94	52.55	61.70	<b>52.18</b>
<b>P<sub>3</sub></b>	50.38	56.00	68.00	77.19	<b>62.89</b>
<b>Mean</b>	<b>44.18</b>	<b>47.62</b>	<b>54.13</b>	<b>61.81</b>	
<b>CD (0.05) P; 0.384</b>		<b>CD (0.05) S; 0.384</b>		<b>CD (0.05) PxS; 0.769</b>	
<b>SE (m) P; 0.132</b>		<b>SE (m) S; 0.132</b>		<b>SE (m) PxS; 0.265</b>	

**Table 99: Effect of application of P and S on nitrogen uptake by plant at harvest stage (kg ha<sup>-1</sup>)**

**T<sub>1</sub>: 46.33**

	S <sub>0</sub>	S <sub>1</sub>	S <sub>2</sub>	S <sub>3</sub>	Mean
P <sub>0</sub>	44.46	48.34	53.82	61.02	51.91
P <sub>1</sub>	48.14	55.48	62.20	65.87	57.92
P <sub>2</sub>	56.62	63.51	68.13	73.23	65.37
P <sub>3</sub>	63.67	68.34	74.98	78.39	71.35
Mean	53.22	58.92	64.78	69.63	
CD (0.05) P; 0.934		CD (0.05) S; 0.934		CD (0.05) PxS; 1.868	
SE (m) P; 0.322		SE (m) S; 0.322		SE (m) PxS; 0.644	

#### 4.6.2 Phosphorus uptake by plant

Uptake of P by plant varied significantly due to different levels of fertilizer application (table no.s 100, 101, 102 and 103). Significantly highest P uptake by plant was noticed in treatment, P<sub>3</sub>S<sub>3</sub> (P, 90 kg ha<sup>-1</sup> and S, 30 kg ha<sup>-1</sup>) at all stages. F statistic for treatments Vs control was calculated and was found to be significantly different.

**Table 100: Effect of application of P and S on phosphorus uptake by plant at flowering stage (kg ha<sup>-1</sup>)**

**T<sub>1</sub>: 1.72**

	S <sub>0</sub>	S <sub>1</sub>	S <sub>2</sub>	S <sub>3</sub>	Mean
P <sub>0</sub>	0.73	1.56	2.05	2.58	1.73
P <sub>1</sub>	0.90	1.61	2.58	3.14	2.06
P <sub>2</sub>	1.04	1.88	2.92	3.57	2.35
P <sub>3</sub>	1.36	2.26	3.59	3.87	2.77
Mean	1.01	1.83	2.78	3.29	
CD (0.05) P; 0.011		CD (0.05) S; 0.011		CD (0.05) PxS; 0.022	
SE (m) P; 0.004		SE (m) S; 0.004		SE (m) PxS; 0.007	

**Table 101: Effect of application of P and S on phosphorus uptake by plant at pegging stage (kg ha<sup>-1</sup>)**

**T<sub>1</sub>: 5.23**

	<b>S<sub>0</sub></b>	<b>S<sub>1</sub></b>	<b>S<sub>2</sub></b>	<b>S<sub>3</sub></b>	<b>Mean</b>
<b>P<sub>0</sub></b>	3.18	3.66	4.76	5.47	<b>4.27</b>
<b>P<sub>1</sub></b>	3.69	4.15	5.41	6.34	<b>4.90</b>
<b>P<sub>2</sub></b>	4.07	5.14	6.15	7.23	<b>5.65</b>
<b>P<sub>3</sub></b>	5.33	5.73	7.35	8.09	<b>6.62</b>
<b>Mean</b>	<b>4.07</b>	<b>4.67</b>	<b>5.92</b>	<b>6.78</b>	
<b>CD (0.05) P; 0.048</b>		<b>CD (0.05) S; 0.048</b>		<b>CD (0.05) PxS; 0.097</b>	
<b>SE (m) P; 0.017</b>		<b>SE (m) S; 0.017</b>		<b>SE (m) PxS; 0.033</b>	

**Table 102: Effect of application of P and S on phosphorus uptake by plant at pod setting stage (kg ha<sup>-1</sup>)**

**T<sub>1</sub>: 7.34**

	<b>S<sub>0</sub></b>	<b>S<sub>1</sub></b>	<b>S<sub>2</sub></b>	<b>S<sub>3</sub></b>	<b>Mean</b>
<b>P<sub>0</sub></b>	3.66	4.15	5.69	6.23	<b>4.93</b>
<b>P<sub>1</sub></b>	4.33	5.62	6.86	7.48	<b>6.07</b>
<b>P<sub>2</sub></b>	4.89	6.40	7.18	8.96	<b>6.86</b>
<b>P<sub>3</sub></b>	6.10	7.28	9.79	11.29	<b>8.62</b>
<b>Mean</b>	<b>4.75</b>	<b>5.86</b>	<b>7.38</b>	<b>8.49</b>	
<b>CD (0.05) P; 0.027</b>		<b>CD (0.05) S; 0.027</b>		<b>CD (0.05) PxS; 0.053</b>	
<b>SE (m) P; 0.009</b>		<b>SE (m) S; 0.009</b>		<b>SE (m) PxS; 0.018</b>	

**Table 103: Effect of application of P and S on phosphorus uptake by plant at harvest stage (kg ha<sup>-1</sup>)**

**T<sub>1</sub>: 7.81**

	S <sub>0</sub>	S <sub>1</sub>	S <sub>2</sub>	S <sub>3</sub>	Mean
P <sub>0</sub>	3.26	5.05	6.61	8.21	<b>5.78</b>
P <sub>1</sub>	4.06	6.02	8.65	9.49	<b>7.05</b>
P <sub>2</sub>	5.56	7.60	8.50	10.76	<b>8.10</b>
P <sub>3</sub>	6.31	8.66	10.27	11.74	<b>9.24</b>
<b>Mean</b>	<b>4.79</b>	<b>6.83</b>	<b>8.51</b>	<b>10.05</b>	
<b>CD (0.05) P; 0.034</b>		<b>CD (0.05) S; 0.034</b>		<b>CD (0.05) PxS; 0.069</b>	
<b>SE (m) P; 0.012</b>		<b>SE (m) S; 0.012</b>		<b>SE (m) PxS; 0.024</b>	

#### 4.6.3 Potassium uptake by plant

Potassium uptake by plant was found to be significant in all treatments due to main effect and interaction effect. The treatment, P<sub>3</sub>S<sub>3</sub> (P, 90 kg ha<sup>-1</sup> and S, 30 kg ha<sup>-1</sup>) showed highest K uptake at flowering stage (20.17 kg ha<sup>-1</sup>) and pod formation stage (64.48 kg ha<sup>-1</sup>). At pegging stage P<sub>3</sub>S<sub>2</sub> (P, 90 kg ha<sup>-1</sup> and S, 20 kg ha<sup>-1</sup>) showed highest K uptake (21.93 kg ha<sup>-1</sup>). At harvesting stage P<sub>2</sub>S<sub>3</sub> (P, 75 kg ha<sup>-1</sup> and S, 30 kg ha<sup>-1</sup>) showed highest K uptake (79.40 kg ha<sup>-1</sup>). F statistic for treatments Vs control was calculated and was found to be significantly different.

**Table 104: Effect of application of P and S on potassium uptake by plant at flowering stage (kg ha<sup>-1</sup>)**

**T<sub>1</sub>: 12.5**

	S <sub>0</sub>	S <sub>1</sub>	S <sub>2</sub>	S <sub>3</sub>	Mean
P <sub>0</sub>	4.38	9.60	11.93	17.91	<b>10.96</b>
P <sub>1</sub>	4.77	9.17	14.27	18.69	<b>11.72</b>
P <sub>2</sub>	4.80	9.18	14.41	19.19	<b>11.89</b>
P <sub>3</sub>	5.44	9.62	17.94	20.17	<b>13.29</b>
Mean	<b>4.85</b>	<b>9.39</b>	<b>14.64</b>	<b>18.99</b>	
CD (0.05) P; 0.125		CD (0.05) S; 0.125		CD (0.05) P x S; 0.25	
SE (m) P; 0.043		SE (m) S; 0.043		SE (m) P x S; 0.086	

**Table 105: Effect of application of P and S on potassium uptake by plant at pegging stage (kg ha<sup>-1</sup>)**

**T<sub>1</sub>: 33.18**

	S <sub>0</sub>	S <sub>1</sub>	S <sub>2</sub>	S <sub>3</sub>	Mean
P <sub>0</sub>	13.64	15.65	16.36	17.30	<b>15.74</b>
P <sub>1</sub>	15.36	17.19	17.49	18.32	<b>17.09</b>
P <sub>2</sub>	16.35	18.97	18.78	20.58	<b>18.67</b>
P <sub>3</sub>	19.18	19.68	21.93	21.86	<b>20.66</b>
Mean	<b>16.13</b>	<b>17.87</b>	<b>18.64</b>	<b>19.52</b>	
CD (0.05) P; 0.001		CD (0.05) S; 0.001		CD (0.05) P x S; 0.003	
SE (m) P; NS		SE (m) S; NS		SE (m) P x S; 0.001	

**Table 106: Effect of application of P and S on potassium uptake by plant at pod setting stage (kg ha<sup>-1</sup>)**

**T<sub>1</sub>: 41.1**

	<b>S<sub>0</sub></b>	<b>S<sub>1</sub></b>	<b>S<sub>2</sub></b>	<b>S<sub>3</sub></b>	<b>Mean</b>
<b>P<sub>0</sub></b>	29.90	37.15	45.37	57.23	<b>42.41</b>
<b>P<sub>1</sub></b>	28.84	36.55	47.56	61.11	<b>43.52</b>
<b>P<sub>2</sub></b>	29.61	38.93	45.01	62.08	<b>43.91</b>
<b>P<sub>3</sub></b>	31.34	40.66	56.94	64.48	<b>48.35</b>
<b>Mean</b>	<b>29.92</b>	<b>38.32</b>	<b>48.72</b>	<b>61.23</b>	
<b>CD (0.05) P; 0.025</b>		<b>CD (0.05) S; 0.025</b>		<b>CD (0.05) PxS; 0.049</b>	
<b>SE (m) P; 0.009</b>		<b>SE (m) S; 0.009</b>		<b>SE (m) PxS; 0.017</b>	

**Table 107: Effect of application of P and S on potassium uptake by plant at harvest stage (kg ha<sup>-1</sup>)**

**T<sub>1</sub>: 45.52**

	<b>S<sub>0</sub></b>	<b>S<sub>1</sub></b>	<b>S<sub>2</sub></b>	<b>S<sub>3</sub></b>	<b>Mean</b>
<b>P<sub>0</sub></b>	38.03	50.23	59.86	77.36	<b>56.37</b>
<b>P<sub>1</sub></b>	37.44	49.21	64.39	78.77	<b>57.45</b>
<b>P<sub>2</sub></b>	36.27	52.76	60.29	79.40	<b>57.18</b>
<b>P<sub>3</sub></b>	40.39	52.72	63.14	76.08	<b>58.08</b>
<b>Mean</b>	<b>38.03</b>	<b>51.23</b>	<b>61.92</b>	<b>77.90</b>	
<b>CD (0.05) P; 0.297</b>		<b>CD (0.05) S; 0.297</b>		<b>CD (0.05) PxS; 0.595</b>	
<b>SE (m) P; 0.102</b>		<b>SE (m) S; 0.102</b>		<b>SE (m) PxS; 0.205</b>	

#### 4.6.4 Calcium uptake by plant

Calcium uptake by plant varied significantly due to main effect and



interaction effect. Highest Ca uptake was noted in P<sub>3</sub>S<sub>3</sub> (P, 90 kg ha<sup>-1</sup> and S, 30 kg ha<sup>-1</sup>) during flowering, pegging, and pod formation stages. Whereas at harvesting stage highest Ca uptake was noted in P<sub>1</sub>S<sub>3</sub> (P, 60 kg ha<sup>-1</sup> and S, 30 kg ha<sup>-1</sup>). Ca uptake increased with plant growth and there was a slight decrease at harvest stage. F statistic for treatments Vs control was calculated and was found to be significantly different.

**Table 108: Effect of application of P and S on calcium uptake by plant at flowering stage (kg ha<sup>-1</sup>)**

T<sub>1</sub>: 10.03

	S <sub>0</sub>	S <sub>1</sub>	S <sub>2</sub>	S <sub>3</sub>	Mean
P <sub>0</sub>	5.31	9.88	12.26	15.94	10.85
P <sub>1</sub>	5.81	9.03	14.83	18.16	11.96
P <sub>2</sub>	5.98	9.29	14.84	18.49	12.15
P <sub>3</sub>	6.60	10.52	17.43	19.27	13.45
Mean	5.93	9.68	14.84	17.96	
CD (0.05) P; 0.029	CD (0.05) S; 0.029			CD (0.05) PxS; 0.058	
SE (m) P; 0.01	SE (m) S; 0.01			SE (m) PxS; 0.02	

**Table 109: Effect of application of P and S on calcium uptake by plant at pegging stage (kg ha<sup>-1</sup>)**

T<sub>1</sub>: 24.74

	S <sub>0</sub>	S <sub>1</sub>	S <sub>2</sub>	S <sub>3</sub>	Mean
P <sub>0</sub>	22.50	25.94	28.87	31.28	27.15
P <sub>1</sub>	24.13	27.86	29.55	32.12	28.42
P <sub>2</sub>	24.75	29.84	31.31	34.84	30.19
P <sub>3</sub>	25.06	29.32	35.16	35.77	31.33
Mean	24.11	28.24	31.23	33.51	
CD (0.05) P; 0.015	CD (0.05) S; 0.015			CD (0.05) PxS; 0.03	
SE (m) P; 0.005	SE (m) S; 0.005			SE (m) PxS; 0.01	

**Table 110: Effect of application of P and S on calcium uptake by plant at pod setting stage (kg ha<sup>-1</sup>)**

T<sub>1</sub>: 27.67

	S <sub>0</sub>	S <sub>1</sub>	S <sub>2</sub>	S <sub>3</sub>	Mean
P <sub>0</sub>	19.48	24.00	29.22	34.28	26.74
P <sub>1</sub>	19.95	23.00	31.04	36.79	27.70
P <sub>2</sub>	20.34	23.64	29.82	37.18	27.75
P <sub>3</sub>	19.13	23.36	32.64	38.62	28.44
Mean	19.73	23.50	30.68	36.72	
CD (0.05) P; 0.016		CD (0.05) S; 0.016		CD (0.05) PxS; 0.031	
SE (m) P; 0.005		SE (m) S; 0.005		SE (m) PxS; 0.011	

**Table 111: Effect of application of P and S on calcium uptake by plant at harvest stage (kg ha<sup>-1</sup>)**

T<sub>1</sub>: 24.79

	S <sub>0</sub>	S <sub>1</sub>	S <sub>2</sub>	S <sub>3</sub>	Mean
P <sub>0</sub>	23.08	26.21	32.18	39.88	30.34
P <sub>1</sub>	23.82	27.97	34.98	40.42	31.80
P <sub>2</sub>	23.83	28.11	32.77	37.34	30.52
P <sub>3</sub>	23.37	27.15	30.72	36.92	29.54
Mean	23.53	27.36	32.66	38.64	
CD (0.05) P; 0.015		CD (0.05) S; 0.015		CD (0.05) PxS; 0.030	
SE (m) P; 0.005		SE (m) S; 0.005		SE (m) PxS; 0.010	

#### 4.6.5 Magnesium uptake by plant

Application of P and S fertilizers significantly affected Mg uptake by plant. The treatment, P<sub>3</sub>S<sub>3</sub> (P, 90 kg ha<sup>-1</sup> and S, 30 kg ha<sup>-1</sup>) had highest Mg uptake during

flowering, pegging, pod formation and harvesting stages. The main effect and interaction effect was found to be significant at all stages. F statistic for treatments Vs control was calculated and was found to be significantly different.

**Table 112: Effect of application of P and S on magnesium uptake by plant at flowering stage (kg ha<sup>-1</sup>)**

**T<sub>1</sub>: 16.69**

	<b>S<sub>0</sub></b>	<b>S<sub>1</sub></b>	<b>S<sub>2</sub></b>	<b>S<sub>3</sub></b>	<b>Mean</b>
<b>P<sub>0</sub></b>	7.79	15.03	17.67	22.54	<b>15.76</b>
<b>P<sub>1</sub></b>	9.17	14.85	22.33	26.31	<b>18.16</b>
<b>P<sub>2</sub></b>	10.04	16.05	24.28	28.88	<b>19.81</b>
<b>P<sub>3</sub></b>	11.66	18.38	29.62	31.20	<b>22.72</b>
<b>Mean</b>	<b>9.66</b>	<b>16.08</b>	<b>23.48</b>	<b>27.23</b>	
<b>CD (0.05) P; 0.005</b>	<b>CD (0.05) S; 0.005</b>		<b>CD (0.05) PxS; 0.009</b>		
<b>SE (m) P; 0.002</b>	<b>SE (m) S; 0.002</b>		<b>SE (m) PxS; 0.003</b>		

**Table 113: Effect of application of P and S on magnesium uptake by plant at pegging stage (kg ha<sup>-1</sup>)**

**T<sub>1</sub>: 33.26**

	<b>S<sub>0</sub></b>	<b>S<sub>1</sub></b>	<b>S<sub>2</sub></b>	<b>S<sub>3</sub></b>	<b>Mean</b>
<b>P<sub>0</sub></b>	24.87	29.22	31.60	34.78	<b>30.12</b>
<b>P<sub>1</sub></b>	27.96	32.35	34.73	38.42	<b>33.37</b>
<b>P<sub>2</sub></b>	31.06	36.79	38.31	43.39	<b>37.39</b>
<b>P<sub>3</sub></b>	36.87	39.44	45.08	47.14	<b>42.13</b>
<b>Mean</b>	<b>30.19</b>	<b>34.45</b>	<b>37.43</b>	<b>40.93</b>	
<b>CD (0.05) P; 0.263</b>	<b>CD (0.05) S; 0.263</b>		<b>CD (0.05) PxS; 0.526</b>		
<b>SE (m) P; 0.091</b>	<b>SE (m) S; 0.091</b>		<b>SE (m) PxS; 0.181</b>		



Table 114: Effect of application of P and S on magnesium uptake by plant at pod setting stage ( $\text{kg ha}^{-1}$ )

$T_1$ : 24.74

	$S_0$	$S_1$	$S_2$	$S_3$	Mean
$P_0$	15.86	16.61	21.27	24.65	19.60
$P_1$	16.58	20.19	25.06	28.06	22.47
$P_2$	19.83	23.36	27.77	34.20	26.29
$P_3$	22.46	28.22	38.08	39.34	32.03
Mean	18.68	22.10	28.05	31.56	
CD (0.05) P; 0.008		CD (0.05) S; 0.008		CD (0.05) PxS; 0.016	
SE (m) P; 0.003		SE (m) S; 0.003		SE (m) PxS; 0.006	

Table 115: Effect of application of P and S on magnesium uptake by plant at harvest stage ( $\text{kg ha}^{-1}$ )

$T_1$ : 33.71

	$S_0$	$S_1$	$S_2$	$S_3$	Mean
$P_0$	19.54	22.11	27.33	33.24	25.55
$P_1$	20.42	27.59	33.80	36.70	29.63
$P_2$	26.94	31.53	38.03	43.68	35.05
$P_3$	29.67	36.13	41.81	47.60	38.80
Mean	24.14	29.34	35.24	40.31	
CD (0.05) P; 0.263		CD (0.05) S; 0.263		CD (0.05) PxS; 0.526	
SE (m) P; 0.091		SE (m) S; 0.091		SE (m) PxS; 0.181	

#### 4.6.6 Sulphur uptake by plant

Application of P and S fertilizers significantly affected S uptake by plant. The treatment,  $P_0S_3$  (P,  $0 \text{ kg ha}^{-1}$  and S,  $30 \text{ kg ha}^{-1}$ ) had highest S uptake during flowering

stage. At pegging, pod setting and harvesting stages, highest S uptake was noted in P<sub>3</sub>S<sub>3</sub> (P, 90 kg ha<sup>-1</sup> and S, 30 kg ha<sup>-1</sup>). The main effect and interaction effect was found to be significant at all stages. F statistic for treatments Vs control was calculated and was found to be significantly different.

**Table 116: Effect of application of P and S on sulphur uptake by plant at flowering stage (kg ha<sup>-1</sup>)**

**T<sub>1</sub>: 0.724**

	S <sub>0</sub>	S <sub>1</sub>	S <sub>2</sub>	S <sub>3</sub>	Mean
P <sub>0</sub>	0.597	1.267	2.088	5.152	2.276
P <sub>1</sub>	0.686	1.219	2.176	3.047	1.782
P <sub>2</sub>	0.463	1.352	2.059	2.868	1.686
P <sub>3</sub>	0.619	1.399	2.565	3.560	2.036
Mean	0.591	1.309	2.222	3.657	
CD (0.05) P; 0.002	CD (0.05) S; 0.002		CD (0.05) PxS; 0.004		
SE (m) P; 0.001	SE (m) S; 0.001		SE (m) PxS; 0.001		

**Table 117: Effect of application of P and S on sulphur uptake by plant at pegging stage (kg ha<sup>-1</sup>)**

**T<sub>1</sub>: 2.13**

	S <sub>0</sub>	S <sub>1</sub>	S <sub>2</sub>	S <sub>3</sub>	Mean
P <sub>0</sub>	1.914	2.906	3.325	6.124	3.567
P <sub>1</sub>	1.963	2.666	3.052	3.915	2.899
P <sub>2</sub>	1.499	2.903	3.073	3.875	2.838
P <sub>3</sub>	1.955	2.513	4.042	6.178	3.672
Mean	1.833	2.747	3.373	5.023	
CD (0.05) P; 0.045	CD (0.05) S; 0.045		CD (0.05) PxS; 0.09		
SE (m) P; 0.015	SE (m) S; 0.015		SE (m) PxS; 0.031		

**Table 118: Effect of application of P and S on sulphur uptake by plant at pod setting stage (kg ha<sup>-1</sup>)**

**T<sub>1</sub>: 3.140**

	<b>S<sub>0</sub></b>	<b>S<sub>1</sub></b>	<b>S<sub>2</sub></b>	<b>S<sub>3</sub></b>	<b>Mean</b>
<b>P<sub>0</sub></b>	2.092	3.261	4.137	6.676	<b>4.042</b>
<b>P<sub>1</sub></b>	1.560	1.696	3.160	4.275	<b>2.673</b>
<b>P<sub>2</sub></b>	2.265	3.242	3.810	4.249	<b>3.391</b>
<b>P<sub>3</sub></b>	2.335	2.552	5.323	6.862	<b>4.268</b>
<b>Mean</b>	<b>2.063</b>	<b>2.688</b>	<b>4.108</b>	<b>5.515</b>	
<b>CD (0.05) P; 0.274</b>		<b>CD (0.05) S; 0.274</b>		<b>CD (0.05) PxS; 0.547</b>	
<b>SE (m) P; 0.094</b>		<b>SE (m) S; 0.094</b>		<b>SE (m) PxS; 0.189</b>	

**Table 119: Effect of application of P and S on sulphur uptake by plant at harvest stage (kg ha<sup>-1</sup>)**

**T<sub>1</sub>: 3.04**

	<b>S<sub>0</sub></b>	<b>S<sub>1</sub></b>	<b>S<sub>2</sub></b>	<b>S<sub>3</sub></b>	<b>Mean</b>
<b>P<sub>0</sub></b>	0.54	0.58	0.96	3.64	<b>1.43</b>
<b>P<sub>1</sub></b>	0.48	0.66	0.73	1.07	<b>0.73</b>
<b>P<sub>2</sub></b>	0.76	1.05	1.25	2.50	<b>1.39</b>
<b>P<sub>3</sub></b>	1.60	2.05	4.08	5.85	<b>3.39</b>
<b>Mean</b>	<b>0.84</b>	<b>1.09</b>	<b>1.75</b>	<b>3.27</b>	
<b>CD (0.05) P; 0.009</b>		<b>CD (0.05) S; 0.009</b>		<b>CD (0.05) PxS; 0.018</b>	
<b>SE (m) P; 0.003</b>		<b>SE (m) S; 0.003</b>		<b>SE (m) PxS; 0.006</b>	

#### **4.6.7 Iron uptake by plant**

Uptake of Fe by plant at different stages is shown in the tables 120, 121, 122 and 123. Uptake of Fe varied significantly due to interaction effect and main effect.

P<sub>3</sub>S<sub>3</sub> (P, 90 kg ha<sup>-1</sup> and S, 30 kg ha<sup>-1</sup>) showed highest Fe uptake in all stages. F statistic for treatments Vs control was calculated and was found to be significantly different.

**Table 120: Effect of application of P and S on iron uptake by plant at flowering stage (g ha<sup>-1</sup>)**

T<sub>1</sub>: 310.02

	S <sub>0</sub>	S <sub>1</sub>	S <sub>2</sub>	S <sub>3</sub>	Mean
P <sub>0</sub>	201.67	317.33	372.50	511.69	<b>350.80</b>
P <sub>1</sub>	190.17	364.92	423.69	506.45	<b>371.31</b>
P <sub>2</sub>	216.24	383.52	454.05	533.50	<b>396.83</b>
P <sub>3</sub>	190.89	382.89	529.04	617.60	<b>430.10</b>
Mean	<b>199.74</b>	<b>362.16</b>	<b>444.82</b>	<b>542.31</b>	
CD (0.05) P; 0.181		CD (0.05) S; 0.181		CD (0.05) PxS; 0.362	
SE (m) P; 0.062		SE (m) S; 0.062		SE (m) PxS; 0.125	

**Table 121: Effect of application of P and S on iron uptake by plant at pegging stage (g ha<sup>-1</sup>)**

T<sub>1</sub>: 425.12

	S <sub>0</sub>	S <sub>1</sub>	S <sub>2</sub>	S <sub>3</sub>	Mean
P <sub>0</sub>	219.69	339.57	377.30	497.87	<b>358.61</b>
P <sub>1</sub>	171.65	385.18	439.09	529.14	<b>381.27</b>
P <sub>2</sub>	235.26	391.28	482.35	570.70	<b>419.90</b>
P <sub>3</sub>	209.08	412.10	552.82	667.13	<b>460.28</b>
Mean	<b>208.92</b>	<b>382.03</b>	<b>462.89</b>	<b>566.21</b>	
CD (0.05) P; 0.294		CD (0.05) S; 0.294		CD (0.05) PxS; 0.587	
SE (m) P; 0.101		SE (m) S; 0.101		SE (m) PxS; 0.202	

**Table 122: Effect of application of P and S on iron uptake by plant at pod setting stage ( $\text{g ha}^{-1}$ )**

**T<sub>1</sub>: 547.32**

	S <sub>0</sub>	S <sub>1</sub>	S <sub>2</sub>	S <sub>3</sub>	Mean
P <sub>0</sub>	229.17	360.35	401.38	532.97	<b>380.97</b>
P <sub>1</sub>	186.61	407.85	454.39	555.28	<b>401.03</b>
P <sub>2</sub>	227.88	427.12	475.25	582.46	<b>428.18</b>
P <sub>3</sub>	230.76	445.27	547.21	655.85	<b>469.77</b>
Mean	<b>218.60</b>	<b>410.15</b>	<b>469.56</b>	<b>581.64</b>	
CD (0.05) P; 0.227		CD (0.05) S; 0.227		CD (0.05) PxS; 0.454	
SE (m) P; 0.078		SE (m) S; 0.078		SE (m) PxS; 0.157	

**Table 123: Effect of application of P and S on iron uptake by plant at harvest stage ( $\text{g ha}^{-1}$ )**

**T<sub>1</sub>: 697.30**

	S <sub>0</sub>	S <sub>1</sub>	S <sub>2</sub>	S <sub>3</sub>	Mean
P <sub>0</sub>	264.03	424.87	477.22	636.67	<b>450.70</b>
P <sub>1</sub>	228.67	480.73	531.50	674.22	<b>478.78</b>
P <sub>2</sub>	276.30	519.18	576.27	708.40	<b>520.04</b>
P <sub>3</sub>	281.62	544.43	649.64	794.35	<b>567.51</b>
Mean	<b>262.65</b>	<b>492.30</b>	<b>558.66</b>	<b>703.41</b>	
CD (0.05) P; 0.233		CD (0.05) S; 0.233		CD (0.05) PxS; 0.465	
SE (m) P; 0.08		SE (m) S; 0.08		SE (m) PxS; 0.16	

#### 4.6.8 Manganese uptake by plants

Manganese uptake was significantly influenced by main effect and interaction effect. The treatment, P<sub>3</sub>S<sub>3</sub> (P, 90 kg ha<sup>-1</sup> and S, 30 kg ha<sup>-1</sup>) showed highest Mn



uptake in all stages. F statistic for treatments Vs control was calculated and was found to be significantly different.

**Table 124: Effect of application of P and S on manganese uptake by plant at flowering stage ( $\text{g ha}^{-1}$ )**

**T<sub>1</sub>: 26.33**

	S <sub>0</sub>	S <sub>1</sub>	S <sub>2</sub>	S <sub>3</sub>	Mean
P <sub>0</sub>	16.60	27.08	38.99	56.00	34.67
P <sub>1</sub>	19.44	29.50	43.52	58.40	37.71
P <sub>2</sub>	22.83	31.28	46.15	59.20	39.87
P <sub>3</sub>	22.74	32.85	43.99	61.99	40.39
Mean	20.40	30.18	43.16	58.90	
CD (0.05) P; 0.021	CD (0.05) S; 0.021		CD (0.05) PxS; 0.042		
SE (m) P; 0.007	SE (m) S; 0.007		SE (m) PxS; 0.014		

**Table 125: Effect of application of P and S on manganese uptake by plant at pegging stage ( $\text{g ha}^{-1}$ )**

**T<sub>1</sub>: 27.15**

	S <sub>0</sub>	S <sub>1</sub>	S <sub>2</sub>	S <sub>3</sub>	Mean
P <sub>0</sub>	17.16	28.80	41.41	60.05	36.86
P <sub>1</sub>	20.70	31.35	46.57	62.81	40.36
P <sub>2</sub>	22.81	32.69	49.28	63.74	42.13
P <sub>3</sub>	24.62	34.69	45.47	65.67	42.61
Mean	21.323	31.882	45.682	63.066	
CD (0.05) P; 0.023	CD (0.05) S; 0.023		CD (0.05) PxS; 0.046		
SE (m) P; 0.008	SE (m) S; 0.008		SE (m) PxS; 0.016		

**Table 126: Effect of application of P and S on manganese uptake by plant at pod setting stage (g ha<sup>-1</sup>)**

**T<sub>1</sub>: 29.98**

	S <sub>0</sub>	S <sub>1</sub>	S <sub>2</sub>	S <sub>3</sub>	Mean
P <sub>0</sub>	18.29	30.17	44.60	64.69	39.44
P <sub>1</sub>	21.90	33.34	49.72	66.86	42.95
P <sub>2</sub>	23.56	33.08	52.23	67.58	44.11
P <sub>3</sub>	25.77	35.95	47.91	68.85	44.62
Mean	22.38	33.14	48.61	66.99	
CD (0.05) P; 0.216		CD (0.05) S; 0.216		CD (0.05) PxS; 0.432	
SE (m) P; 0.074		SE (m) S; 0.074		SE (m) PxS; 0.0149	

**Table 127: Effect of application of P and S on manganese uptake by plant at harvest stage (g ha<sup>-1</sup>)**

**T<sub>1</sub>: 34.56**

	S <sub>0</sub>	S <sub>1</sub>	S <sub>2</sub>	S <sub>3</sub>	Mean
P <sub>0</sub>	21.51	35.49	53.18	77.66	46.96
P <sub>1</sub>	25.85	39.03	59.81	80.35	51.26
P <sub>2</sub>	27.35	39.88	61.34	78.03	51.65
P <sub>3</sub>	30.52	41.83	56.40	81.50	52.57
Mean	26.31	39.06	57.68	79.39	
CD (0.05) P; 0.172		CD (0.05) S; 0.172		CD (0.05) PxS; 0.344	
SE (m) P; 0.059		SE (m) S; 0.059		SE (m) PxS; 0.118	

#### 4.6.9 Zinc uptake by plants

Zinc uptake varied significantly due to application of P and S fertilizers. The treatment P<sub>3</sub>S<sub>3</sub> (P, 90 kg ha<sup>-1</sup> and S, 30 kg ha<sup>-1</sup>) showed highest zinc uptake at

flowering and pod formation stages. Whereas at pegging and harvest stages, P<sub>3</sub>S<sub>2</sub> (P, 90 kg ha<sup>-1</sup> and S, 20 kg ha<sup>-1</sup>) showed highest Zn uptake. F statistic for treatments Vs control was calculated and was found to be significantly different.

**Table 128: Effect of application of P and S on zinc uptake by plant at flowering stage (g ha<sup>-1</sup>)**

T<sub>1</sub>: 27.64

	S <sub>0</sub>	S <sub>1</sub>	S <sub>2</sub>	S <sub>3</sub>	Mean
P <sub>0</sub>	21.27	26.73	28.50	32.06	27.14
P <sub>1</sub>	25.30	30.15	31.68	33.54	30.17
P <sub>2</sub>	30.60	34.07	33.87	34.93	33.37
P <sub>3</sub>	31.63	36.43	38.60	39.67	36.58
Mean	27.20	31.84	33.16	35.05	
CD (0.05) P; 0.173		CD (0.05) S; 0.173		CD (0.05) PxS; 0.346	
SE (m) P; 0.06		SE (m) S; 0.06		SE (m) PxS; 0.119	

**Table 129: Effect of application of P and S on zinc uptake by plant at pegging stage (g ha<sup>-1</sup>)**

T<sub>1</sub>: 29.23

	S <sub>0</sub>	S <sub>1</sub>	S <sub>2</sub>	S <sub>3</sub>	Mean
P <sub>0</sub>	23.00	28.95	30.05	33.16	28.79
P <sub>1</sub>	27.30	32.71	33.30	35.54	32.21
P <sub>2</sub>	32.58	36.76	35.54	36.06	35.23
P <sub>3</sub>	33.82	37.90	41.76	41.68	38.69
Mean	29.17	34.08	35.06	36.61	
CD (0.05) P; 0.121		CD (0.05) S; 0.121		CD (0.05) PxS; 0.243	
SE (m) P; 0.042		SE (m) S; 0.042		SE (m) PxS; 0.084	

**Table 130: Effect of application of P and S on zinc uptake by plant at pod setting stage (g ha<sup>-1</sup>)**

**T<sub>1</sub> : 29.56**

	S <sub>0</sub>	S <sub>1</sub>	S <sub>2</sub>	S <sub>3</sub>	Mean
P <sub>0</sub>	25.22	30.05	32.18	35.47	30.73
P <sub>1</sub>	27.94	33.70	35.62	37.27	33.63
P <sub>2</sub>	33.70	37.93	38.40	38.95	37.25
P <sub>3</sub>	35.37	40.01	44.29	44.63	41.08
Mean	30.56	35.42	37.62	39.08	
CD (0.05) P; 0.026		CD (0.05) S; 0.026		CD (0.05) PxS; 0.052	
SE (m) P; 0.009		SE (m) S; 0.009		SE (m) PxS; 0.018	

**Table 131: Effect of application of P and S on zinc uptake by plant at harvest stage (g ha<sup>-1</sup>)**

**T<sub>1</sub> : 34.56**

	S <sub>0</sub>	S <sub>1</sub>	S <sub>2</sub>	S <sub>3</sub>	Mean
P <sub>0</sub>	29.92	36.26	38.88	42.62	36.92
P <sub>1</sub>	33.48	40.18	41.38	43.75	39.70
P <sub>2</sub>	40.42	45.29	45.37	46.18	44.31
P <sub>3</sub>	41.86	47.20	52.92	52.79	48.44
Mean	36.42	42.23	44.39	46.34	
CD (0.05) P; 0.178		CD (0.05) S; 0.178		CD (0.05) PxS; 0.355	
SE (m) P; 0.061		SE (m) S; 0.061		SE (m) PxS; 0.122	

#### 4.6.10 Copper uptake by groundnut plant

Uptake of Cu was significant at all the stages. The treatment, P<sub>3</sub>S<sub>3</sub> (P, 90 kg ha<sup>-1</sup> and S, 30 kg ha<sup>-1</sup>) showed highest uptake at flowering, pegging and pod setting stages. Highest uptake during harvest stage was noted in P<sub>3</sub>S<sub>2</sub> (P, 90 kg ha<sup>-1</sup>

and S, 20 kg ha<sup>-1</sup>). F statistic for treatments Vs control was calculated and was found to be significantly different.

**Table 132: Effect of application of P and S on copper uptake by plant at flowering stage (g ha<sup>-1</sup>)**

**T<sub>1</sub> : 9.53**

	<b>S<sub>0</sub></b>	<b>S<sub>1</sub></b>	<b>S<sub>2</sub></b>	<b>S<sub>3</sub></b>	<b>Mean</b>
<b>P<sub>0</sub></b>	8.15	9.56	10.77	12.03	<b>10.13</b>
<b>P<sub>1</sub></b>	8.30	9.91	10.85	12.16	<b>10.31</b>
<b>P<sub>2</sub></b>	9.03	10.00	10.58	11.25	<b>10.21</b>
<b>P<sub>3</sub></b>	8.88	10.59	12.78	13.47	<b>11.43</b>
<b>Mean</b>	<b>8.59</b>	<b>10.01</b>	<b>11.25</b>	<b>12.23</b>	
<b>CD (0.05) P; 0.123</b>		<b>CD (0.05) S; 0.123</b>		<b>CD (0.05) PxS; 0.246</b>	
<b>SE (m) P; 0.042</b>		<b>SE (m) S; 0.042</b>		<b>SE (m) PxS; 0.085</b>	

**Table 133: Effect of application of P and S on copper uptake by plant at pegging stage (g ha<sup>-1</sup>)**

**T<sub>1</sub> : 9.77**

	<b>S<sub>0</sub></b>	<b>S<sub>1</sub></b>	<b>S<sub>2</sub></b>	<b>S<sub>3</sub></b>	<b>Mean</b>
<b>P<sub>0</sub></b>	7.57	9.72	11.17	12.88	<b>10.34</b>
<b>P<sub>1</sub></b>	8.87	10.12	10.84	12.17	<b>10.50</b>
<b>P<sub>2</sub></b>	9.80	10.70	11.01	12.21	<b>10.93</b>
<b>P<sub>3</sub></b>	8.94	10.84	12.23	13.59	<b>11.40</b>
<b>Mean</b>	<b>8.80</b>	<b>10.35</b>	<b>11.31</b>	<b>12.71</b>	
<b>CD (0.05) P; 0.003</b>		<b>CD (0.05) S; 0.003</b>		<b>CD (0.05) PxS; 0.006</b>	
<b>SE (m) P; 0.001</b>		<b>SE (m) S; 0.001</b>		<b>SE (m) PxS; 0.002</b>	

**Table 134: Effect of application of P and S on copper uptake by plant at pod setting stage (g ha<sup>-1</sup>)**

**T<sub>1</sub> : 9.89**

	S <sub>0</sub>	S <sub>1</sub>	S <sub>2</sub>	S <sub>3</sub>	Mean
P <sub>0</sub>	7.37	9.46	10.25	11.93	<b>9.75</b>
P <sub>1</sub>	8.25	10.28	10.92	11.57	<b>10.25</b>
P <sub>2</sub>	9.78	11.44	11.25	11.91	<b>11.10</b>
P <sub>3</sub>	9.53	10.78	12.15	13.07	<b>11.38</b>
<b>Mean</b>	<b>8.73</b>	<b>10.49</b>	<b>11.14</b>	<b>12.12</b>	
<b>CD (0.05) P; 0.175</b>		<b>CD (0.05) S; 0.175</b>		<b>CD (0.05) PxS; 0.349</b>	
<b>SE (m) P; 0.060</b>		<b>SE (m) S; 0.060</b>		<b>SE (m) PxS; 0.120</b>	

**Table 135: Effect of application of P and S on copper uptake by plant at harvest stage (g ha<sup>-1</sup>)**

**T<sub>1</sub> : 10.11**

	S <sub>0</sub>	S <sub>1</sub>	S <sub>2</sub>	S <sub>3</sub>	Mean
P <sub>0</sub>	7.59	9.12	9.93	11.42	<b>9.52</b>
P <sub>1</sub>	8.28	10.36	11.57	11.88	<b>10.52</b>
P <sub>2</sub>	10.00	11.73	12.35	12.43	<b>11.63</b>
P <sub>3</sub>	10.58	12.48	15.44	14.82	<b>13.33</b>
<b>Mean</b>	<b>9.11</b>	<b>10.92</b>	<b>12.32</b>	<b>12.64</b>	
<b>CD (0.05) P; 0.175</b>		<b>CD (0.05) S; 0.175</b>		<b>CD (0.05) PxS; 0.349</b>	
<b>SE (m) P; 0.060</b>		<b>SE (m) S; 0.060</b>		<b>SE (m) PxS; 0.120</b>	

#### **4.6.11 Boron uptake by plant**

Uptake of B was significantly influenced by main effect and interaction effect. Treatment, P<sub>3</sub>S<sub>3</sub> (P, 90 kg ha<sup>-1</sup> and S, 30 kg ha<sup>-1</sup>) showed highest uptake at flowering,

pegging and pod formation stages. Treatment, P<sub>3</sub>S<sub>2</sub> (P, 90 kg ha<sup>-1</sup> and S, 20 kg ha<sup>-1</sup>) showed highest uptake at harvesting stage. F statistic for treatments Vs control was calculated and was found to be significantly different.

**Table 136: Effect of application of P and S on boron uptake by plant at flowering stage (g ha<sup>-1</sup>)**

**T<sub>1</sub>: 11.31**

	S <sub>0</sub>	S <sub>1</sub>	S <sub>2</sub>	S <sub>3</sub>	Mean
P <sub>0</sub>	7.25	9.31	10.13	11.59	<b>9.57</b>
P <sub>1</sub>	8.30	10.18	11.20	11.89	<b>10.39</b>
P <sub>2</sub>	9.70	11.08	11.43	11.96	<b>11.04</b>
P <sub>3</sub>	9.70	11.17	12.60	13.31	<b>11.70</b>
Mean	<b>8.74</b>	<b>10.44</b>	<b>11.34</b>	<b>12.19</b>	
CD (0.05) P; 0.245		CD (0.05) S; 0.245		CD (0.05) PxS; 0.490	
SE (m) P; 0.084		SE (m) S; 0.084		SE (m) PxS; 0.169	

**Table 137: Effect of application of P and S on boron uptake by plant at pegging stage (g ha<sup>-1</sup>)**

**T<sub>1</sub>: 11.22**

	S <sub>0</sub>	S <sub>1</sub>	S <sub>2</sub>	S <sub>3</sub>	Mean
P <sub>0</sub>	7.18	9.17	10.05	11.47	<b>9.46</b>
P <sub>1</sub>	8.18	10.00	10.77	11.62	<b>10.14</b>
P <sub>2</sub>	9.52	10.86	11.42	11.94	<b>10.94</b>
P <sub>3</sub>	9.34	11.00	12.11	12.86	<b>11.33</b>
Mean	<b>8.56</b>	<b>10.26</b>	<b>11.09</b>	<b>11.97</b>	
CD (0.05) P; 0.03		CD (0.05) S; 0.03		CD (0.05) PxS; 0.06	
SE (m) P; 0.01		SE (m) S; 0.01		SE (m) PxS; 0.021	

**Table 138: Effect of application of P and S on boron uptake by plant at pod setting stage ( $\text{g ha}^{-1}$ )**

**T<sub>1</sub>: 10.84**

	S <sub>0</sub>	S <sub>1</sub>	S <sub>2</sub>	S <sub>3</sub>	Mean
P <sub>0</sub>	6.95	8.81	9.59	10.96	<b>9.08</b>
P <sub>1</sub>	7.63	9.55	10.29	10.95	<b>9.61</b>
P <sub>2</sub>	9.35	10.76	11.21	11.70	<b>10.76</b>
P <sub>3</sub>	9.38	10.81	12.23	12.80	<b>11.30</b>
Mean	<b>8.33</b>	<b>9.98</b>	<b>10.83</b>	<b>11.60</b>	
CD (0.05) P; 0.008		CD (0.05) S; 0.008		CD (0.05) PxS; 0.016	
SE (m) P; 0.003		SE (m) S; 0.003		SE (m) PxS; 0.005	

**Table 139: Effect of application of P and S on boron uptake by plant at harvest stage ( $\text{g ha}^{-1}$ )**

**T<sub>1</sub>: 11.26**

	S <sub>0</sub>	S <sub>1</sub>	S <sub>2</sub>	S <sub>3</sub>	Mean
P <sub>0</sub>	7.40	9.49	10.43	11.81	<b>9.78</b>
P <sub>1</sub>	8.14	10.07	10.80	11.54	<b>10.14</b>
P <sub>2</sub>	9.80	11.30	11.81	12.32	<b>11.31</b>
P <sub>3</sub>	9.88	11.25	12.96	12.66	<b>11.94</b>
Mean	<b>8.81</b>	<b>10.53</b>	<b>11.50</b>	<b>12.33</b>	
CD (0.05) P; 0.056		CD (0.05) S; 0.056		CD (0.05) PxS; 0.113	
SE (m) P; 0.019		SE (m) S; 0.019		SE (m) PxS; 0.039	

#### 4.7 NUTRIENT UPTAKE BY POD

##### 4.7.1 Nitrogen uptake by kernel

Data (table 140) on N uptake by kernels showed that, N uptake varied



significantly due to application of different levels of P and S. The treatment, P<sub>0</sub>S<sub>2</sub> (P, 0 kg ha<sup>-1</sup> and S, 20 kg ha<sup>-1</sup>) showed highest N uptake (42.97 kg ha<sup>-1</sup>). F statistic for treatments Vs control was calculated and was found to be significantly different.

#### 4.7.2 Phosphorus uptake by kernel

Uptake of P by kernel was shown in the table 141. P uptake by kernel varied significantly due to application of different levels of P and S. The treatment, P<sub>3</sub>S<sub>2</sub> (P, 30 kg ha<sup>-1</sup> and S, 20 kg ha<sup>-1</sup>) showed highest P uptake (4.33 kg ha<sup>-1</sup>). F statistic for treatments Vs control was calculated and was found to be significantly different.

**Table 140: Effect of application of P and S on nitrogen uptake by kernel (kg ha<sup>-1</sup>)**

T<sub>1</sub>: 33.41

	S <sub>0</sub>	S <sub>1</sub>	S <sub>2</sub>	S <sub>3</sub>	Mean
P <sub>0</sub>	31.09	40.18	42.97	41.18	38.86
P <sub>1</sub>	38.28	41.32	39.82	37.86	39.32
P <sub>2</sub>	34.71	34.76	37.12	37.48	36.02
P <sub>3</sub>	36.02	37.56	40.59	39.18	38.34
Mean	35.02	38.46	40.13	38.93	
CD (0.05) P; 0.559	CD (0.05) S; 0.559			CD (0.05) PxS; 1.117	
SE (m) P; 0.192	SE (m) S; 0.192			SE (m) PxS; 0.385	

**Table 141: Effect of application of P and S on phosphorus uptake by kernel (kg ha<sup>-1</sup>)**

T<sub>1</sub>: 4.01

	S <sub>0</sub>	S <sub>1</sub>	S <sub>2</sub>	S <sub>3</sub>	Mean
P <sub>0</sub>	3.28	4.24	4.32	4.16	4.01
P <sub>1</sub>	3.99	4.15	4.01	3.92	4.02
P <sub>2</sub>	3.62	3.54	3.83	4.03	3.75
P <sub>3</sub>	3.64	3.85	4.33	4.24	4.01
Mean	3.63	3.95	4.13	4.09	
CD (0.05) P; 0.028	CD (0.05) S; 0.028			CD (0.05) PxS; 0.056	
SE (m) P; 0.01	SE (m) S; 0.01			SE (m) PxS; 0.019	

#### 4.7.3 Potassium uptake by kernel

Potassium uptake varied significantly due to main effect and interaction effect (table 142). The treatment, P<sub>3</sub>S<sub>2</sub> (P, 90 kg ha<sup>-1</sup> and S, 20 kg ha<sup>-1</sup>) showed highest K uptake (18.94 kg ha<sup>-1</sup>). F statistic for treatments Vs control was calculated and was found to be significantly different.

#### 4.7.4 Calcium uptake by kernel

Calcium uptake varied significantly due to main effect and interaction effect (table 143). The treatment, P<sub>3</sub>S<sub>3</sub> (P, 90 kg ha<sup>-1</sup> and S, 30 kg ha<sup>-1</sup>) showed highest Ca uptake . F statistic for treatments Vs control was calculated and was found to be significantly different.

#### 4.7.5 Magnesium uptake by kernel

Uptake of Mg by plant varied significantly due to different levels of fertilizer application (table 144). Significantly highest Mg uptake by kernel was noticed in treatment, P<sub>3</sub>S<sub>3</sub> (P, 90 kg ha<sup>-1</sup> and S, 30 kg ha<sup>-1</sup>). F statistic for treatments Vs control was calculated and was found to be significantly different.

#### 4.7.6 Sulphur uptake by kernel

Uptake of S by pod varied significantly due to different levels of fertilizer application (table 145). Significantly highest S uptake by kernel was noticed in treatment, P<sub>3</sub>S<sub>3</sub> (P, 90 kg ha<sup>-1</sup> and S, 30 kg ha<sup>-1</sup>). F statistic for treatments Vs control was calculated and was found to be significantly different.

**Table 142: Effect of application of P and S on potassium uptake by kernel (kg ha<sup>-1</sup>)**

T<sub>1</sub>: 14.65

	S <sub>0</sub>	S <sub>1</sub>	S <sub>2</sub>	S <sub>3</sub>	Mean
P <sub>0</sub>	12.03	12.51	12.80	12.48	12.45
P <sub>1</sub>	13.23	13.32	14.35	13.47	13.59
P <sub>2</sub>	14.02	14.94	15.54	16.19	15.17
P <sub>3</sub>	16.32	17.66	18.94	18.56	17.87
Mean	13.90	14.61	15.41	15.17	
CD (0.05) P; 0.121	CD (0.05) S; 0.121		CD (0.05) PxS; 0.242		
SE (m) P; 0.085	SE (m) S; 0.085		SE (m) PxS; 0.171		

**Table 143: Effect of application of P and S on Ca uptake by kernel (kg ha<sup>-1</sup>)**

**T<sub>1</sub>: 2.12**

	S <sub>0</sub>	S <sub>1</sub>	S <sub>2</sub>	S <sub>3</sub>	Mean
P <sub>0</sub>	1.48	2.04	2.40	2.52	2.11
P <sub>1</sub>	1.68	2.19	2.39	2.82	2.27
P <sub>2</sub>	1.72	2.45	3.10	3.06	2.58
P <sub>3</sub>	2.00	2.46	3.56	4.87	3.22
Mean	1.72	2.28	2.86	3.32	
CD (0.05) P; 0.659		CD (0.05) S; 0.659		CD (0.05) PxS; NS	
SE (m) P; 0.227		SE (m) S; 0.227		SE (m) PxS; 0.454	

**Table 144: Effect of application of P and S on Mg uptake by kernel (kg ha<sup>-1</sup>)**

**T<sub>1</sub>: 2.47**

	S <sub>0</sub>	S <sub>1</sub>	S <sub>2</sub>	S <sub>3</sub>	Mean
P <sub>0</sub>	2.27	2.49	2.47	2.52	2.44
P <sub>1</sub>	2.60	2.68	2.76	2.85	2.72
P <sub>2</sub>	3.62	3.38	3.42	3.48	3.47
P <sub>3</sub>	3.49	3.58	3.89	3.94	3.72
Mean	2.99	3.03	3.14	3.20	
CD (0.05) P; 0.012		CD (0.05) S; 0.012		CD (0.05) PxS; 0.024	
SE (m) P; 0.004		SE (m) S; 0.004		SE (m) PxS; 0.008	

**Table 145: Effect of application of P and S on S uptake by kernel (kg ha<sup>-1</sup>)**

**T<sub>1</sub>: 3.45**

	S <sub>0</sub>	S <sub>1</sub>	S <sub>2</sub>	S <sub>3</sub>	Mean
P <sub>0</sub>	1.45	5.41	7.20	7.15	<b>5.30</b>
P <sub>1</sub>	1.28	3.70	6.40	7.46	<b>4.71</b>
P <sub>2</sub>	1.63	3.90	6.28	7.49	<b>4.82</b>
P <sub>3</sub>	3.88	3.20	6.49	8.16	<b>5.43</b>
<b>Mean</b>	<b>2.06</b>	<b>4.05</b>	<b>6.59</b>	<b>7.57</b>	
<b>CD (0.05) P; 0.139</b>		<b>CD (0.05) S; 0.139</b>		<b>CD (0.05) PxS; 0.279</b>	
<b>SE (m) P; 0.048</b>		<b>SE (m) S; 0.048</b>		<b>SE (m) PxS; 0.096</b>	

#### 4.7.7 Iron uptake by kernel

Iron uptake by kernel was found to be significantly different due to various levels of P and S (table no. 146). Compared to other micro nutrients Fe was present in highest amount. The treatment, P<sub>3</sub>S<sub>3</sub> (P, 90 kg ha<sup>-1</sup> and S, 30 kg ha<sup>-1</sup>) showed highest Fe uptake. F statistic for treatments Vs control was calculated and was found to be significantly different.

#### 4.7.8 Manganese uptake by kernel

Uptake of Mn by kernel varied significantly due to different levels of fertilizer application (table no. 147). Significantly highest Mn uptake by kernel was noticed in treatment, P<sub>3</sub>S<sub>3</sub> (P, 90 kg ha<sup>-1</sup> and S, 30 kg ha<sup>-1</sup>). F statistic for treatments Vs control was calculated and was found to be significantly different.

#### 4.7.9 Zinc uptake by kernel

Zinc uptake varied significantly due to main effect and interaction effect (table no. 148). The treatment, P<sub>3</sub>S<sub>3</sub> (P, 90 kg ha<sup>-1</sup> and S, 30 kg ha<sup>-1</sup>) showed highest Zn uptake. F statistic for treatments Vs control was calculated and was found to be significantly different.

**Table 146: Effect of application of P and S on Fe uptake by kernel (g ha<sup>-1</sup>)**

**T<sub>1</sub>: 308.21**

	S <sub>0</sub>	S <sub>1</sub>	S <sub>2</sub>	S <sub>3</sub>	Mean
P <sub>0</sub>	235.17	316.68	397.94	563.37	<b>378.29</b>
P <sub>1</sub>	342.35	383.16	492.45	635.39	<b>463.34</b>
P <sub>2</sub>	369.46	438.60	576.05	729.24	<b>528.34</b>
P <sub>3</sub>	420.24	556.22	703.15	813.96	<b>623.39</b>
Mean	<b>341.81</b>	<b>423.67</b>	<b>542.40</b>	<b>685.49</b>	
<b>CD (0.05) P; 0.423</b>		<b>CD (0.05) S; 0.423</b>		<b>CD (0.05) PxS; 0.845</b>	
<b>SE (m) P; 0.146</b>		<b>SE (m) S; 0.146</b>		<b>SE (m) PxS; 0.291</b>	

**Table 147: Effect of application of P and S on Mn uptake by kernel (g ha<sup>-1</sup>)**

**T<sub>1</sub>: 32.78**

	S <sub>0</sub>	S <sub>1</sub>	S <sub>2</sub>	S <sub>3</sub>	Mean
P <sub>0</sub>	30.60	32.99	34.84	35.98	<b>33.60</b>
P <sub>1</sub>	36.62	36.90	38.44	38.76	<b>37.68</b>
P <sub>2</sub>	40.10	41.20	43.98	44.56	<b>42.46</b>
P <sub>3</sub>	45.23	48.72	49.09	54.90	<b>49.48</b>
Mean	<b>38.14</b>	<b>39.95</b>	<b>41.59</b>	<b>43.55</b>	
<b>CD (0.05) P; 0.161</b>		<b>CD (0.05) S; 0.161</b>		<b>CD (0.05) PxS; 0.321</b>	
<b>SE (m) P; 0.055</b>		<b>SE (m) S; 0.055</b>		<b>SE (m) PxS; 0.111</b>	

**Table 148: Effect of application of P and S on Zn uptake by kernel ( $\text{g ha}^{-1}$ )**

**T<sub>1</sub>: 99.87**

	<b>S<sub>0</sub></b>	<b>S<sub>1</sub></b>	<b>S<sub>2</sub></b>	<b>S<sub>3</sub></b>	<b>Mean</b>
<b>P<sub>0</sub></b>	92.82	95.84	93.84	95.92	<b>94.60</b>
<b>P<sub>1</sub></b>	97.34	98.73	99.96	104.04	<b>100.02</b>
<b>P<sub>2</sub></b>	108.10	108.12	109.04	110.73	<b>109.00</b>
<b>P<sub>3</sub></b>	111.10	114.80	117.03	121.61	<b>116.13</b>
<b>Mean</b>	<b>102.34</b>	<b>104.37</b>	<b>104.97</b>	<b>108.07</b>	
<b>CD (0.05) P; 0.111</b>		<b>CD (0.05) S; 0.111</b>		<b>CD (0.05) PxS; 0.222</b>	
<b>SE (m) P; 0.038</b>		<b>SE (m) S; 0.038</b>		<b>SE (m) PxS; 0.076</b>	

#### 4.7.10 Copper uptake by kernel

Copper uptake varied significantly due to main effect and interaction effect (table no. 149). The treatment, P<sub>3</sub>S<sub>3</sub> (P, 90 kg ha<sup>-1</sup> and S, 30 kg ha<sup>-1</sup>) showed highest Cu uptake. F statistic for treatments Vs control was calculated and was found to be significantly different.

#### 4.7.11 Boron uptake by kernel

Uptake of B by kernel at different stages is shown in the table 150. B uptake by groundnut kernel varied significantly due to different treatments applied. Significantly highest uptake was noted in treatment having high P and S. F statistic for treatments Vs control was calculated and was found to be significantly different.

**Table 149: Effect of application of P and S on Cu uptake by kernel (g ha<sup>-1</sup>)**

**T<sub>1</sub>: 25.21**

	S <sub>0</sub>	S <sub>1</sub>	S <sub>2</sub>	S <sub>3</sub>	Mean
P <sub>0</sub>	19.48	20.73	20.40	20.81	<b>20.35</b>
P <sub>1</sub>	21.53	21.82	23.69	22.95	<b>22.50</b>
P <sub>2</sub>	24.36	24.72	25.13	26.14	<b>25.09</b>
P <sub>3</sub>	26.39	27.14	27.78	28.56	<b>27.47</b>
<b>Mean</b>	<b>22.94</b>	<b>23.60</b>	<b>24.25</b>	<b>24.61</b>	
<b>CD (0.05) P; 0.122</b>		<b>CD (0.05) S; 0.122</b>		<b>CD (0.05) PxS; 0.244</b>	
<b>SE (m) P; 0.042</b>		<b>SE (m) S; 0.042</b>		<b>SE (m) PxS; 0.084</b>	

**Table 150: Effect of application of P and S on B uptake by kernel (g ha<sup>-1</sup>)**

**T<sub>1</sub>: 3.56**

	S <sub>0</sub>	S <sub>1</sub>	S <sub>2</sub>	S <sub>3</sub>	Mean
P <sub>0</sub>	3.18	3.19	3.18	3.20	<b>3.19</b>
P <sub>1</sub>	3.35	3.50	3.58	3.71	<b>3.54</b>
P <sub>2</sub>	3.71	3.76	3.69	3.96	<b>3.78</b>
P <sub>3</sub>	4.65	4.16	4.77	4.92	<b>4.63</b>
<b>Mean</b>	<b>3.72</b>	<b>3.65</b>	<b>3.81</b>	<b>3.95</b>	
<b>CD (0.05) P; 0.05</b>		<b>CD (0.05) S; 0.05</b>		<b>CD (0.05) PxS; 0.10</b>	
<b>SE (m) P; 0.017</b>		<b>SE (m) S; 0.017</b>		<b>SE (m) PxS; 0.034</b>	

#### 4.7.12 Nitrogen uptake by shell

Data (table 151) on N uptake by shell showed that, N uptake by shell varied significantly due to application of different levels of P and S. The treatment, P<sub>3</sub>S<sub>3</sub> (P,

90 kg ha<sup>-1</sup> and S, 30 kg ha<sup>-1</sup>) showed highest N uptake. F statistic for treatments Vs control was calculated and was found to be significantly different.

#### 4.7.13 Phosphorus uptake by shell

Uptake of P by shell was shown in the table 152. P uptake by shell varied significantly due to application of different levels of P and S. The treatment, P<sub>3</sub>S<sub>3</sub> (P, 30 kg ha<sup>-1</sup> and S, 30 kg ha<sup>-1</sup>) showed highest P uptake. F statistic for treatments Vs control was calculated and was found to be significantly different.

**Table 151: Effect of application of P and S on N uptake by shell (kg ha<sup>-1</sup>)**

T<sub>1</sub>: 1.02

	S <sub>0</sub>	S <sub>1</sub>	S <sub>2</sub>	S <sub>3</sub>	Mean
P <sub>0</sub>	0.97	0.98	1.04	1.07	1.02
P <sub>1</sub>	0.98	1.07	1.08	1.08	1.05
P <sub>2</sub>	1.06	1.10	1.12	1.16	1.11
P <sub>3</sub>	1.09	1.12	1.17	1.19	1.14
Mean	1.03	1.07	1.10	1.13	
CD (0.05) P; 0.022		CD (0.05) S; 0.022		CD (0.05) PxS; 0.044	
SE (m) P; 0.08		SE (m) S; 0.08		SE (m) PxS; 0.15	

**Table 152: Effect of application of P and S on P uptake by shell (kg ha<sup>-1</sup>)**

T<sub>1</sub>: 3.88

	S <sub>0</sub>	S <sub>1</sub>	S <sub>2</sub>	S <sub>3</sub>	Mean
P <sub>0</sub>	0.25	0.41	0.45	0.45	0.39
P <sub>1</sub>	0.38	0.44	0.64	0.70	0.54
P <sub>2</sub>	0.29	0.51	0.63	0.71	0.54
P <sub>3</sub>	0.54	0.60	0.78	0.94	0.71
Mean	0.37	0.49	0.63	0.70	
CD (0.05) P; 0.02		CD (0.05) S; 0.02		CD (0.05) PxS; 0.03	
SE (m) P; 0.01		SE (m) S; 0.01		SE (m) PxS; 0.01	



#### 4.7.14 Potassium uptake by shell

Potassium uptake varied significantly due to main effect and interaction effect (table 153). The treatment, P<sub>3</sub>S<sub>3</sub> (P, 90 kg ha<sup>-1</sup> and S, 30 kg ha<sup>-1</sup>) showed highest K uptake. F statistic for treatments Vs control was calculated and was found to be significantly different.

#### 4.7.15 Calcium uptake by shell

Calcium uptake varied significantly due to main effect and interaction effect (table 154). The treatment, P<sub>1</sub>S<sub>3</sub> (P, 60 kg ha<sup>-1</sup> and S, 30 kg ha<sup>-1</sup>) showed highest Ca uptake. F statistic for treatments Vs control was calculated and was found to be significantly different.

#### 4.7.16 Magnesium uptake by shell

Uptake of Mg by shell varied significantly due to different levels of fertilizer application (table 155). Significantly highest Mg uptake by plant was noticed in treatment, P<sub>3</sub>S<sub>3</sub> (P, 90 kg ha<sup>-1</sup> and S, 30 kg ha<sup>-1</sup>). F statistic for treatments Vs control was calculated and was found to be significantly different.

#### 4.7.17 Sulphur uptake by shell

Uptake of S by pod varied significantly due to different levels of fertilizer application (table 156). Significantly highest S uptake by shell was noticed in treatment, P<sub>3</sub>S<sub>3</sub> (P, 90 kg ha<sup>-1</sup> and S, 30 kg ha<sup>-1</sup>). F statistic for treatments Vs control was calculated and was found to be significantly different.

**Table 153: Effect of application of P and S on K uptake by shell (kg ha<sup>-1</sup>)**

T<sub>1</sub>: 1.95

	S <sub>0</sub>	S <sub>1</sub>	S <sub>2</sub>	S <sub>3</sub>	Mean
P <sub>0</sub>	1.97	1.58	1.69	1.71	1.74
P <sub>1</sub>	1.74	1.69	1.82	1.92	1.79
P <sub>2</sub>	1.97	2.07	2.22	2.14	2.10
P <sub>3</sub>	2.26	2.32	2.31	2.87	2.44
Mean	1.98	1.92	2.01	2.16	
CD (0.05) P; 0.13	CD (0.05) S; 0.13			CD (0.05) P x S; 0.27	
SE (m) P; 0.07	SE (m) S; 0.07			SE (m) P x S; 0.15	

**Table 154: Effect of application of P and S on Ca uptake by shell (kg ha<sup>-1</sup>)**

**T<sub>1</sub>: 5.31**

	<b>S<sub>0</sub></b>	<b>S<sub>1</sub></b>	<b>S<sub>2</sub></b>	<b>S<sub>3</sub></b>	<b>Mean</b>
<b>P<sub>0</sub></b>	3.28	4.39	4.58	8.44	5.17
<b>P<sub>1</sub></b>	5.28	4.44	8.06	9.02	6.70
<b>P<sub>2</sub></b>	3.60	6.62	5.96	7.90	6.02
<b>P<sub>3</sub></b>	5.51	5.25	6.46	7.45	6.17
<b>Mean</b>	4.42	5.18	6.27	8.20	
<b>CD (0.05) P; 0.087</b>		<b>CD (0.05) S; 0.087</b>		<b>CD (0.05) PxS; 0.175</b>	
<b>SE (m) P; 0.301</b>		<b>SE (m) S; 0.301</b>		<b>SE (m) PxS; 0.603</b>	

**Table 155: Effect of application of P and S on Mg uptake by shell (kg ha<sup>-1</sup>)**

**T<sub>1</sub>: 5.24**

	<b>S<sub>0</sub></b>	<b>S<sub>1</sub></b>	<b>S<sub>2</sub></b>	<b>S<sub>3</sub></b>	<b>Mean</b>
<b>P<sub>0</sub></b>	3.50	4.10	4.44	4.11	4.04
<b>P<sub>1</sub></b>	6.06	6.20	6.24	6.41	6.23
<b>P<sub>2</sub></b>	6.91	7.07	7.17	7.19	7.08
<b>P<sub>3</sub></b>	7.33	7.42	7.62	7.91	7.57
<b>Mean</b>	5.95	6.20	6.37	6.40	
<b>CD (0.05) P; 0.05</b>		<b>CD (0.05) S; 0.05</b>		<b>CD (0.05) PxS; 0.099</b>	
<b>SE (m) P; 0.17</b>		<b>SE (m) S; 0.17</b>		<b>SE (m) PxS; 0.34</b>	

**Table 156: Effect of application of P and S on S uptake by shell (kg ha<sup>-1</sup>)**

**T<sub>1</sub>: 2.25**

	S <sub>0</sub>	S <sub>1</sub>	S <sub>2</sub>	S <sub>3</sub>	Mean
P <sub>0</sub>	1.50	1.60	1.77	1.77	1.66
P <sub>1</sub>	1.80	1.86	1.96	2.09	1.93
P <sub>2</sub>	2.10	2.17	2.25	2.26	2.19
P <sub>3</sub>	2.27	2.31	2.40	2.61	2.40
Mean	1.92	1.99	2.09	2.18	
CD (0.05) P; 0.048		CD (0.05) S; 0.048		CD (0.05) PxS; 0.095	
SE (m) P; 0.016		SE (m) S; 0.016		SE (m) PxS; 0.033	

#### 4.7.18 Iron uptake by shell

Iron uptake by shell was found to be significantly influenced by different levels of P and S. Compared to other micro nutrients, Fe was present in highest amount in shell. The treatment, P<sub>3</sub>S<sub>3</sub> (P, 90 kg ha<sup>-1</sup> and S, 30 kg ha<sup>-1</sup>) showed highest Fe uptake. F statistic for treatments Vs control was calculated and was found to be significantly different.

#### 4.7.19 Manganese uptake by shell

Uptake of Mn by shell varied significantly due to different levels of fertilizer application. Significantly highest Mn uptake by pod was noticed in treatment, P<sub>3</sub>S<sub>3</sub> (P, 90 kg ha<sup>-1</sup> and S, 30 kg ha<sup>-1</sup>). F statistic for treatments Vs control was calculated and was found to be significantly different.

#### 4.7.20 Zinc uptake by shell

Zinc uptake varied significantly due to main effect and interaction effect. The treatment, P<sub>3</sub>S<sub>3</sub> (P, 90 kg ha<sup>-1</sup> and S, 30 kg ha<sup>-1</sup>) showed highest Zn uptake. F statistic for treatments Vs control was calculated and was found to be significantly different.

**Table 157: Effect of application of P and S on Fe uptake by shell (g ha<sup>-1</sup>)**

**T<sub>1</sub>: 325.24**

	S <sub>0</sub>	S <sub>1</sub>	S <sub>2</sub>	S <sub>3</sub>	Mean
P <sub>0</sub>	219.42	290.29	367.64	442.68	<b>330.01</b>
P <sub>1</sub>	280.85	379.37	444.21	475.02	<b>394.86</b>
P <sub>2</sub>	314.65	416.16	485.85	531.48	<b>437.04</b>
P <sub>3</sub>	359.04	460.81	518.98	559.29	<b>474.53</b>
<b>Mean</b>	<b>293.49</b>	<b>386.66</b>	<b>454.17</b>	<b>502.12</b>	
<b>CD (0.05) P; 0.37</b>		<b>CD (0.05) S; 0.37</b>		<b>CD (0.05) PxS; 0.74</b>	
<b>SE (m) P; 0.13</b>		<b>SE (m) S; 0.13</b>		<b>SE (m) PxS; 0.25</b>	

**Table 158: Effect of application of P and S on Mn uptake by shell (g ha<sup>-1</sup>)**

**T<sub>1</sub>: 40.22**

	S <sub>0</sub>	S <sub>1</sub>	S <sub>2</sub>	S <sub>3</sub>	Mean
P <sub>0</sub>	22.91	30.47	33.09	33.37	<b>29.96</b>
P <sub>1</sub>	33.44	35.75	38.23	38.40	<b>36.46</b>
P <sub>2</sub>	38.63	39.45	41.11	41.94	<b>40.28</b>
P <sub>3</sub>	42.33	45.37	45.96	48.96	<b>45.66</b>
<b>Mean</b>	<b>34.33</b>	<b>37.76</b>	<b>39.59</b>	<b>40.67</b>	
<b>CD (0.05) P; 0.329</b>		<b>CD (0.05) S; 0.329</b>		<b>CD (0.05) PxS; 0.658</b>	
<b>SE (m) P; 0.133</b>		<b>SE (m) S; 0.133</b>		<b>SE (m) PxS; 0.227</b>	

**Table 159: Effect of application of P and S on Zn uptake by shell (g ha<sup>-1</sup>)**

**T<sub>1</sub>: 69.23**

	S <sub>0</sub>	S <sub>1</sub>	S <sub>2</sub>	S <sub>3</sub>	Mean
P <sub>0</sub>	59.77	67.16	70.32	71.86	<b>67.28</b>
P <sub>1</sub>	77.93	75.92	78.31	79.97	<b>78.03</b>
P <sub>2</sub>	80.60	80.78	84.03	84.65	<b>82.52</b>
P <sub>3</sub>	85.08	86.68	88.56	90.45	<b>87.69</b>
Mean	<b>75.85</b>	<b>77.64</b>	<b>80.30</b>	<b>81.73</b>	
CD (0.05) P; 0.044			CD (0.05) S; 0.044		CD (0.05) PxS; 0.088
SE (m) P; 0.08		SE (m) S; 0.08		SE (m) PxS; 0.17	

#### 4.7.21 Copper uptake by shell

Copper uptake varied significantly due to main effect and interaction effect. The treatment, P<sub>3</sub>S<sub>2</sub> (P, 90 kg ha<sup>-1</sup> and S, 20 kg ha<sup>-1</sup>) showed highest Cu uptake. F statistic for treatments Vs control was calculated and was found to be significantly different.

#### 4.7.22 Boron uptake by shell

Uptake of B by shell is shown in the table 161. B uptake by groundnut shell varied significantly due to different treatments applied. Significantly highest uptake was in treatment P<sub>3</sub> S<sub>2</sub> (P, 90 kg ha<sup>-1</sup> and S, 20 kg ha<sup>-1</sup>) and P<sub>3</sub> S<sub>3</sub> (P, 90 kg ha<sup>-1</sup> and S, 30 kg ha<sup>-1</sup>). F statistic for treatments Vs control was calculated and was found to be significantly different.

**Table 160: Effect of application of P and S on Cu uptake by shell (g ha<sup>-1</sup>)**

**T<sub>1</sub>: 22.30**

	S <sub>0</sub>	S <sub>1</sub>	S <sub>2</sub>	S <sub>3</sub>	Mean
P <sub>0</sub>	18.27	18.66	18.97	19.99	<b>18.97</b>
P <sub>1</sub>	21.83	21.71	21.84	21.92	<b>21.82</b>
P <sub>2</sub>	22.13	22.87	22.35	22.96	<b>22.58</b>
P <sub>3</sub>	23.26	23.35	23.52	23.24	<b>23.59</b>
Mean	<b>21.37</b>	<b>21.64</b>	<b>21.67</b>	<b>22.28</b>	
CD (0.05) P; 0.22		CD (0.05) S; 0.22		CD (0.05) PxS; 0.43	
SE (m) P; 0.08		SE (m) S; 0.08		SE (m) PxS; 0.15	

**Table 161: Effect of application of P and S on B uptake by shell (g ha<sup>-1</sup>)**

**T<sub>1</sub>: 2.25**

	S <sub>0</sub>	S <sub>1</sub>	S <sub>2</sub>	S <sub>3</sub>	Mean
P <sub>0</sub>	1.33	1.10	1.10	1.29	<b>1.21</b>
P <sub>1</sub>	1.36	1.66	1.70	1.76	<b>1.62</b>
P <sub>2</sub>	1.76	1.72	1.89	1.96	<b>1.83</b>
P <sub>3</sub>	2.35	2.34	2.69	2.69	<b>2.52</b>
Mean	<b>1.71</b>	<b>1.70</b>	<b>1.84</b>	<b>1.92</b>	
CD (0.05) P; 0.101		CD (0.05) S; 0.101		CD (0.05) PxS; 0.203	
SE (m) P; 0.035		SE (m) S; 0.035		SE (m) PxS; 0.07	

#### 4.8 PROTEIN CONTENT IN KERNEL

Data on protein content in groundnut kernel is shown in the table 162. Protein content was significantly influenced by main effect but interaction effect of P and S was found to be non-significant. Highest protein content was noted in P<sub>3</sub>S<sub>3</sub> (P, 90 kg ha<sup>-1</sup> and S, 30 kg ha<sup>-1</sup>). F statistic for treatments Vs control was calculated and was found to be significantly different.

**Table 162: Effect of application of P and S on protein content (g)**

**T<sub>1</sub>: 19.69**

	S <sub>0</sub>	S <sub>1</sub>	S <sub>2</sub>	S <sub>3</sub>	Mean
P <sub>0</sub>	19.69	20.33	21.83	22.48	<b>21.08</b>
P <sub>1</sub>	20.50	21.52	21.83	23.08	<b>21.73</b>
P <sub>2</sub>	21.06	21.77	22.81	22.85	<b>22.13</b>
P <sub>3</sub>	21.71	22.33	22.98	23.94	<b>22.74</b>
Mean	<b>20.74</b>	<b>21.49</b>	<b>22.37</b>	<b>23.09</b>	
<b>CD (0.05) P; 0.34</b>		<b>CD (0.05) S; 0.34</b>		<b>CD (0.05) PxS; NS</b>	
<b>SE (m) P; 0.12</b>		<b>SE (m) S; 0.12</b>		<b>SE (m) PxS; 0.23</b>	

#### 4.9 OIL CONTENT IN KERNEL

Data on oil content in groundnut kernel is shown in the table 163. Oil content was significantly influenced by main effect and interaction effect of P and S. Oil content increased with increased dose of both P and S. Highest oil content was noted in P<sub>3</sub>S<sub>3</sub> (P, 90 kg ha<sup>-1</sup> and S, 30 kg ha<sup>-1</sup>). It is on par with treatment P<sub>2</sub>S<sub>3</sub> (P, 75 kg ha<sup>-1</sup> and S, 30 kg ha<sup>-1</sup>). F statistic for treatments Vs control was calculated and was found to be significantly different.

**Table 163: Effect of application of P and S on oil content (%)**

**T<sub>1</sub>: 37.66**

	S <sub>0</sub>	S <sub>1</sub>	S <sub>2</sub>	S <sub>3</sub>	Mean
P <sub>0</sub>	32.57	34.03	39.67	49.63	<b>38.98</b>
P <sub>1</sub>	32.90	34.03	40.32	52.80	<b>40.01</b>
P <sub>2</sub>	33.17	37.07	42.10	56.80	<b>42.28</b>
P <sub>3</sub>	34.05	36.20	45.00	57.40	<b>43.16</b>
Mean	<b>33.17</b>	<b>35.33</b>	<b>41.77</b>	<b>54.16</b>	
<b>CD (0.05) P; 0.41</b>		<b>CD (0.05) S; 0.41</b>		<b>CD (0.05) PxS; 0.82</b>	
<b>SE (m) P; 0.14</b>		<b>SE (m) S; 0.14</b>		<b>SE (m) PxS; 0.28</b>	

## 4.10 ECONOMICS

### 4.10.1 Benefit: Cost Ratio

Application of different levels of P and S significantly influenced the benefit: cost ratio (Table 164). Highest benefit: cost ratio was noted in P<sub>3</sub>S<sub>3</sub> (P, 90 kg ha<sup>-1</sup> and S, 30 kg ha<sup>-1</sup>). It was on par with treatment P<sub>3</sub>S<sub>2</sub> (P, 90 kg ha<sup>-1</sup> and S, 20 kg ha<sup>-1</sup>). F statistic for treatments Vs control was calculated and was found to be significantly different.

**Table 164: Effect of application of P and S on benefit cost ratio**

**T<sub>1</sub>: 2.20**

	S <sub>0</sub>	S <sub>1</sub>	S <sub>2</sub>	S <sub>3</sub>	Mean
P <sub>0</sub>	1.99	1.99	2.04	2.11	<b>2.03</b>
P <sub>1</sub>	2.05	2.06	2.08	2.15	<b>2.09</b>
P <sub>2</sub>	2.10	2.12	2.16	2.18	<b>2.14</b>
P <sub>3</sub>	2.19	2.24	2.31	2.35	<b>2.27</b>
<b>Mean</b>	<b>2.08</b>	<b>2.10</b>	<b>2.15</b>	<b>2.20</b>	
<b>CD (0.05) P; 0.05</b>	<b>CD (0.05) S; 0.05</b>			<b>CD (0.05) PxS; NS</b>	
<b>SE (m) P; 0.11</b>	<b>SE (m) S; 0.11</b>			<b>SE (m) PxS; 0.22</b>	



# Discussion

## 5. DISCUSSION

The results of study entitled "Interaction of phosphorus and sulphur in black cotton soils of Palakkad (AEU: 23) under groundnut (*Arachis hypogaea* L.) cultivation" presented in chapter 4 are discussed here with supporting studies conducted elsewhere and based on available literature.

### 5.1 GROWTH AND YIELD ATTRIBUTES OF GROUNDNUT

#### 5.1.1 Growth parameters

##### 5.1.1.1 Plant height and number of leaves

Plant height and number of leaves at flowering, pegging, pod formation and harvesting stages were significantly influenced by application of different levels of P and S fertilizers. The treatment,  $P_3S_3$  (P, 90 kg ha<sup>-1</sup> and S, 30 kg ha<sup>-1</sup>) showed the highest plant height at flowering (22.13 cm) and harvesting stages (41.30 cm). At pegging stage (35.00cm), the treatment  $T_{16}$ ,  $P_3S_2$  (P, 90kg ha<sup>-1</sup> and S, 20 kg ha<sup>-1</sup>) showed highest plant height. Whereas in pod formation (36.67 cm) stage, both the treatments,  $T_{16}$  and  $T_{17}$  recorded highest plant height.

The treatment,  $P_3S_3$  (P, 90kg ha<sup>-1</sup> and S, 30 kg ha<sup>-1</sup>) showed highest number of leaves at pegging (53.81), pod development (57.93) and harvesting (55.06) stages, whereas during flowering stage, interaction effect was found to be non significant. Phosphorus application increased plant height and number of leaves. Application of soluble phosphorus increased the availability of soluble phosphate to plants and enhanced their root development and there by enhanced nutrient uptake and resulted in improved plant growth. Higher level of P is important for formation and growth of root and N fixation (Lakshamma and Raj, 1997). Similar results were reported by Punnoose (1968), Sebale and Khuspe (1980), Patel *et al.* (1981) and Juan *et al.* (1986). Higher soil P content increased N fixation which in turn enhanced the plant growth (Balasubramanian and Palaniappan, 1991). This was in conformity with the findings of Punnoose (1968), Singh and Ahuja (1985) and Rayar (1986). Basha and Rao (1980) observed reduction in plant height and shoot length under P deficiency in groundnut plant.

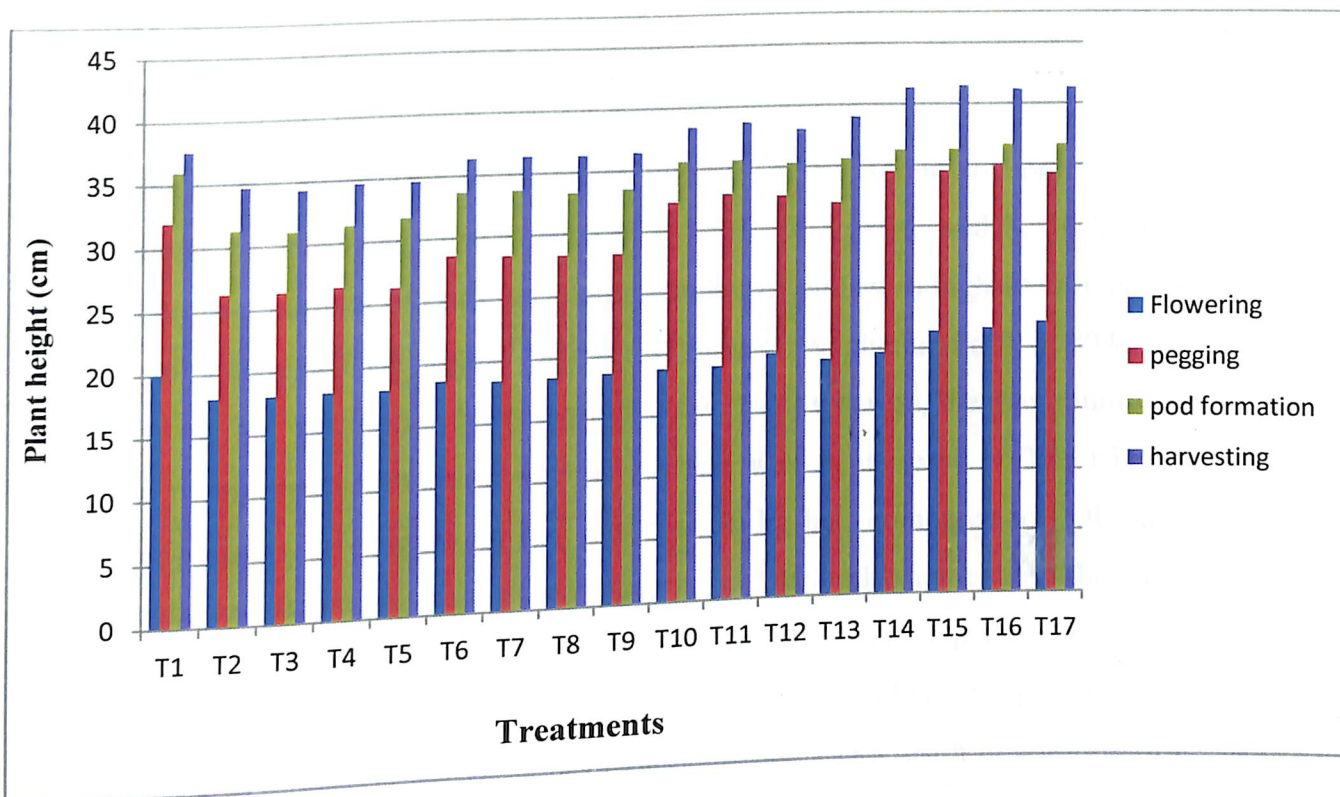


Fig 1: Effect of application of P and S on plant height at different stages

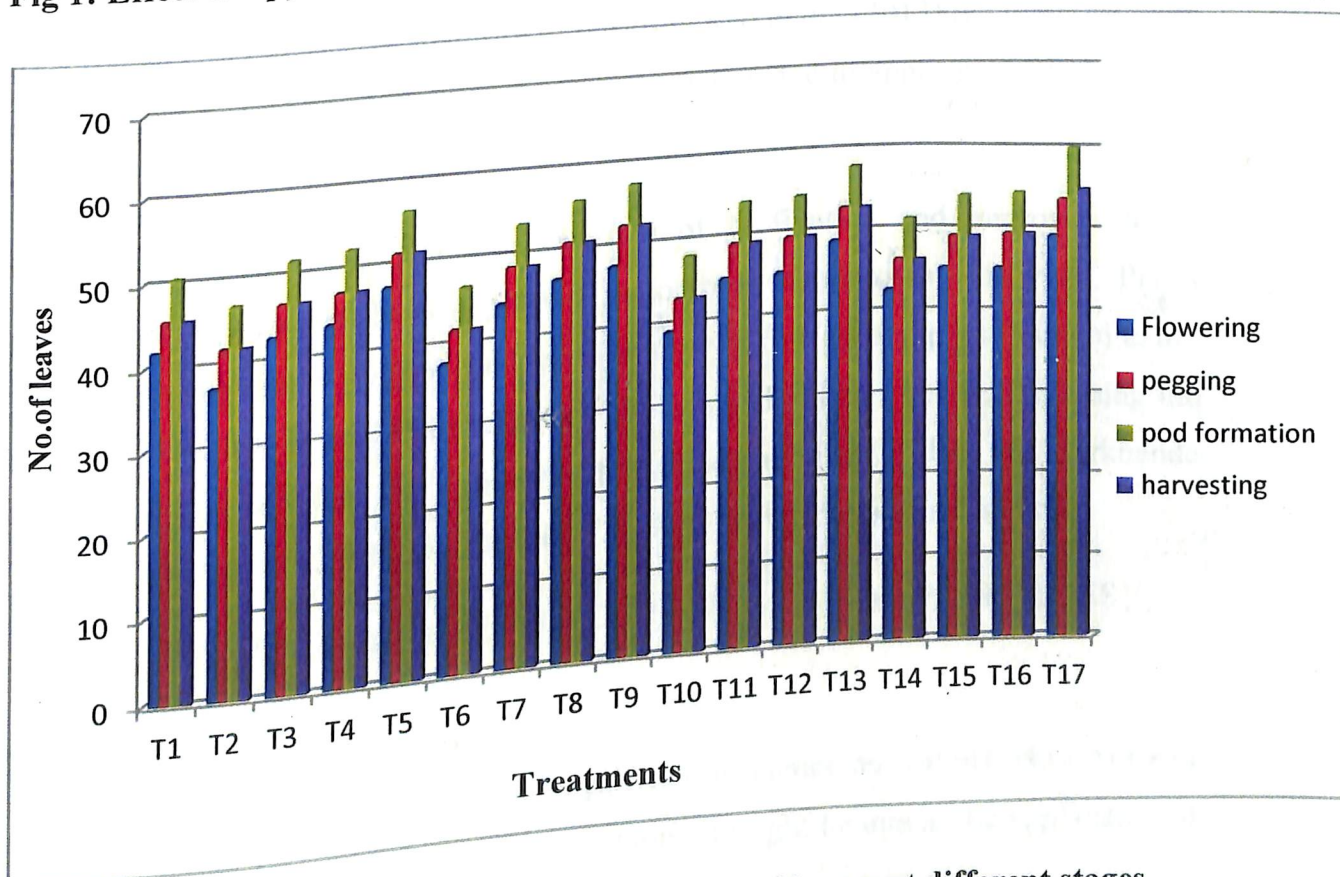


Fig 2: Effect of application of P and S on number of leaves at different stages

## 5.1.2 Yield parameters

### 5.1.2.1 Number of pods per plant and yield

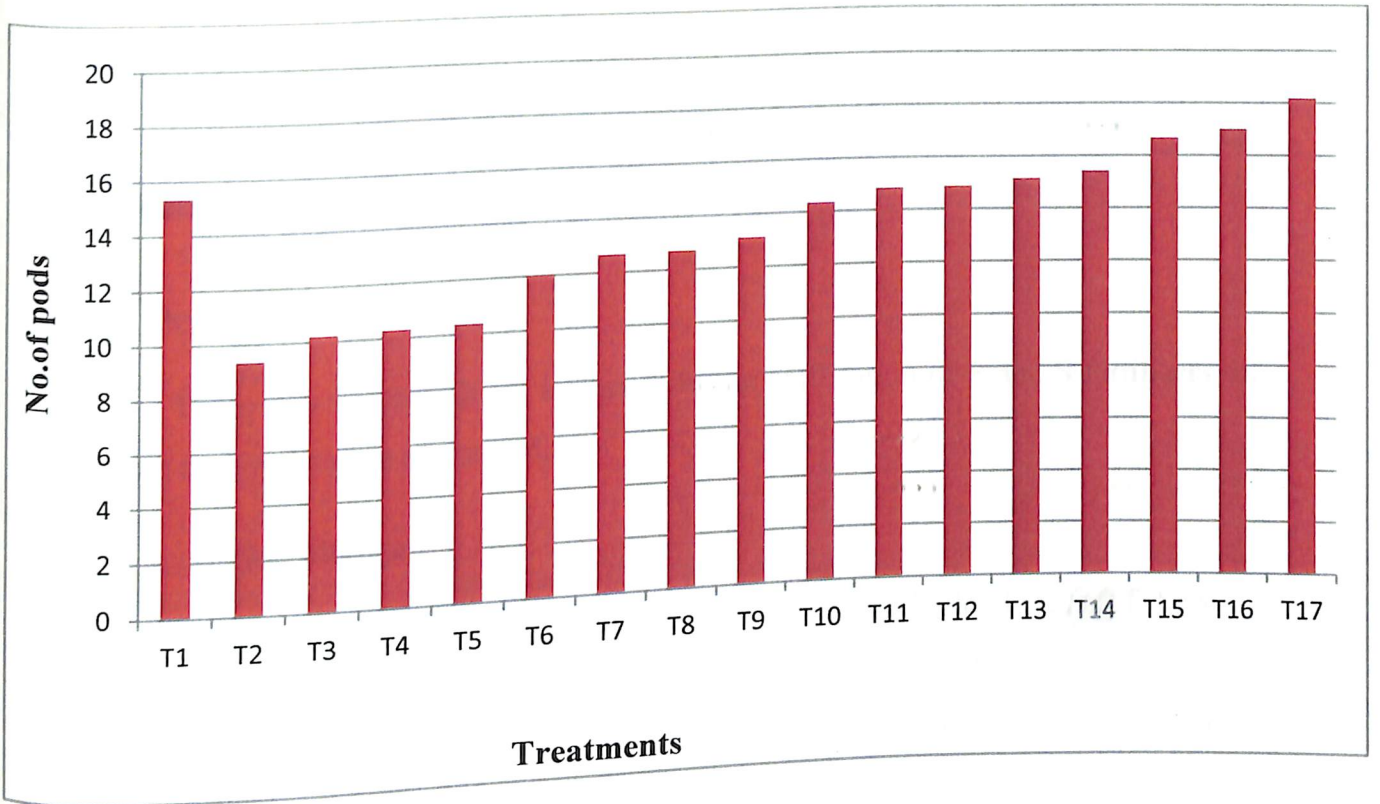
There was significant variation in number of pods due to application of various levels of fertilizers. The treatment,  $P_3S_3$  (P, 90 kg ha<sup>-1</sup> and S, 30 kg ha<sup>-1</sup>) produced highest number of pods per plant. Number of pods was highest in treatment receiving high dose of P (90 kg ha<sup>-1</sup>) and S (30 kg ha<sup>-1</sup>). Application of P at 90 kg ha<sup>-1</sup> and S at 30 kg ha<sup>-1</sup> resulted in highest yield (3.68 t ha<sup>-1</sup>) whereas, the maximum yield from POP recommendation was 3.45 t ha<sup>-1</sup>. There was an increase of 0.23 t ha<sup>-1</sup> in yield on application of P at 90 kg ha<sup>-1</sup> and S at 30 kg ha<sup>-1</sup> compared to POP based recommendation. Banerjee *et al.* (1967) and Puri (1969) also concluded the same. They found that application of phosphorus increased number of pods and yield in groundnut. Application of 90 kg P<sub>2</sub>O<sub>5</sub> ha<sup>-1</sup> recorded highest yield of dry pod. Samtana *et al.* (1994) also reported improvement in yield attributes as a result of addition of P. This was due to formation and proliferation of new roots and improvement in their functional activity. P stimulates setting of pods, decreases the number of unfilled pods (pops) and hastens the maturity of the crop. Rao *et al.*, (2013) reported that there was increase in yield and yield attributing characters due to application of elemental sulphur.

Application of P increases the rate of N fixation and improves the N availability. Increase in N also plays a major role in improving the yield. Plants having high N availability produced more number of flowers and pegs (Saradhi *et al.*, 1990) and resulted in an increased yield. N plays a significant role in increasing the number of pods per plant and number of filled pods per plant. Jadhar and Narkhede (1980), Reddy *et al.* (1984) and Patel *et al.* (1994) also concluded the same.

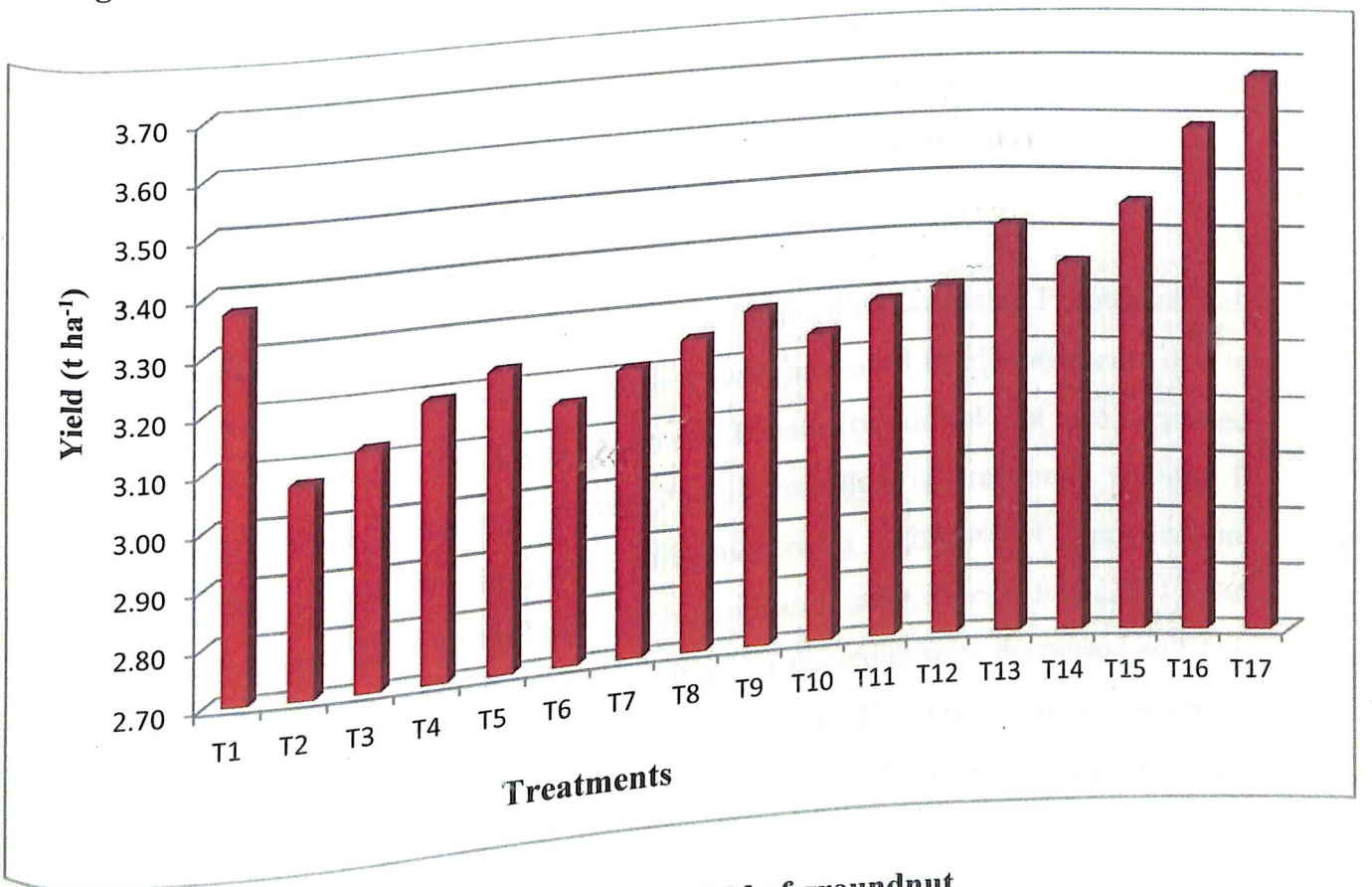
## 5.2 EFFECT OF DIFFERENT LEVELS OF P AND S ON SOIL PROPERTIES

### 5.2.1 Soil pH

Application of P and S had no significant influence on soil pH. However soil pH decreased slightly compared to initial status. It might be due to the application of acid forming fertilizers, urea and elemental sulphur.



**Fig 3: Effect of application of P and S on number of pods per plant**



**Fig 4: Effect of application of P and S on yield of groundnut**

### 5.2.2 EC

There was a slight increase in electrical conductivity of soil due to the application of soluble fertilizers.

### 5.2.3 Organic carbon

There was no significant influence in the application of P and S fertilizers on organic carbon content of the soil.

### 5.2.4 Available nitrogen

Application of P and S significantly influenced available N status of the soil. The requirement of P in nodulating legumes is higher compared to non-nodulating crops as it plays a significant role in nodule formation and fixation of atmospheric N (Brady and Weil, 2002). Available N status of soil increased with increased dose of application of P. P has a positive interaction with N and plant development (Sumner and Farina, 1986). This might be due to the increased N fixation. Balasubramanian and Palaniappan (1991) reported that higher the concentration of P, the higher will be the amount of N fixed. Available N has increased with increased dose of S also. Fazili *et al.* (2008) reported that S addition increases the N availability.

### 5.2.5 Available phosphorus

Application of P and S significantly influenced the available P status of soil. Compared to initial available P. Available P was increased in all treatments due to application of phosphatic fertilizers. Available P status of control plot also increased compared to initial P status. Lowest P was recorded in treatment without P application and highest P was noted in treatment with high dose of P application. Available P status of soil was affected by S application also. Wiedenfeld (2011) also reported that application of elemental sulphur in calcareous soils decreased soil pH, thereby increasing the available P content in soil. The microbial oxidation of elemental sulphur produced protons which dissolved P and increased the plant available P.



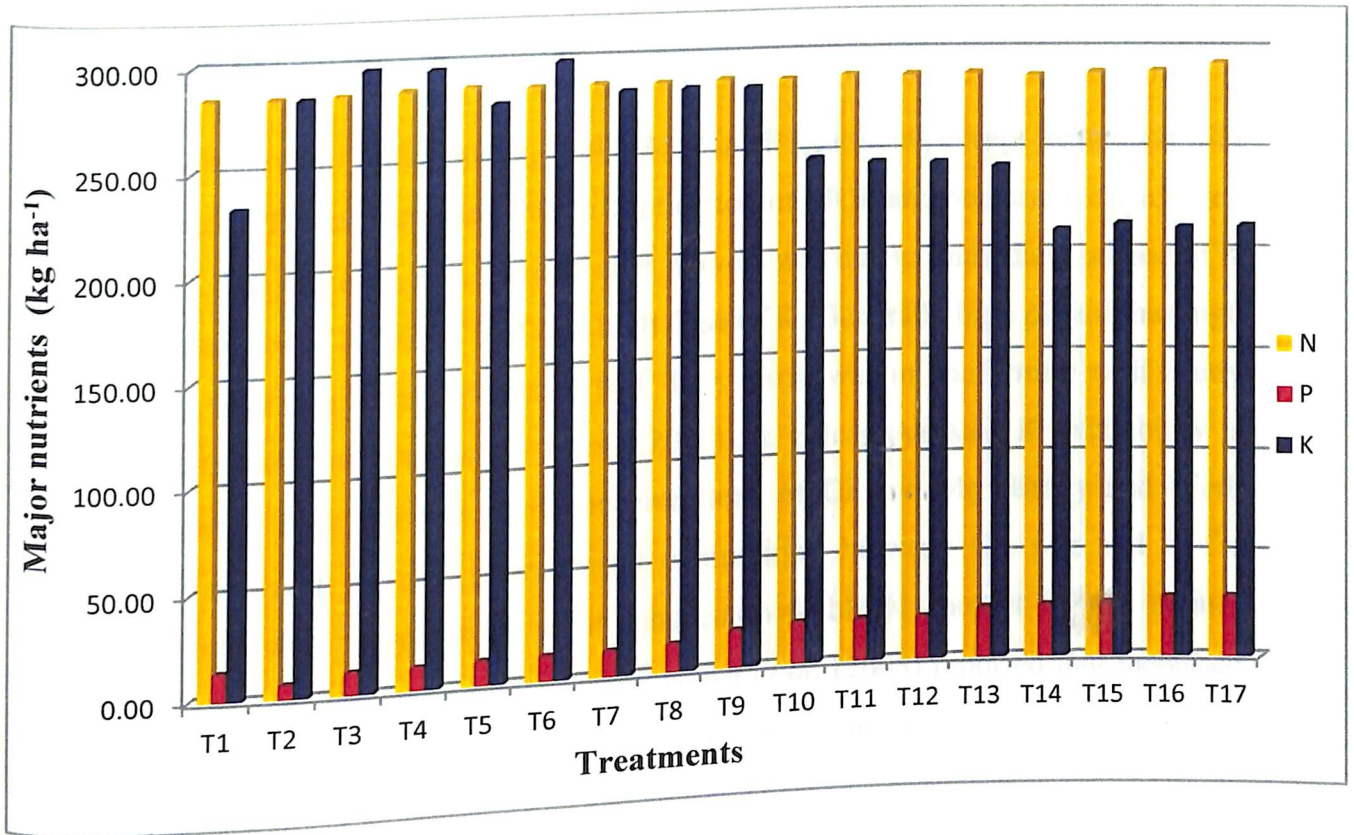


Fig 5: Effect of application of P and S on major nutrients in soil

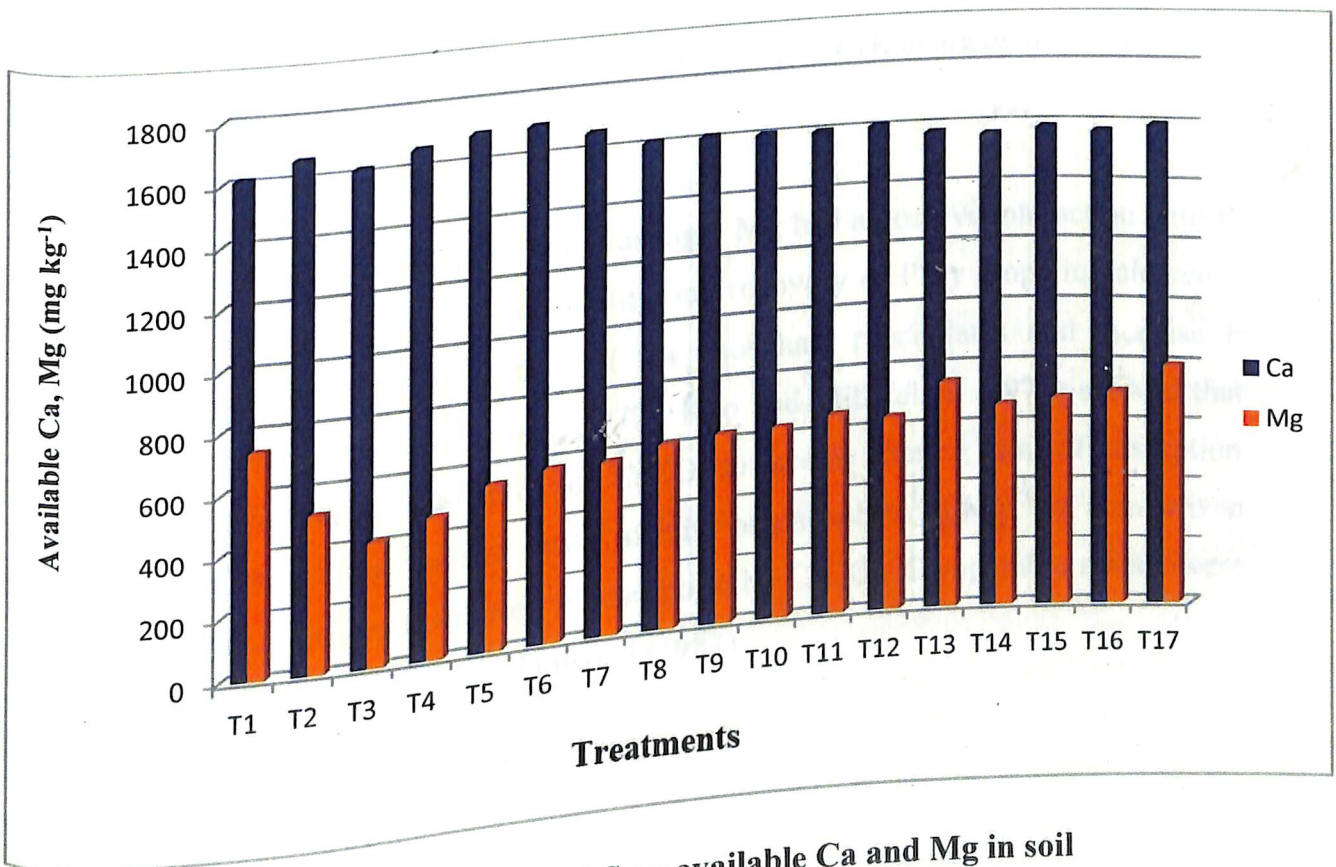


Fig 6: Effect of application of P and S on available Ca and Mg in soil

### 5.2.6 Available potassium

Compared to initial status, available K in all treatments except treatments with higher dose of P was increased due to application of MOP and potassium dihydrogen phosphate. The increase in K content after harvest was due to immediate release of K to available pool of K from FYM and high dose of K. Generally high pH soil has high Mg which decreases the K status of soil. This result was in conformity with Brady and Weil (2002) and Daliparthy *et al.* (2008). In calcareous soil, K availability is limited due to high pH and high concentrations of Ca and Mg (Brady and Weil, 2002). This might be due to increased competition between Ca, Mg and K for exchangeable sites. Application of P reduced available K content in soil mainly because of enhanced plant uptake. Skwierawska *et al.* (2008) reported that increased level of sulphur application reduced available K content in soil.

### 5.2.7 Available calcium

Experimental field had high Ca content since it is a calcareous soil with high soil pH. There was a decrease in available Ca with increased dose of P application. This might be due to the formation of calcium phosphate (Kamara *et al.*, 2011).

### 5.2.8 Available magnesium

The initial Mg content of soil was high. Mg had a positive interaction with P. Mg plays a positive role in P availability and recovery of P by crops in calcareous soils. Mg can alter the formation of Ca phosphate precipitates and increase P availability (Marion and Babbcock, 1977). Kuo and Milkelsen (1979) showed that Mg may interfere with P adsorption on  $\text{CaCO}_3$  surface by altering some of adsorption sites on  $\text{CaCO}_3$  surface, due to lower affinity of phosphate to  $\text{Mg}^{2+}$  in comparison with  $\text{Ca}^{2+}$  which causes decreased P adsorption by  $\text{CaCO}_3$ . Comparable results were reported by Al-Lami (1999) and Dowood (1982).



### 5.2.9 Available sulphur

There was a noticeable increase in available S from T<sub>1</sub> to T<sub>17</sub> due to increased dose of S application as elemental sulphur.

### 5.2.10 Available iron

Application of P and S significantly influenced Fe status of soil. Treatment having low P and high S showed highest available Fe. Heavy application of P reduced availability of Fe. This might be due to the formation of Fe-phosphate which reduced the Fe availability. This was in conformity with result of Ayed (1970). Loneragan *et al.* (1979) reported that application of large amount of P increased P: Fe ratio and led to deficiency of these nutrients.

### 5.2.11 Available manganese

Available Mn was high in treatment with low P. Application of P reduced available Mn status of soil. It may be due to the formation of insoluble Mn-phosphate. High available P induced the deficiency of Mn in maize (Adriano and Murphy, 1970) and in potato (Adriano *et al.*, 1971).

### 5.2.12 Available zinc

Application of P and S significantly affected Zn content. Highest Zn was noticed in treatment, T<sub>4</sub> (P<sub>0</sub>S<sub>2</sub> - P, 0 kg ha<sup>-1</sup> and S, 20 kg ha<sup>-1</sup>) and lowest was in treatment, T<sub>16</sub> (P<sub>3</sub>S<sub>2</sub>- P, 90 kg ha<sup>-1</sup> and S, 20 kg ha<sup>-1</sup>). Available Zn decreased with increase in P. P forms chemical bond with Zn at high levels of P and P binds large quantity of Zn resulting in P induced Zn deficiency that leads to reduced shoot growth. The findings of Adriano and Murphy (1970) in maize and Adriano *et al.* (1971) in potato were in conformity with the results obtained. They found that high P induced Zn deficiency. This result was in conformity with Modaihsh *et al.* (1989) in calcareous soils.

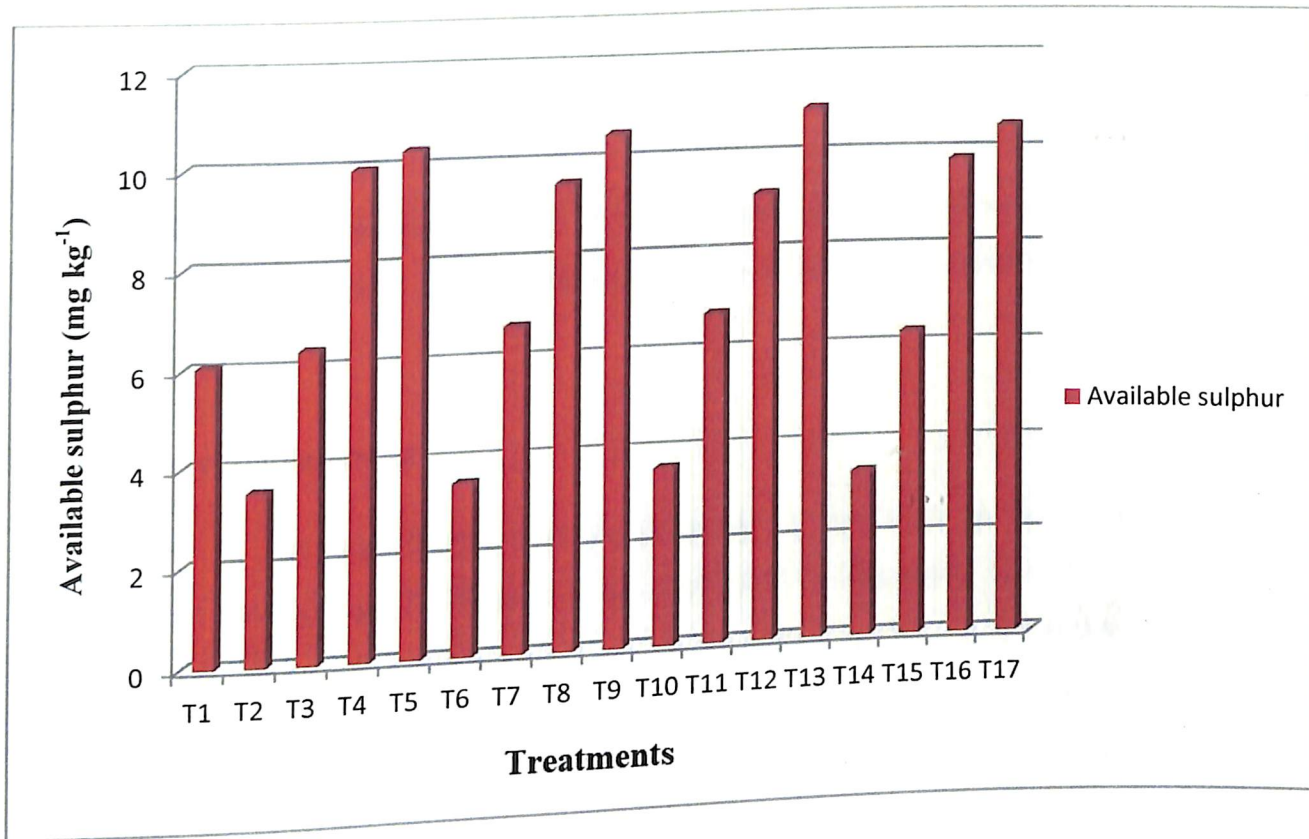


Fig 7: Effect of application of P and S on available S in soil

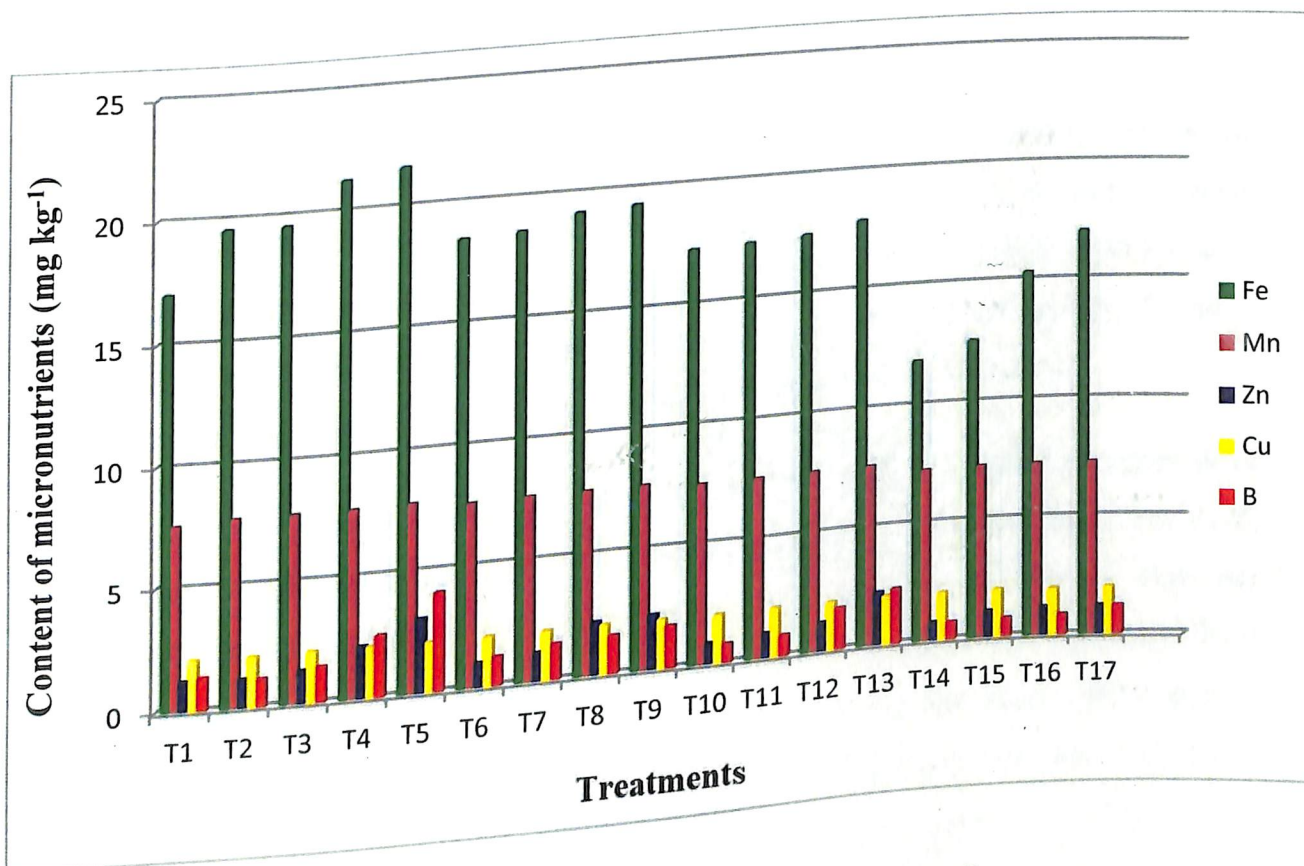


Fig 8: Effect of application of P and S on micronutrients in soil

### 5.2.13 Available copper

Effect of available Cu was non-significant for main effect and interaction effect. This result was in conformity with Modaihsh *et al.* (1989) in calcareous soils.

### 5.2.14 Available boron

A significant positive decrease in available B was noticed in treatment having high P. Highest B was observed in treatment, T<sub>5</sub> (P<sub>0</sub>S<sub>3</sub>: P, 0 kg ha<sup>-1</sup> and S, 30 kg ha<sup>-1</sup>) and lowest was in treatment, T<sub>14</sub> (P<sub>3</sub>S<sub>0</sub>: P, 90 kg ha<sup>-1</sup> and S, 0 kg ha<sup>-1</sup>). There was antagonistic interaction between B and P. This was in conformity with results of Bingham *et al.* (1958) in citrus and May and Pritts (1993) in strawberry.

## 5.3 EFFECT OF APPLICATION OF P AND S ON PLANT NUTRIENT CONTENT AND UPTAKE

### 5.3.1 Nutrient content

Nutrient content in groundnut was significantly influenced by the application of different levels of P and S. Significantly highest N, P, and S content were observed in treatments with high dose of P (90 kg ha<sup>-1</sup>) and S (30 kg ha<sup>-1</sup>). Dhage *et al.* (2014) reported that application of P and S increased N, P and S contents in groundnut plant. Highest K content was noted in T<sub>5</sub> (P<sub>0</sub>S<sub>3</sub>).

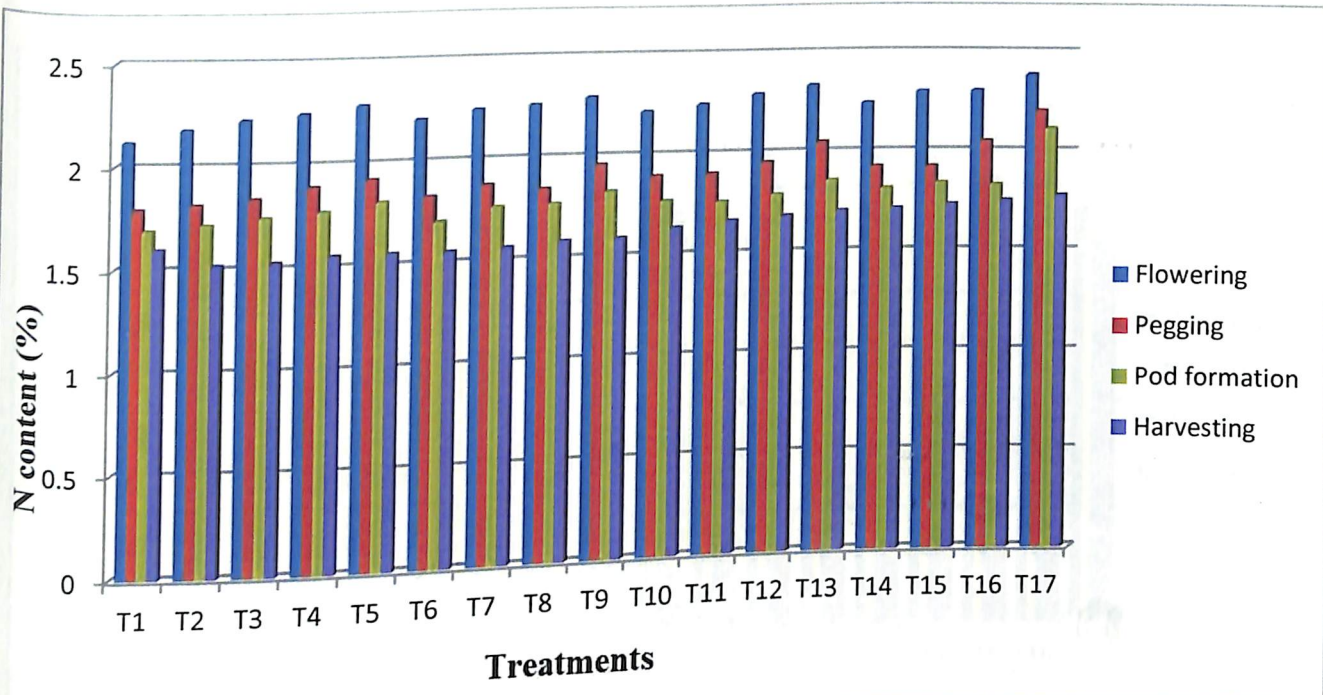
P<sub>1</sub>S<sub>0</sub> (P, 60 kg ha<sup>-1</sup> and S, 0 kg ha<sup>-1</sup>) showed highest Ca content at flowering (1.99 per cent) stage. At pegging stage (1.62 per cent) treatment P<sub>0</sub>S<sub>0</sub> showed highest Ca content. At pod formation (1.21 per cent) and harvest (1.03 per cent) stages, P<sub>0</sub>S<sub>3</sub> showed highest Ca content. Low P status of soil decreased N, P and Ca contents in 30 day old groundnut plants (Basha and Rao, 1980). Higher content of N, P, S and K in these treatments helped root growth and enhanced uptake of nutrients and increased dry matter production and yield. The findings of Nakagawa *et al.* (1981) and Patel *et al.* (1981) also supported the result. Highest Mg

content was observed in T<sub>17</sub> (P<sub>3</sub>S<sub>3</sub>). Mg content in plant increased with increased dose of sulphur. This is in conformity with Barczak (2010). He also reported increase in total plant Mg content with increased dose of S application. Mg content also showed an increasing trend with increased dose of P. Mg showed positive interaction with N and P (Ranade and Malvi, 2011). The haulm retains magnesium accumulated during vegetative growth indicating its utilization for structural and developmental processes, with less translocation of magnesium towards reproductive parts; hence kernels contain least amount of magnesium compared to haulm (Babu *et al.*, 2007). Plant S content increased with increased dose of S application. Dhage *et al.* (2014) reported that S application increased the plant S content in soybean.

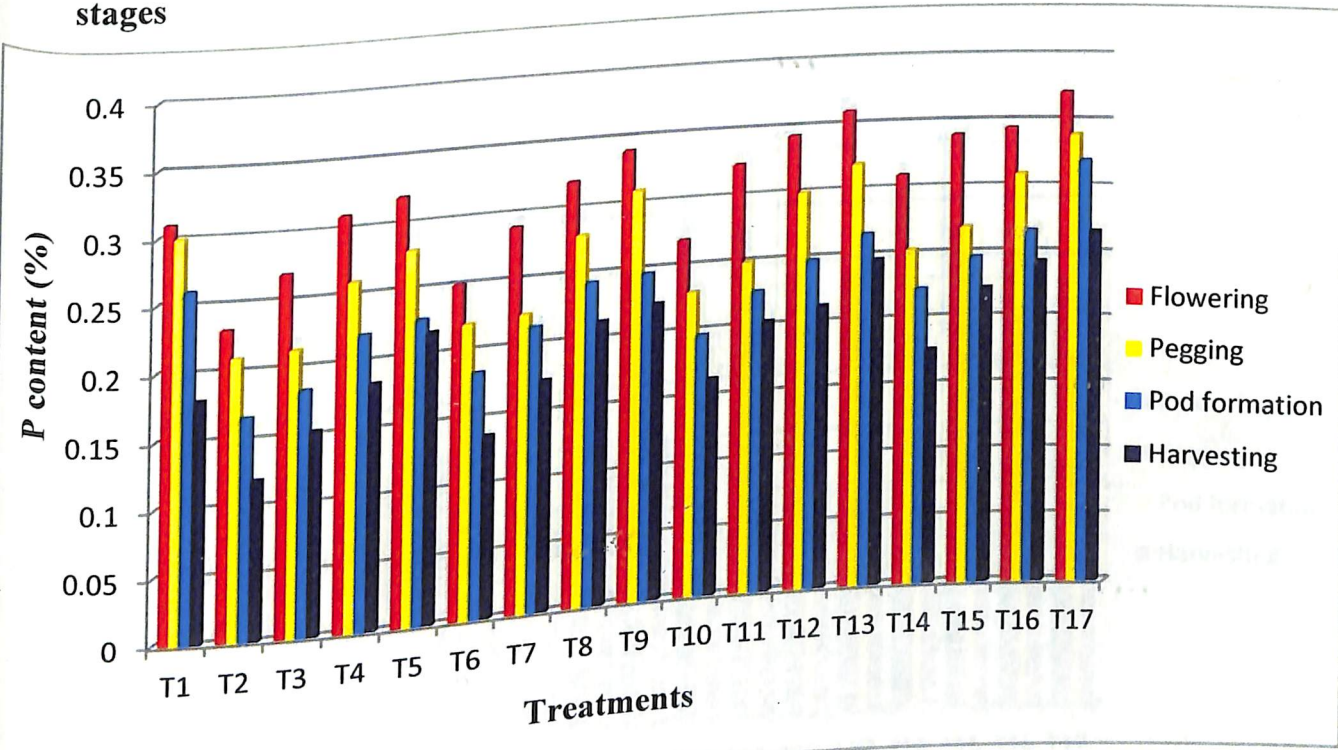
T<sub>5</sub> (P<sub>0</sub>S<sub>3</sub>) showed highest Fe, Mn, and Zn content. Soliman *et al.*, (1992) reported that application of sulphur increased Mn content in corn plants grown in calcareous soils. Cu content was highest in T<sub>2</sub> (P<sub>0</sub>S<sub>0</sub>) at flowering stage and in T<sub>5</sub> (P<sub>0</sub>S<sub>3</sub>) at pegging and pod formation stages. There was no significant effect for the application of P and S on plant Cu content at harvesting stage.

Nutrient content in plant decreased from flowering to harvest stage. Highest nutrient content was noticed in flowering stage and it gradually decreased towards harvest stage. The findings of Chahal *et al.* (1983) were in conformity with this result. Reddy and Murthy (1985) observed that plant N content decreased as crop grew older.

N, P and S content by the kernel increased with increased dose of P and S. The result was in conformity with Das (2017). Ca content in shell increased with increased dose of P. This result was in conformity with Amruth *et al.* (2018). The content of Fe, Mn and Zn in kernel and shell increased with increased dose of sulphur. This result was in conformity with the findings of Kharol *et al.* (2014). Jankowski *et al.* (2014) also concluded that application of S increased Zn and Mn content in rapeseed.



**Fig 9: Effect of application of P and S on nitrogen content in plant at different stages**



**Fig 10: Effect of application of P and S on phosphorus content in plant at different stages**



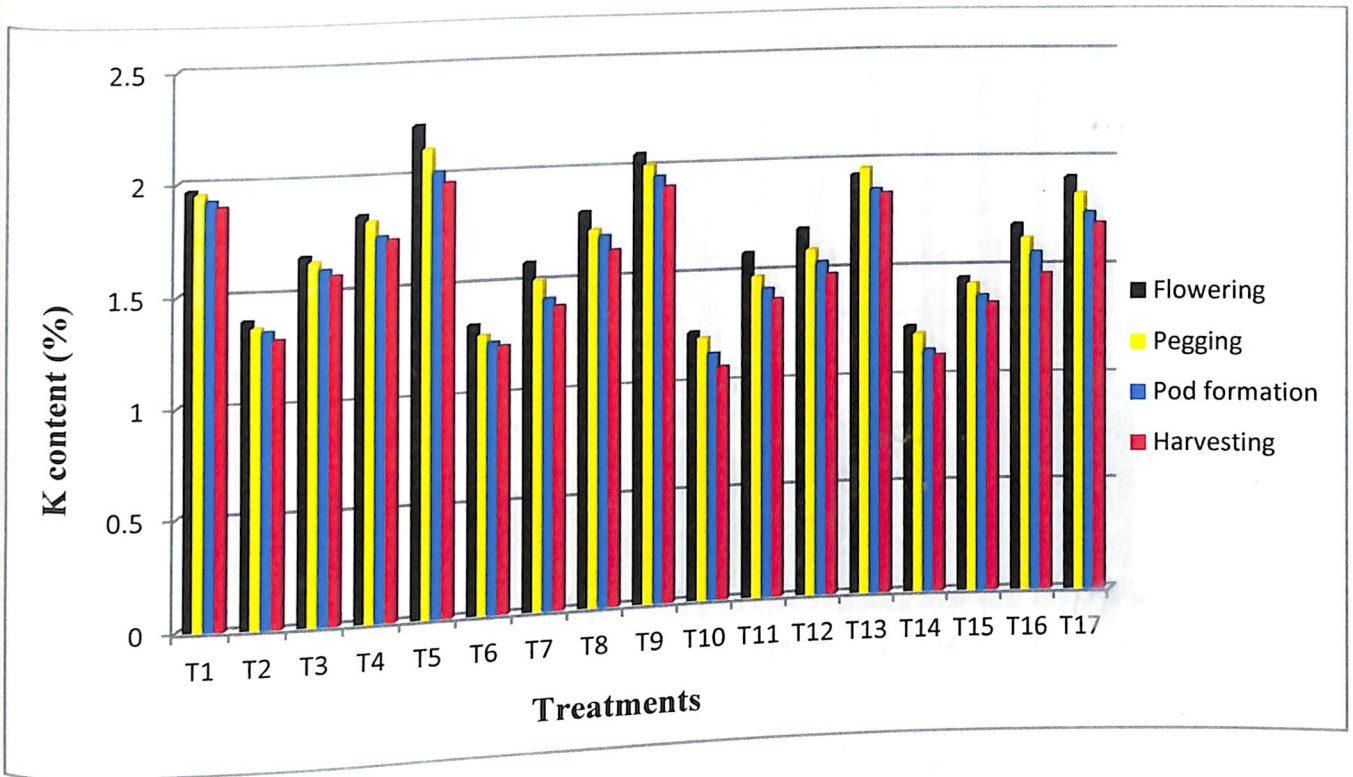


Fig 11: Effect of application of P and S on potassium content in plant at different stages

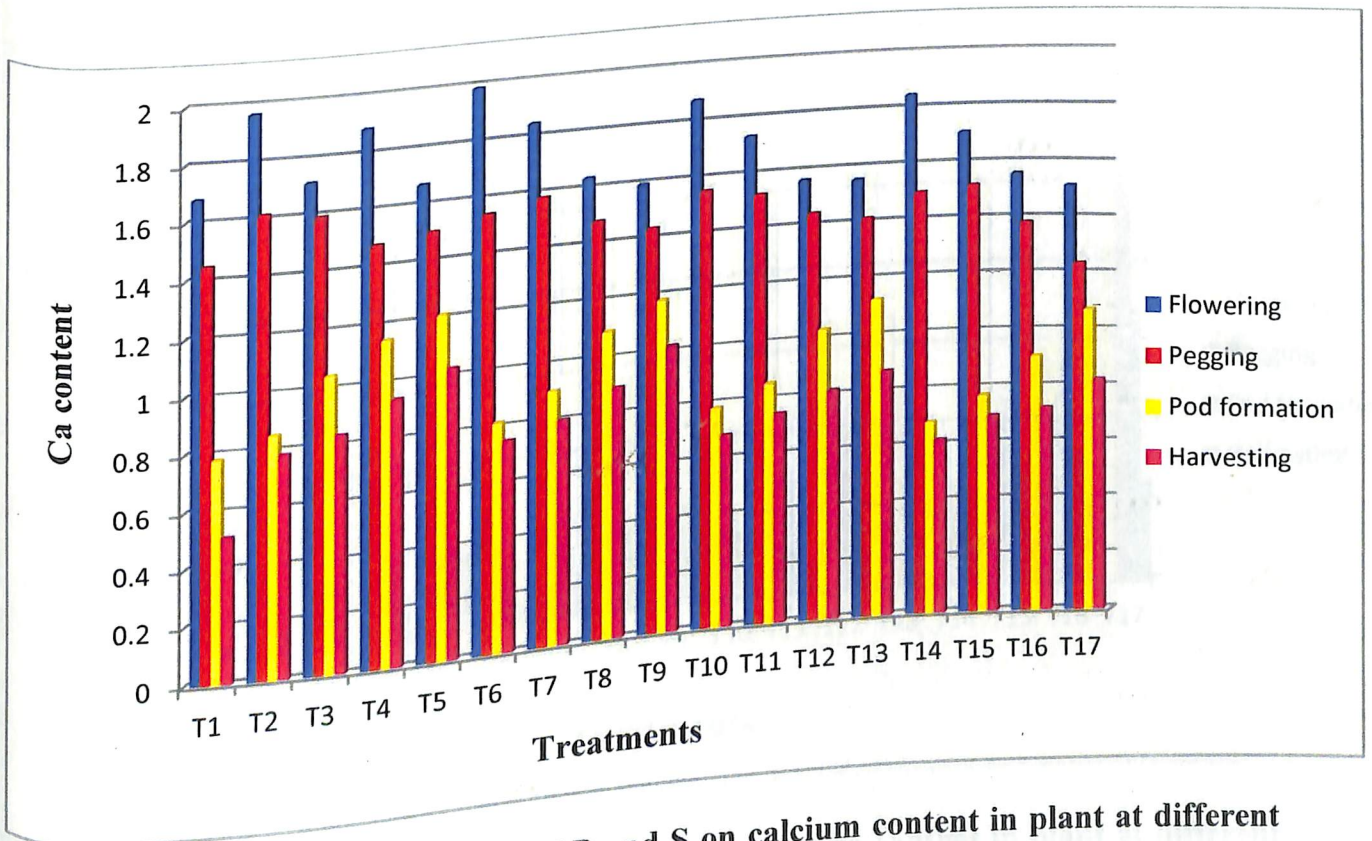
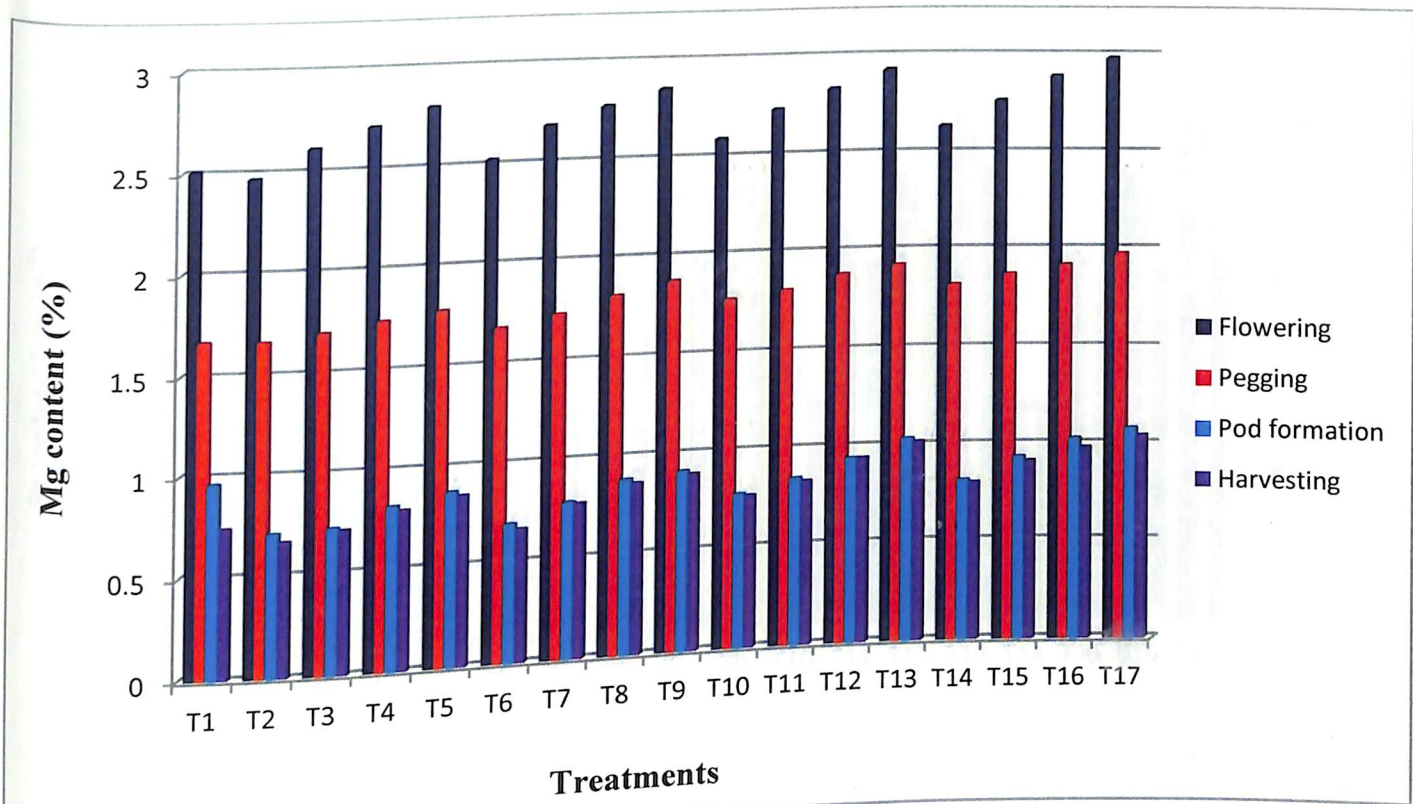
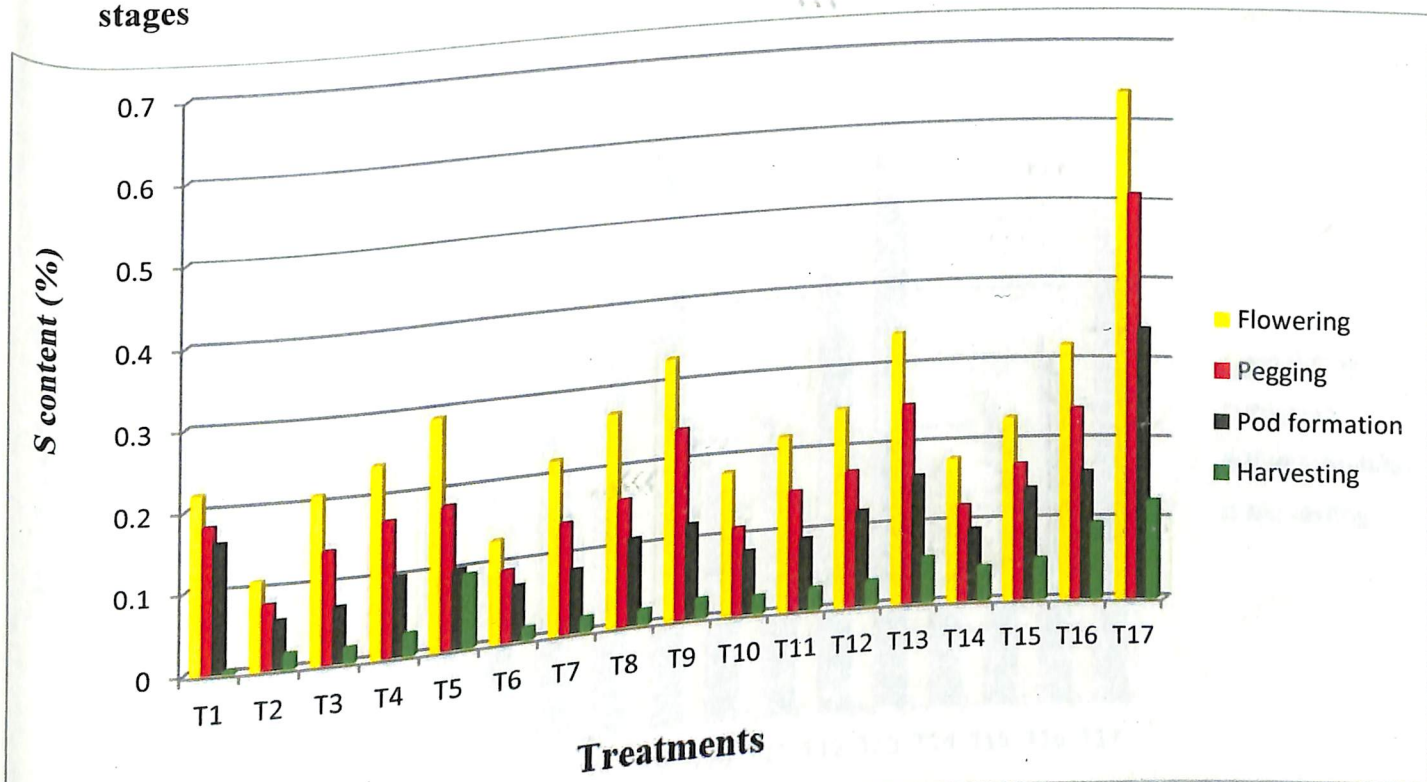


Fig 12: Effect of application of P and S on calcium content in plant at different stages



**Fig 13: Effect of application of P and S on magnesium content in plant at different stages**



**Fig 14: Effect of application of P and S on sulphur content in plant at different stages**



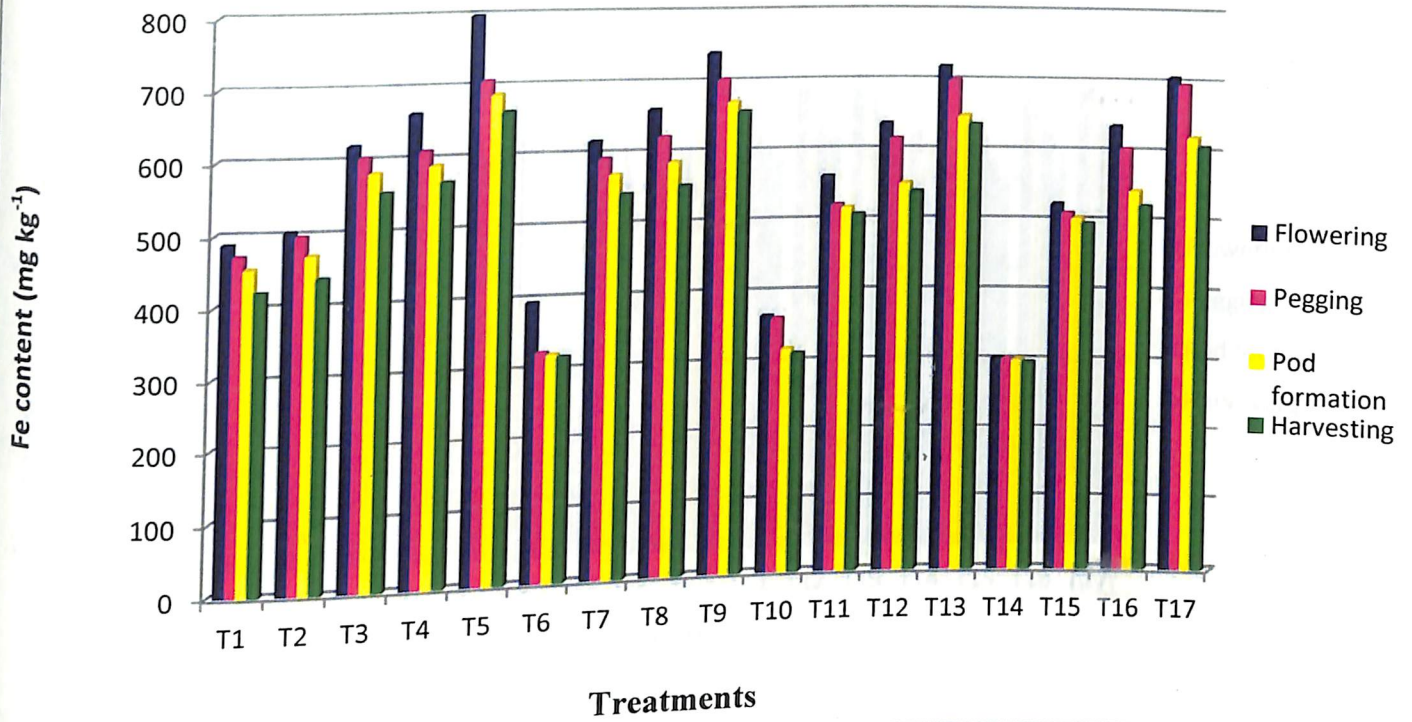


Fig 15: Effect of application of P and S on iron content in plant at different stages

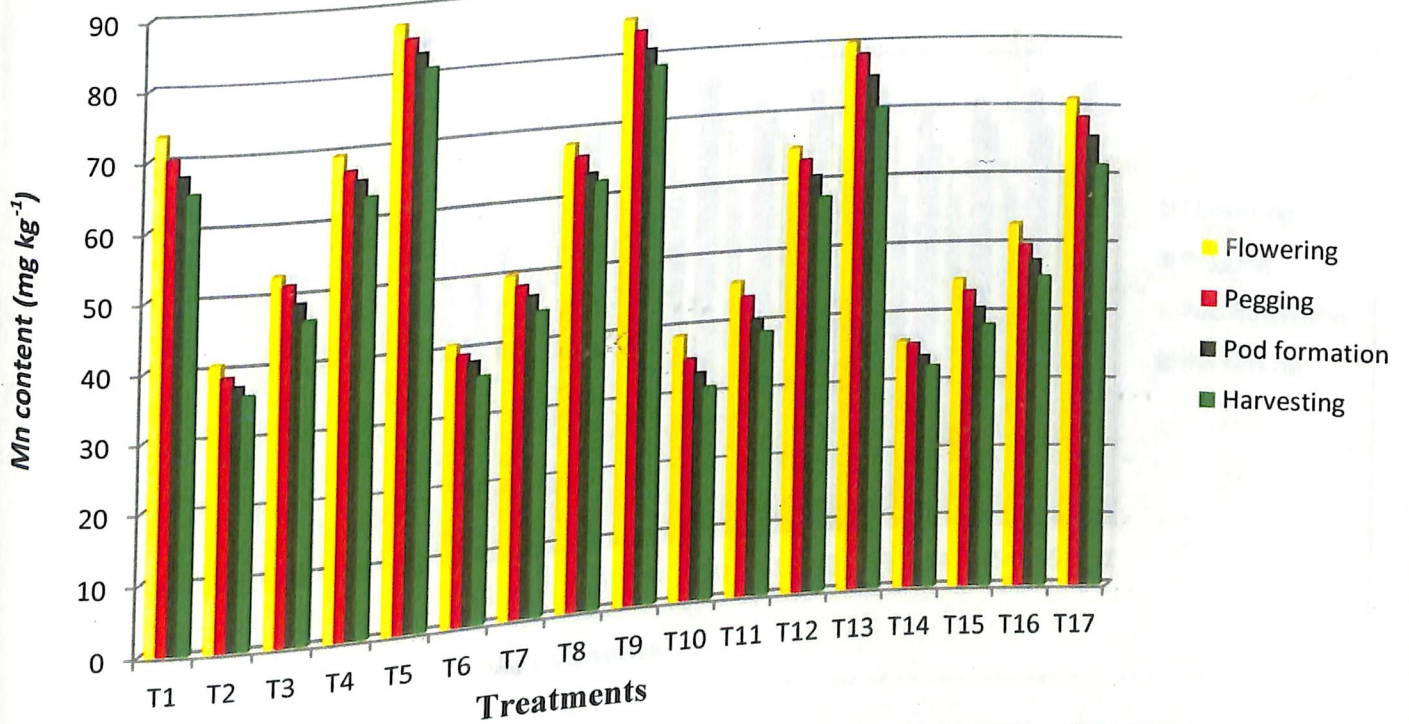


Fig 16: Effect of application of P and S on manganese content in plant at different stages



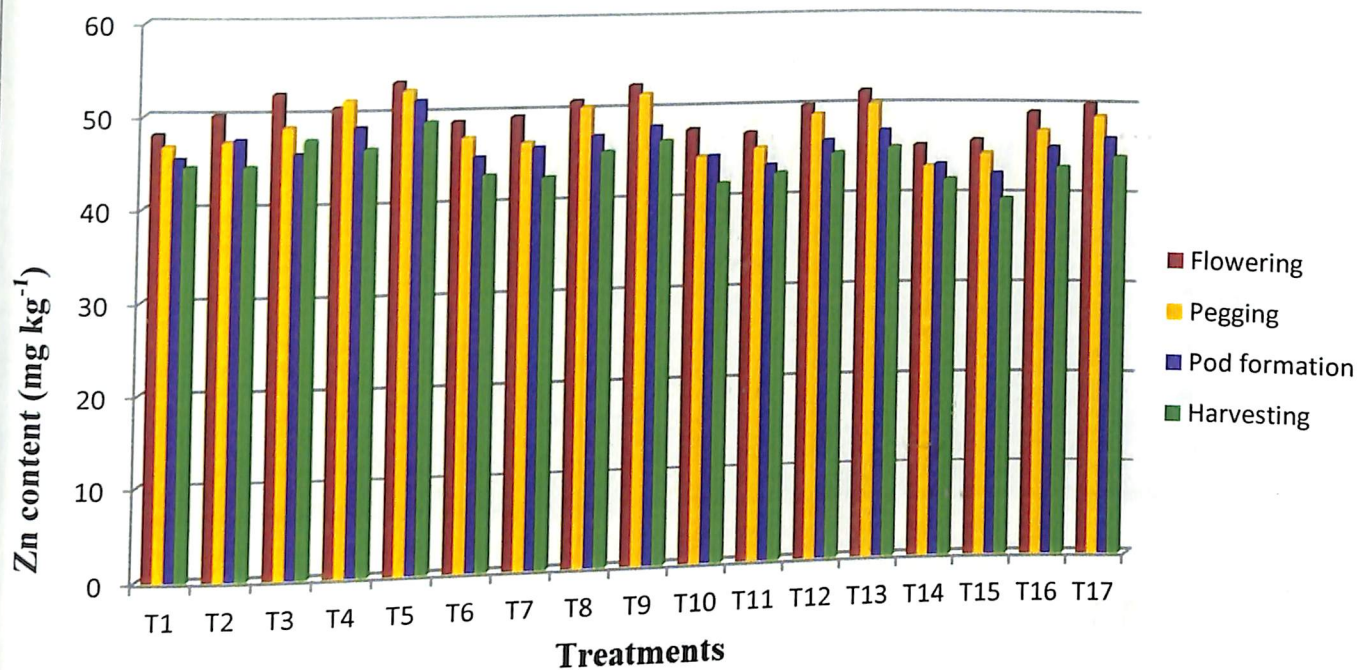


Fig 17: Effect of application of P and S on zinc content in plant at different stages

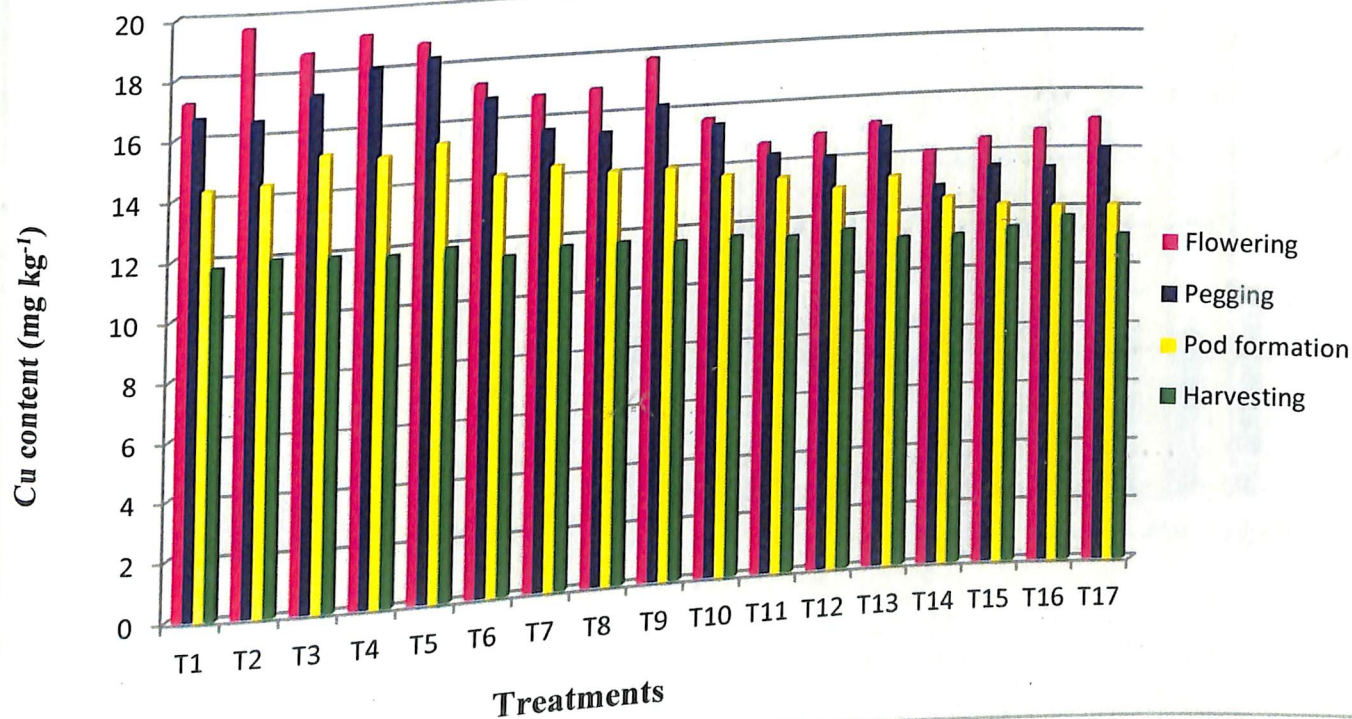


Fig 18: Effect of application of P and S on copper content in plant at different stages

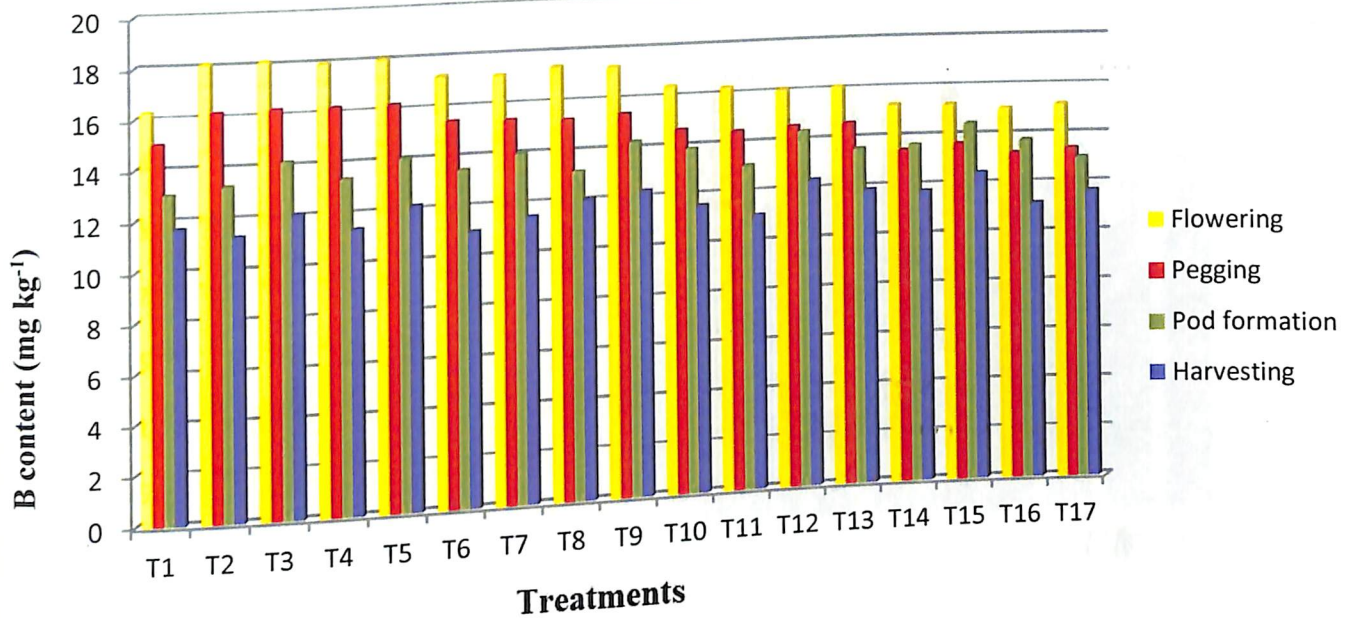


Fig 19: Effect of application of P and S on boron content in plant at different stages

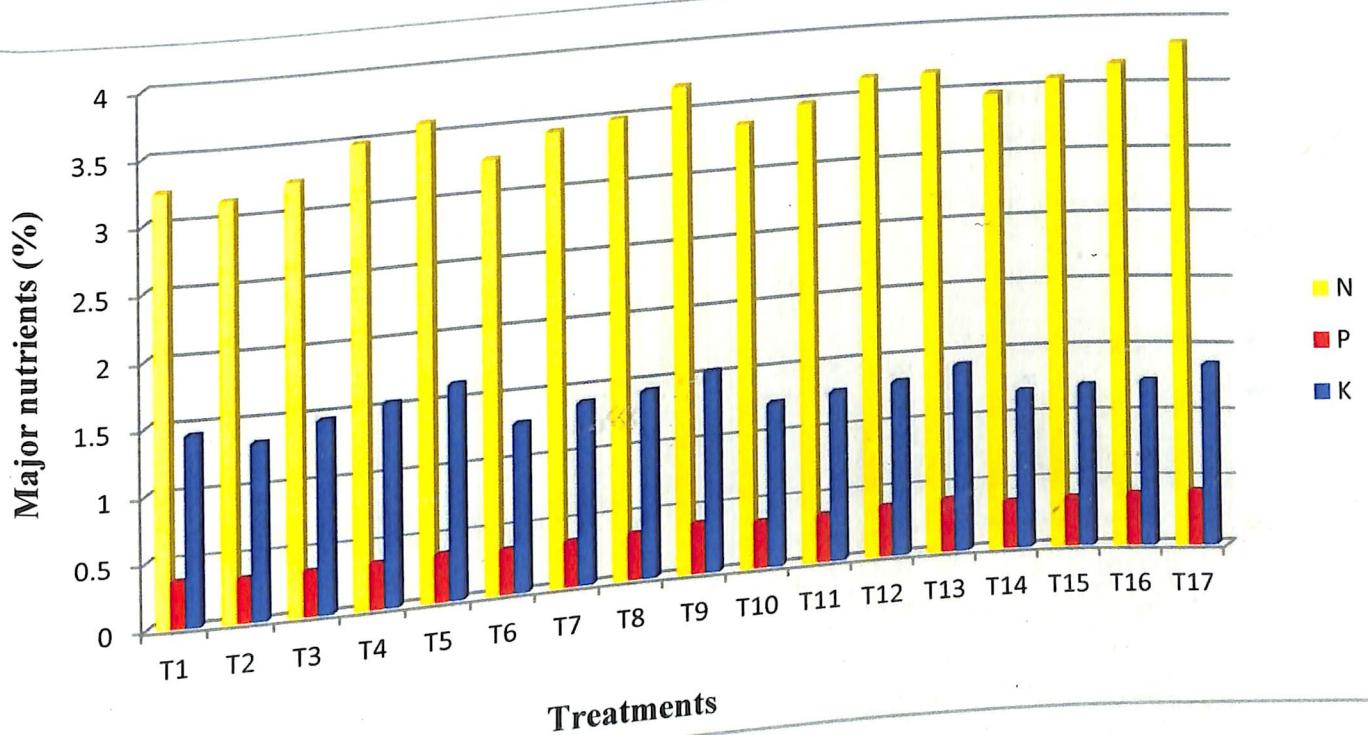


Fig 20: Effect of application of P and S on content of major nutrients in kernel



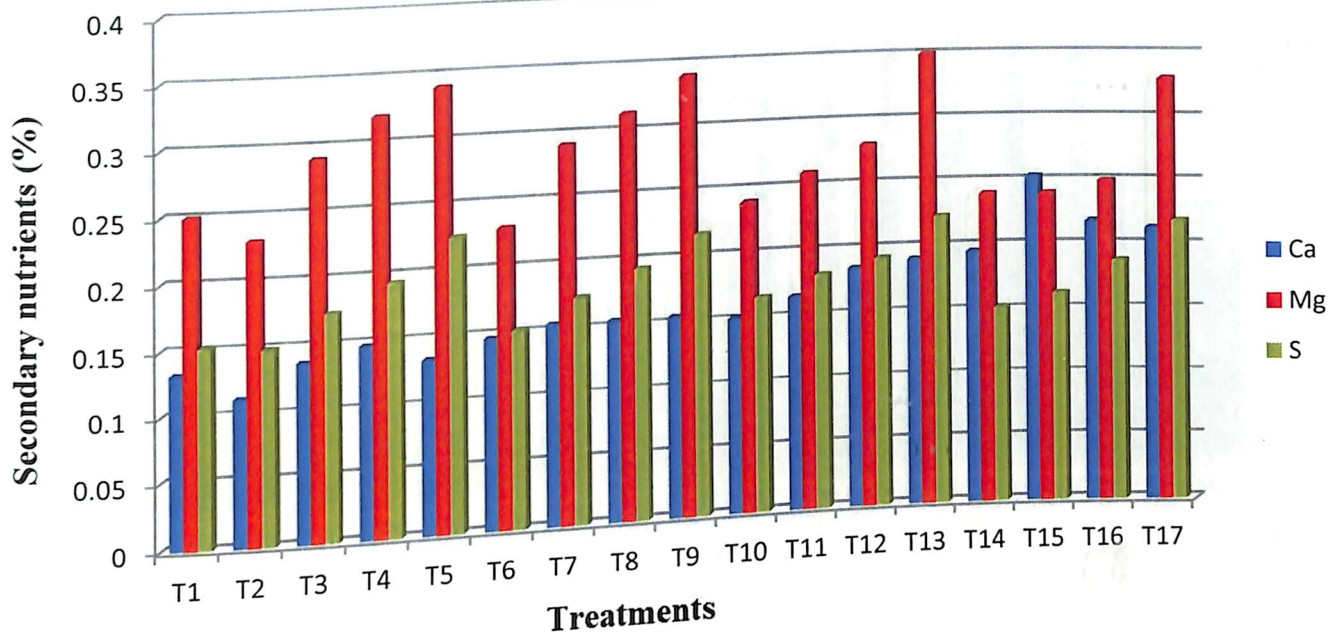


Fig 21: Effect of application of P and S on content of secondary nutrients in kernel

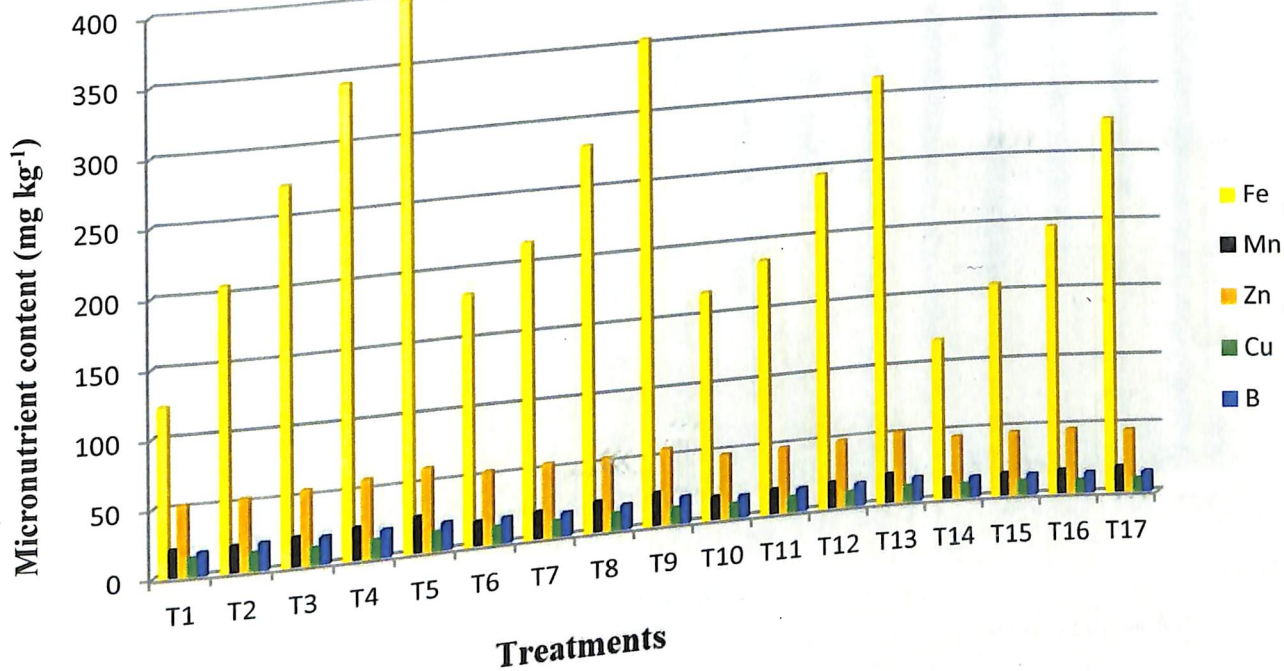


Fig 22: Effect of application of P and S on content of micro nutrients in kernel

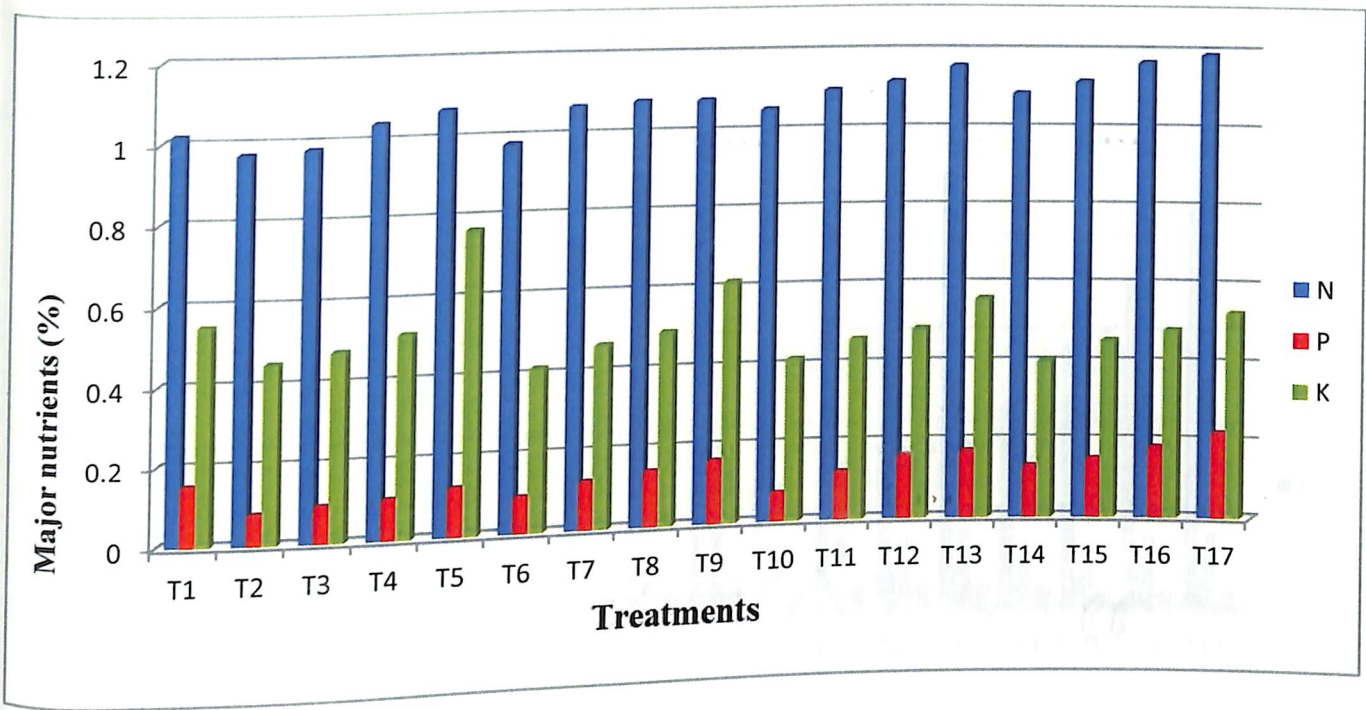


Fig 23: Effect of application of P and S on content of major nutrients in shell

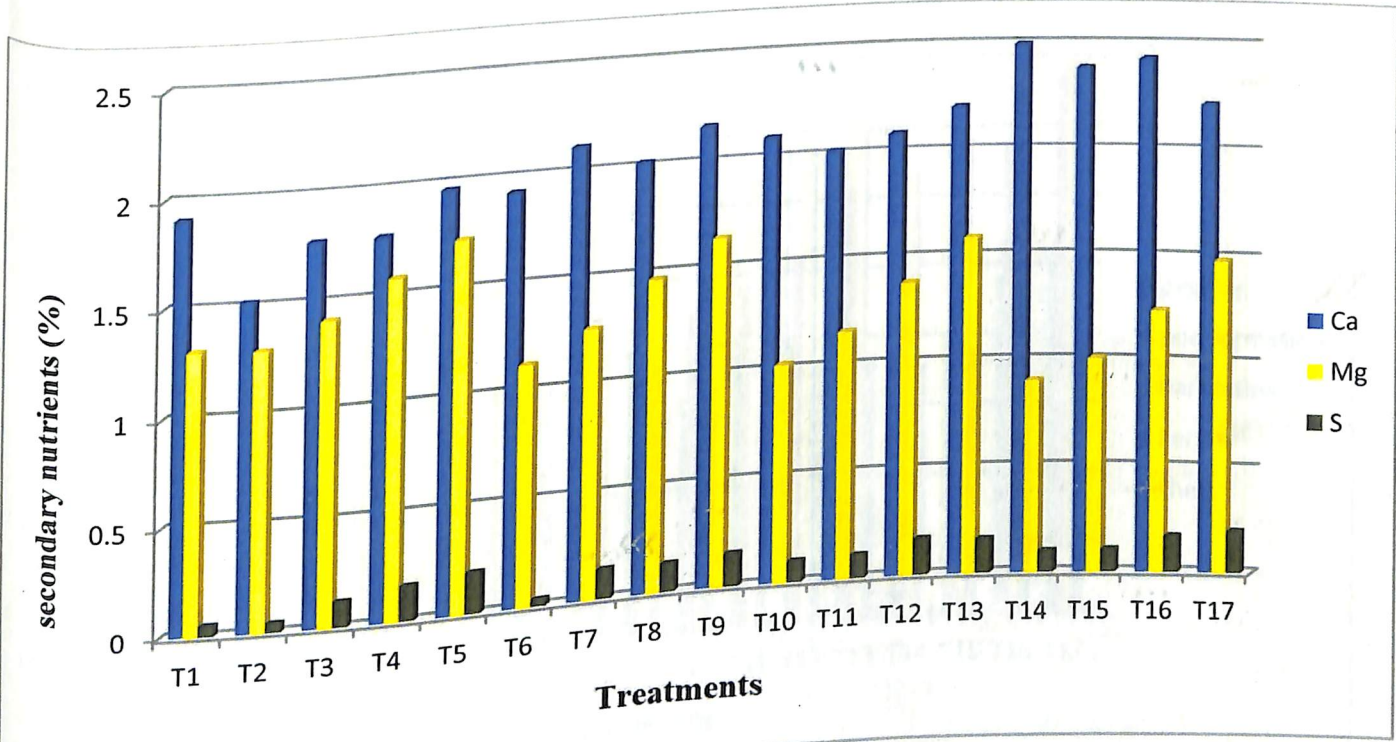


Fig 24: Effect of application of P and S on content of secondary nutrients in shell



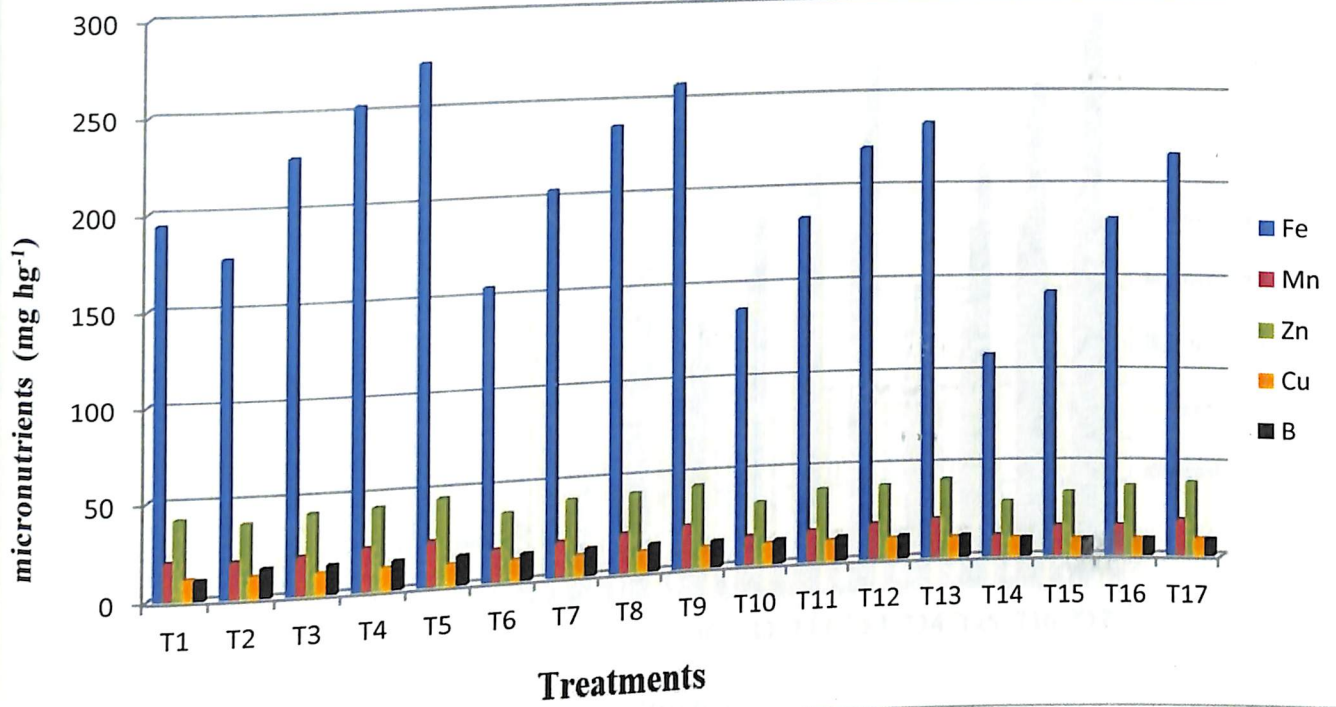


Fig 25: Effect of application of P and S on content of micro nutrients in shell

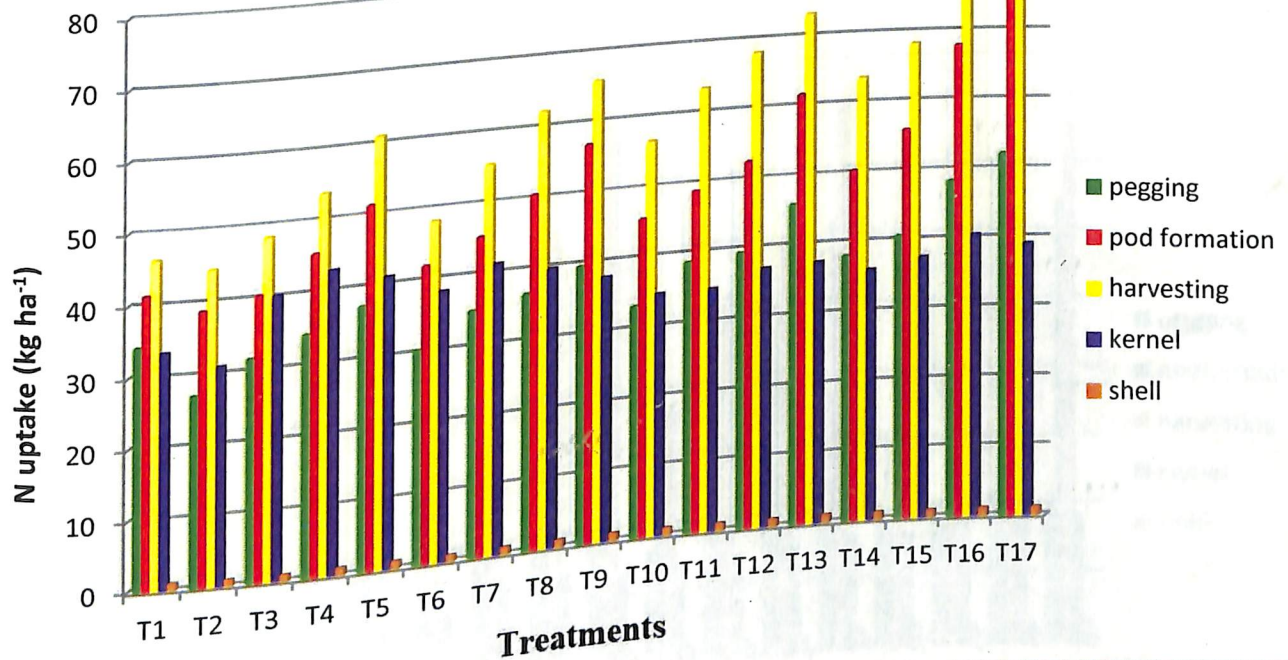
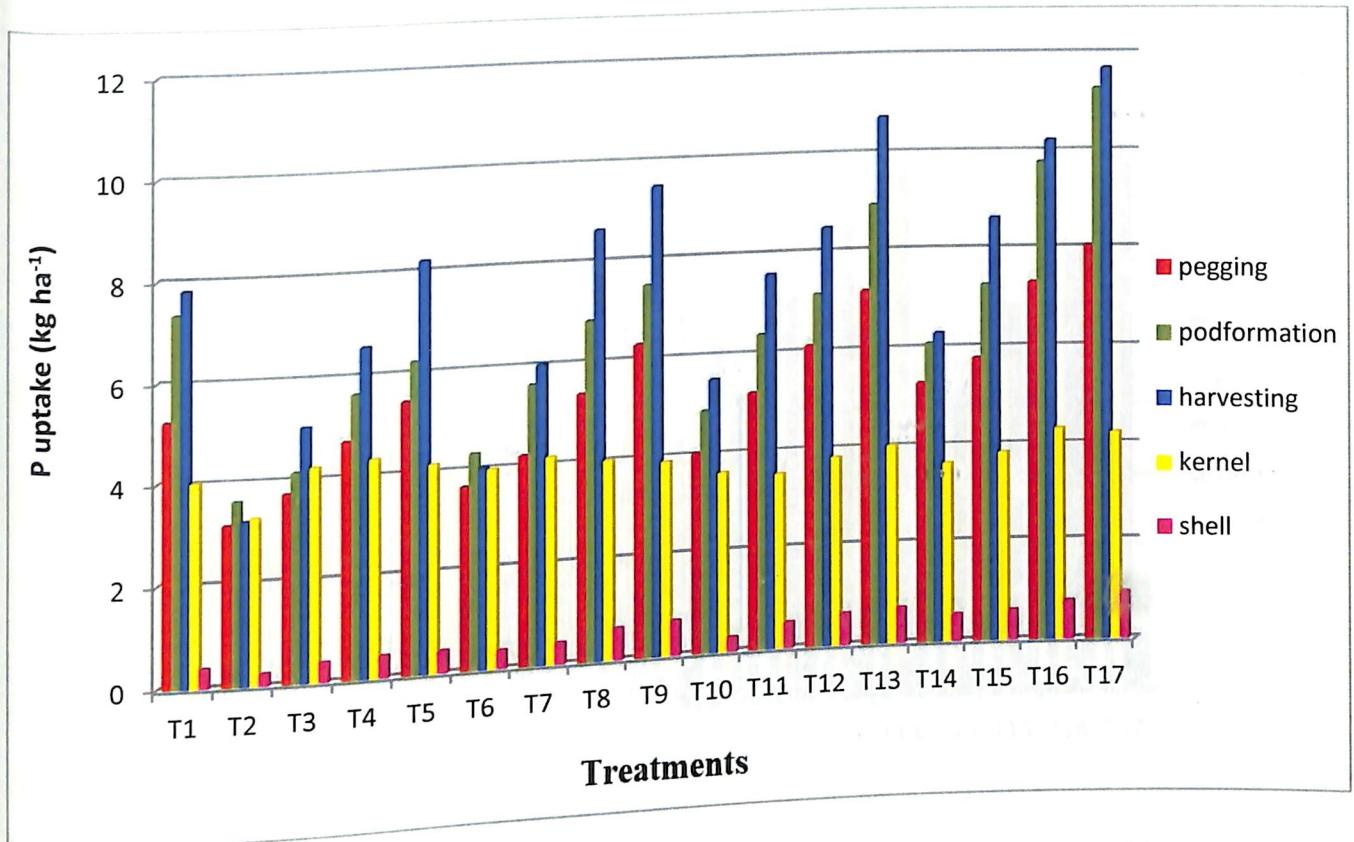
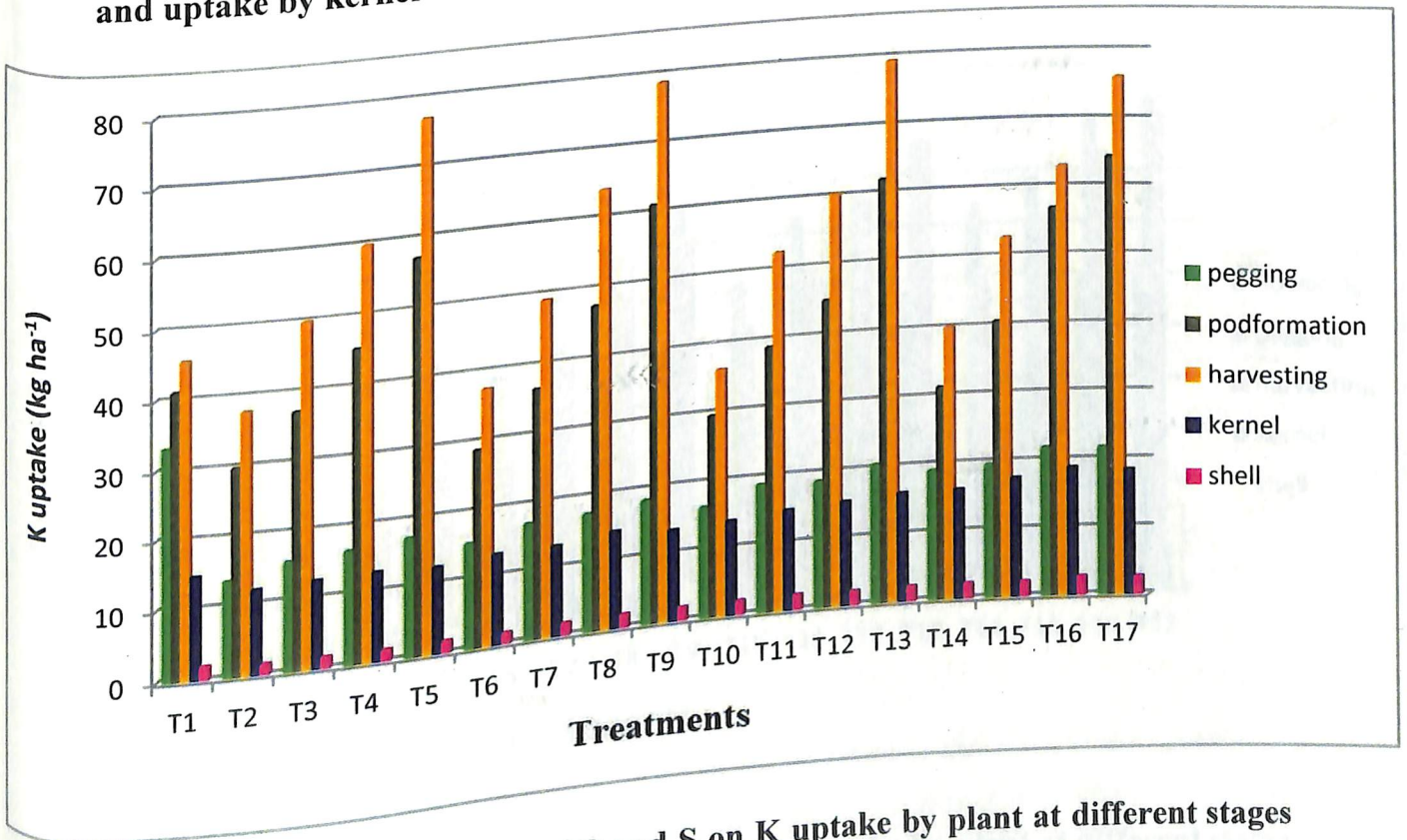


Fig 26: Effect of application of P and S on N uptake by plant at different stages and uptake by kernel and shell

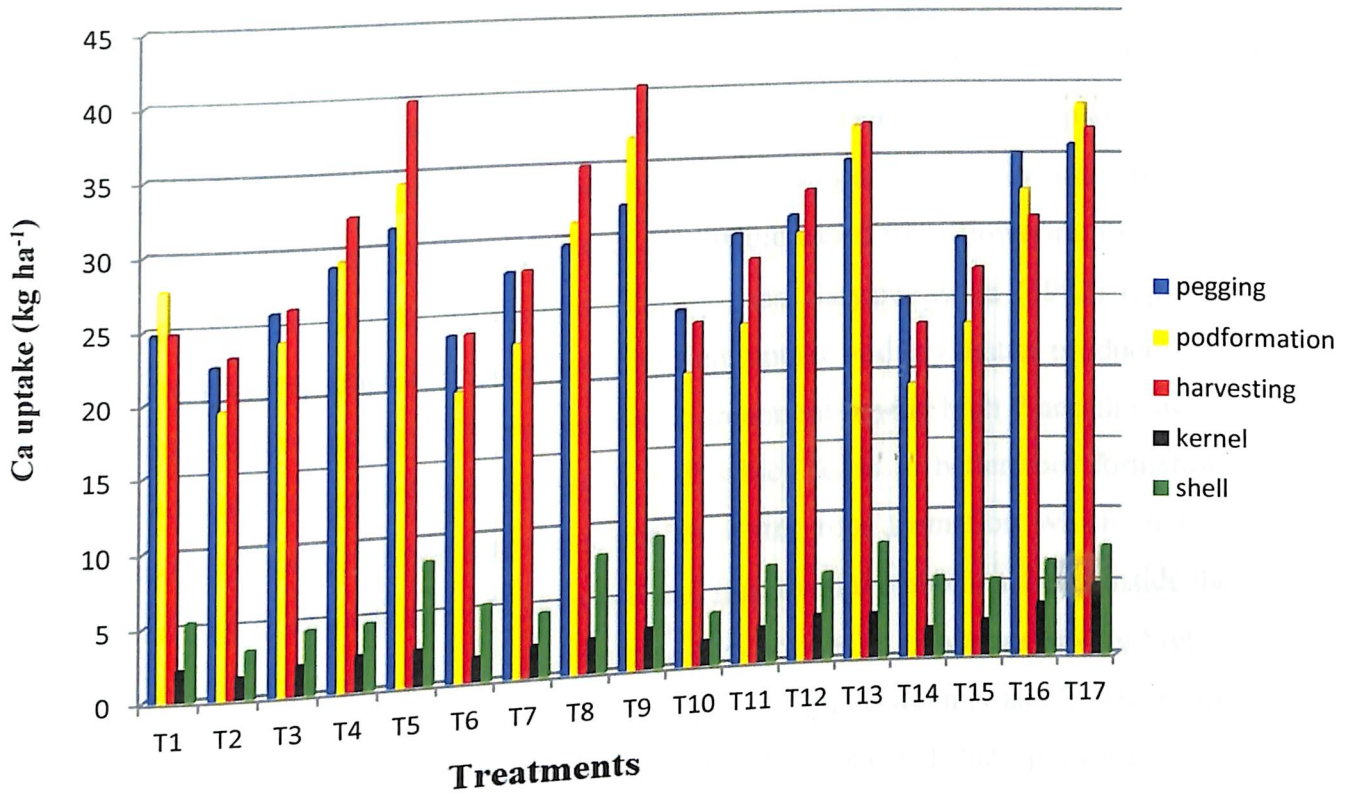


**Fig 27: Effect of application of P and S on P uptake by plant at different stages and uptake by kernel and shell**

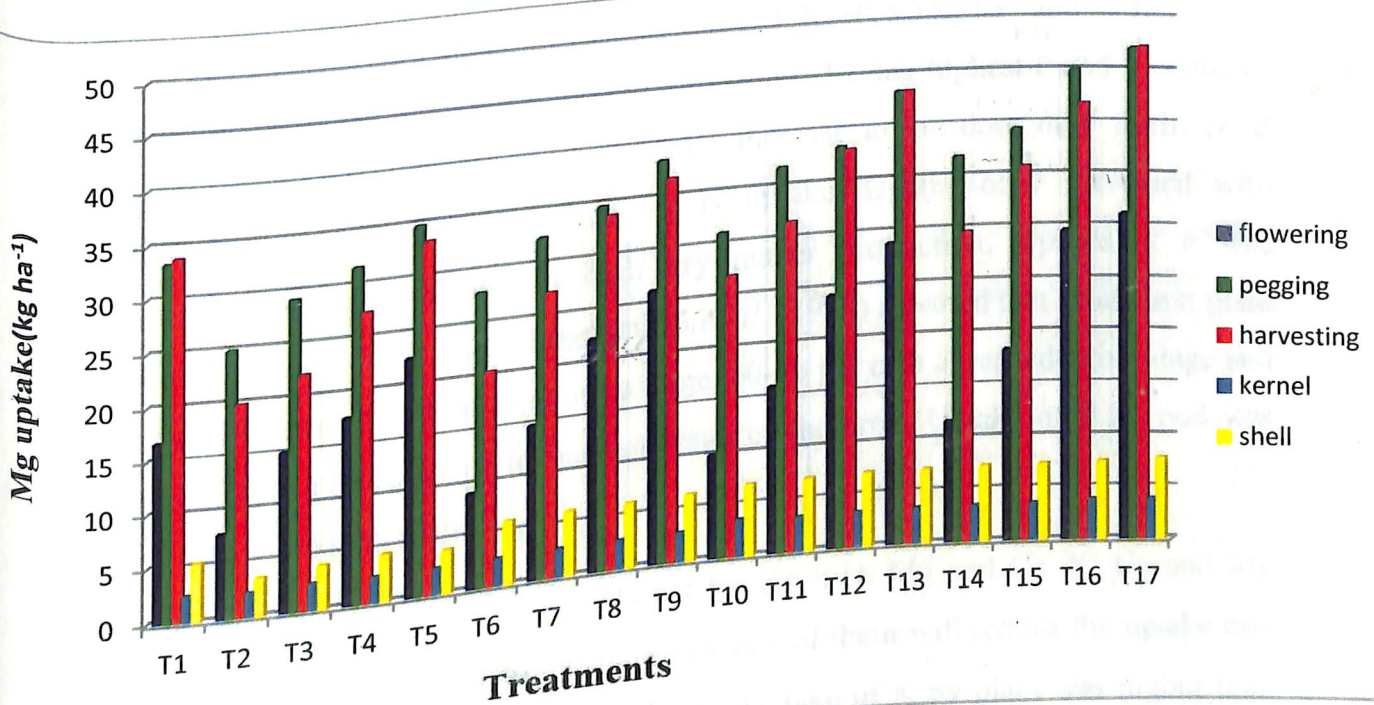


**Fig 28: Effect of application of P and S on K uptake by plant at different stages and uptake by kernel and shell**





**Fig 29: Effect of application of P and S on Ca uptake by plant at different stages and uptake by kernel and shell**



**Fig 30: Effect of application of P and S on Mg uptake by plant at different stages and uptake by kernel and shell**

### 5.3.2 Nutrient uptake

The uptake of nutrients is associated with the metabolic activities of the plant and depends on the concentration and distribution of nutrient ions in the plant system (Manasa *et al.*, 2015). Uptake of nutrients at various stages was significantly affected by application of P and S. The uptake of N was increased from flowering to harvest with increased dry matter production. Yakadri and Satyanarayana (1992) reported that there is a close relationship between nutrient uptake and dry matter production in groundnut. Higher uptake of N was noticed in treatment having high P and this might be due to the fixation of N in presence of P which leads to better root formation. Increased dry matter production was due to better root formation which in turn enhanced higher absorption of N from soil and improved metabolic activity inside the plant (Laxminarayana, 2004). Higher uptake was noticed in treatment having high S application. This might be due to synergistic interaction between N and S. The result is in conformity with Nasreen and Huq (2002). They reported that application of S significantly increased N uptake in sunflower. N, P and S uptake by the kernel increased with increased dose of P and S. The result was in conformity with Das (2017).

Uptake of P by groundnut was highest in  $P_3S_3$  (P, 90 kg ha<sup>-1</sup> and S, 30 kg ha<sup>-1</sup>) at all stages. Highest uptake was noted in treatment having highest P and S contents. The availability of nutrients increased with increase in the dose of P fertilizer. P uptake by the plant was less than the N uptake. Uptake of P increased with advancement in growth due to high dry matter production. Uptake of P was maximum at pod setting stage. Loganathan *et al.* (1996) reported that groundnut plant absorbed 10 per cent of P at vegetative stage, 40-50 per cent at reproductive stage and remaining P at reproductive to harvest stage of the crop. Uptake of P by pod was higher than uptake by plant.

Uptake of K was less in treatment having high Mg and Ca. K, Ca and Mg compete with each other and addition of any one of them will reduce the uptake rate of the other two (Ranade and Malvi, 2011). Uptake of K by plant was higher than uptake by pods. The haulm retains major part of K accumulated during vegetative

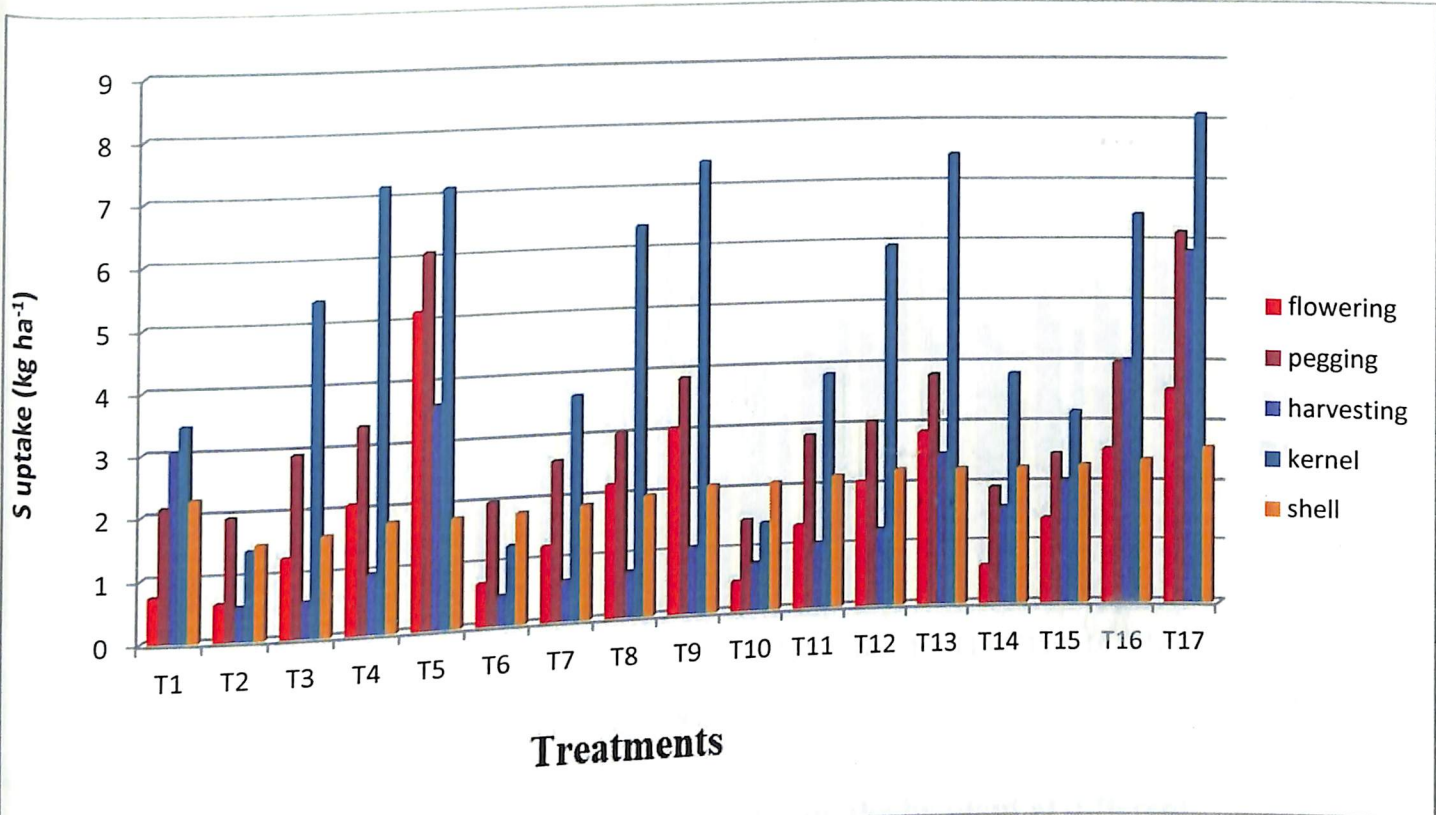


growth indicating its utilization for structural and developmental processes and allowed little translocation of potassium towards reproductive parts and hence kernel contains less amount of K (Yakadri and Satyanarayana, 1992).

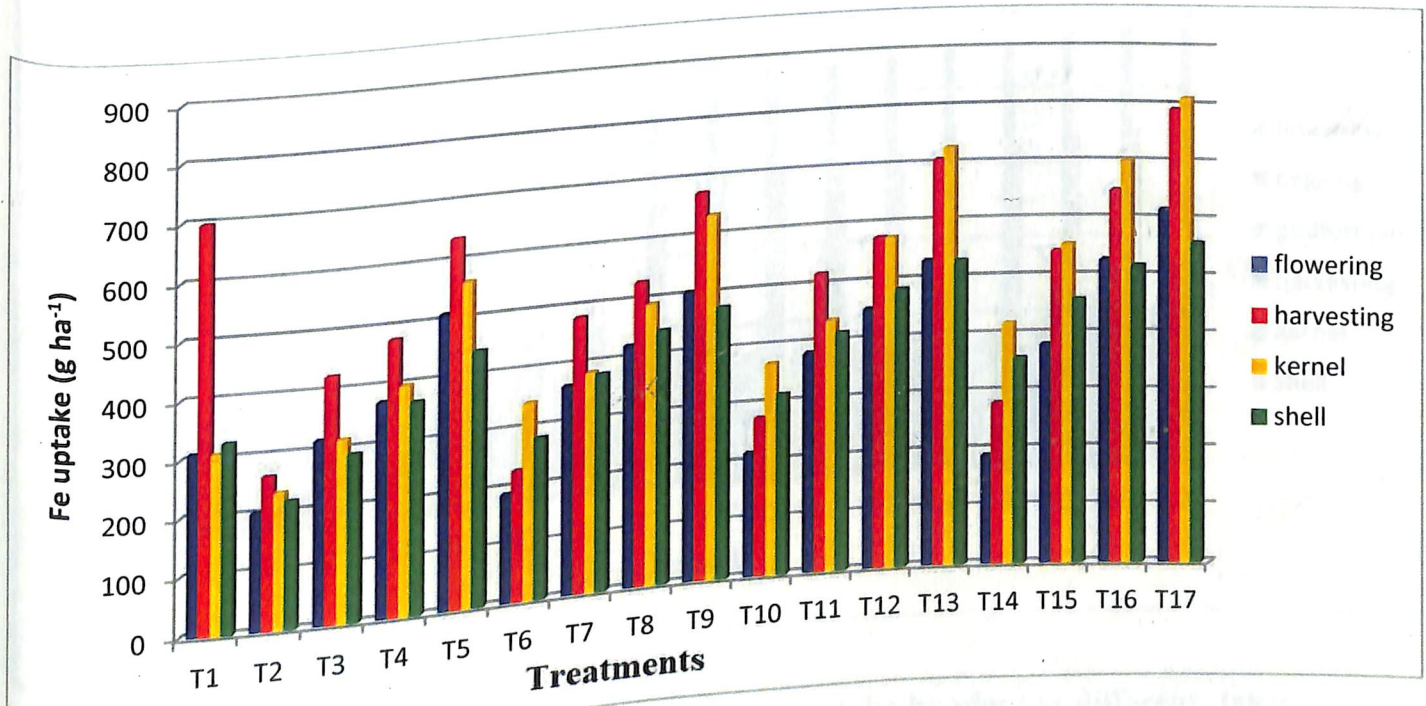
Uptake of Ca and Mg increased with advance in the age of the crop due to the increased dry matter production. Groundnut crop was a heavy feeder of Ca. (Sarkar *et al.*, 1999). At harvest, Ca and Mg uptake was more in plant than in pod. Ca taken up by the plant will remain in the leaf tissues, and will not move from the leaves to the developing pods where its requirement was high (Ca is immobile in plant system). So the uptake by pods was lesser compared to plant (Meena *et al.*, 2007).

Dhage *et al.* (2014) reported that S application increased plant S uptake in soybean. The results revealed that the uptake of S by groundnut increased with the advancement in the age of the crop. This was mainly due to increased dry matter production. S uptake was more in pod than in plant and this might be due to role of S in amino acids, proteins and oil synthesis in pod. Uptake of Fe, Mn and Zn increased with increased dose of sulphur. This result was in conformity with the findings of Kharol *et al.* (2014).

Rate of micro nutrient uptake by plant increased from 30 to 90 DAS, but lowest uptake was noticed during harvest. At harvest, iron, zinc, copper and boron uptake was more in pod than in haulm, whereas uptake was more in haulm than in pod in case of manganese. Uptake of zinc by pod was higher than that of plant and this might be due to involvement of zinc in metabolism of amino acids and protein in pod. Since B is immobile in plant, B uptake was more by plant than pod (Mahajan *et al.*, 1994).



**Fig 31: Effect of application of P and S on S uptake by plant at different stages and uptake by kernel and shell**



**Fig 32: Effect of application of P and S on Fe uptake by plant at different stages and uptake by kernel and shell**

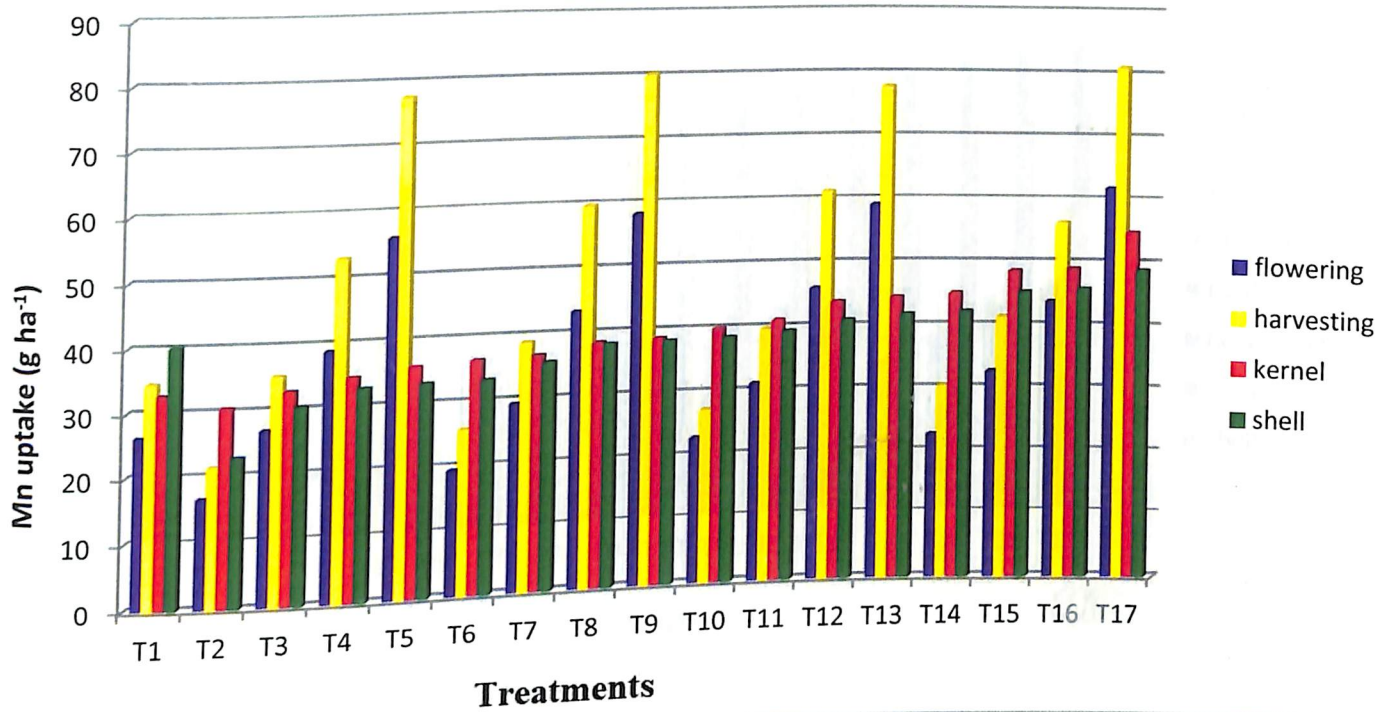


Fig 33: Effect of application of P and S on Mn uptake by plant at different stages and uptake by kernel and shell

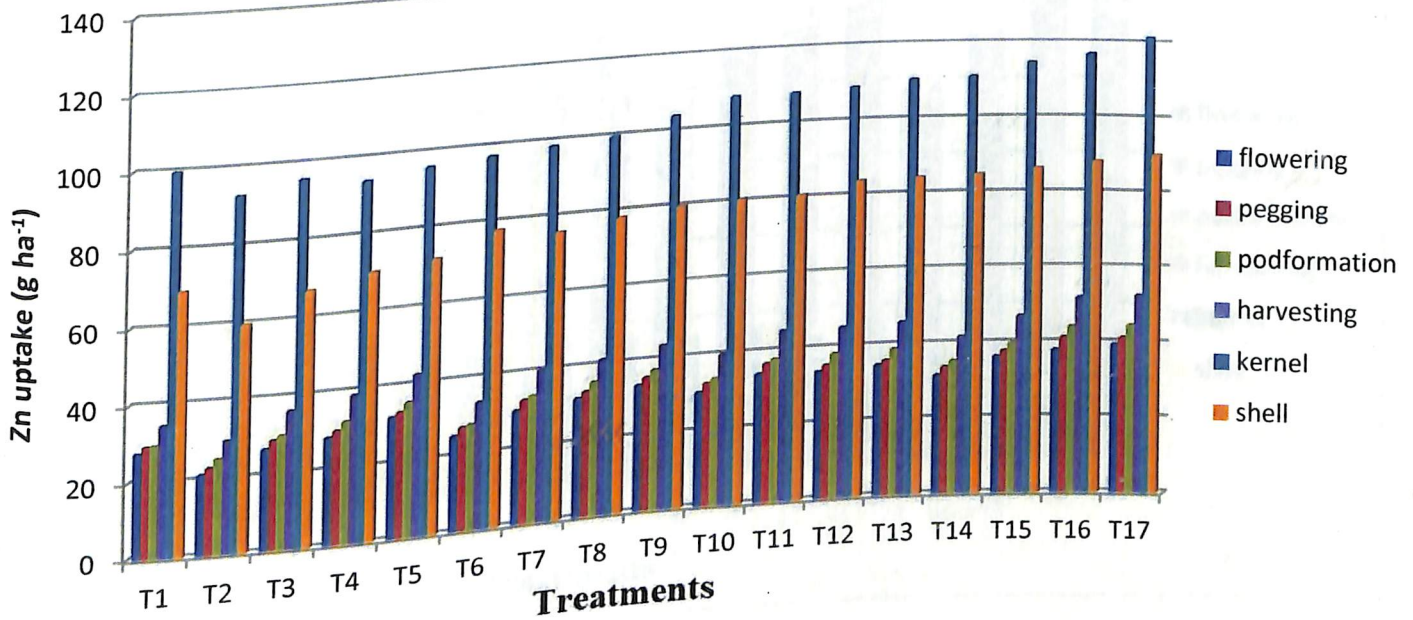
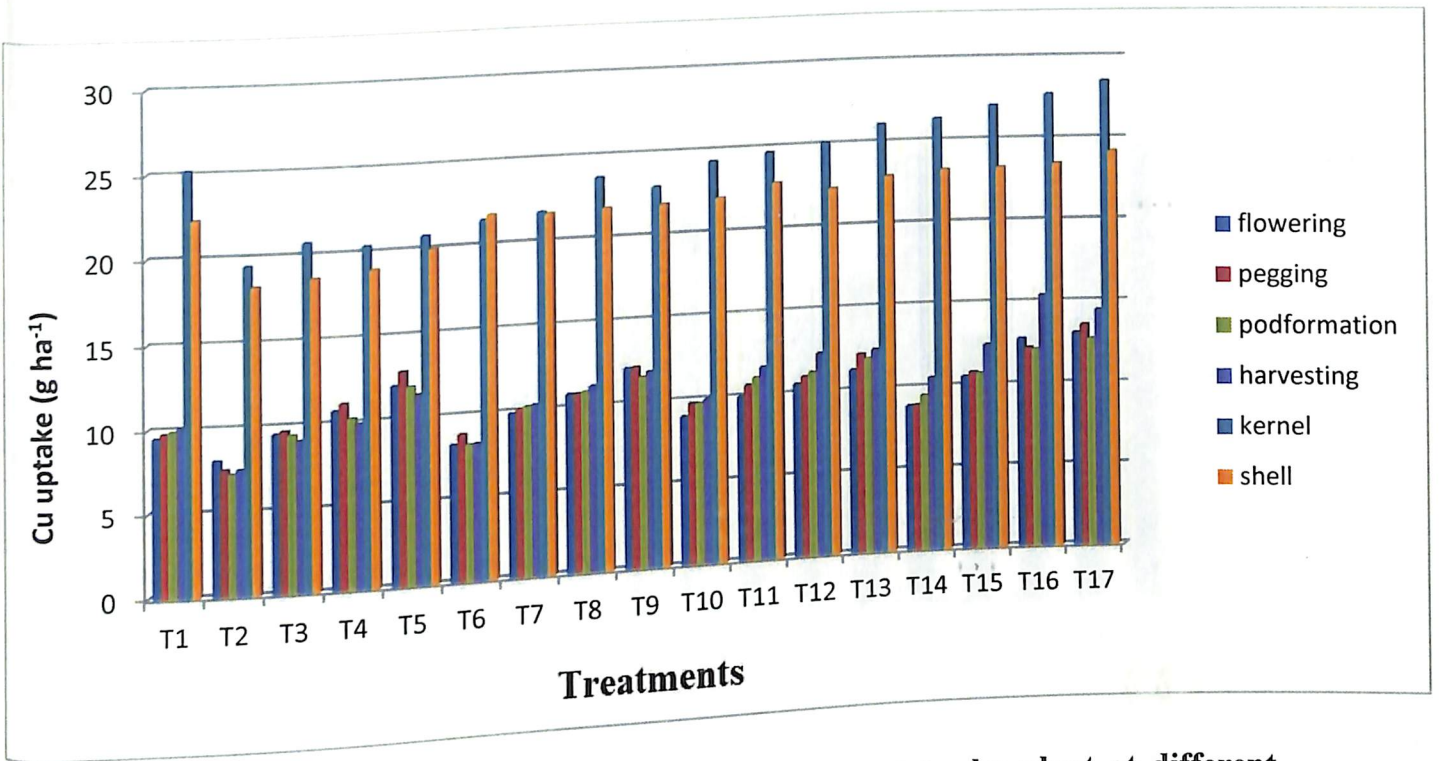
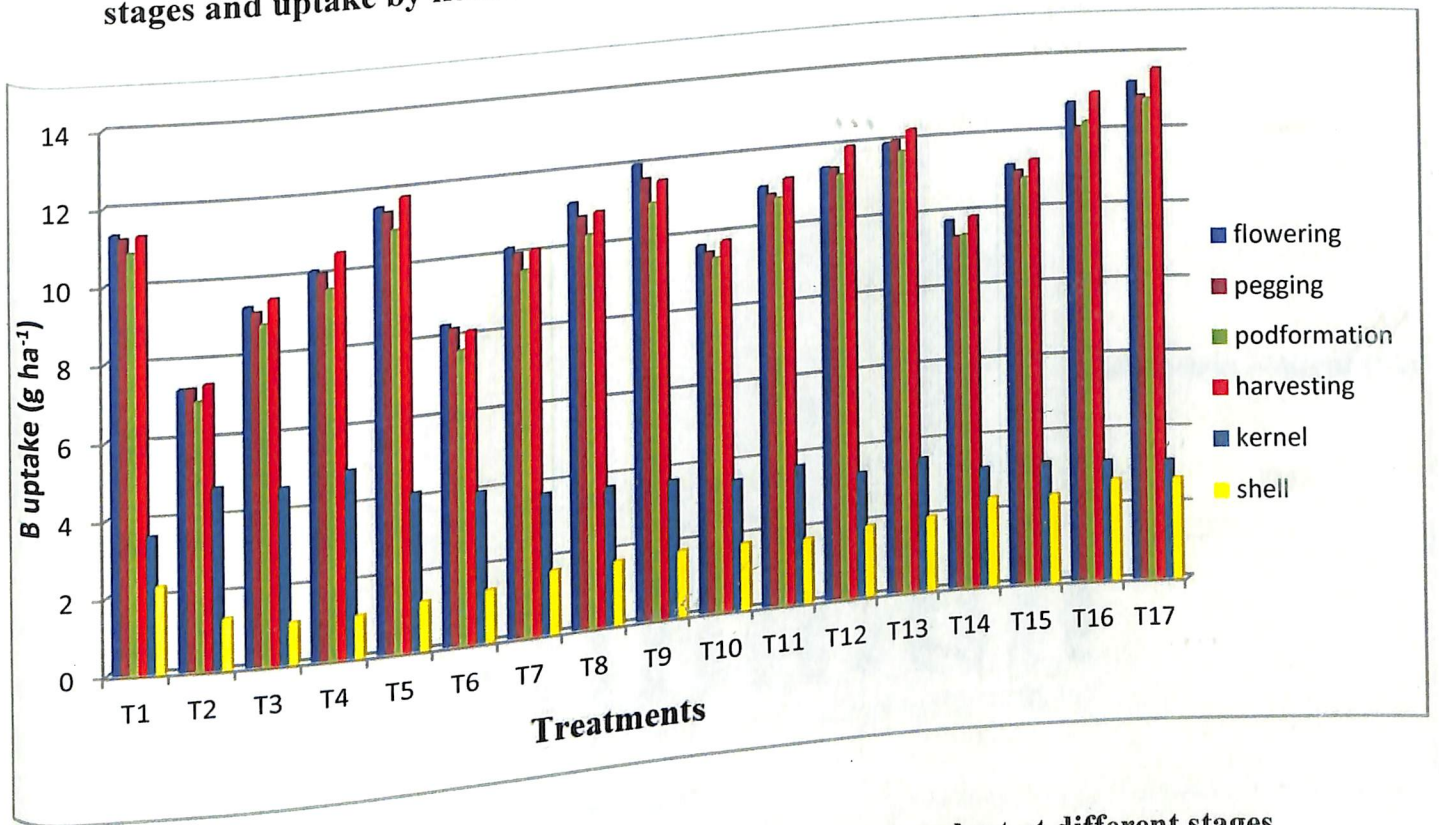


Fig 34: Effect of application of P and S on Zn uptake by plant at different stages and uptake by kernel and shell

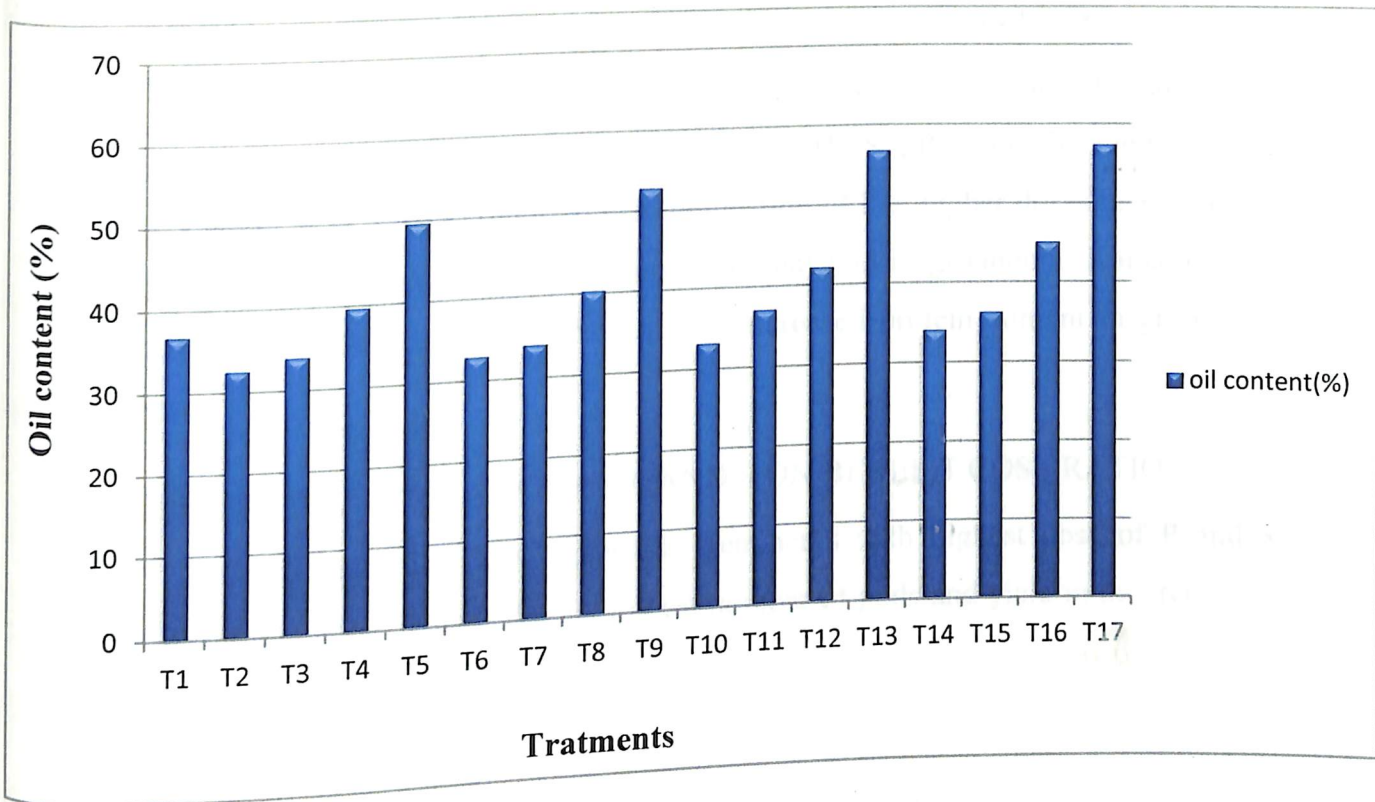




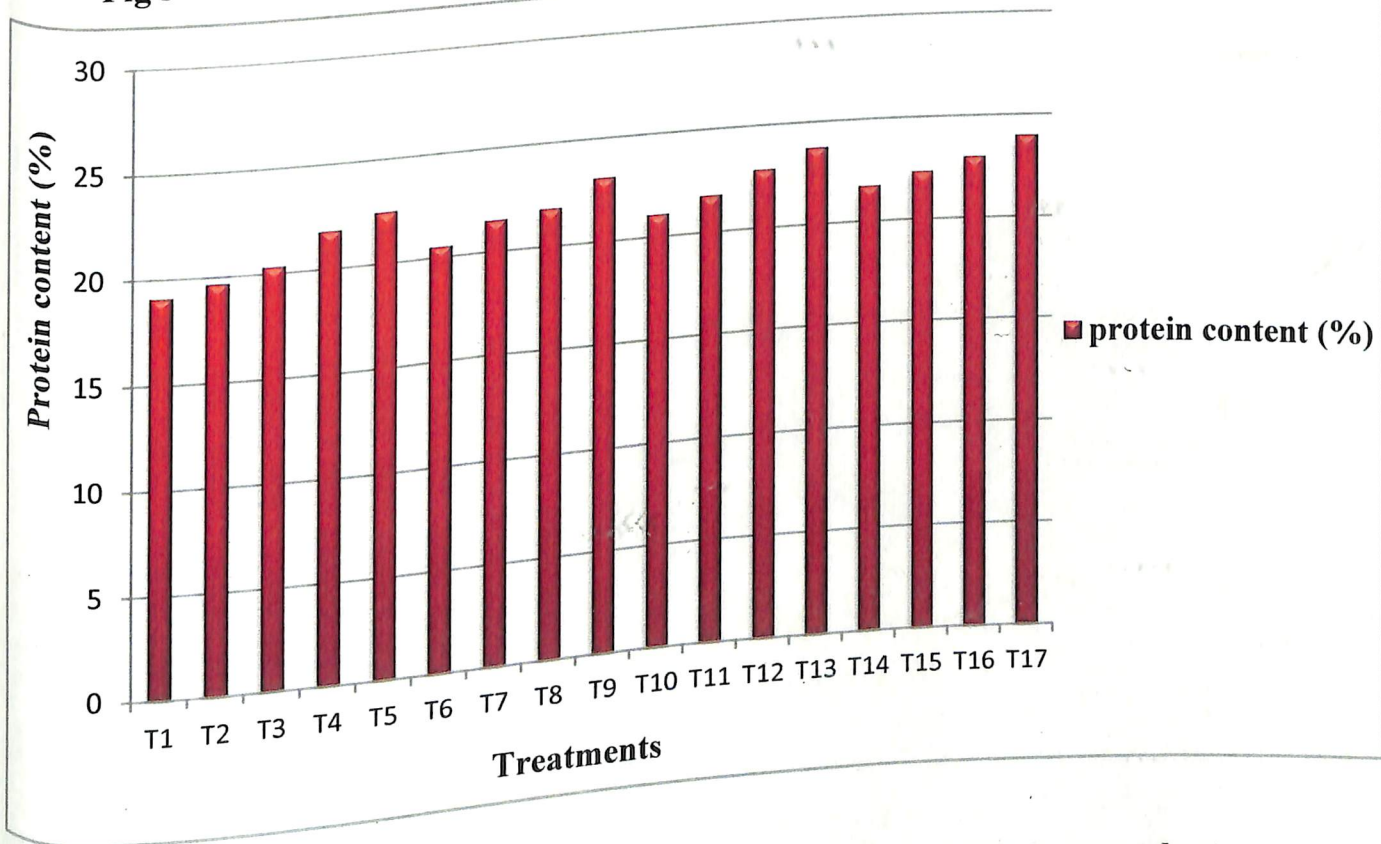
**Fig 35: Effect of application of P and S on Cu uptake by plant at different stages and uptake by kernel and shell**



**Fig 36: Effect of application of P and S on B uptake by plant at different stages and uptake by kernel and shell**



**Fig 37: Effect of application of P and S on oil content (%) in groundnut kernel**



**Fig 38: Effect of application of P and S on protein content (%) in groundnut kernel**

#### 5.4 EFFECT OF APPLICATION OF P AND S ON QUALITY PARAMETERS

Oil and protein content increased with increased dose of both P and S. The highest protein and oil content was recorded in T<sub>17</sub> (P<sub>3</sub> S<sub>3</sub>; P : 90 kg ha<sup>-1</sup> and S: 30 kg ha<sup>-1</sup>). Kadam *et al.* (2018) reported that application of P at higher doses increased the quality parameters such as oil and protein content (%) in groundnut. Kamara *et al.* (2011) also concluded that application of P increased protein content in groundnut kernels.

#### 5.5 EFFECT OF APPLICATION OF P AND S ON BENEFIT COST RATIO

Benefit cost ratio was highest in treatments with highest dose of P and S fertilizer application. This is due to highest number of pods and yield in the treatment with highest dose of P and S.

# Summary

## 6. SUMMARY

In Kerala, black soils are located in Chittur taluk of Palakkad district occupying an area of 2000 ha. These soils are dark, low in organic matter, calcareous, neutral to alkaline (pH 7.0 to 8.5), high in clay content and CEC. The texture of soil ranges from clay loam to clay. Even though these soils are fertile, the nutrient imbalances and poor physical conditions may adversely affect the yield of the crop. These are deficient in phosphorus (P) and sulphur (S). So the availability of P and S is one of the yield limiting factors in this soil. Finding the interaction between S and P in black soils would help to understand the factors determining the availability of these nutrients to crops.

Soil samples were collected from different locations of Chittur and analyzed for available P and S. Field experiment was conducted in a field at Nellimed that is deficient in both P and S. The study consisted of a field experiment with groundnut variety, K-6 followed by analysis of soil, plant, kernel and shell. Experiment was laid out in factorial RBD with 17 treatments and 3 replications.

Treatment combinations were made with four levels of P and four levels of S with soil test based recommendations as control. N and K levels were kept same (based on POP recommendations of KAU) for all treatments except for the first treatment where soil test based recommendations was given.  $P_0$  - 0 kg ha<sup>-1</sup>,  $P_1$  - 60 kg ha<sup>-1</sup>,  $P_2$  - 75 kg ha<sup>-1</sup> and  $P_3$  - 90 kg ha<sup>-1</sup> were the four levels of P and  $S_0$  - 0 kg ha<sup>-1</sup>,  $S_1$  - 10 kg ha<sup>-1</sup>,  $S_2$  - 20 kg ha<sup>-1</sup> and  $S_3$  - 30 kg ha<sup>-1</sup> were the four levels of sulphur. Potassium dihydrogenphosphate was used as source of P and elemental sulphur was used as source of sulphur.

Application of P at 90 kg ha<sup>-1</sup> and S at 30 kg ha<sup>-1</sup> increased plant height and number of leaves per plant. Application of P at 90 kg ha<sup>-1</sup> and S at 30 kg ha<sup>-1</sup> resulted in highest number of pods as well as yield (3.68 t ha<sup>-1</sup>). Effect of application of different doses of P and S was non-significant for pH, EC and OC. N content in soil increased by increasing levels of P and S. Application of P



increased soil S and vice versa. P and S showed positive interaction. Available K content in soil decreased with increased dose of P. Ca content in soil decreased with increased levels of P due to complex formation. Mg showed synergistic interaction with P.

Application of P decreased B content in soil due to antagonistic interaction, whereas Available B content in soil increased with increased dose of S. Application of P reduced the availability of micronutrients such as Fe, Mn and Zn in soil due to the formation of insoluble compounds like Fe-P, Mn-P and Zn-P. Whereas available micronutrient content in soil increased with increased dose of S fertilizer application. Increased available micronutrient status of soil with increased dose of S may be due to slight reduction in pH of the soil as a result of application of acid forming fertilizers and also due to soluble sulphate formation. Available Cu content in soil was non-significant to the addition of different levels of P and S.

Plant nutrient content showed a decreasing trend from flowering to harvesting stage. Content of N, P, Mg and S in plant, kernel and shell was increased with increased levels of P and S due to enhanced availability of these nutrients from soil as well as increased vegetative growth and vigour of the plant. Content of K in plant, kernel and shell decreased with increased dose of P and increased with increased dose of S. Content of Ca in plants decreased with increased dose of P as a result of decreased availability in soil due to insoluble complex formation. Content of Fe, Mn, Zn, Cu and B in plants, kernel and shell decreased with increased dose of P due to reduced availability and competition and increased with increased dose of S due to increased availability in soil.

Uptake of nutrients followed increasing trend from flowering to harvesting stage. Uptake of nutrients by plant, kernel and shell increased with increased dose of P and S fertilizer application. It may be due to increased vegetative growth of plants followed by increased dry matter production. Uptake of S, Fe, Zn and Cu were higher in kernel than plant. Oil content and protein content in kernels increased with increased dose of both P and S. Application of P

at 90 kg ha<sup>-1</sup> and S at 30 kg ha<sup>-1</sup> resulted in highest protein and oil contents. Benefit: cost ratio was calculated, and it was found that application of P at 90 kg ha<sup>-1</sup> and S at 30 kg ha<sup>-1</sup> resulted in highest benefit: cost ratio.

## **Future line of work**

- Conduct OFT in different location before recommending for the farmers for adoption.
- Fractionation of phosphorus and sulphur in Vertisols of Kerala.
- Influence of phosphorus and sulphur application on soil biology of black soils of Kerala.
- Interaction study of phosphorus and sulphur in deficient black soils using different sources of nutrients.

# References

## REFERENCES

- Abdin, M. Z., Ahmad, A., Khan, N., Khan, I., Jamal, A., and Iqbal, M. 2003. Sulphur interaction with other nutrients. *Sulphur in Plants* 2: 359–374.
- Adriano, D. C. and Murphy, L. S. 1970. Effects of ammonium polyphosphates on yield and chemical composition of irrigated corn. *J. Agron.* 62(5): 561- 567.
- Adriano, D. C., Paulsen, G. M., and Murphy, L. S. 1971. Phosphorus-iron and phosphorus-zinc relationships in corn (*zea mays* l.) seedlings as affected by mineral nutrition. *J. Agron.* 63(1): 36-39.
- Ae, N., Otani, T., Makino, T., and Tazawa, J. 1996. Role of cell wall of groundnut roots in solubilizing sparingly soluble phosphorus in soil. *Plant Soil* 186(2): 197-204.
- Agasimani, C. A. and Hosmani, M. M. 1989. Response of groundnut crop to stand geometry in rice fallows in coastal sandy soils of Uttara Kannoda district in Karnataka. *J. Agric. Sci.* 2(1-2): 7-11.
- Akhtar, S., Bangash, N., Shahzad, A., Fatima, S., Nayab, D., Arshad, M., Iqbal, M. S., Akbar, M., Khalil, T., and Hassan, F. 2018. Foliar applications of FeSO<sub>4</sub> alone and in combination with citric acid can reduce iron deficiency induced chlorosis in two Pakistani peanut (*Arachis hypogaea* L.) varieties. *Applied Ecol. Environ. Res.* 16(3): 2873-2884.
- Al-lami, A.S.J. 1999. Evaluation of magnesium supplying power in plastic house soils. Ph. D Dissertation, Baghdad University, 136p.

- Amruth, G.N.B., Thippeshappa, K.T., Gurmurthy., and Chidanandappa, H. M. 2018. Effect of phosphorus levels through integrated nutrient management (INM) packages on nutrient content in various parts of the crop. *Int. J. Curr. Microbiol. App. Sci.* 7(3): 2080-2087.
- Aulakh, M. S., Pasricha, N. S., and Azad, A. S. 1990. Phosphorus - sulphur interrelationships for soybean on P and S deficient soil. *Soil Sci.* 150: 194-198.
- Ayed, I. A. 1970. A study of the mobilization of iron in tomato roots by chelate treatments. *Plant Soil* 32(1-3): 18-26.
- Babu, K. G., Munaswamy, V., John, K., and Raju, P. A. 2007. Effect of applied nutrients on major (N, P, K) and secondary (Ca, Mg, S) nutrient concentration at different growth stages of rainfed groundnut (*Arachis hypogaea* L.) in Alfisols. *J. Oilseeds Res.* 24(1): 84-87.
- Bala, H.M.B., Ogunlela, V.B., Kuchinda, N.C., and Tanimu, B. 2011. Response of two groundnut (*Arachis hypogaea* L.) varieties to sowing date and NPK fertilizer rate in a semi-arid environment: yield and yield attributes. *Asian J. Crop Sci.* 3(3): 130-134.
- Balasubramanian, P. and Palaniappan, S. 1991. Effect of population density, fertilizer levels and time of application on rice (*Oryza sativa*) – groundnut (*Arachis hypogaea* L.). *Indian J. Agron.* 36: 218-221.
- Bandopadhyay, K. K., Misra, A. K., Ghosh, P. K., Hati, K. M., and Mandal, K. G. 2003. Effect of integrated use of farmyard manure and inorganic fertilizers on soil water dynamics, root growth, crop yield and water expense efficiency of rainfed soybean in a Vertisol. *J. Agric. Physic.* 3: 49-55.

- Banerjee, H. T., Das, M. and Bhattacharjee, T. K. 1967. Nutrition of laterite zone groundnut in West Bengal. *Fert. News* 12 (9): 41-42.
- Barczak, B. 2010. *Sulphur as a nutrient determining the yield, size and quality of selected crops*. Monograph 144, Publ. UTP Bydgoszcz (in Polish).
- Basha, S. M. and Rao, G. R. 1980. Effect of potassium deficiency on growth and metabolism of peanut (*Arachis hypogaea* L.) plants. *Proceedings: Plant Sci.* 89(5): 415-420.
- Bell, M.J. 1985. Phosphorus nutrition of peanut (*Arachis hypogaea* L.) on cockatoo sandy soils of the ord river irrigation area. *Aust. J. Exp. Agric.* 25: 649-653.
- Bharthi, B., Sonawane, P., Nawalkar, S. and Patil, V.D. 2010. Effect of micronutrients on growth and yield of groundnut. *J. Soils Crops* 20(2): 269-273.
- Bingham, F. T., Martin, J. P., and Chastain, J. A. 1958. Effect of phosphorus fertilization of California soils on minor element nutrition of citrus. *Soil Sci.* 86: 24-31.
- Black, C. A. 1965. *Method of Soil Analysis part 11 Agronomy Monograph No.9*, American Society of Agronomy, Madison, Wisconsin, 148p.
- Boote, K. J., Graw, R. L., and Mc. Cloud, D. E. 1985. Seasonal trends in carbohydrate and N concentrations of plant parts of Florunner peanut and Bragg soyabean. *Proceedings, Am. Peanut Res. Educ. Soc.* 17 : 70.

- Brady, N.C. and Weil, R.R. 2002. The nature and properties of soils. Prentice Hall, Upper Saddle River, NJ. *The nature and properties of soils. 13th ed. Prentice Hall, Upper Saddle River, NJ.*
- Brennan, R. and Bolland, M. D. 2007. Influence of potassium and nitrogen fertilizer on yield, protein and oil concentration of canola (*Brassica napus L.*) grain harvested in south-western Australia. *Australian J. Experimental Agric.* 47: 976-983.
- Chahal, R.S., Singh, S. and Singh, M. 1983. Nitrogen, phosphorus and potassium uptake in groundnut at various stages of plant growth and levels of phosphorus. *Indian J. Plant Physiol.* 26(2): 220-225.
- Chauhan, Y.S., Jain, V.K., Khandedar, M.P. and Jain, P.C. 1987. Response of groundnut varieties to phosphorus fertilization. *Madras Agric. J.* 74: 261-264.
- Chavan, L.S. and Kalra, G.S. 1983. Effect of phosphorus and potassium levels under varying row spacing on yield, quality and nutrient uptake by groundnut (*Arachis hypogaea L*) variety TG-1 under high rainfall conditions of Konkan region of Maharashtra. *Indian J. Agric. Res.* 17 (1 & 2): 62-68.
- Cheema, M. A., Malik, M. A., Hussain, A., Shah, S. H., and Basra, S. M. A. 2001. Effects of time and rate of nitrogen and phosphorus application on the growth and the seed and oil yields of canola (*Brassica napus L.*). *J. Agron. Crop Sci.* 186(2): 12-15.
- Choudhary, C. H. V. S. K. 1979. Studies on the effect of different levels of nitrogen, phosphorus and potash on growth, yield and quality of irrigated groundnut. *Thesis abstract 3*: 30-31.



- Cox, F.R., Adams, J.F. and Tucker, B.B. 1982. Liming, fertilization and mineral nutrition. In: H.E. Pette and C.T. Young (Eds.) *Peanut Science and Technology*, American Peanut Research and Education Society Inc. Yoakum, Texas, U.S.A. pp. 139-163.
- Csinos, A. S. and Gaines, T. P. 1986. Peanut pod rot complex: A geocarposphere nutrient imbalance. *Plant Dis.* 70(6): 525-529.
- Dahatonde, B. N. 1982. Effect of nitrogen and phosphate fertilization on yield and yield contributory characters of groundnut (*Arachis hypogaea* L.). *Punjab Krishi Vidyapeeth Res. J.* 6(1): 31-33.
- Daliparthi, J., Barker, A.V. and Mondal, S.S. 2008. Potassium fractions with other nutrients in crops: a review focusing on the tropics. *J. Plant Nutr.* 17 (11): 1859-1886.
- Dart, P. J., Mc Donald, D., and Gibbons, R. W. 1983. Groundnut production systems with some implication from recent research, Paper presented at Indonesia National Planning Seminar on Palawija, Yogyakarta, March.
- Das, H. N. 1982. Effect of nitrogen, phosphorus, potassium and calcium on growth and yield of groundnut. *J. Res. Assam Agric. Univ.* 3(1): 15-18.
- Das, S.K. 2017. Effect of phosphorus and sulphur on yield attributes, yield, nodulation and nutrient uptake of green gram (*Vigna radiata* L.) Wilczek]. *Legume Res.* 40: 138-143.
- Dhage, S. J., Patil, V. D., and Patange, M. J. 2014. Effect of various levels of phosphorus and sulphur on yield, plant nutrient content, uptake and availability of nutrients at harvest stages of soybean [*Glycine max* (L.)]. *Int.J.Curr.Microbiol.App.Sci.* 3(12): 833-844.

- Dholaria, S.J., Joshi, S.N. and Kabaria, M.M. 1972. Correlation of yield and yield-contributory characters in groundnut grown under high and low fertility levels. *Indian J. Agric. Sci.* 42(6): 467-470.
- Dowood, M. A. 1982. Magnesium and phosphorus studies in dohuk and alhawler soil and its relation to grass tetany. M. Sc. Dissertation, University of Mosul, 129p.
- Dubey, S. K. 1997. Co-inoculation of phosphorus bacteria with *Bradyrhizobium japonicum* to increase phosphate availability to rainfed soybean in Vertisol. *J. Indian Soc. Soil Sci.* 45: 506-509.
- Dutta, D. and Patra, B. C. 2005. Response of groundnut (*Arachis hypogaea L.*) to sources and levels of sulphur fertilization in alluvial soils of West Bengal. *J. Inter Academia.* 9(1): 45-48.
- Elliott, G. C. and Laeuchli, A. 1985. Phosphorus efficiency and phosphate-iron interaction in maize. *J. Agron.* 77(3): 399-403.
- Fageria, N. K. 1983. Ionic interactions in rice plants from dilute solutions. *Plant Soil* 70(3): 309-316.
- Farhad, I. S. M., Islam, M. N., Hoque, S., and Buiyan, M. S. I. 2010. Role of potassium and sulphur on the growth, yield and oil content of soybean (*Glycine max L.*). *Acad. J. Plant Sci.* 3(2): 99-103.
- Fazili, A. I., Jamal, S., Ahmad, Khan., and Abdin, M. Z. 2008. Interactive effect of sulfur and nitrogen on nitrogen accumulation and harvest in oilseed crops differing in nitrogen assimilation potential. *J. Plant Nutrition* 31(7): 1203-1220.

- Friesen, D. K., Miller, M. H., and Juo, A. S. R. 1980. Liming and lime-phosphorus-zinc interactions in two Nigerian Ultisols- effects on maize root and shoot growth. *Soil Sci. Soc. Am. J.* 44: 1127-1232.
- Geethalakshmi, V., Lourduraj, A. C., Joel, A. J., and Rajamanickam, K. 1993. Nutrient management in groundnut. *Madras Agric. J.* 80(7): 412-414.
- Ghadekar, S. R., Wankhede, R. G., Miskin, R. B., and Das, S. N. 1993. Performance of summer groundnut under different levels of irrigation and fertilizers. *J. Soils Crops* 3(2): 121-124.
- Giri, U., Mohammad H. S. A., Nanda, M. K. and Bandyopadhyay, P. 2014. Productivity and nutrient uptake of summer groundnut (*Arachis hypogaea* L.) towards different levels of irrigation and sulphur. *J. Crop Weed.* 10(2): 248-251.
- Gomez, K.A. and Gomez, A. A. 1984. *Statistical Procedures for Agricultural Research* (2<sup>nd</sup> Ed.). John Wiley and Sons Inc., New York, 56p.
- Gupta, V.K. and Raj, H. 1983. Response of groundnut genotypes to different levels of zinc. *J. Crop Physiol.* 1(1): 55-67.
- Hadwani, G.J. and Gundalia, J.D. 2005. Effect of N, P and K levels on yield, nutrient content, uptake and quality of summer groundnut grown on Typic Haplustepts. *J. Indian Soc. Soil Sci.* 53(1): 125-128.
- Hallock, D. L. and Garren, K. H. 1968. Pod breakdown, yield, and grade of virginia type peanuts as affected by Ca, Mg, and K sulfates . *J. Agron.* 60(3): 253-257.

- Hasan, M. 2018. Effect of rhizobium inoculation with phosphorus and nitrogen fertilizer on physico-chemical properties of the groundnut soil. *Environ. Ecosystem Sci.* 2(1): 4-6.
- Islam, M.S. and Noor, S. 1982. Performance of groundnut under different levels of phosphate fertilization in grey floodplain of Jamalpur. *Bangladesh J. Agric. Res.* 1: 35-37.
- Jackson, M. L. 1973. *Soil Chemical Analysis*, Prentice Hall of India, Pvt. Ltd., New Delhi, 498p.
- Jadhar, A. S. and Narkhende, B. N. 1980. Influence of nitrogen and phosphorus fertilization on growth, yield and quality of groundnut. *Madras Agric. J.* 67: 131-135.
- Jakobsen, S. T. 1979. Interaction between phosphate and calcium in nutrient uptake by plant roots. *Commun. Soil Sci. Plant Anal.* 10: 141-152.
- Jankowski, K., Kijewski, Ł., Skwierawska, M., Krzebietke, S., and Mackiewicz-Walec, E. 2014. Effect of sulfur fertilization on the concentrations of copper, zinc and manganese in the roots, straw and cake of rapeseed (*Brassica napus L. ssp. oleifera Metzg.*). *J. Elementology* 19: 433-446.
- Juan, A. R., Curayag, L. J., and Pava, H. M. 1986. Influence of phosphorus fertilization on pod yield and seed quality of three peanut varieties (*Arachis hypogea L.*). *J. Agric. Food Nutri.* 8(1): 33-62.
- Kachot, M. A., Patel, J. C., and Malavia, D. D. 1984. Response of summer groundnut to irrigation scheduling based on IW/CPE ratio under varying levels of nitrogen and phosphorus. *Indian J. Agron.* 67 (5): 234-236.

- Kadam, D.V., Indulkar, B.S., Kadam, V.S., Jadhav, L.S., and Sonune, P.N. 2018. Effect of phosphorus and zinc on yield and quality of groundnut (*Arachis hypogea L.*) in inceptisol. *Int. J. Pure App. Biosci.* 6(1): 105-110.
- Kalaiyarasan, C., Vaiyapuri, V., and Chandrasekharan, M. V. S. 2003. Effect of sulphur sources and levels on the nutrient uptake, crop quality and sulphur use efficiency in groundnut. *Ann. Agric. Res. New Series* 24(3): 478-480.
- Kamalakaran, P. and Ravichandran, M. 2013. Effect of organic and inorganic sources of nutrients on growth and yield of groundnut in two different textured soils. *Asian J. Soil Sci.* 8(1): 88-93.
- Kamara, E.G., Olympio, N.S. and Asibuo, J.Y. 2011. Effect of calcium and phosphorus fertilizer on the growth and yield of groundnut (*Arachis hypogaea L.*). *Int. Res. J. Agric. Sci. Soil Sci.* 1(8): 326-331.
- Keisling, T. C., Lauer, D. A., Walker, M. E., and Henning, R. J. 1983. Visual, tissue, and soil factors associated with Zn toxicity of peanuts. *J. Agron.* 69: 765-769.
- Kharol, S., Sharma, M., Lai, M., and Sumeriya, H. K. 2014. Productivity of chickpea (*Cicer arietinum L.*) as influenced by sulphur and zinc under agroclimatic zone IV- A of Rajasthan. *Ann. Biol.* 30 (4): 676-680.
- Krishnakumar, A. K. 1978. Study of physico-chemical characteristics of the poonthalpadam soils of Kerala. M.Sc. (Ag) thesis, Kerala Agricultural University. p.120
- Koch, K. and Mengal, K. 1977. Effect of K and N on utilization by spring wheat during grain protein formation. *J. Agron.* 69: 477-480.

- Kuchanwar, O. D., Matte, D. B., and Kene, D. R. 1997. Evaluation of graded doses of fly-ash and fertilizers on nutrient content and uptake of groundnut grown on Vertisol. *J. Soils Crops*. 7: 1-3.
- Kuo, S. and Mikkelsen, D. S. 1979. Effect of magnesium phosphate adsorption by calcium carbonate. *Soil Sci*. 127: 65-69.
- Kvien, C. S., Branch, W. D., Sumner, M. E., and Csinos, A. S. 1988. Pod characteristics influencing calcium concentrations in the seed and hull of peanut. *Crop Sci*. 28(4): 666-671.
- Lakshamma, P. and Raj, A. S. 1997. Effect of cobalt, phosphorus and potassium on groundnut (*Arachis hypogaea* L.). *Legume Res*. 2: 20-160.
- Laxminarayana, K. 2004. Effect of organic and inorganic manures on yield and nutrient uptake of groundnut (*Arachis hypogaea* L.) in Ultisols of Mizoram. *J. Oilseeds Res*. 21(2): 280-283.
- Lickfett, T. 1999. Seed yield, oil and phytate concentration in the seeds of two oilseed rape cultivars as affected by different phosphorus supply. *European J. Agron*. 11(3-4): 23-25.
- Lindsay, W.L. and Norvell, W.A. 1978. Development of a DTPA soil test for zinc, iron, manganese, and copper. *Soil Sci. Soc. Am. J.* 42(3): 421-428.
- Loganathan, S. and Krishnamoorthy, V. S. 1980. Potash application to groundnut. *Madras Agric J.* 67(9): 610-612.

- Loganathan, S., Helkiah, L. J., and Thangavelu, S. 1996. Effect of FYM, nitrogen and potash on yield and uptake of nutrients in groundnut. *Madras Agric. J.* 83(8): 484-487.
- Loneragan, J. F., Grove, T. S., Robson, A. D., and Snowball, K. 1979. Phosphorus toxicity as a factor in zinc-phosphorus interactions in plants. *Soil Sci. Soc. Am. J.* 43(5): 966-972.
- Loneragan, J. F., Grunes, D. L., Welch, R. M., Aduayi, E. A., Tengah, A., Lazar, V. A., and Cary, E. E. 1982. Phosphorus accumulation and toxicity in leaves in relation to zinc supply. *Soil Sci. Soc. Am. J.* 46(2): 345-352.
- Lundergardh, H. 1934. Mineral nutrition of plants. *Annu. Rev. Biochem.* 1934(3): 485-500.
- Mahajan, T. S., Chavan, A. S., and Dongale, J. H. 1994. Effect of boron on yield and quality of groundnut (*Arachis hypogaea* L.) on lateritic soil. *Indian J. Agric. Sci.* 64(8): 532-535.
- Mahapatra, I. C., Singh, K. N., Pillai, K. G. and Bapat, S. R. 1985. Rice soils and their management. *Indian J. Agron.* 30(1): 1-41.
- Mahmoud, M., Shaaban, A.F., El-Sayed, A., El-Nour, E.A.M., Aly, E.S., and Mohamed, A.K. 2006. Boron/nitrogen interaction effect on growth and yield of faba bean plants grown under sandy soil conditions. *Int. J. Agric. Res.* 1(4): 322-330.
- Maini, N. S. and Bhandar, D. S. 1965. Studies on the resistance of oleiferous *Brassica* to *Alternaria* blight. *Indian Oilseeds J.* 9(1): 58.

- Mallikarjuna, G., Sudhir, K., Srikanth, K., and Srinivasamurthy, C. A. 2003. Phosphorus fixation capacity and its relationship with the soil characteristics in laterite soils of Karnataka. *J. Indian. Soc. Soil Sci.* 51(1): 23-25.
- Manasa, V., Hebsur, N. S., Malligawad, L. H., Shiva Kumar, L., and Ramakrishna, B. 2015. Effect of water soluble fertilizers on uptake of major and micro nutrients by groundnut and post-harvest nutrient status in a Vertisol of northern transition zone of Karnataka. *The Ecoscan.* 9(1&2): 01-05.
- Mandal, S., Samui, R.C., and Mondal, A. 2005. Growth, yield and yield attributes of groundnut (*Arachis hypogaea* L.) cultivars as influenced by gypsum application. *Legume Res.* 28 (2): 119-121.
- Marion, G. M. and Babbcock, K. L. 1977. The solubility of carbonates and phosphates in calcareous soil suspension. *Soil Sci. Soc. Am. J.* 41: 724-728.
- Mathew, J., Nair, K. P. M., and Kuriakose, T. C. 1983. The response of groundnut to phosphorus and potassium under different water management practices. *Agric. Res. J. Kerala* 21(2): 27-31.
- May, G. M. and Pritts, M. P. 1993. Phosphorus, zinc, and boron influence yield components in Earliglow strawberry. *J. Am. Soc. Hortic. Sci.* 118(1): 43-49.
- Meena, S., Malarkodi, M., and Senthilvalavan, P. 2007. Secondary and micronutrients for groundnut – A Review. *Agric. Reviews.* 28(4): 295-300.
- Modaihsh, A.S., Al-Mustafa, W.A., and Metwally, A.I. 1989. Effect of elemental sulphur on chemical changes and nutrient availability in calcareous soils. *Plant Soil* 116: 95.
- Moraghan, J. T. and Mascagni, H. J. 1991. Environmental and soil factors affecting micronutrient deficiencies and toxicities. *Micronutr. Agric.* 2: 371-425.



- Nadaf, S.A. and Chidanandappa, H.M. 2015. Effect of zinc and boron application on distribution and contribution of zinc fractions to the total uptake of zinc by groundnut (*Arachis hypogaea* L.) in sandy loam soils of Karnataka, India. *Legume Res. Int. J.* 38(5): 598-602.
- Nair, N. P. and Sadanandan, N. 1981. Quality of kernal in groundnut varieties: TMV-2 and TMV-9 as influenced by phosphorus and potassium fertilization. *Agric. Res. J. Kerala.* 19(3): 25-28.
- Nair, N.P., Sadanandan, N., Mohamedkunju, U. and Nair, K.P.M. 1981. Potash fertilization and higher yields of bunch groundnut in Kerala. *Indian Pot. J.* 6: 14-17.
- Nakagawa, J., Machedo, J. R., and Toledo, F. F. De. 1981. The effect of increasing rates of phosphorus fertilizer on groundnut (*Arachis hypogaea* L.) crops. *Cientifica* 9 (2): 227-234.
- Nasreen, S. and Huq, S. M. I. 2002. Effect of sulphur fertilizer on yield and nutrient uptake of sunflower crop in an albaquept soil. *Pakistan J. Biol. Sci.* 5: 533-536.
- Nijhawan, H. L. 1962. Effect of application of manures on the composition of groundnut crop. *Indian Oilseeds J.* 6: 123-129.
- Nijhawan, H.L. and Maini, N.S. 1966. The effect of calcium on groundnut yields. *Indian Farming* 16: 27-29.
- Nurezannat., Sarkar, M. A. R., Uddin, M. R., Sarkar, U. K., Kaysar, M. S., and Saha, P. K. 2019. Effect of variety and sulphur on yield and yield components of groundnut. *J. Bagladesh. Agric. Univ.* 17(1): 1-8.

- Padmaja, P., Geethakumari, V. L., Nair, H. K., Chinnamma, N. P., Sashidharan, N. K., and Rajan, K. C. 1994. *A glimpse to problem soils of Kerala*. Kerala Agricultural University, Vellanikkara, Thrissur, Kerala. pp. 104-111.
- Page, A. L., Miller, R. H. and Keeney, D. R. (Ed. 2). 1982. *Methods of soil analysis; Chemical and microbiological properties*. Madison, Wisconsin, USA.
- Pant, L.M. and Katiyar, A.K. 1996. Response of groundnut of fertilizer N, FYM and *Bradyrhizobium* inoculation. *Agric. Sci. Digest* 16: 52-54.
- Parvathi, E., Venkaiah, K., Naidu, M. V. S., Munaswami, V., and Reddy, K. B. 2015. Long term effect of manure and fertilizers on yield and nutrient uptake of groundnut grown in Alfisols of Chittoor district in Andhra Pradesh, India. *Thai J. Agric. Sci.* 48(1): 23-27.
- Patel, G. N., Patel, P. T., Patel, P. H., Patel, D. M., Patel, D. K., and Patel, R. M. 2009. Yield attributes, yield, quality and uptake of nutrients by summer groundnut (*Arachis hypogaea* L.) as influenced by sources and levels of sulphur under varying irrigation schedules. *J. Oilseeds Res.* 26(2): 119-122.
- Patel, M. S., Golaka, P. R., and Shobhana, H. K. 1981. Growth and nutrient uptake by groundnut (*Arachis hypogaea* L.) as influenced by levels of phosphorus and soil moisture. *Indian J. Agric. Chem.* 14 (1): 33-37.
- Patel, Z.G., Solanki, N.P., and Patel, R.S. 1994. Effect of varying levels of seeds, N and phosphatic fertilizer on the growth and yield of summer groundnut (*Arachis hypogaea* L.). *Gujarat Agric. Univ. Res. J.* 20: 148-150.
- Pearson, D. 1981. *The chemical analysis of food*, Churchill livingstone, Edinburgh, 504-530.

Piper, C.S. 1966. *Soil and Plant Analysis*. Hans Publishers, Bombay, 368p.

Pradhan, A.C. and Das, A.D. 1989. Study on fertilizer response of groundnut at Singthum district of Bihar under rainfed condition. *Indian Agric. St.* 33: 159-162.

Punnoose, K. I. 1968. *Studies on the effect of nitrogen and phosphorus on the growth, yield and quality of groundnut in Red loam soils of Kerala*. M. Sc. (Ag) thesis, Kerala Agricultural University, Thrissur, 127p.

Puntamkar, S.S. and Bathkal, B.G. 1967. Influence of N, P and K fertilizers on composition, growth and yield of groundnut. *Indian J. Agron.* 12(4): 344-350.

Puri, D. N. 1969. Groundnut responds well to super phosphate. *Fert. News* 14(7): 46-47.

Ramanathan, K.M., Bruchholz, H., Hanora, J.F. and Subbiah, S. 1982. Influence of nitrogen and potassium on yield and quality of groundnut var. POL-2. *Indian Pot. J.* 7: 6-10.

Rana, D. S., Kalsi, H. S., Kapur, M. L. and Bhandari, A. L. 1984. Response of groundnut (*Arachis hypogaea* L.) and raya (*Brassica juncea* L.) to fertilizer application. *Indian J. Agron.* 29(1): 122-124.

Ranade, U. and Malvi. 2011. Interaction of micronutrients with major nutrients with special reference to potassium. *Karnataka J. Agric. Sci.* 24(1): 106-109.

- Rao, I.V.S., Babu, M.D., and Reddy, P.R. 1984. Effect of phosphorus and Rhizobium on groundnut. *Andhra Agric. J.* 31: 253-254.
- Rao, K.T., Rao, A.U., and Sekhar, D. 2013. Effect of sources and levels of sulphur on groundnut. *J. Academia and Industrial Res.* 2(5): 268-270.
- Rao, S. R. 1979. Studies on the effect of potassium, calcium, magnesium on growth and yield of irrigated groundnut (TMV-2). M. Sc. (Ag) thesis, S. V Agricultural College, Tirupathy, 117p.
- Rao, S. S. and Shaktawat, M. S. 2002. Effect of organic manure, phosphorus and gypsum on groundnut (*Arachis hypogaea* L.) production under rainfed condition. *Indian J. Agron.* 47(2): 234-241.
- Ravichandra, K., Jyothi, C.N., Sigh, B.J., Dawson, J. and Krupakar, A. 2015. Growth of groundnut (*Arachis hypogaea* L.) and its yield as influenced by foliar spray of boron along with *Rhizobium* inoculation. *Indian J. Dryland Agric. Res. Dev.* 30(1): 60-63.
- Rayar, A. J. 1986. Response of groundnut (*Arachis hypogaea* L.) to application of FYM, N and P on light sandy loam Savanna soils of northern Nigeria. *Int. J. Trop. Agri.* 4(1): 46-54.
- Reddy, B.B., Ramesh, S., Raju, M.S., and Rao, C.M. 1984. Response of groundnut to nitrogen fertilizer and plant densities in relation to rhizobial inoculation. *J. Oilseed Res.* 1: 233-235.
- Reddy, G. P. and Rao, S. C. 1965. Fertilizers response in groundnut. *Indian Oilseeds J.* 9:274-279.

- Reddy, K.C. and Murthy, P.K. 1985. Distribution of N, P, K, Ca, Mg, S, Zn, Fe, Mn and Cu in groundnut. *Plant Soil* 84(2): 269-273.
- Reddy, R., Krishnamoorthy, K., and Malewar, G. U. 1983. Response of groundnut to the application of nitrogen, phosphorus and potassium both in the presence and absence of FYM through soil and foliar spray. *Indian J. Agron.* 21(4): 321-326.
- Reddy, T. S., Reddy, S. D., and Reddy, P. G. 2011. Fertilizer management for maximizing productivity and profitability of export oriented groundnut (*Arachis hypogaea* L.). *J. Res. ANGRAU.* 39(4): 83-85.
- Saeed, M. and Fox, R. L. 1979. Influence of phosphate fertilization on zinc adsorption by tropical soils. *Soil Sci. Soc. Am. J.* 43(4): 683-686.
- Saini, J. S. and Tripathi, H. P. 1973. Effect of nitrogen and phosphorus application on the yield of spreading and erect varieties of groundnut (*Arachis hypogaea* L.). *Indian J. Agron.* 48(2): 116-118.
- Samtana, M., Bhadoria, P.B.S., and Ghosh, B. 1994. Effect of lime and phosphorus on phosphate availability and yield of groundnut (*Arachis hypogaea* L.) in Oxisol. *Indian J. Agron.* 39: 692-693.
- Sanchez, L. F. S. and Owen, E. J. B. 1978. Influence of liming and fertilization with N, P and K on yields of peanut (*Arachis hypogaea* L.) grown in soils of the high terraces of the Eastern plains of Colombia. *Fert. Abstr.* 13: 190-193.
- Saradhi, K., Subbiah, G., Raghavulu, P., and Rao, G.V.H. 1990. Effect of graded levels of nitrogen and phosphorus on growth and yield of groundnut on sandy soils. *Andhra Agric. J.* 37: 407-409.

- Sarkar, R.K., Chakraborty, A., and Saha, A. 1999. Effect of foliar application of potassium nitrate and calcium nitrate on groundnut (*Arachis hypogaea* L.). *Indian J. Agron.* 44(4): 809-812.
- Sebale, R.N. and Khuspe, V.S. 1986. Effects of moisture, phosphate and antitranspirants on growth, dry matter and yield of summer groundnut. *J. Maharashtra Agric. Universities* 11:13-16.
- Shelke, D. K. and Khuspe, V. S. 1980. Response of groundnut (*Arachis hypogaea* Linn.) to varying levels of irrigation, phosphorus and antitranspirant in summer. *J. Maharashtra Agric. Univ.* 5(2): 149-153.
- Shipkule, M.R., 2008. Nutrient requirement of summer groundnut by conjoint use of organic manure and inorganic fertilizers based on STCRC approach. Doctoral dissertation, Mahatma Phule Krishi Vidyaapeeth, Rahuri, 138p.
- Singh, A.L. and Chaudhari, V. 2008. Macronutrient requirement of groundnut: effects on growth and yield components. *Indian J. Plant Physiol.* 11(4): 401-402.
- Singh, F. and Oswalt, D. L. 1995. Groundnut production practices. *Skill Development Series* 1(3): 5-7.
- Singh, K.P. and Ahuja, K.N. 1985. Dry matter accumulation, oil content and nutrient uptake in groundnut (*Arachis hypogaea* L.) cv. T 64 as affected by fertilizers and plant density. *Indian J. Agron.* 25: 111-115.
- Singh, Lal, A., Jat, R. S., and Misra, J.B. 2009. Boron fertilization is a must to enhance peanut production in India. *The proceedings of the international plant nutrition colloquium.*

- Sireesha, P. V. G., Padmaja, G., and Babu, V. S. 2017. Effect of organic and inorganic sources of nutrients on available N, P, K and yield of rainfed groundnut. *Environ. Ecol.* 35. (1): 3446-3449.
- Skelton, B. J. and Shear, G. M. 1971. Calcium translocation in the peanut (*Arachis hypogaea* L.). *Indian J. Agron.* 63(3): 409-412.
- Skwierawska, M., Zawartka, L., and Zawadzki, B. 2008. The effect of different rates and forms of sulphur applied on changes of soil agro chemical properties. *Plant Soil Environ.*, 54: 171-177.
- Soliman, M.F., Kostandi, S.F., and Beusichem, M.L. 1992. Influence of sulfur and nitrogen fertilizer on the uptake of iron, manganese, and zinc by corn plants grown in calcareous soil. *Commun. Soil Sci. Plant Anal.* 23:1289- 1300.
- Subbiah, B. V. and Asija, G. L. 1956. A rapid procedure for the estimation of available nitrogen in soils. *Curr. Sci.* 25: 259- 260.
- Subrahmaniyan, K., Kalaiselvan, P., and Arulmozhi, N. 2001. Response of confectionery groundnut to micronutrients. *Legume Res.* 24(2): 139-140.
- Sumner, M. E. and Farina, M. P. 1986. Phosphorus interactions with other nutrients and lime in field cropping systems. *Adv. Soil Sci.* Springer, New York, pp.201-236.
- Survase, D.N., Dongale, J.H. and Kadrekar, S.B. 1986. Growth, yield, quality and composition of groundnut as influenced by FYM, calcium, sulphur and boron in lateritic soil. *J. Maharashtra Agric. Univ.* 11(1): 49-51.
- Teotia, U.S., Mehta, V.S., Ghosh, D., and Srivastava, P.C. 2000. Phosphorus-sulphur interaction in moongbean (*Vigna radiata* L.): I. Yield, phosphorus and sulphur contents. *Legume Res.* 23: 106-109.

- Thimmegowda, S. 1993. Effect of fertilizer levels on growth and yield of groundnut after kharif rice. *J. Res.* 21(42): 36-38.
- Tisdale, S.L., Nelson, W.L., Beaton, J.N., and Havlin, J.L. 1993. *Soil Fertility and Fertilizers* (Indian Reprint, 1997). Prentice Hall of India Private Ltd., New Delhi, 634p.
- Tripathi, S. B. and Hazra, C. R. 2003. Sulphur requirement of groundnut (*Arachis hypogaea*) under wheat (*Triticum aestivum*) based cropping system. *Indian J. Agric. Sci.* 73(7): 368-372.
- Umadevi, M., Munaswamy, V., Santaiah, V., and Rao, P. A. 1999. Uptake of major and secondary nutrients by groundnut as affected by sulphur levels under farmer's field conditions. *Leg. Res.* 22(1): 31-35.
- Umadevi, M., Munaswamy, V., Santaiah, V., and Rao, P. A. 1999. Field verification of critical available sulphur or sulphur availability index for groundnut in red soils. *J. Oilseeds Res.* 17(1): 168-170.
- Veerabhadrapa, B. H. and Yeledhalli, N. A. 2005. Effect of soil and foliar application of nutrients on growth and yield of groundnut. *Karnataka J. Agric. Sci.* 18(4): 814.
- Venkateswarlau, M. S. and Nath, V. S. 1989. N, P and K requirement of groundnut during kharif and rabi seasons in Chittoor district (AP). *J. Res. APAU*, 17(1): 45-46.
- Walker, M.E. 1975. Calcium requirements for peanuts. *Communications in Soil Sci. and Plant Analysis* 6(3): 299-313.



- Walkley, A. J. and Black, C. A. 1934. An examination of the degtjareff method for determining soil organic matter and a proposed modification of the chromic acid titration method. *Soil Sci.* 37: 29-38.
- Watanabe, F.S. and Olsen, S.R. 1965. Test of an ascorbic acid method for determining phosphorus in water and NaHCO<sub>3</sub> extracts from soil. *Soil Sci. Soc. Am. J.* 29(6): 677-678.
- Wiedenfeld, B. 2011. Sulfur application effects on soil properties in a calcareous soil and on sugarcane growth and yield. *J. Plant Nutrition* 34 (7): 1003-1013.
- Williams, J. H. 1979. The Physiology of Groundnuts (*Arachis hypogaea* L.) cv. Egret, nitrogen accumulation and distribution. *Rhodesian J. Agric. Res.* 17(1): 49-55.
- Yadav, G.L. 1990. Fertilizer requirement of groundnut (*Arachis hypogaea* L.) under clay land conditions on cultivators fields. *J. Oilseed Res.* 7: 133-135.
- Yakadri, M. and Satyanarayana, V. 1992. Response of rainfed groundnut (*Arachis hypogaea*) to potassium with varying levels of nitrogen and phosphorus. *Indian J. Agron.* 37: 202-202.
- York, E.T Jr. and Colwell, W.E. 1951. Soil properties, fertilization and maintenance of soil fertility. In *Symposium the peanut, the unpredictable legume*. Washington, pp. 122-171.

**INTERACTION OF PHOSPHORUS AND SULPHUR IN  
BLACK COTTON SOILS OF PALAKKAD (AEU: 23)  
UNDER GROUNDNUT (*Arachis hypogaea* L.)  
CULTIVATION**

**SHAHANA, C. K.  
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## ABSTRACT

Supply of nutrients in a balanced manner is one of the most important factors determining crop yield. Sometimes due to interaction between nutrients in soil, the applied nutrients may not be available for plant use. When the supply of one nutrient element affects the absorption and utilization of other nutrient element, the elements are said to be in interaction and interactions may be negative or positive. In Kerala, black cotton soils are seen in Chittur taluk of Palakkad district occupying an area of approximately 2000 ha. These soils are sandy clay loam, dark, calcareous, neutral to alkaline (pH 7.0 to 8.5), high in clay content and CEC. These soils are deficient in phosphorus (P) and sulphur (S). Finding the interaction between P and S in these soils will help to understand the factors determining the availability of these nutrients to crops.

The present study was carried out at Nellimed in Chittur taluk of Palakkad district during 2017-19. The objectives of the study were to find out the interaction of P and S in black cotton soils of Palakkad and to assess the treatment level of phosphorus and sulphur for maximizing the yield. The study consisted of a field experiment with groundnut variety, K-6 in black cotton soils of Chittur, Palakkad followed by analysis of soil, plant and pod samples taken from the experimental field. Soil samples were collected from different locations of Chittur and analyzed for available P and S. Field experiment was carried out in the field deficient in both P and S.

Experiment was laid out in factorial RBD with 17 treatments and 3 replications. Soil test based recommendation was taken as  $T_1$  and treatment combinations were made with four levels of P and four levels of S. The levels of N and K applications are kept same (based on POP recommendations of KAU) for all treatments, except for the first treatment where soil test based recommendation was given.  $P_0$  - 0 kg ha<sup>-1</sup>,  $P_1$  - 60 kg ha<sup>-1</sup>,  $P_2$  - 75 kg ha<sup>-1</sup> and  $P_3$  - 90 kg ha<sup>-1</sup> were the four levels of P and  $S_0$  - 0 kg ha<sup>-1</sup>,  $S_1$  - 10 kg ha<sup>-1</sup>,  $S_2$  - 20 kg ha<sup>-1</sup> and  $S_3$  - 30 kg ha<sup>-1</sup> were the four levels of sulphur.

Physical characteristics of soil viz., texture and bulk density were analyzed before experiment and chemical characteristics viz., pH, EC, organic carbon, N, P, K, Ca, Mg, S, Fe, Mn, Zn, Cu and B were analyzed before and after the experiment. Growth parameters such as plant height and number of leaves per plant were recorded at flowering, pegging, pod formation and

harvesting stages and yield parameters like number of pods per plant and yield were recorded at harvesting stage. The nutrient contents in plant were analyzed and uptake was computed in critical growth stages. The nutrient content, protein and oil content in pod were analyzed.

Soil nutrient status, plant nutrient content and uptake of nutrients were influenced by main effect and interaction effect of P and S. Application of P at  $90 \text{ kg ha}^{-1}$  and S at  $30 \text{ kg ha}^{-1}$  resulted in highest plant height and number of leaves per plant. Application of P at  $90 \text{ kg ha}^{-1}$  and S at  $30 \text{ kg ha}^{-1}$  resulted in highest number of pods per plant and yield. Highest protein and oil content were recorded by application of P at  $90 \text{ kg ha}^{-1}$  and S at  $30 \text{ kg ha}^{-1}$ .

Application of increased dose of P increased the availability of S. Application of P reduced the availability of Fe, Mn and Zn in soil due to the formation of insoluble compounds like Fe-P, Mn-P and Zn-P. Application of P enhanced the availability of N. The K content in soil was reduced with increased dose of P application. Ca content in soil was reduced due to P fertilizer application.

Plant nutrient content showed a decreasing trend from flowering to harvest stage. Application of P and S enhanced content of P in plant and pod. Content of N, P, Mg and S was increased with increased levels of P and content of Fe, Mn, Zn, Cu and B was reduced with increased dose of P. The uptake of nutrients followed an increasing trend from flowering to harvest due to increased dry matter production. Uptake of S, Fe, Zn and Cu by kernel is higher compared to plant.

P and S showed a positive interaction and P at  $90 \text{ kg ha}^{-1}$  and S at  $30 \text{ kg ha}^{-1}$  was found to be the best treatment in black cotton soils of Palakkad for high yield, protein and oil content in groundnut.

