SEED INVIGORATION WITH NANOPARTICLES FOR SEED YIELD AND QUALITY IN CHILLI

(Capsicum annuum L.)

by

RIYA MARY MATHEW

(2018 - 11 - 154)



DEPARTMENT OF SEED SCIENCE AND TECHNOLOGY COLLEGE OF HORTICULTURE VELLANIKKARA, THRISSUR - 680 656 KERALA, INDIA

2020

SEED INVIGORATION WITH NANOPARTICLES FOR SEED YIELD AND QUALITY IN CHILLI

(Capsicum annuum L.)

by

RIYA MARY MATHEW

(2018 - 11 - 154)

THESIS

Submitted in partial fulfilment of the requirements for the degree of

Master of Science in Agriculture

Faculty of Agriculture Kerala Agricultural University



DEPARTMENT OF SEED SCIENCE AND TECHNOLOGY COLLEGE OF HORTICULTURE VELLANIKKARA, THRISSUR - 680 656 KERALA, INDIA 2020

DECLARATION

I hereby declare that this thesis entitled "Seed invigoration with nanoparticles for seed yield and quality in chilli (*Capsicum annuum* L.)" is a bonafide record of research work done by me during the course of research and the thesis has not previously formed the basis for the award to me of any degree, diploma, associateship, fellowship or other similar title, of any other University or Society.

Vellanikkara

Riya Mary Mathew

(2018-11-154)

Date:

CERTIFICATE

Certified that this thesis entitled "Seed invigoration with nanoparticles for seed yield and quality in chilli (*Capsicum annuum* L.)" is a record of research work done independently by Ms. Riya Mary Mathew (2018-11-154) under my guidance and supervision and that it has not previously formed the basis for the award of any degree, diploma, fellowship or associateship to her.

Debalm Dr. Dijee Bastian

Mellianikkara

(Major Advisor, Advisory Committee) Professor (Plant Breeding and Genetics) Department of Seed Science and Technology College of Horticulture Vellanikkara

Dene

CERTIFICATE

We, the undersigned members of the advisory committee of Ms. Riya Mary Mathew (2018-11-154), a candidate for the degree of Master of Science in Agriculture with major field in Seed Science and Technology, agree that the mesis entitled "Seed invigoration with nanoparticles for seed yield and quality in chilli (*Capsicum annuum* L.)" may be submitted by Ms. Riya Mary Mathew, in partial fulfilment of the requirement for the degree.

Dr. Dijee Bastian

(Chairman, Advisory Committee) Professor (Plant Breeding and Genetics) Dept. of Seed Science and Technology College of Horticulture Vellanikkara

Dr. Rose Mary Francies Member, Advisory Committee) Professor and Head Dept. of Seed Science and Technology College of Horticulture Vellanikkara

Dr. Anita Cherian. K (Member, Advisory Committee) Professor and Head Department of Plant Pathology College of Horticulture Vellanikkara

Dr. K. Raja (Member, Advisory Committee) Assistant Professor (Seed Science and Technology) Dept. of Nano Science and Technology Tamil Nadu Agricultural University Coimbatore

<u>Acknowledgement</u>

ACKNOWLEDGEMENT

As a prelude to my thanksgiving, I wish to bow my head before the Almighty God for showering me the strength, knowledge, ability and making me confident and optimistic throughout my journey and enabled me to complete the thesis work successfully on time.

At this juncture, I would like to thank with lots of love, my godparents, Achacha, & Amma for their profound love, boundless affection and incessant inspiration which supported me to stay on track and I thankfully remember Ammichi, who has been showering blessings and care upon me from the heavens.

With immense pleasure I avail this opportunity to express my deep sense of whole hearted gratitude and indebtedness to my major advisor **Dr. Dijee Bastian**, Professor (Plant Breeding and Genetics), Department of Seed Science & Technology and Chairperson of my Advisory Committee for her precious suggestions, constant care and support, extreme patience and untiring assistance and also constructive criticism throughout the course of study starting from its preparatory stage to submission of thesis. This work would not have been possible without her valuable guidance and support. I consider myself extremely fortunate in having the privilege of being able to be guided by her.

I would like to express my extreme indebtedness and obligation to **Dr. Rose Mary Francies**, Professor and Head, Department of Seed Science & Technology and member of my advisory committee for her meticulous help, unwavering encouragement, forbearance, timely support and critical examination of the manuscript that has helped a lot for the improvement and preparation of the thesis.

I sincerely thank **Dr. Anita Cherian, K.** Professor and Head, Department of Plant Pathology for her support, ever willing help, scholarly suggestions and expert advice and help in taking the observations on seed microflora and interpretation of results. I express my heartiest gratitude to **Dr. K. Raja** Professor of Seed Science and Technology, Department of Nanoscience & Technology, TNAU, Coimbatore and member of my advisory committee for his encouragement, timely support, valuable guidance and creative suggestions throughout the period of my study.

It gives me great pleasure in acknowledging the support and help of all the nonteaching staff members of Department of seed Science and Technology, **Smitha chechi**, **Jeana chechi**, **Divya chechy** and **Bilha** for their sincere co-operation, timely help and assistance during the research work. It would be an omission, should I forget to mention and acknowledge the help extended by the farm laboures of Department of seed Science and Technology, **Abhilash chettan**, **Uthaman chettan**, **Sajeesh chettan**, **Omana chechy**, **Shantha chechy**, **Kunjumol chechy** without whose help it would have been difficult to conduct the study. I also thank Academic cell staffs, **Indira chechy** and **Naveen chettan** for all the help rendered.

I am particularly thankful for the support and co-operation of my classmates Jyothish, Milu and Harsha throughout the duration of my MSc. work. I also extend my heartfelt thanks to my seniors Gayathri chechy, Athulya chechy, Agina chechy, Bennet chettan, Nagendra chettan, Nishidha itha and Adharsh chettan. I wish to express my sincere gratitude to my dear juniors Sneha, Vaisakh, Vaishnavi and Olivia for their whole hearted support.

I would take this opportunity to express my heartfelt gratitude to **Rashi** for being the pillar of encouragement and strength during every time of need. I am particularly thankful for the heartfelt support, help and critical evaluation of **Jyothish**, **Nayana**, **Ashwinettan** and **Sarin** throughout the period of research.

It gives me great pleasure in acknowledging the love and gratitude of my beloved friends **Beegam**, **Akhil K.P**, **Vinu**, **Shuhda**, **Abinsha**, **Abhaya**, **Pooja**, **Neha**, **Reshma Ravi**, **Harya**, **Nidhin**, **Nibin**, **Sali** and **all of Taurus '14**. A special mention of thanks to **Lakshmi** and **Reshma shaji** and all others of **RASPUTINS' 18** for their cooperation, time and presence in my research. I am in dearth of words to express my gratitude, appreciation and indebtedness to my father, Mr. Mathew Scaria, mother, Smt. Teena Mathew and my little brother, Joeku for being inexhaustible source of inspiration, encouragement, support, blessing and love. I also thank my other family members especially Rithu chechy, Mercyamma, Mekha, Bindu aunty, Rani aunty and Meettu chechy for their prayers and support to complete my thesis successfully. Without the unconditional love, sacrifices, time and support of my family this two-year course would not have happened.

I thankfully remember the services rendered by all the staff members of Student's computer club, Library, Office of COH and Central Library, KAU.

A word of apology to those I have not mentioned in person and a note of thanks to everyone who helped me for the successful completion of this endeavor.

Above all, I thank the power that drives me all along.

Riva Mary Mathew

CONTENTS

| Chapter | Title | Page No. |
|---------|-----------------------------|----------|
| 1 | INTRODUCTION | 1-3 |
| 2 | REVIEW OF LITERATURE | 5-25 |
| 3 | MATERIALS AND METHODS | 27-38 |
| 4 | RESULTS | 39-58 |
| 5 | DISCUSSION | 59-69 |
| 6 | SUMMARY | 71-73 |
| | REFERENCES | i-xxii |
| | APPENDICES | |
| | ABSTRACT | |

LIST OF TABLES

| Table No. | Title | Page No. |
|-----------|--|----------|
| 1. | Details of seed treatment used for dry-dressing of chilli seeds | 28 |
| 2. | Details of seed treatment used for wet-dressing of chilli seeds. | 30 |
| 3. | Analysis of variance of seed yield and fruit yield attributes in chilli | 40 |
| 4. | Influence of normal grade and nano grade ZnO and TiO_2 on yield attributes in chilli | 41 |
| 5. | Influence of normal grade and nano grade ZnO and TiO_2 on seed yield and seed weight in chilli | 44 |
| 6. | Seed quality of chilly before storage | 46 |
| 7. | Influence of nano particles of ZnO and TiO_2 on germination of chilli | 47 |
| 8. | Influence of nano particles of ZnO and TiO ₂ on shoot length (cm) of chilli | 50 |
| 9. | Influence of nano particles of ZnO and TiO ₂ on root length (cm) of chilli | 51 |
| 10. | Influence of nano particles of ZnO and TiO_2 on dry weight (mg) of chilli | 52 |
| 11. | Influence of nano particles of ZnO and TiO ₂ on Vigour Index I of chilli | 54 |
| 12. | Influence of nano particles of ZnO and TiO ₂ on Vigour Index II of chilli | 55 |
| 13. | Influence of nano particles of ZnO and TiO_2 on electrical conductivity of seed leachate (μ Scm ⁻¹) of chilli | |
| 14. | Influence of nano particles of ZnO and TiO_2 on seed moisture and seed infection at the end of storage in chilli | 57 |

LIST OF FIGURES

| Figure No. | Title | Between |
|------------|--|---------|
| | | pages |
| 1. | Influence of normal grade and nano grade ZnO and TiO_2 on plant height in chilli | 60-61 |
| 2. | Influence of normal grade and nano grade ZnO and TiO_2 on days to harvest in chilli | 60-61 |
| 3. | Influence of normal grade and nano grade ZnO and TiO_2 on number of fruits per plant in chilli | 60-61 |
| 4. | Influence of normal grade and nano grade ZnO and TiO_2 on fruit length | 60-61 |
| 5. | Influence of normal grade and nano grade ZnO and TiO_2 on fruit weight at maturity in chilli | 62-63 |
| 6. | Influence of normal grade and nano grade ZnO and TiO_2 on fruit yield in chilli | 62-63 |
| 7. | Influence of normal grade and nano grade ZnO and TiO_2 on number of seeds per fruit in chilli | 62-63 |
| 8. | Influence of normal grade and nano grade ZnO and TiO_2 on seed yield per plant in chilli | 62-63 |
| 9. | Influence of nano grade ZnO and TiO ₂ in maintaining seed germination after seven months of storage | 66-67 |
| 10. | Influence of nano grade ZnO and TiO ₂ on vigour index I after seven months of storage | 66-67 |
| 11. | Influence of nano grade ZnO and TiO ₂ on vigour index II after seven months of storage | 68-69 |
| 12. | Influence of nano grade ZnO and TiO_2 on electrical conductivity after seven months of storage | 68-69 |

LIST OF PLATES

| Plate No. | Title | Between pages |
|-----------|---|------------------|
| 1. | Dry seed treatments in chilli | 28-29 |
| 2. | Field performance of dry dressed chilli seeds | 28-29 |
| 3. | Wet seed treatment using nanoparticles | 30-31 |
| 4. | Germination test using between paper method | 34-35 |
| 5. | Testing seeds for identification of seed microflora | 36-37 |
| 6. | Microorganisms observed after seven months of storage | 68-69 |

LIST OF APPENDICES

| Appendix No. | Title |
|-----------------|--|
| Ι | Monthly meteorological data from June 2019 to march 2020 |

LIST OF ABBREVIATIONS

| ISTA | : International Seed Testing Association |
|-------------------------|--|
| mL | : milli litre |
| h | : hours |
| EC | : electrical conductivity |
| mg | : milli gram |
| kg | : kilo gram |
| $\mu S \text{ cm}^{-1}$ | : micro Siemens per centimetre |
| ppm | : Parts per million |
| mm | : milli metre |
| nm | : nano metre |
| m | : metre |
| μm | : micro metre |
| °C | : degree Celsius |
| Pa | : Pascal |
| L | : Litre |
| ha | : Hectare |
| сс | : cubic centimeters |
| ANOVA | : Analysis of Variance |

| ZnO | : zinc oxide |
|-------------------|--|
| nZnO | : nano-zinc oxide |
| TiO ₂ | : titanium dioxide |
| nTiO ₂ | : nano-titanium dioxide |
| SE _m | : standard error of error mean sum of squares |
| CD | : critical difference |
| var. | : variety |
| CV. | : cultivar |
| NP | : nano particle |
| IMSCS | : Indian Minimum Seed Certification Standard |
| mg/kg | : milligram per kilogram |
| °N | : degrees north |
| °E | : degrees east |
| G | : gauge |
| DAT | : Days after transplanting |
| MAS | : months after storage |
| % | : per cent |
| et al. | : et alia (Latin: 'and others') |
| V.I. | : vigour index |
| PDA | : Potato Dextrose Agar |
| VFPCK | : Vegetable and fruit promotion council Kerala |

<u>Introductíon</u>

1. INTRODUCTION

Chilli (*Capsicum annuum* L.) is the largest produced spice crop in Asia. India is the largest producer followed by China. In India, Andhra Pradesh (49%), Karnataka (15%), Maharashtra (6%) and Tamil Nadu (3%) constitute nearly 75 per cent of the total area under chilli production. During 2018-19, in India, chilli was grown over an area of 364 thousand ha with a production of 3720 thousand MT (NHB, 2019).

In Kerala the net cropped area was 1.42 thousand ha with a production of 10.95 thousand MT and Palakkad, Thiruvananthapuram, Kollam were the major chilli producing districts during 2018. Chilli seed like all other agricultural commodities, invariably contains a high moisture content (60-85%) at the time of harvest, which must be brought down to 8-12 per cent. Poor seed storage is one of the major constraints in chilli production. The high temperature (25-37°C) and high humidity (70-90 %) prevailing in Kerala, makes storage, a still more difficult task.

Conservation of seed vigour and viability during storage is crucial. Seed qualities like germination and vigour can be considerably affected at any stage of storage. Storage and preservation of quality seed stocks till the next season is as important as producing quality seeds (Singh and Gill, 1996). Quality seed production begins from the time of harvest to planting. The importance of seed storage has been recognized ever since humans began to domesticate plants. Seeds are stored to facilitate planting in the following season and this practice is advantageous to seed growers and farmers to carry over seeds for more years. This results in accumulating supplies of desired seed stocks for use in years of low production (USDA, 1978). Seeds confirming to the prescribed standards in terms of high genetic purity, physical purity, germination per cent, low moisture content and good health are required to raise productivity.

Storage potential of seeds varies with the crop, variety and environment. Seeds with greater storage potential possess higher vigour and can be stored for longer time. The loss of viability impairs the biological value or function of seeds, which is to protect and nourish the living cells of the embryo, until seedling is established. Seed deterioration is inevitable and the best that can be done is to control its rate. Seed treatments can be defined as operations which aim to mitigate seed deterioration, to increase productivity through improving seed quality, good seedling establishment and minimizing crop loss by managing pest and diseases.

Seed deterioration can be alleviated with seed invigoration. Seed invigoration with nanoparticles is an emerging area and is successful in many crops. "Nanotech", is the study of manipulating matter on an atomic and molecular scale. The term 'nano' can be defined as the molecular aggregates with at least one dimension between 1 and 100 nm .The positive effects of nanoparticles include enhanced germination per cent, length of root and shoot and vegetative biomass of seedlings in many crop plants (Agarwal and Rathore, 2014). Nanotechnology has the potential to protect plants, monitor plant growth, detect plant diseases, weed management, stress tolerance, increase global food production, enhance food quality and reduce waste for sustainable intensification. Nano particles are being developed for slow release of fertilizers for plants. This makes nutrients more available to nanoscale plant pores and therefore result in efficient nutrient use (Suppan 2013). The mechanism by which nano particles alter biological activities is dependent on their shape, size, surface properties and composition. Growth of microorganism and attack of insects on seeds causes detrimental effects on germination due to loss of protein, total oil content, increased fatty acid and carbohydrates. These effects causes biochemical changes in seeds. The knowledge about the use of nanoparticles as antimicrobial agent will help in commercializing nanoparticle as a safer method for improving seed health. (Jo and Kim, 2009)

Treatment with zinc oxide nanoparticles (ZnO) and titanium dioxide (TiO₂) nanoparticles have been reported to have a positive impact on seedling vigour and storability. The most commonly used and widely applied types of nano particles are Zn NPs and they are known to have antibiotic properties. Zinc (Zn) is considered as an essential micronutrient for both animals and plants and this metal is extremely essential for enzyme system of plants as it acts as metal components, cofactors and other regulatory factors of many enzymes (Prasad *et al.* 2012). Zinc is essential for pollen function, chlorophyll production, germination and fertilization. Zinc NPs are toxic to several cell lines hence it is utilized for treating cancer cells and many other bacterial

cells. Moreover, Zn NPs are utilized in sensors, solar cells, photocatalytic purposes etc. (Varseem *et al.*, 2010)

TiO₂ NPs are having high stability, photo-catalytic activity and low costs. They are eco-friendly and safe for human. At lower concentrations, TiO₂ NPs promotes early seedling growth and seed germination. Nano-TiO₂ treatments could markedly promote aged seeds vigour and chlorophyll biosynthesis particularly, the ribulose-1,5-bisphosphate carboxylase/ oxygenase (Rubisco) activity and the photosynthesis efficiency (Gao *et al.*, 2006).The reason for the increased germination per cent could be the generation of hydroxide and superoxide anions by nano - TiO₂ that increased intake of oxygen and water needed for quick germination.

In consideration of the above, the present study entitled 'Seed invigoration with nanoparticles for seed yield and quality in chilli (*Capsicum annuum* L.)', was conducted with the following objectives.

- i. To standardize the optimum dose of nano ZnO and nano TiO₂ for seed treatment to increase yield in chilli.
- To standardize the optimum dose of nano ZnO and nano TiO₂ for seed treatment to improve viability and prolong seed longevity during storage
- iii. To compare the efficacy of titanium dioxide and zinc oxide based on their particle size (normal-size and nano-sized powder) on field performances.

<u>Revíew of Líterature</u>

2. REVIEW OF LITERATURE

Seed invigoration is any treatment which is physical, chemical or biological, applied to seed for improving the physiological status of the seed and thereby results in improved seed quality, better storage life and field performance. In any crop, seeds have to be invariably stored. During storage, seeds being hygroscopic in nature, absorbs moisture from the atmosphere and accelerates the ageing process with consequent loss of quality of seeds (Teckrony and Egli, 1997). It is seen that substandard seed quality generally delays seed germination and produces unhealthy seedlings which subsequently impairs the crop yield. Hence, seed invigoration is inevitable for maintaining qualities like germination, viability, vigour and yield potential of the seed. Seed treatments using nanoparticles is one such seed treatment practice which is currently gaining importance.

Nanotechnology is an emerging discipline with novel applications in agriculture. The nanoparticles whose size is 100 nm (or less than 100nm) in one or more dimensions have unique properties and they have the potential to improve plant metabolism. Zinc oxide nanoparticles increases biomass accumulation, maintains membrane integrity and helps in the functioning of several enzymes (Burman. *et al.*, 2013). Nano TiO₂ plays a key role in absorption of inorganic nutrients and breakdown of organic substances. It also helps to remove oxygen free radicles and thus increases photosynthetic rate (Khot *et al.*, 2012)

In this context, effect of seed invigoration with nanoparticles has been briefly reviewed in this chapter as,

- 2.1 Impact of nanoparticle seed treatment on crop growth and yield
- 2.2 Impact of nanoparticle seed treatment on seed quality
- 2.3 Seed infection in chilli

2.1 Impact of nanoparticle seed treatment on crop growth and yield

2.1.1 Plant height

| Treatment used | Сгор | Details of the experiment | Reference |
|----------------------------------|--------|--|---------------------------------|
| Bulk and nano grade ZnO | Peanut | Seeds of variety K-134 was treated with doses of bulk and normal ZnO. Chelated bulk ZnSO ₄ was used as a primary source of Zn. The seed treatments were given at the concentrations of $2g / 15$ Land $13g / 15$ L along with NPK (30-40-50). Nano scale treatments showed superior results than bulk treatments. Seeds treated with nano ZnO @ $2g / 15$ L recorded taller plants than other treatments (43.80 cm). | Prasad <i>et al.</i> , 2012 |
| Normal and nano TiO ₂ | Wheat | Seeds treated with 0.01%, 0.02%, 0.03 % titanium nanoparticle and bulk titanium showed significant differences in plant height. | Jaberzadeh <i>et al.</i> , 2013 |
| Nano ZnO and TiO ₂ | Tomato | ZnO @ 250 mg /kg recorded an increase in plant height by 24.5 % while the same dose of $TiO_2(250 \text{ mg /kg})$ did not have any significant effect | Raliya <i>et al.</i> , 2015 |
| Nano TiO ₂ | Barley | Application of nano TiO_2 at 500 and 1000 mg kg ⁻¹ resulted in a gradual increase in plant height with an increase in dosage. | Mattiello and Marchio, 2017 |

| ZnO nanoparticles | Wheat | Wheat seeds of variety Lassani-2008 were treated with different concentrations of ZnO @ 0, 25, 50, 75, and 100 mg L ⁻¹ . The crop was raised in greenhouse conditions and height of each plant recorded. The plant height increased by 37 per cent when the seeds were treated with | Rizwan <i>et al.</i> , 2018 |
|----------------------|--------|---|-----------------------------|
| ZnO | Red | ZnO @ 100 mg L ⁻¹ . Application of ZnO and FeO at 50 ppm and 60ppm respectively on red | Mahmoud <i>et al.</i> , |
| nanoparticles | radish | radish variety 'Champion' recorded in an increase in plant height. | 2019 |

2.1.2 Fruits per plant

| Treatment used | Сгор | Details of the experiment | Reference |
|-----------------------|-------------|--|---------------------------------|
| Bulk TiO ₂ | Cowpea | Treatments with TiO_2 at 125 cc/ha (double dose) exhibited more number of pods per plant (33.33) which was followed by single dose of TiO2 at 125 cc/ha (27.33), double dose (25) and single dose of TiO2 at 62 cc/ha (24). | Owalde <i>et al.</i> , 2008 |
| Nano Zn | Pomegranate | Nanoparticles of Zn at 120mg L^{-1} was found to be efficient for obtaining increased number of fruits per branch | Davarpanah <i>et al.</i> , 2017 |
| Nano ZnO | Mango | Treatments with Nano zinc @ 0.5 g/l and 1 g/l recorded highest fruit yield and it caused indirect effect on number of fruits per plant. | Zagzog et al., 2017 |

2.1.3 Fruit length

| Treatment used | Сгор | Details of the experiment | Reference |
|---------------------------------|--------|---|-----------------------------------|
| Bulk TiO ₂ | Cowpea | Treatments with TiO_2 at 125 cc/ha exhibited the highest pod length of 18.33 cm. | Owalde <i>et al.</i> , 2008 |
| Nano ZnO | Maize | Seeds treated with nano ZnO @ 400 ppm recorded the highest cob length of 16.40 cm, which was 18% more than control. | Subbaiah, 2014 |
| Nano Zn and TiO ₂ | Barley | Higher spike length was seen when treated with ZnO nanoparticle and TiO ₂ nanoparticle. | Janmohammadi <i>et al.</i> , 2016 |
| ZnO NP and FeO NP | Carrot | Plant material used for the treatment was Pusa Rudhira. Nanoparticles were applied in different doses (50ppm, 100 ppm and 150 ppm)of ZnO and FeO separately and in combined form. The results revealed that the highest root length of 19.75 cm were observed in treatment with nano ZnO @1000 ppm along with nano FeO @ 50 ppm. | Elizabath <i>et al</i> , 2017 |
| Nano TiO ₂ | Rice | Rice seeds were treated with nano TiO_2 @ 0, 10, 20, 50, 80 and 100 ppm in three replications. An increase in panicle length of 3.2 per cent was observed when treated with nano TiO_2 @ 50 ppm than the control. But at higher (100 ppm) doses the treatments showed negative effects on panicle length. | Debnath et al., 2020 |

| Treatment used | Сгор | Details of the experiment | Reference |
|-----------------------------------|--------|--|------------------------------------|
| Nano TiO ₂ | Barley | Nano particles were given at the rate of 0.01 %, 0.02 %, and 0.03 %. The results revealed that weight of spikelets increased when treated with 0.03 % nano TiO ₂ (6.7t/ha). | Moaveni <i>et al.</i> , 2011 |
| Nano and bulk TiO ₂ | Wheat | Wheat variety 'Pishtaz' were treated with 0.01% , 0.02% , 0.03% bulkand nano grade TiO2. The highest ear weight was obtained whentreated with nano TiO2 @ 0.02 % and least ear weight when treatedwith TiO2 @ 0.01% . | Jabersadeh <i>et al.</i> , 2013 |
| ZnO NP and FeO NP | Carrot | Plant material used for the treatment was Pusa Rudhira. Nanoparticles were given as foliar spray in different doses of 50ppm, 100 ppm and 150 ppm of ZnO and FeO separately and in combined form. The results revealed that the highest fruit weight of 72.33 kg was observed in treatment with nano ZnO @1000 ppm along with nano FeO @ 50 ppm. | Elizabath <i>et al</i> , 2017 |

2.1.4 Fruit weight at maturity

2.1.5 Fruit yield

| Treatment used | Сгор | Details of the experiment | Reference |
|-----------------------|--------|---|---------------------|
| Bulk TiO ₂ | Cowpea | Treatments with TiO ₂ at 125 cc/ha (double dose) exhibited the highest | Owalde et al., 2008 |
| | | yield of 948.90 kg. | |

| Nano TiO ₂ | Barley | Treatments were given with different doses of nano TiO ₂ and bulk | Moaveni et al., 2011 |
|-----------------------|-----------|--|-----------------------|
| | | TiO_2 . The results revealed that treatments at 0.03% recorded the | |
| | | highest yield of 5.7 t/ha. | |
| Bulk and | Peanut | Seeds of variety K-134 was treated with doses of bulk and normal | Prasad et al., 2012 |
| normal ZnO | | ZnO. The seed treatments were effeced at the concentrations of $2g / 151$ | |
| | | and 13g /15l along with NPK (30-40-50). Nano scale treatments | |
| | | showed superior results than bulk treatments. Seeds treated with nano | |
| | | ZnO @ 2g / 15 l recorded more yield (3121.54 kg/ ha) than bulk | |
| | | treatments. | |
| Nano titanium | Coriander | Coriander plants were treated twice with different doses of nano | Khater, 2015 |
| dioxide | | titanium dioxide at 2, 4 and 6 ppm at 30 DAP and 60 DAP. The | |
| | | treatment exhibited significant increase in growth parameters. The | |
| | | highest yield of 106.5 g was obtained when seeds were treated with | |
| | | nanoTiO ₂ at 6 ppm. | |
| Nano ZnO | Carrot | Nanoparticles were applied in Pusa Rudhira as foliar spray at different | Elizabath et al, 2017 |
| | | doses of 50ppm, 100 ppm and 150 ppm of ZnO and FeO separately and | |
| | | in combined form. The results revealed that maximum fruit yield was | |

| | | obtained when treated with nano ZnO @1000 ppm along with nano FeO @ 50 ppm. Along with individual effect, interaction effect was also found to be significant. | |
|------------------------|-------|--|-----------------------------|
| Nano Zn | Mango | Number of fruits per plant was recorded after 48 days of full bloom in mango trees. Treatments with Nano zinc @ 1 g/l showed increased fruit yield by 37.28 per cent than control. | Zagzog et al., 2017 |
| Bulk and normal ZnO | Rice | Seeds of variety Tarom Hashemi were treated without Zn, bulk Zn and nano Zn. The yield increased in both bulk and nano Zn seed treatments, but it was not significantly different. Nano Zn recorded the highest grain yield than control by 12.6 per cent and bulk Zn recorded 9.2 per cent. | Kheyri <i>et al.</i> , 2019 |

2.1.6 Seeds per fruit

| Treatment used | Crop | Details of the experiment | Reference |
|-----------------------|--------|---|---------------------|
| Bulk TiO ₂ | Cowpea | Treatments with TiO_2 at 125 cc/ha (double dose) exhibited more number of | Owalde et al., 2008 |
| | | seeds per pod of 18.67. | |
| Nano ZnO | Maize | Nano ZnO @ 400 ppm showed more number of grains per row (38.50) | Subbaiah, 2014 |
| | | which was 36% more than to control treatment. | |
| Nano ZnO | Rice | Higher numbers of seeds (294) were recorded in 5 g L^{-1} of nano ZnO treatment | Bala et al., 2019 |
| | | when compared to other nanoparticle treatments. | |

| Bulk and nano | Rice | The results showed that nano Zn recorded more number of filled grains | Kheyri et al., 2019 |
|---------------|------|--|---------------------|
| ZnO | | per panicle (175.1), while the control showed the least value (88.3 filled | |
| | | grains). | |

2.1.7 Seed yield per plant

| Treatment used | Crop | Details of the experiment | Reference |
|---------------------------|--------|---|--------------------------|
| Bulk and nano | Pearl | The variety HHB 67 was utilized for the experiment. Normal grade zinc | Tarafdar <i>et al.</i> , |
| ZnO | millet | oxide and nano grade zinc oxide was sprayed over germinated seedlings @ | 2014 |
| | | 10 mg L ⁻¹ . Grain yield was found to be increased by 37.7 per cent over | |
| | | control when treated with nanoparticles. | |
| Nano TiO ₂ and | Barley | Seeds were treated with nano iron -chelate, nano zinc and foliar application | Janmohammadi <i>et</i> |
| Zn | | was given with nano- TiO2 at 2000 ppm along with control. Nano | <i>al.</i> , 2016 |
| | | treatments were applied at the time of initiation of the tillering stage, | |
| | | booting, and milky stage. The results revealed that nano particles showed | |
| | | greater potential in obtaining higher yield. Nano Zn along with nano TiO ₂ | |
| | | @ 2000 ppm recorded the highest yield. | |
| Nano TiO ₂ | Rice | Rice seeds were treated with nanoTiO ₂ $@$ 0, 10, 20, 50, 80 and 100 ppm. | Debnath <i>et al.</i> , |
| | | The treated seeds exhibited higher values for several growth parameters. | 2020 |

| | | The results revealed that seed treatment with 20 ppm of nano TiO2 | |
|----------|---------|---|-------------|
| | | recorded increased seed yield by 19.22 per cent than control. | |
| Nano ZnO | Soybean | Nano ZnO @ 160 mg/kg showed the highest seed yield when compared to | |
| | | all other treatments. | et al.,2020 |

2.1.8 100 seed weight

| Treatment used | Crop | Details of the experiment | Reference |
|-----------------------|--------|--|--------------------|
| Bulk TiO ₂ | Cowpea | The treatments irrespective of number of doses, and concentration showed | Owalde et al., |
| | | significant differences. Treatments with TiO ₂ at 125 cc/ha exhibited the | 2008 |
| | | highest 100 seed weight (18.47 g) | |
| Bulk and nano | Peanut | Seeds of variety K-134 was treated with various doses of bulk and nano | Prasad et al.,2012 |
| ZnO | | ZnO. The seed treatments were given at the concentrations of $2g / 15 L$ and | |
| | | 13g /15 L along with NPK (30-40-50). Nano scale treatments showed | |
| | | superior results than bulk treatments. Seeds treated with nano ZnO $@2g/$ | |
| | | 15 L recorded highest 100 kernal weight of 36.25g. | |

2.2 Impact of nanoparticle seed treatment on seed quality

2.2.1 Germination

| Treatment used | Crop | Details of the experiment | Reference |
|----------------|---------|--|-----------------|
| Nano ZnO | Soybean | Seeds treated with nano ZnO at 1000 mg L ⁻¹ increased seed germination. | Lopez-Moreno et |
| | | | al., 2010 |

| Nano ZnO | Rice | Irrespective of dose of treatments, application of nano ZnO (10-1000 | Boonyanitipong <i>et</i> |
|-----------------------|----------|---|--------------------------|
| | | mgL ⁻¹) recorded 100 per cent germination. The results revealed that seed | al. 2011 |
| | | treatments did not adversely affect seedling growth. | |
| Nano and bulk | Wheat | Seed were treated with both nano and bulk TiO_2 at 0, 5, 20, 40, 60 and 80 | Feizi et al., 2013 |
| titanium dioxide | | mgL ⁻¹ . It was observed that treatment with nano TiO ₂ @ 60ppm (76 per | |
| | | cent) showed greater potential in improving germination and least values | |
| | | (41%) were recorded in bulk TiO ₂ @ 60 ppm. It was seen that, mean | |
| | | germination time was improved by 31.8 per cent when treated with nano | |
| | | TiO ₂ at 40 ppm when compared to control. | |
| Nano TiO ₂ | Tomato | Seeds were incorporated with nano TiO_2 at 0, 100, 200, and 400 mg L ⁻¹ . | Haghighi <i>et al.</i> , |
| | Onion | Treatments with 100 and 200 mg L ⁻¹ showed superior effects on | 2014 |
| | | germination. Nano TiO ₂ @ 100 mg L^{-1} recorded the highest values in | |
| | | tomato (100%) and onion (30%). | |
| Nano TiO ₂ | Radish | Seeds were treated with different doses of nano TiO_2 (0, 100, 200, and | Haghighi <i>et al.</i> , |
| | | 400 mg L^{-1}). At 400 mg L^{-1} , the highest germination (100%) was | 2014 |
| | | observed. | |
| Nano ZnO | Mungbean | ZnO nanoparticles for seed treatment was prepared by dissolving | Jayarambabu et al., |
| | | nanoparticles in distilled water by placing on sonicator for 15 minutes. | 2014 |
| | | Seeds were treated with 20, 40, 60 and 100mg nano ZnO and reported an | |
| | | increase in germination. Treated seeds showed the highest germination at | |

| | | 20 mg (100%) followed by 40mg (95 %) | |
|--------------------------|-----------|---|----------------------|
| Nano ZnO | Maize, | Seeds were treated with nano-scale ZnO at 25 mg and 50 mg. The results | Adhikari et al.,2016 |
| | soyabean, | of germination test revealed that treated seeds showed better germination | |
| | pigeon | of $98 - 100$ per cent compared to control. | |
| | pea, okra | | |
| Nano ZnO and | Cowpea | Nano ZnO (0, 2, 4, 8, 10, 15 ppm) and nano TiO2 (0, 10, 20, 30, 40, 50 | Priya, 2016 |
| TiO2 | | ppm) was incorporated into cowpea seeds. Highest germination per cent | |
| | | was recorded when treated with 20 ppm nano TiO ₂ (98) and 2 ppm nano | |
| | | ZnO (98). An increase of germination by 22.91 per cent was observed | |
| | | when treated with both the nanoparticles. | |
| Nano ZnO | Chilly | Seeds were treated with nano ZnO at 0.0, 0.25, 0.50 and 0.75g. Highest | Afrayeem and |
| | | seed germination (65.7 %) was observed when treated with 0.75 g nano | Chaurasia, 2017 |
| | | particle. | |
| Nano TiO ₂ | Chickpea | Seeds of variety PBG-7 was wet dressed with nano ZnO and TiO_2 at 100, | Hajra and Mondal, |
| | | 500 and 1000 ppm. Nano TiO_2 at 500 ppm showed highest germination. | 2017 |
| Nano ZnO | Maize | Nano zinc oxide treatments (800 ppm, 1000 ppm, 1200 ppm 1400 ppm | Meena et al., 2017 |
| | | and 1600 ppm) showed superior performances. Treatments with 1000 and | |
| | | 1200 ppm recorded 100 per cent germination. | |
| Bulk and nano | Chilly | Seeds were dry dressesd with nano and bulk ZnO and TiO ₂ each at 750, | Kumar, 2019 |
| ZnO and TiO ₂ | | 1000 and 1250 mg kg ⁻¹ . Nano ZnO at 1000 mg kg ⁻¹ showed highest | |

| Bulk and nano ZnO | Chilly | germination of 75 per cent and least values (66%) was observed in TiO2at 750 mg kg ⁻¹ .Seeds treated with nanoparticles of ZnO at 500 mg/kg and bulk ZnO at1300 mg/kg showed superior performances in maintaining germinationabove 60 per cent till the end of ten months of storage. | Gayathri, 2019 |
|-----------------------------------|--------|---|------------------------------|
| Bulk and nano TiO ₂ | Chilly | Seed treatments with nano-TiO2 @ 500 mg/kg, 900 mg/kg, 1300 mg/kg and 500 mg/kg retained 60 per cent germination at the end of ten months of storage. | Gayathri, 2019 |
| Nano ZnO | Wheat | Wheat variety Lok-1 was treated with various doses of nano ZnO (10000, 8000, 6000, 4000 and 2000 ppm). Seeds treated with nano ZnO @ 2000 ppm exhibited 100 per cent germination. A gradual reduction in germination was observed at higher concentrations. The shoot length and root lengths are decreased with increase in concentrations of nanoparticles | Bagawade and Jagtap, 2020 |

| Rice | Rice variety CN-1794-2 were treated with six different doses of nano | Debnath | et | al., |
|------|--|--|--|--|
| | TiO ₂ at 0, 10, 20, 50, 80 and 100 ppm for 48 hours. Significantly higher | 2020 | | |
| | differences were found in all the treatments irrespective of concentration | | | |
| | of nanoparticles. Seed germination was improved by 85 per cent when | | | |
| | treated with 20ppm nanoparticle solution when compared to control. | | | |
| | Higher germination of 98 per cent and 98 per cent was recorded in | | | |
| | treatments with 20 ppm and 50 ppm. | | | |
| | Rice | TiO ₂ at 0, 10, 20, 50, 80 and 100 ppm for 48 hours. Significantly higher differences were found in all the treatments irrespective of concentration of nanoparticles. Seed germination was improved by 85 per cent when treated with 20ppm nanoparticle solution when compared to control. Higher germination of 98 per cent and 98 per cent was recorded in | TiO2 at 0, 10, 20, 50, 80 and 100 ppm for 48 hours. Significantly higher2020differences were found in all the treatments irrespective of concentrationof nanoparticles. Seed germination was improved by 85 per cent whentreated with 20ppm nanoparticle solution when compared to control.Higher germination of 98 per cent and 98 per cent was recorded in | TiO2 at 0, 10, 20, 50, 80 and 100 ppm for 48 hours. Significantly higher2020differences were found in all the treatments irrespective of concentration2020of nanoparticles. Seed germination was improved by 85 per cent whentreated with 20ppm nanoparticle solution when compared to control.Higher germination of 98 per cent and 98 per cent was recorded in |

2.2.2 Seedling length

| Treatment used | Сгор | Details of the experiment | Reference |
|-----------------------|---------|--|--------------------------|
| Nano ZnO | Rice | At low concentration of 10 mg L ⁻¹ , nano ZnO showed positive effect on | Boonyanitipong et |
| | | root length. | al., 2011 |
| Nano TiO ₂ | Tomato | Seeds treated with nano TiO ₂ (100, 200 and 400 mg L ⁻¹) were grown | Haghighi <i>et al.</i> , |
| | | under greenhouse conditions. At 400 mg L ⁻¹ seedling recorded the highest | 2014 |
| | | root length of 4.5 cm. | |
| Nano TiO ₂ | Raddish | Seeds treated with nano TiO_2 @ 100 mg L ⁻¹ recorded the highest shoot | Haghighi <i>et al.</i> , |
| | | length of 2.21 cm. | 2014 |

| Nano ZnO and | Cowpea | ZnO and TiO ₂ nanoparticles at various doses showed significant effects | Priya, 2016 |
|------------------|----------|--|--------------------|
| nano TiO2 | | on seedling length. Nano ZnO at 2 ppm and nano TiO ₂ at 20 ppm showed | |
| | | superior results. At low concentration, ZnO and TiO2 nanoparticles | |
| | | showed good effect on root and shoot length. An increase of root length | |
| | | by 65.96% and shoot length by 61.82% was observed when treated nano | |
| | | TiO ₂ . | |
| Nano ZnO | Chilly | Seeds were treated with nano ZnO at 0.0, 0.25, 0.50 and 0.75g. The results | Afrayeem and |
| | | showed that at lower concentrations (0.25 and 0.50g) a decrease in shoot | Chaurasia, 2017 |
| | | and root lengths was seen and at 0.75 g an increase in length was | |
| | | observed. | |
| Nano ZnO and | Chickpea | Longer roots of 12.36 cm was produced when treated with nano TiO_2 at | Hajra and Mandal, |
| TiO ₂ | | 1000 ppm when compared to nano ZnO (0.93 cm) treatments. In case of | 2017 |
| | | shoot length, nano ZnO @ 500 ppm produced shoots of 7.6 cm and 9.2 | |
| | | cm by nano TiO ₂ at 1000ppm. | |
| Nano ZnO | Maize | Nano ZnO treatments were given at the rate of 800, 1000, 1200, 1400, | Meena et al., 2017 |
| | | 1600 ppm. Seed treatments at 1200 ppm of nano ZnO exhibited shoot | |
| | | length of 3.9 cm and root length of 6.5 cm. | |

| Normal and nano | Maize | Seeds were wet dressed with normal and nano ZnO at 500 ppm, 1000 | Tiwari, 2017 |
|--------------------------|--------|--|-------------------------|
| ZnO | | ppm and 2000 ppm for 2 and 4 hours each. Seed treatment with nano | |
| | | grades of ZnO @ 1000 ppm recorded higher root length of 6.82 cm and | |
| | | shoot length of 1.94 cm. It was observed that at higher doses of | |
| | | treatment like 2000 ppm root length was reduced. | |
| Bulk and nano | Chilly | Seed treatment with nano ZnO at 1000 mg kg ⁻¹ recorded maximum | Kumar <i>et al.</i> , |
| ZnO and TiO ₂ | | shoot length of 4.3 cm and root length of 11.90 cm. | 2019 |
| Nano ZnO | Wheat | Wheat variety Lok-1 treated with nano ZnO @ 2000 ppm exhibited the | Bagawade and |
| | | highest shoot and root length. | Jagtap, 2020 |
| Nano TiO ₂ | Rice | Significant differences in seedling length was observed when treated | Debnath <i>et al.</i> , |
| | | with nano TiO ₂ at 0, 10, 20, 50, 80 and 100 ppm. Nano seed treatment | 2020 |
| | | @ 50ppm and 80 ppm recorded the highest root length and shoot | |
| | | respectively when compared to other treatments. | |

2.2.3 Dry weight

| Treatment used | Сгор | Details of the experiment | Reference |
|------------------|----------|---|---------------------|
| Nano ZnO | Chickpea | Nano ZnO treated seeds at lower doses (1.5 ppm) improved dry matter | Burman et al., 2013 |
| | | accumulation in seedlings | |
| Nano and bulk | Fennel | At 5 ppm and 80 ppm highest shoot biomass was recorded in bulk ${\rm TiO_2}$ | Feizi et al., 2013 |
| titanium dioxide | | (1.18 mg) and nano TiO ₂ $(1.16 mg)$ respectively. Bulk TiO ₂ at 20 ppm and | |

| | | nano TiO_2 at 5ppm and 20 ppm showed the highest root biomass. | |
|-----------------------|----------|--|--------------------------|
| Nano TiO ₂ | Tomato | Seeds were treated with various doses of nano TiO_2 (100, 200 and 400 mg | Haghighi <i>et al.</i> , |
| | | L^{-1}). Highest seedling dry weight of 0.74 g was observed in seeds treated | 2014 |
| | | with nano titanium dioxide at 400 mg L^{-1} . | |
| Nano TiO ₂ | Raddish | Treatments using nano TiO ₂ at 100 mg L ⁻¹ recorded the highest seedling | Haghighi <i>et al.,</i> |
| | | dry weight of 0.21g. | 2014 |
| Nano ZnO and | Cowpea | Seedling dry weight showed significant differences when treated with | Priya, 2016 |
| nano TiO2 | | various doses of nano ZnO (0, 2, 4, 8, 10, 15 ppm) and nano TiO2 (0, 10, | |
| | | 20, 30, 40, 50 ppm). Nano ZnO at 8 ppm and nano TiO_2 at 20 ppm showed | |
| | | superior results. An increase of dry weight by 118.6% in nano ZnO and | |
| | | 148.8 per cent in nano TiO ₂ was observed. | |
| Nano ZnO | Chickpea | Nano ZnO treatments at 100, 500 and 1000 ppm was incorporated into | Hajra and Mondal, |
| | | chickpea seeds. Among the treated seeds, treatments at 1000 ppm | 2017 |
| | | recorded higher seedling dry weight. | |
| Zinc oxide NPs | Chilli | Seeds were wet dressed with doses of nano ZnO at 0, 100, 200 and 500 | García-López et al., |
| | | ppm. Lower doses recorded higher seedling dry weight. | 2018 |

2.2.4 Vigour index

| Treatment | Сгор | Details of the experiment | Reference |
|------------------|--------|---|--------------------|
| used | | | |
| Nano and bulk | Fennel | Application of nano TiO ₂ exhibited significant improvement in | Feizi et al., 2013 |
| titanium | | vigour index while bulk TiO2 reduced seedling vigour index II. | |
| dioxide | | Nano TiO ₂ @ 5 ppm recorded the higher values for vigour index II | |
| | | compared to other treatments. | |
| Nano ZnO and | Cowpea | Seed treatment with nano ZnO at 2 ppm and nano TiO_2 at 20 ppm | Priya, 2016 |
| nano TiO2 | | showed superior results than control. An observed increase of seed | |
| | | vigour by 100 per cent and 116.45 per cent respectively was seen in | |
| | | nano ZnO and nano TiO ₂ . | |
| Nano ZnO and | Onion | Treatments at 750, 1000, 1250 and 1500 mg kg ⁻¹ was given using | Anandaraj and |
| TiO ₂ | | nano grades of ZnO and TiO ₂ . Seeds treated with nano grades of | Nataraja, 2017 |
| | | ZnO and TiO ₂ at 1000 mg kg ⁻¹ recorded the highest seedling vigour | |
| | | of 998 and 795 respectively. | |
| Nano ZnO | Maize | Nano ZnO treatments were given at the rate of 800, 1000, 1200, | Meena et al., 2017 |
| | | 1400, 1600 ppm. Seed treatments at 1200 ppm of nano ZnO | |
| | | recorded the highest vigour index I of 1040. | |

| Normal and | Maize | Seeds were wet dressed with normal and nano ZnO at 500 ppm, | Tiwari, 2017 |
|--------------------------|--------|--|----------------------------|
| nano ZnO | | 1000 ppm and 2000 ppm for two and four hours each. Seed | |
| | | treatment with nano grades of ZnO @ 1000 ppm recorded the high | |
| | | vigour index I of 861.4 and 845.4 when soaked at two and four | |
| | | hours respectively. | |
| Nano ZnO | Peanut | Nano scale ZnO of 25 mm particle size at a concentration of 1000 | Prasad et al., 2012 |
| | | ppm exhibited the higher seed vigour index I of 1701. | |
| Bulk and nano | Chilly | Vigorous seedlings were observed in treatments with nano ZnO at | Kumar <i>et al.</i> , 2019 |
| ZnO and TiO ₂ | | 1000 mg kg^{-1} (1285) and least values in control (861). | |
| Bulk and nano | Chilly | Seeds treated with nanoparticles of ZnO at 500 mg/kg and bulk ZnO | Gayathri, 2019 |
| ZnO | | at 1300 mg/kg showed superior performances in maintaining seed | |
| | | vigour till the end of ten months of storage. | |
| Bulk and nano | Chilly | Seed treatments with nano-TiO ₂ @ 500 mg/kg and 1300 mg/kg | Gayathri, 2019 |
| TiO ₂ | | recorded high seed vigour. | |
| Nano TiO ₂ | Rice | Rice seeds were treated with nano TiO_2 at 0, 10, 20, 50, 80 and 100 | Debnath et al., 2020 |
| | | ppm showed significant differences in seed vigour over control. | |
| | | Seeds treated with nano TiO_2 @ 20ppm recorded the highest vigour. | |

| 2.2.5 Electrical conductivity | of seed leachate |
|-------------------------------|------------------|
|-------------------------------|------------------|

| Treatment used | Сгор | Details of the experiment | Reference | | | |
|-----------------|-----------|--|-----------------------|--|--|--|
| Nano zinc oxide | Groundnut | ndnut Seeds of variety VRI-2 was treated with 750, 1000, 1250 mg/kg S | | | | |
| | | of seeds of nano ZnO. Seed treatments @ 1000 mg kg ⁻¹ | 2014 | | | |
| | | recorded lower electrical conductivity of 0.347dSm ⁻¹ at the end | | | | |
| | | of 12 months of period of storage. | | | | |
| Nano titanium | Maize | Nano TiO ₂ @ 200 mg kg ⁻¹ exibited lower EC (0.278 dSm ⁻¹) | Vijayalakshmi et al., | | | |
| dioxide | | compared to control when treated with various doses. | 2018 | | | |
| Bulk and nano | Chilly | The nanoparticle seed treatments significantly influenced | Kumar et al., 2019 | | | |
| grade ZnO | | electrical conductivity. Nano ZnO at 1000 mg kg ⁻¹ recorded the | | | | |
| | | lowest EC (0.118 dSm ⁻¹) compared to control (0.149 dSm ⁻¹). | | | | |
| Nano and bulk | Chilly | Nano ZnO at 500 mg/kg and 250 mg/kg reduced leakage of | Gayathri, 2019 | | | |
| Zno | | electrolyte from seeds during the period of storage | | | | |

2.3 Seed infection in chilli

Seed infections are caused by fungi, bacteria and viruses which results in inferior seed quality and reduction in yield. Loss of seeds at the time of storage is the greatest challenge in quality seed production. Fungus causes 70% of diseases in several crops. They invade host plants and establish infection by colonizing from inside.

Temperature, moisture, aeration etc. are the factors contributing growth of microbes during storage. Germinability and vigour of seeds get reduced as pathogens are present with the advancement of storage. The seeds may show discolouration and produce abnormal odours. Several toxins produced by fungus causes negative impact on growth of seedling. Thus, biochemical changes occurs inside the seed due to which nutrients are lost (Aher, 2013). The pathogens causing diseases in chilly are enlisted below.

| Pathogen observed | Reference |
|--|-------------------------------|
| Alternaria alternata, , A. niger, A. flavus, | Sharfun-Nahar <i>et al.</i> , |
| Cephalosporium sp., Bipolaris australiensis, B. | 2004 |
| spicifera, Cladosporium spp., Drechslera spp., | |
| Colletotrichum capsici, Rhizoctonia solani, | |
| Curvularia lunata, Macrophomina phaseolina, | |
| Fusarium moniliforme, F. solani, Penicillium spp., | |
| Verticillium albo-atrum and Trichoderma harzianum,. | |
| Penicillium digitatum, Aspergillus flavus and | Balogun et al., 2005 |
| Aspergillus niger | |
| Aspergillus niger, Rhizopus nigricans, Alternaria | Jogi et al., 2010 |
| alternate, Colletotrichum capsica, Macrophomina | |
| phaseolina, Penicillium citrinum, Fusarium | |
| oxysporum and Curvularia lunata, | |
| Agar plate method:, Aspergillus niger and A. flavus, | Chigoziri and Ekefan, |
| Colletothrichum capsici, Bispora betulina, Humicola | 2013 |

| fuscoatra, Phoma spp, Humicola dimorphospora, , | |
|---|------------------------------|
| Periconia byssoides, Botryotrichum piluferum, | |
| Phomopsis spp, | |
| Colletotrichum capsici, Rhizopus stolonifer, | Alam <i>et al.</i> , 2014 |
| Aspergillus flavus. Fusarium moniliforme and | |
| Curvularia lunata, | |
| Colletotrichum capsici, Alternaria alternate, C. | Machenahalli et al., 2014 |
| gloeosporioides, Fusarium sporotrichioides, C. | |
| acutatum and F. oxysporum. | |
| Aspergillus niger, Pencillium spp., Aspergillus flavus, | Navya, 2016 |
| and Alternaria spp. | |
| Aspergillus spp. Alternariaspp, and Pencilliumspp | Sandhya 2016 |
| Aspergillus flavus, Aspergillus niger, Rhizopus spp., | Chauhan <i>et a</i> l., 2018 |
| Penicillium spp., Colletotrichum capsici, Fusarium | |
| solani | |
| Aspergillus flavus, Alternaria spp Aspergillus niger, | Gayathri, 2019 |
| and Mucor spp. | |
| Curvularia pallescens, Myrothecium sp., Fusarium | Ghyasi <i>et al.</i> , 2020 |
| verticillioides, Cladosporium sphaerospermum, | |
| ,Fusarium solani, Aspergillus flavus, Aspergillus | |
| niger, Penicillium sp., Colletotrichum gloesporioides, | |
| Colletotrichum capsici, Rhizopus sp. And | |
| Macrophomina phaseolina | |

<u>Materíals and Methods</u>

3. Materials and methods

The study entitled 'Seed invigoration with inorganic nanoparticles in chillies (*Capsicum annuum* L.)', was conducted in the Department of Seed Science and Technology, College of Horticulture, Kerala Agricultural University, Vellanikkara from May 2019 to March 2020.

3.1 Experimental site

The field and laboratory experiments were conducted in the Department of Seed Science and Technology, College of Horticulture, Vellanikkara, Thrissur.

3.2 Climatic conditions

Vellanikkara, of Thrissur district, is located 22.25 m above mean sea level and its co-ordinates lies between 10.5452 °N and 76.2740 °E. A hot and humid climate prevails in this region.

3.3 Experimental material

Chilli seeds of variety Anugraha, harvested in May 2019 were procured from Vegetable And Fruit Promotion Council Keralam at Alathur, Palakkad.

3.4 Experimental details

The study was divided into two experiments as listed below:

Experiment 1: Field performance of dry dressed inorganic nanoparticles on seed yield in chilli

Experiment 2 - Seed storage studies with wet dressed inorganic nanoparticles

3.4.1 Experiment 1: Field performance of dry dressed inorganic nano particles on seed yield in Chilli

3.4.1.1 Layout of the experimental field

Design: Randomized Block Design (RBD)

Variety: Anugraha

Treatments: 13

Replication: 3

Total number of plots: 39

Plot size: 5 m X 1.5 m

Spacing: $45 \text{ cm} \times 45 \text{ cm}$

Number of plants per plot: 20

3.4.1.2 Treatment details

Seeds of chilli variety Anugraha were dry dressed with bulk and nano sized zinc oxide (ZnO) and titanium dioxide (TiO₂) particles in a Randomized block Design with thirteen treatments in 3 replications.

3.4.1.3 Dry seed treatment

1g seeds of chilli were dry dressed with normal grade ZnO, nano grade ZnO, normal grade TiO_2 and nano grade TiO_2 in screw capped glass bottles at room temperature using the specified quantity of chemicals detailed in Table 1. The glass bottles along with seed and nanoparticles was shaken gently 5 times for 3 min, at an interval of 3hrs. Untreated seeds served as control.



Untreated seeds



Chemical for seed treatment



Seeds treated with nanoparticle

PLATE 1: Dry seed treatments in chilli

| COLLEGE | OF HORTIC | LTURAL UNIVER ULTURE, VELLA d Science and Technolog | NIKKARA |
|--|--|--|----------------------------|
| Seed invigor seed yield and | ation with inc quality in chi | organic nanoparticl llies (<i>Capsicum an</i> | les on nuum L.) |
| Design: RBD Varie Area: 8.16 cents Spac Treatments: 13 | ety: Anugraha cing: 45 cm × 45 c | Replications: 3 cm Date of transplanti | ng: 17/07/2019 |
| T ₃ ⁻ : nano ZnO | (500 mg/kg) (900 mg/kg) (1300 mg/kg) | F _a : nano TiO ₂ (500 mg) F _a : nano TiO ₂ (900 mg) F ₁₀ : nano TiO ₂ (1300 mg F ₁₁ : TiO ₂ (500 mg/kg) T ₁₂ : TiO ₂ (900 mg/kg) | /kg |
| Τ ₆ : ZnO (900 Τ ₇ : ZnO (1300 | | T ₁₃ :TiO ₂ (1300 mg/kg) | |
| Name of the student : R Admission number : 20 | iya Mary Mathew 018-11-154 | Major Advisor : Dr | Dijee Bastian Professor |



PLATE 2: View of experimental plot

| Treatment | Details |
|-----------------|---|
| T1 | Control (Untreated seeds) |
| T ₂ | 500 mg nano ZnO/ kg of seed |
| T ₃ | 900 mg nano ZnO/ kg of seed |
| T ₄ | 1300 mg nano ZnO/ kg of seed |
| T ₅ | 500 mg ZnO/ kg of seed |
| T ₆ | 900 mg ZnO/kg of seed |
| T ₇ | 1300 mg ZnO/kg of seed |
| T ₈ | 500 mg nano TiO ₂ /kg of seed |
| Т9 | 900 mg nano TiO ₂ /kg of seed |
| T ₁₀ | 1300 mg nano TiO ₂ /kg of seed |
| T ₁₁ | 500 mg TiO ₂ /kg of seed |
| T ₁₂ | 900 mg TiO ₂ /kg of seed |
| T ₁₃ | 1300 mg TiO ₂ /kg of seed |

Table 1: Details of seed treatment used for dry-dressing of chilli seeds

The treated seeds were raised in nursery and transplanted to main field after four weeks. The experimental crop was raised as per Package of practices recommendations of Kerala Agricultural University, to study the field performance of dry dressed inorganic nanoparticles on seed yield.

3.4.2. Experiment 2 - Seed storage studies with wet dressed inorganic nanoparticles

3.4.2.1 Treatment details

Chilli seeds were wet dressed with different doses of nano sized zinc oxide (ZnO) and titanium dioxide (TiO₂) particles.

Design: Completely Randomized Design (CRD)

Variety: Anugraha

Treatments: 13

Replication: 3

3.4.2.2 Wet seed treatment

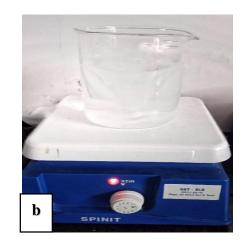
The nanoparticles were dispersed in distilled water by sonicating for 15 min (Kumar, 2019). The chilli seeds were soaked in different solutions as mentioned in Table 2, for three hours. Seeds soaked in water served as control. Soaked seeds were then removed and dried back to a moisture content of less than 8 per cent.

Table 2: Details of seed treatment used for wet-dressing of chilli seeds

| Treatment | Details |
|----------------|-----------------------------|
| T_1 | Control (Untreated seeds) |
| T ₂ | 100 mg nano ZnO/ kg of seed |
| T ₃ | 250 mg nano ZnO/ kg of seed |
| T4 | 500 mg nano ZnO/ kg of seed |



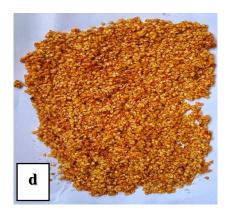
Solution containing nanoparticles



Solution kept for sonicating for dispersing nanoparticles



Seeds soaked in treatment solution



Drying of seeds after wet dressing



Seeds packed after wet dressing for storage

PLATE 3: Wet seed treatment using nanoparticles

| T ₅ | 750 mg nano ZnO/ kg of seed |
|-----------------|--|
| T ₆ | 1000 mg nano ZnO/ kg of seed |
| T ₇ | 1250 mg nano ZnO/ kg of seed |
| T ₈ | 100 mg nano TiO ₂ / kg of seed |
| T9 | 250 mg nano TiO ₂ / kg of seed |
| T ₁₀ | 500 mg nano TiO ₂ / kg of seed |
| T ₁₁ | 750 mg nano TiO ₂ / kg of seed |
| T ₁₂ | 1000 mg nano TiO ₂ / kg of seed |
| T ₁₃ | 1250 mg nano TiO ₂ / kg of seed |

3.4.2.3 Method of seed packing and storage

The treated and untreated seeds in each treatment were packed separately in 700 gauge polythene bags and stored under ambient conditions for a period of seven months. Separate seed lots were maintained in each replication of a treatment for ease of drawing seed samples to record seed quality parameters at monthly intervals as well as reduce the inadvertent imbibition of moisture by the stored seeds during seed sampling.

The packed seeds were stored at ambient conditions and observations like seed germination (%), shoot length (cm), root length (cm), dry weight of seedling (g) and electrical conductivity (EC) of seed leachate were recorded at monthly intervals and seed micro flora (%) and seed moisture (%) were recorded during the start and end of the storage.

3.5 Observations

3.5.1 Experiment 1: Field performance of dry dressed inorganic nano particles on seed yield in Chilli

Five plants in each replication of each treatment were randomly selected. All biometric observations *viz.*, plant spread (cm), plant height (cm), days to harvest, fruits per plant, fruit length (cm), fruit weight (g) at maturity, fruit yield per plant (g), seeds per fruit, seed yield per plant (g) and 100 seed weight (g) were recorded at appropriate growth stages in the tagged plants and averaged to compute the data pertaining to a given replication of a treatment.

3.5.1.1 Plant height (cm)

On the 120th day after transplanting, plant height was measured from ground level to the tip of the main stem and the average was expressed in centimetre.

3.5.1.1 Plant spread (cm)

Plant spread is the maximum width of the plant as measured at its widest part from leaf tip to leaf tip at maturity. The plant spread was measured using a metre scale and the average expressed in centimeters.

3.5.1.2 Days to harvest

The number of days taken from transplanting to first harvest was counted and the average expressed as whole numbers.

3.5.1.3 Fruits per plant

Total number of fruits harvested per plant was counted and the average expressed in numbers.

3.5.1.4 Fruit length (cm)

Ten fruits from each plant were measured from distal end to proximal length using meter scale and the average length was expressed in centimetres.

3.5.1.5 Fruit weight at maturity (g)

Fruits collected from each plant were weighed using weighing balance and average was expressed in grams.

3.5.1.6 Fruit yield per plant (g)

The fruits collected from each plant were weighed, and the yield of fruits per plant was computed and expressed in grams.

3.5.1.7 Seeds per fruit

Seeds from the fruits of the tagged plants were extracted carefully in each replication. The number of seeds per fruit was counted and average number of fruits were worked out.

3.5.1.8 Seed yield per plant (g)

The seeds extracted from the fruits of the tagged plants were weighed using weighing balance and expressed in grams

3.5.1.10 100 seed weight

Three samples of 100 well filled seeds drawn at random from the tagged of a replication were weighed and the average expressed in gram.

3.5.2 Experiment 2: Seed storage studies with wet dressed inorganic nanoparticles

Observations on germination per cent, shoot and root length of the seedling (cm), seedling dry weight, seed moisture content (%), electrical conductivity (E.C.) and Seed microflora infections were assessed as detailed below.

3.5.2.1 Germination (%)

Germination test was conducted using roll paper towel as the substratum following the standard procedure advocated by ISTA (1995). Four replications of hundred seeds each were drawn from each replication of a treatment, for conducting

germination test. The seeds were placed between two layers of germination paper and rolled. The rolled towels were placed in the buckets containing water in a slanting position inside a seed germinator maintained at 25 ± 2 °C temperature and 95 ± 2 per cent relative humidity. The germination per cent was worked out by counting the number of normal seedlings on the 14th day and the average expressed in per cent.

3.5.2.2 Seedling shoot length (cm)

On the 14th day of germination test, ten normal seedlings from each replication of the treatment were selected randomly and the shoot length was measured from the base to the collar region of the seedling and the average was expressed in centimeters.

3.5.2.3 Seedling root length (cm)

On the 14th day, the root length of the seedlings selected for measurement of the shoot length, was measured from the collar region to the tip of the root of the seedling and the mean root length was expressed in centimeters.

3.5.2.4 Seedling dry weight (mg)

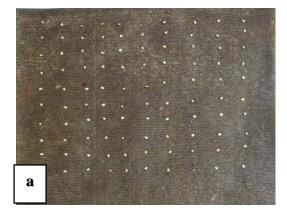
Seedling dry weight estimation was done using the same ten seedlings used for measurement of seedling shoot and root length. The seedlings were placed in butter paper covers and transferred to an oven and dried at 80^o C temperature for 24 h. These were then weighed using an automatic digital balance and the average seedling dry weight computed and expressed in milligrams (mg).

3.5.2.5 Seed Vigour Indices

Seed vigour indices was calculated using the germination percentage obtained in the germination test. The vigor index was calculated adopting the method of Abdul Baki and Anderson (1973).

Vigour index-I = Germination (%) x Seedling length (cm)

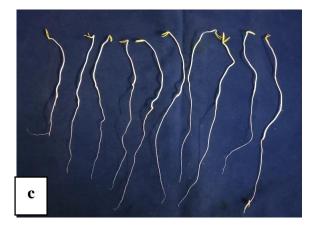
Vigour index-II = Germination (%) x Seedling dry weight (mg)



100 seeds placed on a moistened roll towel paper



Seeds germinated on roll towel at 14th day



10 seedlings selected for measuring shoot and root length

PLATE 4: Germination test using between paper method

3.5.2.6 Electrical conductivity of seed leachate (µS cm-1)

Three replicates of five gram of seeds were drawn from each treatment for estimating electrical conductivity of seed leachate. For surface sterilization, randomly selected seeds were treated in 0.1% mercuric chloride (HgCl2) for one minute. These seeds were thoroughly washed with distilled water several times for removing the residues of treated chemicals and was soaked in 50 ml of distilled water for 24 hours. Electrical conductivity (EC) of the seed leachate collected in a beaker after decanting was measured using digital conductivity meter (EUTECH CON-510) and the mean value was recorded as micro Siemens per centimetre (μ S cm⁻¹).

3.5.2.7 Seed moisture content (%)

Seed moisture content was measured using the low constant temperature procedure advocated by ISTA (1985). Seed samples (5 g each) were drawn from each replication and were evenly placed in a container made up of glass. Weight of the container along with lid before and after filling with the seeds was measured. The samples in the container were placed in hot air oven maintained at 103 ± 2 °C and dried for 17 ± 1 h. After drying, the containers was placed in a dessicator for cooling for 30-45 minutes. The weight of the container along with its lid was taken again after cooling and the seed moisture content in per cent was estimated using the formulae.

Moisture content (%) = $\frac{M2 - M3}{M2 - M1} \times 100$

where,

M1: weight of container with lid

M2: weight of container with lid + seeds before drying

M3: weight of container with lid + seeds after drying

3.5.2.8 Seed infection (%)

Detection of seed infection was conducted using blotter paper method and agar plate method as recommended by ISTA (1999).

3.5.2.8.1 Blotter paper method

The procedure described by Neergard (1979) was followed for detecting seed infection using blotter paper method. Sterilized petriplates with three layers of blotter papers were used in this method and sterilized water was used to soak the blotter papers. Twenty five seeds were placed equidistantly in such a way that, the outer most layer consisted of 16 seeds, eight in the middle and one seed at the centre under aseptic conditions of laminar air flow. Incubation of the petriplates was done under alternate cycle of 12 h darkness and 12 h light at 20±2 ^oC, for seven days. The plates were observed for the presence of seed microflora on the eighth day under stereo binocular microscope and the number of infected seeds were counted and recorded in per cent.

3.5.2.8.2 Agar plate method

In agar plate method, seeds were surface sterilized using 0.1 per cent mercuric chloride and then washed thoroughly three times in sterile water to remove residues of mercuric chloride. The seeds were kept in sterile filter paper to remove excessive water and were placed on petriplates having potato dextrose agar media, under aseptic conditions of laminar air flow. After plating, they were incubated for six days after packing in polyethylene cover, under bell jar. The plates were observed under stereo-binocular microscope and the number of infected seeds were counted and recorded in per cent.

3.6 Statistical analysis

The statistical analysis of the data was performed using Web Agri Stat Package (WASP) developed by Indian Council of Agricultural Research and ranking of significant treatments was done using Duncan's Multiple Range Test (DMRT). For all F- test critical difference was calculated at 5 per cent probability. The zero values present in the data were converted to 1 n value, where 'n' is the number of observations.

Agar plate method



Initial



Final

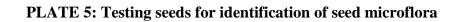
Blotter paper method



Initial



Final



The recorded data in each observation were analyzed using ANOVA table for completely randomized design to test the differences as depicted below:

| Source of variation | Degree of freedom (df) | Sum of Squares (SS) | Mean square MS = SS/df | Computed F |
|------------------------|---------------------------|------------------------|---------------------------|------------|
| Treatment | t-1 | SST | MST | MST/MSE |
| Error | n-t | SSE | MSE | |
| Total | N-1 | SST | | |

Where,

t: Treatments

n: Number of observations

MST: Treatment mean sum of squares

MSE: Error mean sum of squares

3.6.1 DMRT test for ranking

For the evaluation and ranking of all possible Pairs of treatment means, Duncan's Multiple Range Test (DMRT) is applied, particularly for experiments having big numbers of treatments. To classify the difference between any two treatments as significant or non-significant, numerical boundaries are created by DMRT and its value calculation depends primarily on the specific standard error (SEm) of the pair of treatments being compared. The procedure for ranking of data suggested by Gomez and Gomez (1976) is given below:

i. The treatment means are ranked in the increasing or decreasing order according to the order of preference

ii. Calculate standard error of error mean sum of squares (SEm) by using the formula

$$SEm = \sqrt{2MSE}$$

Where,

MSE: error mean sum of squares

R : number of replications.

iii. The (t-1) values of shortest significant ranges was calculated as follows:

$$R_{p}=(r_{p})(SE_{m})$$

$$\sqrt{2}$$

Where,

```
t : number of treatments
```

SEm: standard error obtained in step ii

rp: tabulated value of ranges which are significant

p: difference in rank between the pairs of treatment means to be compared (p = t for the highest and lowest means).

iv. After identifying all the treatment means which do not differ significantly from each other was grouped.

v. The obtained results were presented using alphabetical notation.

<u>Results</u>

4. RESULTS

The results obtained from field and storage studies on seed invigoration with nanoparticle on seed yield and quality in chilli conducted in the Department of Seed Science and Technology, College of Horticulture, Vellanikkara are presented in this chapter. The data obtained with respect to seed quality, plant growth and yield parameters were analysed statistically and presented below.

4.1: Field performance of dry dressed inorganic nanoparticles on seed yield in chilli

4.1.1 Analysis of variance

The analysis of variance of the observations recorded is presented in Table 3. The treatments were highly significant for yield attributes such as plant spread (cm), plant height (cm),days to harvest (days), number of fruits per plant, fruit length (cm), fruit weight (g) at maturity, fruit yield, number of seeds per fruit, seed yield per plant (g) and 100 seed weight (g).

4.1.2 Plant spread (cm)

The treatments with normal and nano grades of ZnO and TiO₂ exhibited significant differences in plant spread (Table 4). Treated seeds produced more plant spread than control (42.66 cm). Among them, nano grade ZnO and TiO₂ performed better than normal grade seed treatments. Seeds treated with T₄: nano ZnO @ 1300 mg kg⁻¹ of seed (60.1 cm) found to have highest plant spread followed by T₁₀- nano TiO₂ @ 1300 mg kg⁻¹ of seed (56cm) and T₉- nano TiO₂ @ 900 mg kg⁻¹ of seed (54 cm). These treatments were found to be on par with each other. The least plant spread was seen in control (42.66 cm) while comparing to other treatments.

4.1.3 Plant height (cm)

The chilli seeds treated with normal and nano grades of ZnO and TiO₂ exhibited significant differences in plant height (Table 4). The treated seeds produced taller plants than control (60.33 cm). Among normal and nano grade seed treatments, seeds treated with nano grade ZnO and TiO₂ were found to have favourable effect on plant height.

| | Mean sum of squares | | | | | | | | | | |
|----------------------|--------------------------|-------------------------|-------------------------|--------------------|---------------------|-------------------------|---------------------------------------|-------------|--------------------|-------------------------------|---------------------------|
| Sources of variation | Degrees of freedom | Plant spread (cm) | Plant height (cm) | Days to harvest | Fruits per plant | Fruit length (cm) | Fruit weight (g) at maturity | Fruit yield | Seeds per fruit | Seed yield per plant(g) | 100 seed weight (g) |
| Replications | 2 | 12.29 | 2.57 | 0.231 | 8.94 | 0.373 | 0.028 | 656.48 | 2.33 | 1.27 | 0.000 |
| Treatments | 12 | 62.19** | 11.79** | 74.63** | 490.38** | 0.227** | 0.385** | 15953.27** | 39.70** | 180.32** | 0.007** |
| Error | 24 | 12.57 | 1.12 | 0.203 | 3.56 | 0.023 | 0.018 | 221.38 | 4.86 | 0.87 | 0.000 |
| SE | | 2.895 | 0.866 | 0.135 | 1.540 | 0.123 | 0.109 | 12.148 | 1.800 | 0.764 | 0.000 |
| CV (%) | | 7.043 | 1.632 | 0.486 | 1.892 | 2.134 | 4.412 | 4.898 | 3.856 | 3.201 | 2.207 |

Table 3: Analysis of variance of seed yield and fruit yield attributes in chilli

Significant at 5 % level * Significant at 1 % level **

| Treatment | Plant spread (cm) | Plant height (cm) | Days to harvest (nos) | Fruits/ plant (nos.) | Fruit length (cm) | Fruit weight at maturity(g) | Fruit yield (g) |
|--|-------------------------|-------------------------|-----------------------------|----------------------------|-------------------------|-----------------------------------|-----------------------|
| T ₁ - Control | 42.66 ^f | 60.33 ^e | 90 ^f | 85.33 ⁱ | 6.43 ^g | 2.30 ^f | 195.5 ^f |
| T ₂ - n ZnO@500 | 48.20 ^{cdef} | 62.00 ^e | 90 ^{ef} | 90.00 ^{gh} | 6.90 ^{ef} | 2.86 ^{cd} | 255.00 ^e |
| T ₃₋ n ZnO@900 | 46.66 ^{ef} | 66.00 ^{bc} | 92° | 88.00 ^{hi} | 6.80 ^f | 2.80 ^d | 246.33 ^e |
| T ₄ -n ZnO@1300 | 60.10 ^a | 67.30 ^a | 92 ^{cd} | 122.00 ^a | 7.40 ^a | 3.46 ^{ab} | 422.70 ^a |
| T ₅₋ ZnO@500 | 50.10 ^{bcde} | 65.33 ^{bcd} | 92° | 98.33 ^e | 7.06 ^{cde} | 2.93 ^{cd} | 288.50 ^d |
| T ₆₋ ZnO@900 | 50.90 ^{bcde} | 66.46 ^{ab} | 91 ^{de} | 103.66 ^d | 7.33 ^{ab} | 3.26 ^b | 338.73° |
| T ₇₋ ZnO@1300 | 46.80 ^{def} | 65.00 ^{bcd} | 90 ^f | 87.66 ^{hi} | 6.90 ^{ef} | 2.83 ^{cd} | 251.30 ^e |
| T ₈₋ nTiO ₂ @500 | 49.30 ^{cde} | 65.80 ^{bcd} | 91 ^{ef} | 93.00 ^{fg} | 7.23 ^{abcd} | 2.56 ^e | 238.70 ^e |
| T ₉₋ n TiO ₂ @900 | 54.00 ^{bc} | 66.33 ^{ab} | 90 ^f | 117.66 ^b | 7.30 ^{abc} | 3.30 ^b | 388.30 ^b |
| T ₁₀₋ nTiO ₂ @1300 | 56.00 ^{ab} | 64.10 ^d | 90 ^f | 114.66 ^{bc} | 7.00 ^{cde} | 3.33 ^{ab} | 382.13 ^b |
| T ₁₁ -TiO ₂ @500 | 52.66 ^{bcd} | 64.50 ^{cd} | 91 ^{ef} | 111.66° | 7.40 ^a | 3.53ª | 394.56 ^b |
| T ₁₂ -TiO ₂ @900 | 49.70 ^{cde} | 65.60 ^{bcd} | 105 ^a | 95.00 ^f | 7.10 ^{bcde} | 3.03° | 288.30 ^d |
| T ₁₃₋ TiO ₂ @1300 | 47.10 ^{def} | 66.23 ^{abc} | 103 ^b | 89.33 ^h | 7.00 ^{def} | 2.90 ^{cd} | 259.10 ^e |
| SEm | 2.895 | 0.866 | 0.135 | 1.540 | 0.123 | 0.109 | 12.148 |
| CD (0.05) | 5.976 | 1.789 | 0.759 | 3.180 | 0.254 | 0.224 | 25.075 |

Table 4: Influence of normal grade and nano grade ZnO and TiO₂ on yield attributes in chilli

Tallest plants were produced by T₄: nano ZnO @ 1300 mg kg⁻¹ of seed (67.3 cm) followed by T₆: ZnO @ 900 mg kg⁻¹ of seed (66.46 cm) ,T₉: nano TiO₂ @ 900 mg kg⁻¹ of seed (66.33 cm) and T₁₃: TiO₂ @ 1300 mg kg⁻¹ of seed (66.23 cm) which were on par with each other. Least plant height was recorded in control (60.33 cm) and among the treated seeds least plant height was recorded in T₂: nano ZnO @ 500 mg kg⁻¹ of seed (62 cm).

4.1.4 Days to harvest (days)

Significant differences were found among the seeds treated with normal and nano grades of ZnO and TiO₂ (Table 4). Days to harvest varied from 90 to 105 days. Irrespective of the doses of various treatments, T₂ -nano ZnO @ 500 mg kg⁻¹ of seed, T₉ -nano TiO₂ @900 mg kg⁻¹ of seed, T₁₀ -nanoTiO₂ @1300 mg kg⁻¹ of seed , T₇ - ZnO @ 1300 mg kg⁻¹ of seed and control were harvested at 90 DAT. Among the treatments, delayed harvest (105 days) was recorded in seeds treated with T₁₃ -TiO₂ @ 1300 mg kg⁻¹ of seed.

4.1.5 Fruits per plant (Nos.)

Chilli seeds treated with ZnO and TiO₂ recorded more number of fruits than untreated seeds (Table 4). Among them, nano grade ZnO and TiO₂ showed good results than normal grade seed treatments. Seed treatments with T₄ -nano ZnO @ 1300 mg kg⁻¹ of seed (122) produced more number of fruits followed by T₉. nano TiO₂ @ 900 mg kg⁻¹ of seed (118) which were on par with T₁₀. nano TiO₂ @ 1300 mg kg⁻¹ of seed (115). Least number of fruits were recorded in T₃- nano ZnO @ 900 mg kg⁻¹ of seed (88), T₇-ZnO @ 1300 mg kg⁻¹ of seed (88) and control (88).

4.1.6 Fruit length (cm)

There existed a significant variation among the treatments for fruit length (Table 4). In general it is observed that TiO_2 was more effective in improving the trait. Treatments T₄-nano ZnO @ 1300 mg kg⁻¹ of seed and T₁₁-TiO₂ @ 500 mg kg⁻¹ of seed recorded the highest value and were on par (7.40cm). Among the treatments, control recorded lowest fruit length (6.4 cm).

4.1.7 Fruit weight at maturity (g)

All treatments recorded higher values than control for fruit weight at maturity with significant differences among themselves (Table 4). Seeds treated with T_{11} -TiO₂ @ 500 mg kg⁻¹of seed recorded highest fruit weight (3.53 g) at maturity followed by T₄ -nano ZnO @ 1300 mg kg⁻¹ (3.46g) and T_{10} - nano TiO₂ @1300 mg kg⁻¹ of seed (3.33 g). These treatments were on par with each other. In general, TiO₂ seed treatments exhibited superiority for this trait.

4.1.8 Fruit yield per plant (g)

Significant differences were found among the seeds treated with normal and nano grades of ZnO and TiO₂ (Table 4). Nano ZnO @ 1300 mg kg⁻¹ of seed (T₄) recorded highest fruit yield per plant (422.70 g). T₁₁-TiO₂ @ 500 mg kg⁻¹ of seed, T₉-nanoTiO₂ @900 mg kg⁻¹ of seed and T₁₀-nanoTiO₂ @1300 mg kg⁻¹ of seed which were on par with each other recorded fruit yield of 394.56 g, 388.30 g, 382.13 g respectively. Least fruit yield per plant (195.50g) was recorded by control.

4.1.9 Number of seeds per fruit

Compared to untreated seeds, number of seeds per fruit was higher in treatments with normal and nano grades of ZnO and TiO₂ (Table 5). Seeds treated with T₆- ZnO 900 mg kg⁻¹ of seed (62) recorded more number of seeds per fruit followed by T₄-nano ZnO @ 1300 mg kg⁻¹ of seed (61), T₁₂-TiO₂ @ 900 mg kg⁻¹ of seed (61), T₁₁-TiO₂ @ 500 mg kg⁻¹ of seed (60), T₉- nano TiO₂ @ 900 mg kg⁻¹ of seed (59) and T₁₀- nano TiO₂ @ 1300 mg kg⁻¹ of seed (59) which were on par with each other compared to control (50). The number of seeds were less in T₈- nano TiO₂@ 500 mg kg⁻¹ of seed (53) among treated seeds.

In general, normal grades of ZnO and TiO₂ exibited superiority in recording more seed yield per fruit than nano grade seed treatments.

| Treatment | Seeds/fruit (nos) | Seed yield/plant (g) | 100 seed weight | |
|--|----------------------|-------------------------|--------------------|--|
| T ₁ - Control | 50.33 ^f | 17.18 ^h | 0.40 ^e | |
| T ₂ . n ZnO@500 | 54.66 ^{de} | 24.62 ^e | 0.50 ^{bc} | |
| T ₃₋ n ZnO@900 | 58.66 ^{abc} | 23.22 ^{ef} | 0.45 ^c | |
| T ₄ -n ZnO@1300 | 61.33ª | 41.14 ^a | 0.55 ^a | |
| T ₅₋ ZnO@500 | 56.33 ^{bcd} | 28.80 ^d | 0.52 ^b | |
| T ₆₋ ZnO@900 | 62.33 ^a | 35.49° | 0.55 ^a | |
| T ₇₋ ZnO@1300 | 54.33 ^{de} | 21.43 ^g | 0.45 ^d | |
| T ₈₋ nTiO ₂ @500 | 52.33 ^{ef} | 24.32 ^e | 0.50 ^{bc} | |
| T ₉₋ n TiO ₂ @900 | 59.00 ^{ab} | 38.17 ^b | 0.55 ^a | |
| T ₁₀₋ nTiO ₂ @1300 | 59.00 ^{ab} | 37.20 ^b | 0.55 ^a | |
| T ₁₁ -TiO ₂ @500 | 60.00 ^{ab} | 36.84 ^{bc} | 0.55 ^a | |
| T ₁₂ -TiO ₂ @900 | 61.00 ^{ab} | 29.62 ^d | 0.52 ^b | |
| T ₁₃₋ TiO ₂ @1300 | 55.00 ^{cde} | 22.10 ^{fg} | 0.45 ^c | |
| SE _m | 1.800 | 0.764 | 0.000 | |
| CD (0.05) | 3.716 | 1.578 | 0.019 | |

Table 5: Influence of normal grade and nano grade ZnO and TiO2 on seed yield and seed weight in chilli

1.1.10 Seed yield per plant (g)

Significant variations were observed for seed yield per plant among the treatments with treatments performing better than control. (Table 5). Seeds treated with T₄-nano ZnO @ 1300 mg kg⁻¹ of seed (41.14 g) recorded highest seed yield compared to control (17.18 g). Treatments with T₉- nano TiO₂ @ 900 mg kg⁻¹ of seed (38.17 g), T₁₀- nano TiO₂ @ 1300 mg kg⁻¹ of seed (37.20) and T₁₁-TiO₂ @ 500 mg kg⁻¹ of seed (36.84) were found to be on par with other. Lowest seed yield was recorded in seeds treated with T₇- ZnO @ 1300 mg kg⁻¹ of seed (21.43g) among treated seeds.

4.1.11 100 seed weight (g)

Significant differences was found among the seeds treated with normal and nano grades of ZnO and TiO₂ (Table 5). 100 seed weight varied from 0.55g in T₉ - nano TiO₂ @ 900 mg kg⁻¹ of seed, T₁₁ -TiO₂ @ 500 mg kg⁻¹ of seed, T₄ -nano ZnO @ 1300 mg kg⁻¹ of seed, T₁₀ - nano TiO₂ @ 1300 mg kg⁻¹ of seed and T₆ -ZnO @ 900 mg kg⁻¹ of seed to 0.40 g in control. Least seed weight was recorded in seeds treated with T₇₋ ZnO @ 1300 mg kg⁻¹ of seed (0.45g) among the treated seeds. In general, nano grades of ZnO and TiO₂ exibited greater potential in recording more seed weight than normal grade seed treatments. Irrespective of various doses of treatments, both ZnO and TiO₂ performed well.

4.2 Seed storage studies with wet dressed inorganic nanoparticles

4.2.1 Initial seed quality

Initial seed quality prior to storage was determined and are presented in the Table 6. Seed quality parameters like germination, shoot length, root length, seedling dry weight, vigour index - I and II, electrical conductivity of seed leachates and seed microflora infestation were recorded significant differences existed among the treatments except for shoot length.

| Treatment | Germination (%) | Shoot length (cm) | Root length (cm) | Dry weight (mg) | EC (µS cm ⁻¹) | VI I | VI II | Moisture content (%) | Microflora (agar plate method) (%) |
|---|--------------------|-------------------------|------------------------|-----------------------|------------------------------|------|-------|-------------------------|---|
| T ₁ - Control | 70.00 | 5.10 | 7.63 | 25.00 | 268.33 | 885 | 1777 | 7.10 | 16.66 |
| T ₂ - ZnO@100 | 71.60 | 5.96 | 9.53 | 26.66 | 221.00 | 1105 | 1894 | 7.00 | 0 |
| T ₃ - ZnO@250 | 72.00 | 6.06 | 9.33 | 26.33 | 204.33 | 1105 | 1901 | 7.00 | 0 |
| T ₄ - ZnO@500 | 69.00 | 5.43 | 9.80 | 26.00 | 237.66 | 1056 | 1800 | 6.30 | 0 |
| T ₅ - ZnO@750 | 70.00 | 5.86 | 10.16 | 26.66 | 249.33 | 1128 | 1861 | 6.30 | 0 |
| T ₆ -ZnO@1000 | 71.66 | 6.10 | 9.83 | 27.33 | 210.33 | 1132 | 1945 | 7.00 | 0 |
| T ₇ -ZnO@1250 | 69.00 | 5.70 | 9.50 | 26.33 | 249.67 | 1048 | 1807 | 7.40 | 0 |
| T ₈ -TiO2@100 | 69.33 | 5.46 | 8.13 | 27.00 | 261.66 | 941 | 1869 | 7.30 | 0 |
| T ₉ -TiO ₂ @250 | 70.66 | 5.20 | 8.33 | 26.33 | 254.33 | 954 | 1848 | 7.60 | 0 |
| T ₁₀ -TiO ₂ @500 | 70.33 | 5.90 | 9.50 | 28.33 | 223.00 | 1085 | 1975 | 6.40 | 0 |
| T ₁₁ -TiO ₂ @750 | 72.00 | 6.03 | 8.93 | 27.66 | 201.66 | 1074 | 2012 | 6.90 | 0 |
| T ₁₂ -TiO ₂ @1000 | 72.00 | 6.10 | 9.23 | 27.33 | 191.66 | 1105 | 1965 | 7.60 | 0 |
| T ₁₃ -TiO2@1250 | 68.66 | 5.20 | 8.43 | 26.33 | 279.67 | 924 | 1788 | 6.30 | 0 |
| CD (0.05) | NS | 0.182 | NS | NS | NS | NS | NS | NS | NS |

 Table 6: Seed quality of chilly before storage

| Tuestuesent | | | Ν | Ionths of storag | ge | | |
|--|----------------------|----------------------|----------------------|---------------------|---------------------|----------------------|---------------------|
| Treatment | M1 | M2 | M3 | M4 | M5 | M6 | M7 |
| T ₁ - Control | 71.66 ^{cd} | 69.66 ° | 67.33 ^d | 65.33 ^d | 62.33 ^f | 60.66 ^f | 58.00 ^f |
| | (57.843) | (56.584) | (55.152) | (53.930) | (52.14) | (51.16) | (49.60) |
| T 7mO@100 | 73.00 ^{abc} | 72.33 ^{ab} | 71.33 ^{abc} | 70.33 ^{ab} | 68.33° | 66.33 ^{bc} | 62.33 ^{cd} |
| T_2 - ZnO@100 | (58.696) | (58.271) | (57.629) | (56.998) | (55.75) | (54.54) | (52.14) |
| T ₃ - ZnO@250 | 74.60 ^a | 73.33 ª | 73.00 ^a | 72.33ª | 70.33 ^{ab} | 69.66 ^a | 65.66 ^a |
| 13 - ZHO(<i>W</i> 250 | (59.781) | (58.910) | (58.696) | (58.266) | (56.99) | (56.58) | (54.13) |
| T ₄ - ZnO@500 | 72.33 ^{bcd} | 70.66 ^{bc} | 69.66 ^{bcd} | 68.33 ^{bc} | 66.33 ^d | 65.66 ^{cd} | 60.66 ^{de} |
| 14-ZIIO@300 | (58.275) | (57.216) | (56.584) | (55.757) | (54.54) | (54.13) | (51.16) |
| T. 7n0@750 | 72.00 ^{bcd} | 71.6 ^{abc} | 69.66 ^{bcd} | 67.33 ^{cd} | 65.66 ^d | 63.66 ^{de} | 59.66 ^{ef} |
| $T_5 - ZnO@750$ | (58.061) | (57.843) | (56.584) | (55.146) | (54.13) | (52.93) | (50.57) |
| T ₆ - ZnO@1000 | 73.33 ^{abc} | 72.00 ^{ab} | 72.66 ^{ab} | 72.00 ^a | 70.66 ^{ab} | 68.33 ^{ab} | 64.00 ^{bc} |
| 16-ZIIO@1000 | (58.910) | (58.061) | (57.845) | (58.061) | (57.20) | (55.75) | (53.13) |
| T ₇ -ZnO@1250 | 71.33 ^{cd} | 70.33 ^{bc} | 69.66 ^{bcd} | 67.33 ^{cd} | 64.66 ^{de} | 62.33 ^{ef} | 59.33 ^{ef} |
| | (57.210) | (57.003) | (56.584) | (55.146) | (53.52) | (52.14) | (50.37) |
| T ₈ -TiO2@100 | 71.66 ^{cd} | 70.66 ^{bc} | 68.33 ^d | 66.66 ^{cd} | 63.66 ^{ef} | 60.66^{f} | 58.66 ^{ef} |
| 18-1102@100 | (57.845) | (57.216) | (55.757) | (54.742) | (52.93) | (51.16) | (49.99) |
| T ₉ -TiO ₂ @250 | 72.00 ^{bcd} | 71.33 ^{abc} | 69.33 ^{cd} | 68.00 ^c | 65.66 ^d | 63.33 ^{de} | 59.66 ^{ef} |
| 19-1102@230 | (58.061) | (57.629) | (56.374) | (55.552) | (54.13) | (52.73) | (50.57) |
| T ₁₀ -TiO ₂ @500 | 72.66 ^{bcd} | 72.33 ^{ab} | 71.00 ^{abc} | 70.30 ^{ab} | 68.33° | 65.66 ^{cd} | 63.66 ^{bc} |
| $1_{10} - 110_2 (2000)$ | (58.266) | (58.266) | (57.424) | (56.998) | (55.75) | (54.13) | (52.93) |
| T ₁₁ -TiO ₂ @750 | 74.00^{ab} | 73.00 ^a | 72.66 ^{ab} | 71.66 ^a | 69.33 ^{bc} | 66.00 ^{cd} | 62.33 ^{cd} |
| 111-1102@750 | (59.345) | (58.696) | (57.845) | (57.843) | (56.37) | (54.13) | (52.14) |
| T ₁₂ . TiO ₂ @1000 | 74.66 ^a | 73.33 ^a | 73.00 ^a | 72.33 ^a | 71.33 ^a | 69.33 ^a | 65.33 ^a |
| | (59.781) | (58.910) | (58.482) | (58.266) | (57.633) | (56.37) | (54.54) |
| T ₁₃ -TiO2@1250 | 70.60^{d} | 70.33 ^{bc} | 69.00 ^{cd} | 66.66 ^{cd} | 62.33 ^f | 60.66^{f} | 58.66 |
| | (57.210) | (58.910) | (56.172) | (54.742) | (52.14) | (51.16) | (49.99) |
| SE _m | 0.66 | 0.69 | 0.71 | 0.66 | 0.55 | 0.72 | 0.65 |
| CD (0.05) | 1.364 | 1.438 | 1.467 | 1.372 | 1.151 | 1.483 | 1.342 |

Table 7: Influence of nano particles of ZnO and TiO₂ on germination of chilli

Highest shoot length of 6.10 cm was recorded in treatments with nano ZnO @ 1000 mg kg⁻¹ of seed (T₆) and nanoTiO₂ @ 1000 mg kg⁻¹ of seed (T₁₂).

4.2.2 Quality of seed during storage

4.2.2.1 Analysis of variance

The analysis of variance of the observations recorded at monthly intervals for seven months of storage revealed that there existed high significant differences for seed quality parameters among the various doses of nano grade ZnO and TiO₂ treatments used to wet dress the seeds.

4.2.2.2 Germination (%)

Seed treatment with various doses of nano grades of ZnO and TiO_2 exibited significant differences in germination per cent throughout the storage period (Table 7). The germinability of seeds were found to decline in both treated and untreated seeds (control) over the period of storage. However, it was seen that, treated seeds recorded higher germination compared to control for most part of the storage period.

At 1MAS germination ranged from 74.66 per cent in nanoTiO₂@1000 mg kg⁻¹ of seed (T₁₂) to 70 per cent in nanoTiO₂ @ 1250 mg kg⁻¹ of seed (T₁₃). All the treatments including control retained germination above the prescribed 60 per cent by IMSCS (Indian Minimum Seed Certification Standards) till sixth month of storage.

Among the treatments $T_{3.}$ nano ZnO 250 mg kg⁻¹ of seed (65.66) recorded maximum germination followed by $T_{12.}$ nano TiO₂₋ 1000 mg kg⁻¹ of seed (65.33) which were on par with each other compared to control (58) at the end of storage *i.e.*, seven MAS. Seeds treated with $T_{3.}$ nano ZnO 250 mg kg⁻¹ of seed (65.66) which was on par with $T_{12.}$ nano TiO₂₋ 1000 mg kg⁻¹ of seed (65.33) and T₆₋ nano ZnO 1000 mg kg⁻¹ of seed (64.00) which was on par with $T_{10.}$ nano TiO₂ 500 mg kg⁻¹ of seed (63.66), T₂ -nano ZnO-100 mg kg⁻¹ of seed (62.33) and T_{11.} nano TiO₂ 750 mg kg⁻¹ of seed (62.33) retained more than 60 per cent germination after seven months of storage period. In general, the seeds treated with nano ZnO @ 250 mg kg⁻¹ of seed(T₃)and nano TiO₂@1000 mg kg⁻¹ of seed (T₁₂) showed superiority in recording highest germination per cent.

4.2.2.3 Shoot length (cm)

Significant influence of storage period and seed treatments were seen in seedling shoot length (Table 8). The shoot length were found to decline in both treated and untreated seeds (control) over the period of storage. However, it was seen that, treated seeds recorded higher shoot length compared to control.

Among the treatments, T_{6-} nano ZnO 1000 mg kg⁻¹ of seed (5.57 cm), T_{3-} nano ZnO 250 mg kg⁻¹ of seed (5.26 cm), T_2 - nano ZnO-100 mg kg⁻¹ of seed (5.21 cm), T_{11-} nano TiO₂ 750 mg kg⁻¹ of seed (5.19 cm) and T_{10-} nano TiO₂ 500 mg kg⁻¹ of seed (5.13 cm) exhibited highest shoot length at seven MAS. The treatments were found to be on par with each other. It was seen that T_{13-} nano TiO₂ -1250 mg kg⁻¹ of seed (4.07 cm) recorded lower shoot length among the seed treatments at the seven MAS.

4.2.5 Root length (cm)

Seed treatment using nano grades of ZnO and TiO_2 had a noticeable effect on seedling root length (Table 9). The treatments were found to be significantly different from each other. Over the period of storage, a gradual decrease was observed in seedling root length.

After 7 MAS, T₅₋ nano ZnO 50 mg kg⁻¹ of seed (9.37 cm) produced longer roots followed by T₆- nano ZnO@1000 (8.89). Among the treated seeds, nano TiO₂@100 mg kg⁻¹ of seed (T₈) recorded shorter root length (7.38 cm).

| Treatment | Months of storage | | | | | | | | |
|---|--------------------|-------------------|--------------------|-------------------|-------------------|-------------------|---------------------|--|--|
| | M1 | M2 | M3 | M4 | M5 | M6 | M7 | | |
| T ₁ - Control | 5.03 ^g | 4.92 ^h | 4.81 ⁱ | 4.76 ⁱ | 4.68 ⁱ | 4.59 ⁱ | 4.44 ^{de} | | |
| $T_2 - ZnO@100$ | 5.80 ^b | 5.69 ^d | 5.53 ^d | 5.40 ^d | 5.38° | 5.32 ^d | 5.21 ^{abc} | | |
| T ₃ - ZnO@250 | 5.83 ^b | 5.71 ^d | 5.61 ^c | 5.53° | 5.48 ^b | 5.37° | 5.26 ^{ab} | | |
| T ₄ - ZnO@500 | 5.38 ^{de} | 5.31 ^f | 5.25 ^{fg} | 5.10 ^f | 5.07 ^f | 4.96 ^f | 4.82 ^{bcd} | | |
| T ₅ - ZnO@750 | 5.51 ^{cd} | 5.33 ^f | 5.18 ^g | 5.08 ^g | 4.92 ^g | 4.83 ^g | 4.72 ^{cd} | | |
| T ₆ - ZnO@1000 | 6.03 ^a | 6.00 ^b | 5.98ª | 5.81ª | 5.71ª | 5.61ª | 5.57ª | | |
| T ₇ -ZnO@1250 | 5.63° | 5.53 ^e | 5.36 ^e | 5.28 ^e | 5.15 ^e | 4.94 ^f | 4.81 ^{bcd} | | |
| T ₈ -TiO2@100 | 5.32 ^e | 5.31 ^f | 5.28 ^f | 5.16 ^f | 5.06 ^f | 4.86 ^g | 4.64 ^d | | |
| T9-TiO2@250 | 5.18 ^f | 5.03 ^g | 4.96 ^h | 4.87 ^h | 4.75 ^h | 4.69 ^h | 4.52 ^{de} | | |
| T_{10} -TiO ₂ @500 | 5.84 ^b | 5.73 ^d | 5.50 ^d | 5.40 ^d | 5.31 ^d | 5.27 ^e | 5.13 ^{abc} | | |
| T ₁₁ -TiO ₂ @750 | 6.00 ^a | 5.87° | 5.72 ^b | 5.63 ^b | 5.47 ^b | 5.31 ^d | 5.19 ^{abc} | | |
| T ₁₂₋ TiO ₂ @1000 | 6.10 ^a | 6.13ª | 6.00ª | 5.83ª | 5.71ª | 5.51 ^b | 4.71 ^{cd} | | |
| T ₁₃ -TiO2@1250 | 5.10 ^{fg} | 4.93 ^h | 4.79 ⁱ | 4.68 ^j | 4.40 ^j | 4.30 ^j | 4.07 ^e | | |
| SE _m | 0.063 | 0.025 | 0.036 | 0.001 | 0.001 | 0.001 | 0.238 | | |
| CD (0.05) | 0.134 | 0.050 | 0.077 | 0.032 | 0.031 | 0.037 | 0.490 | | |

Table 8: Influence of nano particles of ZnO and TiO₂ on shoot length (cm) of chilli

| | Months of storage | | | | | | |
|---|---------------------|--------------------|--------------------|-------------------|-------------------|-------------------|-------------------|
| Treatment | M1 | M2 | M3 | M4 | M5 | M6 | M7 |
| T ₁ - Control | 7.51 ^j | 7.32 ¹ | 7.21^{1} | 7.11 ^k | 7.03 ¹ | 6.91 ^m | 6.82 ¹ |
| T ₂ - ZnO@100 | 9.39 ^c | 9.21 ^e | 9.18 ^d | 9.03 ^d | 8.97° | 8.76 ^d | 8.63 ^d |
| T ₃ - ZnO@250 | 9.20 ^{de} | 9.13 ^f | 9.00 ^e | 8.91 ^e | 8.74 ^e | 8.62 ^f | 8.51 ^e |
| T ₄ - ZnO@500 | 9.10 ^e | 9.02 ^g | 8.91 ^f | 8.72 ^f | 8.62 ^f | 8.55 ^g | 8.35 ^g |
| $T_5 - ZnO@750$ | 10.00 ^a | 10.23 ^a | 10.03 ^a | 9.83 ^a | 9.77 ^a | 9.54 ^a | 9.37 ^a |
| T ₆ - ZnO@1000 | 9.78 ^b | 9.61 ^b | 9.49 ^b | 9.32 ^b | 9.22 ^b | 9.03 ^b | 8.89 ^b |
| T ₇ -ZnO@1250 | 9.22 ^d | 9.00 ^g | 8.81 ^g | 8.72 ^f | 8.51 ^g | 8.43 ^h | 8.33 ^g |
| T ₈ -TiO2@100 | 8.02 ⁱ | 8.00 ^k | 7.89 ^k | 7.71 ^j | 7.69 ^k | 7.53 ¹ | 7.38 ^k |
| T ₉ -TiO ₂ @250 | 8.29 ^h | 8.13 ^j | 8.10 ^j | 7.94 ⁱ | 7.79 ^j | 7.64 ^k | 7.52 ^j |
| T ₁₀ -TiO ₂ @500 | 9.35° | 9.24 ^d | 9.18 ^d | 8.94 ^e | 8.81 ^d | 8.68 ^e | 8.48 ^f |
| T ₁₁ -TiO ₂ @750 | 8.87^{f} | 8.54 ^h | 8.35 ⁱ | 8.25 ^h | 8.03 ⁱ | 7.92 ^j | 7.87 ⁱ |
| T ₁₂₋ TiO ₂ @1000 | 9.78 ^b | 9.53° | 9.31° | 9.25° | 8.96° | 8.85° | 8.71° |
| T ₁₃ -TiO2@1250 | 8.38 ^g | 8.51 ⁱ | 8.44 ^h | 8.32 ^g | 8.22 ^h | 8.07 ⁱ | 7.91 ^h |
| SE _m | 0.025 | 0.001 | 0.036 | 0.025 | 0.001 | 0.025 | 0.001 |
| CD (0.05) | 0.048 | 0.029 | 0.077 | 0.047 | 0.031 | 0.053 | 0.021 |

Table 9: Influence of nano particles of ZnO and TiO₂ on root length (cm) of chilli

| | Months of storage | | | | | | |
|---|---------------------|---------------------|--------------------|--------------------|---------------------|---------------------|--------------------|
| Treatment | M1 | M2 | M3 | M4 | M5 | M6 | M7 |
| T ₁ - Control | 26.13 ^e | 25.83 ^d | 25.00 ⁱ | 23.72 ⁱ | 22.12 ^f | 21.84 ^e | 20.35 ⁱ |
| T ₂ - ZnO@100 | 26.86 ^c | 26.01 ^{cd} | 25.36 ^f | 25.03 ^d | 24.26 ^{bc} | 23.54 ^{ab} | 22.04 ^c |
| T ₃ - ZnO@250 | 26.82 ^c | 26.32° | 25.00 ⁱ | 24.46 ^h | 24.03 ^{cd} | 23.42 ^b | 21.97 ^e |
| T ₄ - ZnO@500 | 26.02 ^e | 25.77 ^d | 25.51 ^e | 24.91 ^e | 23.13 ^e | 22.64 ^d | 21.92 ^d |
| T ₅ - ZnO@750 | 25.51 ^f | 25.25 ^e | 25.13 ^g | 24.82 ^f | 23.12 ^e | 21.95 ^e | 19.13 ^j |
| T ₆ - ZnO@1000 | 26.93° | 26.83 ^b | 26.51ª | 25.63 ^b | 25.01ª | 23.73 ^a | 22.63ª |
| T ₇ -ZnO@1250 | 25.20 ^g | 25.54 ^e | 25.03 ^h | 24.62 ^g | 23.55 ^{de} | 22.05 ^e | 20.74 ^g |
| T ₈ -TiO2@100 | 26.16 ^e | 26.22 ^c | 25.92 ^d | 23.75 ^k | 22.42 ^f | 22.18 ^e | 21.23 ^f |
| T9-TiO2@250 | 25.31 ^{fg} | 25.07 ^e | 24.89 ^j | 23.91 ^j | 22.00 ^f | 21.20 ^f | 20.71 ^h |
| T ₁₀ -TiO ₂ @500 | 27.97 ^a | 27.43 ^a | 26.28 ^b | 25.76 ^a | 24.40 ^{cd} | 22.96° | 21.22 ^f |
| T ₁₁ -TiO ₂ @750 | 26.98 ^b | 26.82 ^b | 25.12 ^g | 24.39 ⁱ | 23.91 ^{cd} | 22.45 ^d | 20.72 ^h |
| T ₁₂₋ TiO ₂ @1000 | 26.51 ^d | 26.22° | 26.12 ^c | 25.51° | 24.76 ^{ab} | 23.32 ^b | 22.41 ^b |
| T ₁₃ -TiO2@1250 | 26.00 ^e | 25.73 ^d | 24.25 ^k | 22.92 ^m | 21.03 ^g | 19.62 ^g | 18.05 ^k |
| SE _m | 0.025 | 0.001 | 0.036 | 0.025 | 0.001 | 0.025 | 0.001 |
| CD (0.05) | 0.245 | 0.317 | 0.014 | 0.013 | 0.539 | 0.268 | 0.019 |

Table 10: Influence of nano particles of ZnO and TiO_2 on dry weight (mg) of chilli

4.2.6 Dry weight (mg)

Seedling dry weight differed significantly throughout the storage period irrespective of seed treatments in a declining trend with the advancement of the period of storage (Table 10). At 7 MAS, Maximum dry matter production was recorded in T₆₋ nano ZnO 1000 mg kg⁻¹ of seed (22.63 mg),T₁₂₋ nano TiO₂₋ 1000 mg kg⁻¹ of seed (22.41 mg),and T₂ - nano ZnO-100 mg kg⁻¹ of seed (22.04 mg) compared to control (20.35 mg) at seven MAS.

4.2.7 Vigour index I

Significant differences due to seed treatments in vigour index I was observed during the period of storage of seven months (Table 11). It was seen that seed vigour declined during the advancement of storage. At the initial month of storage, vigour index I varied between 1133 in nano ZnO @ 1000 mg kg⁻¹ of seed (T₆) and 845 in control.

At seven MAS T₆- nano ZnO @ 1000 mg kg⁻¹ of seed (925.88) retained superiority in maintaining vigour index I followed by T₁₀- nano TiO₂ @ 500 mg kg⁻¹ of seed (867) which was on par with T₁₂-nano TiO₂ @ 1000 mg kg⁻¹ of seed (857.62), T₂ -nano ZnO @ 100 mg kg⁻¹ of seed (844) and T₅- nano ZnO @ 50 mg kg⁻¹ of seed (841) compared to control (652).

4.2.8 Vigour index II

Significant differences due to seed treatments in vigour index II was observed during the period of storage and it declined during the advancement of storage (Table 12). Seeds treated with T_{12} -nano TiO₂ @ 1000 mg kg⁻¹ of seed (1472) which was on par with T_{3-} nano ZnO 250 mg kg⁻¹ of seed (1457) and T₆₋ nano ZnO @ 1000 mg kg⁻¹ of seed (1448) retained superiority in maintaining vigour index II. Among the seed treatments T_{13-} nano TiO₂-1250 mg kg⁻¹ of seed (1059) exhibited lowest vigour index II.

| | Months of storage | | | | | | |
|--|----------------------|--------------------|--------------------|--------------------|-------------------|--------------------|-----------------------|
| Treatment | M1 | M2 | M3 | M4 | M5 | M6 | M7 |
| T ₁ - Control | 845 ^g | 825 ^g | 809 ^g | 799 ^f | 789 ^b | 775 ^h | 652 ^g |
| T ₂ - ZnO@100 | 10701 ^{cde} | 1048 ^{de} | 1023 ^{de} | 1007 ^{cd} | 991 ^a | 981 ^{cde} | 844 ^{bcd} |
| T ₃ - ZnO@250 | 1095 ^{bc} | 1083 ^{bc} | 1067 ^{bc} | 1054 ^{ab} | 1038 ^a | 1022 ^{ab} | 781 ^e |
| T ₄ - ZnO@500 | 1056 ^{de} | 1034 ^{de} | 1015 ^{de} | 1004 ^d | 857 ^b | 962 ^{def} | 821 ^{cde} |
| T ₅ - ZnO@750 | 10823 ^{bcd} | 1084 ^{bc} | 1055 ^{bc} | 1039 ^{bc} | 1023 ^a | 1001 ^{bc} | 841.30 ^{bcd} |
| T ₆ - ZnO@1000 | 1133 ^a | 1119 ^a | 1503ª | 1085 ^a | 1057 ^a | 1049 ^a | 926 ^a |
| T ₇ -ZnO@1250 | 1047 ^e | 1027 ^e | 1013 ^{de} | 997 ^d | 982 ^a | 955 ^{ef} | 798 ^e |
| T ₈ -TiO2@100 | 912 ^f | 909 ^f | 899 ^f | 879 ^e | 864 ^b | 847 ^g | 705 ^f |
| T ₉ -TiO ₂ @250 | 934 ^f | 913 ^f | 906 ^f | 888° | 869 ^b | 855 ^g | 718 ^f |
| T ₁₀ -TiO ₂ @500 | 1079 ^{bcde} | 1064 ^{cd} | 1042 ^{cd} | 1016 ^{cd} | 1006 ^a | 990 ^{cd} | 867 ^b |
| T ₁₁ -TiO ₂ @750 | 1066 ^{cde} | 1033 ^{de} | 1009 ^e | 995 ^d | 973 ^a | 948 ^f | 814 ^{de} |
| T ₁₂ - TiO ₂ @1000 | 1107 ^{ab} | 1101 ^{ab} | 1084 ^b | 1058 ^{ab} | 1041 ^a | 1022 ^{ab} | 858 ^{bc} |
| T ₁₃ -TiO2@1250 | 930 ^f | 928 ^f | 913 ^f | 897° | 873 ^b | 856 ^g | 703 ^f |
| SE _m | 16.98 | 16.56 | 15.53 | 15.83 | 46.11 | 14.86 | 20.47 |
| CD (0.05) | 34.92 | 34.06 | 31.93 | 32.56 | 94.80 | 30.56 | 42.09 |

Table 11: Influence of nano particles of ZnO and TiO2 on Vigour Index I of chilli

| | Months of storage | | | | | | |
|---|---------------------|---------------------|----------------------|--------------------|--------------------|--------------------|--------------------|
| Treatment | M1 | M2 | M3 | M4 | M5 | M6 | M7 |
| T ₁ - Control | 1893° | 1799 ^{de} | 1683 ^g | 1549 ^{hi} | 1379 ^f | 1324 ^f | 1181 ^f |
| T ₂ - ZnO@100 | 1961 ^b | 1881 ^{bc} | 1809 ^{bcd} | 1761 ^{bc} | 1658 ^{bc} | 1561 ^{bc} | 1374 ^b |
| T ₃ - ZnO@250 | 2002 ^{ab} | 1929 ^{ab} | 1825 ^{bc} | 1769 ^{bc} | 1690 ^b | 1631ª | 1457 ^a |
| T ₄ - ZnO@500 | 1882 ^{cd} | 1821 ^{cde} | 1777 ^{cdef} | 1702 ^{de} | 1534 ^d | 1487 ^d | 1330 ^{bc} |
| T ₅ - ZnO@750 | 1837 ^{def} | 1809 ^{de} | 1751 ^{def} | 1671 ^{ef} | 1518 ^d | 1398 ^e | 1142 ^f |
| T ₆ - ZnO@1000 | 1975 ^{ab} | 1932 ^{ab} | 1900 ^a | 1845 ^a | 1767 ^a | 1622 ^{ab} | 1449 ^a |
| T ₇ -ZnO@1250 | 1797 ^f | 1773 ^e | 1744 ^{ef} | 1657 ^{ef} | 1523 ^d | 1375 ^{ef} | 1230 ^e |
| T ₈ -TiO2@100 | 1875 ^{cde} | 1853 ^{cd} | 1771 ^{cdef} | 1583 ^{gh} | 1427 ^{ef} | 1326 ^f | 1246 ^{de} |
| T ₉ -TiO ₂ @250 | 1822 ^{ef} | 1788 ^{de} | 1726 ^{fg} | 1626 ^{fg} | 1451 ^e | 1343 ^{ef} | 1236 ^e |
| T ₁₀ -TiO ₂ @500 | 2024 ^a | 1984 ^a | 1866 ^{ab} | 1812 ^{ab} | 1623° | 1507 ^{cd} | 1351 ^b |
| T ₁₁ -TiO ₂ @750 | 2029 ^a | 1957 ^a | 1800 ^{cde} | 1748 ^{cd} | 1657 ^{bc} | 1474 ^d | 1291 ^{cd} |
| T ₁₂ -TiO ₂ @1000 | 1979 ^{ab} | 1923 ^{ab} | 1898 ^a | 1845 ^a | 1766 ^a | 1616 ^{ab} | 1472 ^a |
| T ₁₃ -TiO2@1250 | 1838 ^{def} | 1809 ^{de} | 1673 ^g | 1528 ⁱ | 1311 ^g | 1190 ^g | 1059 ^g |
| SEm | 26.45 | 32.63 | 28.96 | 26.37 | 31.61 | 29.78 | 23.28 |
| CD (0.05) | 54.315 | 67.089 | 59.558 | 54.231 | 64.992 | 61.248 | 47.882 |

Table 12: Influence of nano particles of ZnO and TiO₂ on Vigour Index II of chilli

| | Months of storage | | | | | | |
|---|---------------------|---------------------|---------------------|---------------------|----------------------|---------------------|----------------------|
| Treatment | M1 | M2 | M3 | M4 | M5 | M6 | M7 |
| T ₁ - Control | 295.66ª | 316.33ª | 334.67ª | 362.33ª | 381.00 ^a | 401.33 ^a | 404.66 ^a |
| T ₂ - ZnO@100 | 234.66 ^h | 247.00 ^h | 261.33 ^f | 286.67 ^g | 297.33 ^f | 315.33 ^h | 319.67 ^f |
| T ₃ - ZnO@250 | 215.33 ^j | 222.00 ^k | 234.33 ⁱ | 262.67 ^k | 281.66 ^{hi} | 283.66 ¹ | 295.00 ⁱ |
| T ₄ - ZnO@500 | 254.67 ^f | 270.66 ^f | 295.66 ^d | 306.33 ^f | 320.66 ^e | 333.67 ^f | 347.00 ^d |
| T ₅ - ZnO@750 | 283.67 ^c | 296.66° | 314.66° | 320.33 ^d | 343.00 ^c | 350.67 ^e | 372.00 ^c |
| T ₆ -ZnO@1000 | 217.00 ^j | 226.33 ^j | 246.66 ^h | 267.33 ^j | 286.33 ^{gh} | 293.66 ^j | 301.00 ^{gh} |
| T ₇ -ZnO@1250 | 261.33 ^e | 227.00 ^j | 255.00 ^g | 272.00 ⁱ | 294.33 ^{fg} | 319.33 ^g | 331.00 ^e |
| T ₈ -TiO2@100 | 285.33 ^c | 290.00 ^d | 323.33 ^b | 342.00 ^c | 363.33 ^b | 371.00 ^c | 386.67 ^b |
| T ₉ -TiO ₂ @250 | 272.33 ^d | 287.00 ^e | 294.00 ^d | 316.33 ^e | 330.67 ^d | 353.33 ^d | 372.66° |
| T ₁₀ -TiO ₂ @500 | 246.66 ^g | 261.33 ^g | 272.33 ^e | 283.67 ^h | 296.33 ⁱ | 305.66 ⁱ | 308.66 ^g |
| T ₁₁ -TiO ₂ @750 | 221.00 ⁱ | 234.67 ⁱ | 254.66 ^g | 266.00 ^j | 281.66 ^{hi} | 285.33 ^k | 290.33 ⁱ |
| T ₁₂₋ TiO ₂ @1000 | 203.00 ^k | 215.33 ¹ | 229.66 ^j | 243.67 ¹ | 274.33 ⁱ | 292.66 ^j | 299.33 ^h |
| T ₁₃ -TiO2@1250 | 290.00 ^b | 311.00 ^b | 322.66 ^b | 349.66 ^b | 368.33 ^b | 383.66 ^b | 397.67 ^a |
| SEm | 0.987 | 1.169 | 1.029 | 1.339 | 3.960 | 0.750 | 3.814 |
| CD (0.05) | 2.029 | 2.404 | 2.117 | 2.754 | 8.153 | 1.544 | 7.842 |

Table 13: Influence of nano particles of ZnO and TiO_2 on electrical conductivity of seed leachate (μ Scm⁻¹) of chilli

| | | Seed infection | | | | |
|---|----------------------|----------------------|-------------------|--|--|--|
| Treatment | Moisture content (%) | Blotter paper method | Agar plate method | | | |
| T ₁ - Control | 7.13 | 3.33 | 13.33 | | | |
| T ₂ - ZnO@100 | 7.16 | 0 | 10.00 | | | |
| T ₃ - ZnO@250 | 7.03 | 0 | 0 | | | |
| T ₄ - ZnO@500 | 6.70 | 0 | 0 | | | |
| T ₅ - ZnO@750 | 6.93 | 0 | 0 | | | |
| T ₆ -ZnO@1000 | 7.23 | 0 | 0 | | | |
| T ₇ -ZnO@1250 | 7.50 | 0 | 0 | | | |
| T ₈ -TiO2@100 | 7.43 | 0 | 0 | | | |
| T ₉ -TiO ₂ @250 | 7.76 | 0 | 10.00 | | | |
| T ₁₀ -TiO ₂ @500 | 7.06 | 0 | 0 | | | |
| T ₁₁ -TiO ₂ @750 | 7.06 | 0 | 0 | | | |
| T ₁₂₋ TiO ₂ @1000 | 7.70 | 0 | 0 | | | |
| T ₁₃ -TiO2@1250 | 6.43 | 0 | 0 | | | |
| SEm | 0.48 | 1.30 | 4.71 | | | |
| CD (0.05) | NS | NS | 9.69 | | | |

Table 14: Influence of nano particles of ZnO and TiO₂ on seed moisture and seed infection at the end of storage in chilli

4.2.9 Electrical conductivity of seed leachates (µS cm⁻¹)

Electrical conductivity of seed leachates of seeds treated with various doses of nano grades of ZnO and TiO_2 was significantly increasing throughout the storage period irrespective of the treatments (Table 13). It was seen that treated seeds recorded lower values while comparing with control.

Seeds treated with T₃₋ nano ZnO @ 250 mg kg⁻¹ of seed (295 μ Scm⁻¹) which was on par with T₁₁₋ nano TiO₂ @ 750 mg kg⁻¹ of seed (290.33 μ Scm⁻¹) had least electrical conductivity of seed leachates compared to control (404.66 μ Scm⁻¹) at seven MAS. Among the treated seeds T₁₃₋ nano TiO₂ @ 1250 mg kg⁻¹ of seed (397.67 μ Scm⁻¹) recorded highest value at seven MAS.

4.2.10 Seed moisture content (per cent)

A marginal increase in moisture content was noticed during the advancement of storage period (Table 14). Insignificant differences was found when seeds treated with both nano ZnO and TiO₂.

4.2.11 Seed microflora (per cent)

Seed treatments with both nano ZnO and TiO₂ showed significant difference in seed infection (Table 14). The control seeds showed higher infection in both agar plate method and blotter paper method irrespective of the treatments.

In case of agar plate method, control showed an infection of 13.3% and in nano ZnO @ 100 mg kg⁻¹ of seeds (T₂) and nano TiO₂ @ 250 mg kg⁻¹ of seeds (T₉), 10 % seed infection was observed. Only control seeds showed an infection of 3.33% in blotter paper method. The presence of *Aspergillus flavus* and *Aspergillus niger* was observed in both methods.

<u>Díscussíon</u>

5. Discussion

Seed is the vital input for sustainable agriculture since most of the food crops are produced from seeds. To feed the ever growing population, new management practices that provides high productivity, healthy ecosystem and cost effectiveness along with good quality seeds are required (Kumar, 2012). Seed treatment with nanoparticle is one such practice. Rapid seed germination and uniform crop stand are crucial for economic sustainability in agriculture. Nanoparticles promotes plant growth and development thereby enhancing crop yield.

Hence the present study was conducted to assess the impact of nanoparticle treatments of zinc oxide and titanium dioxide on yield and seed quality in chilli. The findings of the study are discussed in this chapter.

5.1 Impact of seed invigoration using inorganic nanoparticles of zinc oxide and titanium dioxide on yield in chilli

5.1.1 Effect of Nano grade zinc oxide on yield attributes

Regardless of the treatments all the yield parameters recorded superior results in treatments with nano grades of ZnO. The efficacy of nano treatments in enhancing plant characters was reported by Salama (2012) in maize and bean.

Application of nano ZnO@ 1300 mg kg⁻¹ of seed (T₄) obtained highest plant spread (60.1 cm), number of fruits per plant (122), fruit length (7.40cm), fruit weight (3.53 g), fruit yield (422.7 g), 100 seed weight (0.55 g) and seed yield (41.14 g). While nanoparticles of ZnO @ 900 mg kg⁻¹ of seed (T₃) recorded highest plant height. In case of number of days to harvest, T₂ (500 mg kg⁻¹ of seed) recorded least days (90 DAT).

Subbaiah (2014) by using nano ZnO at 400 ppm obtained highly significant results for yield, plant height, cob length and number of grains per cob when compared to control in maize. Raliya *et al.* (2015) in tomato, and García-López *et al.* (2018) in pepper reported that plant growth increased significantly throughout the growth stages when using nanoparticles for seed treatments. The impact of seed treatment with nanoparticles on fruit yield attributes was noticed by Khanm *et al.* (2017) in tomato

Mahdieh *et al.* (2018) in bean and Sadak and Bakry (2020) in flax. Similar results on seed yield attributes was noticed by Rezaei *et al.* (2015) in soyabean, and Poornima and Koti (2019) in sorghum and Sadak and Bakry (2020) in flax.

5.1.2 Effect of Normal grade zinc oxide on yield attributes

Normal grade ZnO treatments was effective for improving traits like number of seeds per fruit (ZnO @ 1300 mg kg⁻¹ of seed) and 100 seed weight (ZnO @ 900 mg kg⁻¹ of seed).

Potarzycki and Grzebisz (2009) inferred that when sufficient Zn was applied, seed yield in maize could be significantly increased. This increment was due to the increase in yield attributes like cob length, thousand kernel weight, number of kernels per cob and number of rows per cob and the enhancement in nitrogen use efficiency by the application of Zn. Ziaeyan and Rajaie (2009) and Vazin (2012) in maize reported similar effects of normal grade Zn

Zinc is an important factor in synthesis of indole-acetic acid (IAA), proteins, chlorophyll, carbohydrates etc. It also helps in biosynthesis of cytochromes, detoxification of reactive oxygen species (ROS) and reduces cadmium (heavy metal) uptake by plants (Buchanan *et al.*, 2000). When Chickpea seeds were treated with nano ZnO, an increment in IAA levels was observed in roots which in turn increased plant growth (Avinash *et al.*, 2010).

Agronomic efficiency of Zn treatments are influenced by particle size. When particle size is reduced, number of particles per unit weight is increased and thus surface area is also increased (Mortvedt, 1992). This enhances uptake of Zn. These nanoparticles have good catalytic surface, high activity, adsorb more water and are rapidly dispersible (Khanm *et al.*, 2017). As the Zn content in seeds are higher, it will hinder the pathogen invasion during germination and growth of seedling thus ensures good crop stand and better yield (Marschner,1995). When Zn shortage occurs, plants make physiological adjustments to maintain homeostasis (Grusak, 2002). These processes may adversely affect yield and quality.

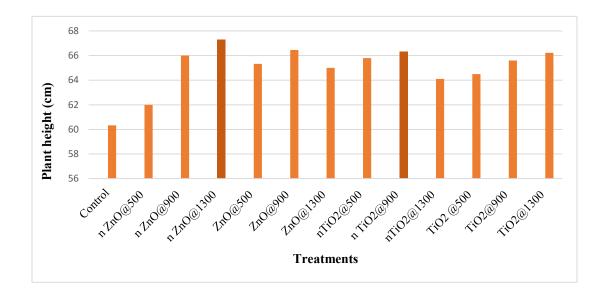


Fig. 1. Influence of normal grade and nano grade and TiO₂ on plant height in chilli

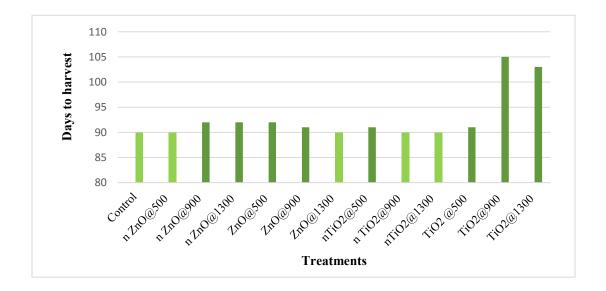


Fig. 2. Influence of normal grade and nano grade ZnO and TiO₂ on days to harvest in chilli

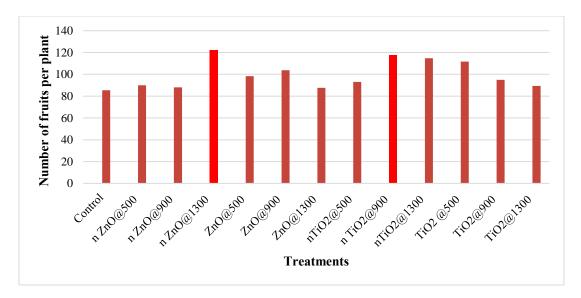


Fig. 3. Influence of normal grade and nano grade ZnO and TiO₂ on number of fruits per plant in chilli

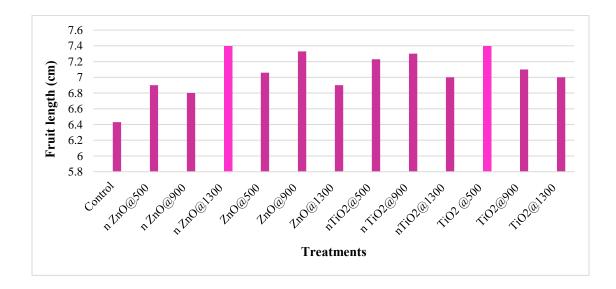


Fig. 4. Influence of normal grade and nano grade ZnO and TiO_2 on fruit length in chilli

5.1.4 Effect of Nano grade titanium dioxide on yield attributes

Significant variations was seen in seeds treated with nano grades of TiO₂. Among the TiO₂ treatments, nanoTiO₂ @ 900 mg kg⁻¹ of seed (T₉) recorded highest plant height (66.33 cm), number of fruits per plant (118), seed yield (38.17), 100 seed weight (0.55g) and the least days to harvest (90 DAT). Nano TiO₂ @ 1300 mg kg⁻¹ of seed (T₁₀) recorded highest plant spread (56 cm) and 100 seed weight (0.55 g).

An increase in yield and yield promoting traits was observed by Debnath *et al.*, (2020) in rice when treated with nano TiO₂ at 500, 1000 and 2000 mg kg⁻¹. Impact of nano TiO₂ on plant characters was observed by Jaberzadeh *et al.* (2013) in wheat and Mattiello and Marchio (2017) in barley. Similar findings on fruit and seed yield attributes was reported by Owolade *et al.*, (2008) in cowpea, Moaveni *et al.* (2011) in barley and Khater (2015) in coriander.

5.1.4 Effect of normal grade titanium dioxide on yield attributes

Normal grades of TiO₂ also reported significant effects on yield characters. TiO₂ @ 500 mg kg⁻¹ of seed (T₁₁) recorded longest fruits (7.40 cm), fruit yield (394.56), fruit weight (3.53 g) and TiO₂ @ 900 mg kg⁻¹ of seed (T₁₂) reported more number of seeds (61) among TiO₂ treatments. This was in confirmation with the observations of Grajkowski and Ochmian, (2007) in raspberry and Haghighi *et al.*, (2014) in tomato.

Titanium is involved in plant metabolism by improving absorption of other nutrients like Iron and Magnesium. The major impact of titanium are enhancement of chlorophyll content, improvement of essential elements in tissues and yield in chilli (Hruby *et al.*, 2002). Enhancement in plant growth and yield might be due to the action of titanium in several cellular mechanisms. For instance, increase in photosynthetic rate is one of the major reason. (Jabersadeh *et al.*, 2013). The photosynthetic efficiency is increased when light energy is transformed to active electrons in chloroplast by Ti nano particles and thus stimulates Rubisco activase complex. This amplification of photosynthesis promotes yield (Moaveni *et al.*, 2011). By the application of nano TiO₂, Rubisco carboxylase activity increased by 2.67 times over control (Gao *et al.*, 2006).

Titanium is not an essential element for growth of plants as per criteria for essentiality by Arnon and Stout (1939) but several studies agrees that Ti have positive impact on plant growth and development. Hong *et al.* (2005) stated that, redox reactions that occur when nano TiO_2 enters the chloroplast, might trigger oxygen evolution and electron transport.

5.1.6 Influence of nanoparticles over bulk treatments

Nanoparticles interact with the living cells at molecular level. They act on plant metabolism by regulating genes, interferes with oxidative process or by providing micronutrients. Nanoparticles have the ability to penetrate the seed coat and results in increased water absorption. Thus seed germination is improved (Hatami *et al.*, 2014)

In the above experiment, nanoparticle treatments were found to perform better than normal grade treatments for many of the traits. According to Poornima and Koti (2019), the reason may be due to the increased uptake and translocation efficiency of nano particles over bulk forms. The bulk particles have high solubility and low retention time inside the plant system. Hence, the bioavailability of particles for the uptake are reduced.

As the retention time of nanoparticles are increased inside the plant system, they are translocated to all plant parts. Hence, the contents of the nanoparticles will be available for an extended period to the plant. This stimulates physiological and biochemical processes of plant system (Subbaiah, 2014). A gradual increase of nutrient uptake can be seen as granular size is reduced. Liscano *et al.* (2000) inferred that weight of granules of 1.5 mm is less than 2 mm or 2.5 mm. The use of smaller sized granules for same weight results in better distribution of nutrients and thus better nutrient uptake.

5.2. Influence of nanoparticles on storage life of chilli

Nanoparticles possess smaller particle sizes, higher surface area and increased proportion of reactive surface atoms when compared to bulk particles (Wigginton *et al.*, 2007). Such unique properties led to wide range of application in the different fields. Seed treatment with various nanoparticles have the potential to meet crop requirements.

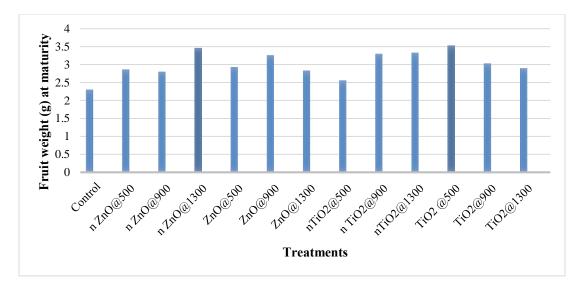


Fig. 5. Influence of normal grade and nano grade ZnO and TiO₂ on fruit weight at maturity in chilli

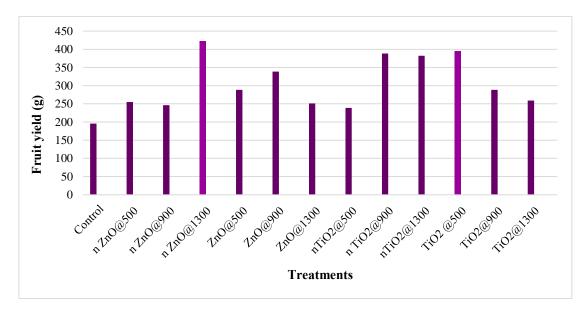


Fig. 6. Influence of normal grade and nano grade ZnO and TiO_2 on fruit yield in chilli

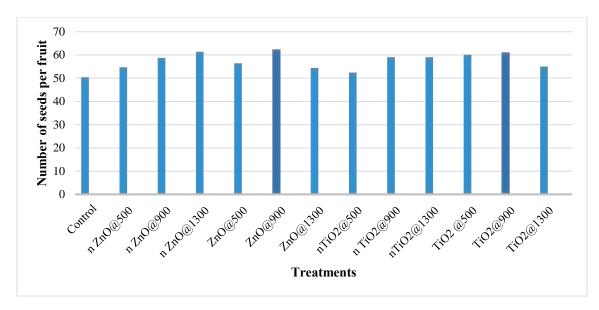


Fig. 7. Influence of normal grade and nano grade ZnO and TiO_2 on number of seeds per fruit in chilli

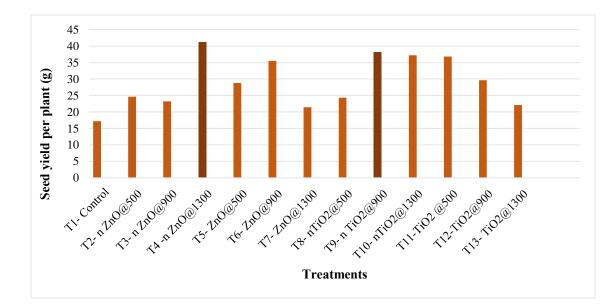


Fig. 8. Influence of normal grade and nano grade ZnO and TiO_2 on seed yield per plant in chilli

It can be applied either by soaking in solution containing specific concentration for specific time period (seed priming) or as seed coating. Seed deterioration is the loss of viability, quality and vigor of seeds either due to adverse environmental conditions or aging of seeds.

The exact reason for loss of viability is unknown, but according to Chiub *et al.* (1995) the primary reason is that, the non-enzymatic peroxidation of free radicles damages the membrane of seeds during storage. Even though seed quality is mainly governed by genetic makeup, seeds may get deteriorated during storage. This occurs due to mechanical damages, changes in temperature and humidity, presence of storage insects and pathogen, seed moisture content etc. during storage. Relative humidity is an important factor that directly influences the moisture content of seeds during storage. Thus physiological and biochemical changes occurs inside the seed and hence seed quality is reduced.

Fluctuating relative humidity and temperature are the major constraints for storage in chilli seeds. High relative humidity increase moisture content in seeds. High temperature along with increased moisture will enhance respiration which triggers deterioration. This results in reduced germination per cent and loss of vigour in seeds at the end of storage period (Barua *et al.*, 2009). Increased moisture may harbour storage pests and pathogens which in turn makes seeds unusable. In case of chilly seeds, it loses viability faster due to free fatty acid accumulation. So it is important to invigorate seeds to maintain viability and vigour throughout the period of storage.

5.3 Impact of seed invigoration using inorganic nanoparticles of zinc oxide and titanium dioxide on seed quality in chilly

5.3.1 Germination per cent

Various doses of nano grades of ZnO and TiO₂ exhibited significant differences in germination per cent throughout the storage period (Table 7). Irrespective of the treatments a decline in germination was seen over the period of storage. These results was in consonance with findings of Nagendra *et al.* (2017) in oriental pickling melon, Athmaja *et al.* (2019) in ash gourd, Navya (2016), Sandhya (2016) and Gayathri (2019) in chilli. The deterioration of seed quality was less in treated seeds compared to untreated seeds. According to Farooq *et al.* (2009) primed seeds show synchronized germination than control.

An increase in germination was seen in the initial month than the readings taken before storage. Gayathri (2019) recorded similar results and explained that the increment is due to the dormancy present in chilli during initial stages of growth.

In the seven months of storage period, all the treatments including control maintained the prescribed 60 percent germination by IMSCS (Indian Minimum Seed Certification Standards) till sixth month. At the end of storage ZnO@100 mg kg⁻¹ of seed(T₂), ZnO@250 mg kg⁻¹ of seed (T₃), ZnO@500 mg kg⁻¹ of seed (T₄), ZnO@1000 mg kg⁻¹ of seed (T₆), TiO₂@500 mg kg⁻¹ of seed (T₁₀), TiO₂@750 mg kg⁻¹ of seed (T₁₁) and TiO₂@1000 mg kg⁻¹ of seed (T₁₂) retained germination above 60 % (Figure 9).

The highest germination of 65.66 % was recorded by nano ZnO @ 250 mg kg⁻¹ of seed (T₃). Khanm *et al.* (2017) noticed significant germination (93.33 %) in tomato seeds when treated with nano ZnO at 400 ppm. Nano ZnO may affect the antioxidant system of plants which is important in mitigating the negative impact of reactive oxygen species (ROS) on photorespiration and photosynthesis (Burman *et al.*, 2013). It also increases indole acetic acid levels in roots thus growth rate is increased (Lu *et al.*, 2002).

Nano TiO₂ @ 1000 mg kg⁻¹ of seed (T₁₂) recorded highest germination (65.33) at the end of storage period. When nano-TiO₂ enter into cells, redox reactions occurs through superoxide ion radical and free radicles will be removed from the germinating seeds. The oxygen thus formed will be used for respiration and germination of seed is promoted (Zheng *et al.*, 2005).

Several studies reported the impact of ZnO and TiO₂ seed treatments in different crops. Meena *et al.* (2017) in maize and Baddar and Unrine (2018) in wheat reported impact of ZnO and Mahmoodzadeh *et al.* (2013) in canola, Haghighi *et al.* (2014) in Tomato, onion and radish and Debnath *et al.* (2020) in rice reported impact of TiO₂.

5.3.2 Seedling length (cm)

Seedling length was significantly influenced by storage period and seed treatments (Table 8). Treated seeds recorded highest seedling length than untreated seeds. Both shoot and root lengths were found to decline in all the treatments throughout the period of storage. Similar findings were reported by Sandhya (2016) and Gayathri (2019) in chilli variety Anugraha.

In case of shoot length, T₆- nano ZnO 1000 mg kg⁻¹ of seed (5.57 cm) which wqas on par with T₃- nano ZnO 250 mg kg⁻¹ of seed (5.26 cm), T₂ - nano ZnO-100 mg kg⁻¹ of seed (5.21 cm), T₁₁- nano TiO₂ 750 mg kg⁻¹ of seed (5.19 cm) and T₁₀- nano TiO₂ 500 mg kg⁻¹ of seed (5.13 cm) recorded higher shoot length at the end of storage (7 months). Similar observations on effect of ZnO nanoparticles was reported by Narendhran *et al.* (2016) in sesame, Afrayeem and Chaurasia, (2017) in chilli, and Raju and Rai, (2017) in pigeon pea. The impact of titanium dioxide nanoparticles was observed in crops like canola (Mahmoodzadeh *et al.*, 2013), radish (Haghighi *et al.*, 2014) and maize (Vijayalakshmi *et al.*, 2018).

Radicle growth was higher in nano ZnO @ 750 mg kg⁻¹ of seed (T₅) (9.37 cm) followed by T₆- nano ZnO@1000 (8.89) at the end of storage period. This was in confirmation with the observations of Pokhrel and Dubey, (2013) in maize, Segatto *et al.* (2018) in maize and sesame and Maity *et al.* (2018) in sorghum on impact of ZnO treatments. Laware and Raskar (2014) in onion and Hajra and Mondal (2017) in chickpea noticed the action of TiO₂ on radicle development.

Mahajan *et al.* (2011) observed that, seedling growth declined at higher concentrations of ZnO. At 20 ppm both root and shoot growth was at its maximum in mung bean and in gram, at 1ppm significant increase in root and shoot growth by 53.13% and 6.38% was recorded respectively. Similarly, Maity *et al.* (2018) also revealed that ZnO at lower doses of 750 mg kg⁻¹ was beneficial for growth in sorghum over higher dose of 1000 and 1250 mg kg⁻¹.

Zheng *et al.* (2005) noticed the impact of nano TiO_2 on seedling growth and reported that due to smaller particle size of nano TiO_2 it can be easily penetrated into

the seed and enhances the growth in spinach and Clément *et al.* (2013) noted that increment in root growth of flax seeds was due to antimicrobial activity of nano TiO₂.

5.3.3 Dry weight (mg)

Significant differences was seen in dry matter production when treated with nanoparticles of ZnO and TiO_2 in a declining trend. In chilli, Gayathri (2019) reported similar trend when treated with nanoparticles of ZnO and TiO_2 .

At the end of storage period, higher dry weight of 22.63 mg 10^{-1} seedlings was recorded in T₆-nano ZnO 1000 mg kg⁻¹ of seed. Similar observations was reported in pigeon pea (Raju and Rai, 2017), chickpea (Burman *et al.*, 2013) and wheat (Rizwan *et al.*, 2018).

In general, irrespective of treatments, as concentration of nanoparticle increased a decline in dry matter production was reported (Burman *et al.*, 2013). According to Zheng *et al.* (2005) as the chlorophyll content, rubisco activity and photosynthetic rate increased dry matter accumulation increased. He concluded that, the particle size and physiology of plants are related.

5.3.4 Vigour indices

Vigour indices of stored seeds declined during the advancement of storage (Table 11, 12). According to Kavitha (2002), when enzyme and amino acid synthesis decreases, loss of seed vigour might occur.

Higher seed vigour index I was recorded by nano ZnO @ 1000 mg kg⁻¹ of seed (925.88) followed by T_{10-} nano TiO₂ @ 500 mg kg⁻¹ of seed (866.93) at the end of storage (Figure 10). Nano TiO₂ @ 1000 mg kg⁻¹ of seed (T₁₂) (1471.81) which was on par with ZnO treatments like nano ZnO@ 250 mg kg⁻¹ of seed (T₃) and T₆₋ nano ZnO @ 1000 mg kg⁻¹ of seed (1448.75) maintained higher seed vigour index II at the end of period of storage (Figure 11). Several authors reported similar effects in seeds treated with ZnO and TiO₂ nanoparticles.

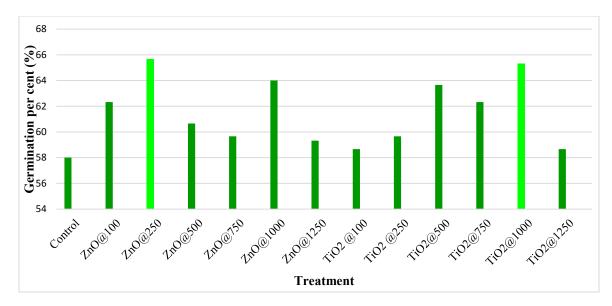


Fig. 9. Influence of nano grade ZnO and TiO₂ in maintaining seed germination after seven months of storage

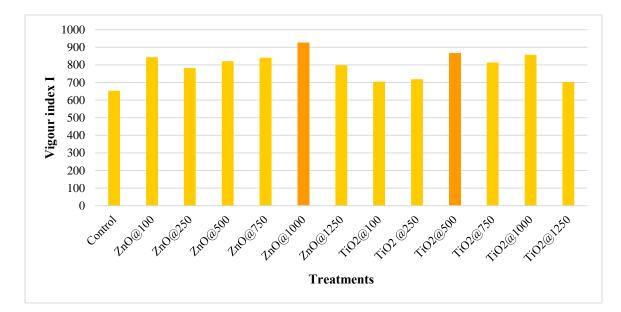


Fig. 10. Influence of nano grade ZnO and TiO₂on vigour index I after seven months of storage

Shyla and Natarajan (2014) in groundnut, Jalill and Yousef (2015) in rice and Maity *et al.* (2018) in cowpea reported the effects of nanoTiO₂ and Korishettar *et al.* (2016) in pigeon pea, Subbaiah *et al.* (2016) in maize and García-López *et al.* (2018) in chilli published the effects of nano ZnO treatments.

According to Kumar *et al.* (2019), nanoparticles enter into seeds through the cracks on surfaces of seeds. These particles will react with the free radicles present in the seeds and thus enhances viability and seed vigour. The reason for loss in seed vigour can be lipid peroxidation, accumulation of reactive oxygen species (ROS) (Bailly, 2004) or increase in free radicle concentration (Khan *et al.*, 1996). The ROS interrupts metabolism of seeds by making damage to proteins and lipids causing physiological and biochemical changes (Mittler *et al.*, 2004).

5.3.5 Electrical conductivity

Electrical conductivity is increased when there is electrolytic leakage due to loss in membrane integrity (Ratajczak and Pukacka, 2005). Significant increase in electrical conductivity was seen throughout the storage period in both ZnO and TiO₂ treatments (Table 13). Among the treatments control recorded highest electrical conductivity (404.66 μ Scm⁻¹) (Figure 12). Similar observations was recorded in seeds of chilli variety Anugraha by Sandhya (2016) and Gayathri (2019). Irrespective of the treatments and doses nanoparticles acts as a protector from seed deterioration.

According to Murali *et al.* (2002) germination and electrical conductivity are related. With the advancement of storage electrical conductivity increases which germination declines. As the permeability of membranes are lost, leaching of organic acids and sugars increases the conductivity of seeds (Sujatha and Srimathi, 2006). This leakage is more in aged seeds.

5.3.6 Seed moisture content (per cent)

Seed moisture content recorded insignificant differences among the treatments at the end of storage (Table 14). Only a marginal increment in moisture was recorded since 700 gauge polyethylene, moisture and vapour impervious packing was used.

5.3.7 Seed microflora (per cent)

Invasion of microorganisms play a major role in seed deterioration. Seed infection was higher at 7th month of storage than in the beginning of storage. Hence, we can say that pathogens have a role in determining period of storage.

Irrespective of the methods, control seeds showed more infection (Table 14). All the treatments except nano ZnO @ 100 mg kg⁻¹ of seeds (T₂), nano TiO₂ @ 250 mg kg⁻¹ of seeds (T₉) and control didn't exhibit any infection in agar plate method. While in blotter paper method, only control seeds showed seed infection. *Aspergillus flavus* and *Aspergillus niger* was identified from both agar plate and blotter paper method. Presence of *Aspergillus sp*. in stored seeds was observed in chilli by Sandhya (2016), Navya (2016), Chauhan *et al.* (2018) and Gayathri (2019). The pathogens might cause changes in protein, lipids, carbohydrate and vitamins. Reduction in total oil content, increased free fatty acid, discoloration, abnormal odours and caking of seeds also occurs and resulting in loss of germination.

Both ZnO and TiO₂ nanoparticles are effective in controlling pathogen infections. When the nanoparticles come in contact with the fungal membrane, it hinder the metabolic processes which eventually causes fatality of the pathogen. ZnO nanoparticles disrupt cellular functions and it results in deformation of fungal hyphae (He *et al.*, 2011) and also creates structural changes which causes cytoplasm leakage and death in bacteria (Brayner *et al.*, 2010). TiO₂ nanoparticles causes disintegration and deformation of fungal spores and hyphae (Argawy *et al.*, 2017). They also have anti-bacterial properties similar to ZnO particles (Frazer, 2001). According to Perez-de-Luque and Rubiales (2009) these inhibitory actions are due to the release of metabolites and extracellular enzymes.

In general, wet seed dressing with TiO_2 and ZnO nanoparticles were found to be effective in maintaining seed quality throughout the storage period. Higher doses (1250 mg kg⁻¹ of seed) were found to be highly effective. Feizi *et al.*, (2012) noticed that TiO_2 nanoparticles showed promontory effects for seedling length at 2ppm and 10 ppm on wheat and at higher doses neutral effects was observed.

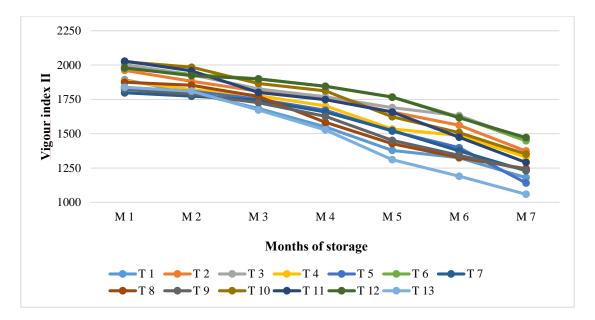


Fig. 11. Influence of nano grade ZnO and TiO₂on vigour index II after seven months of storage

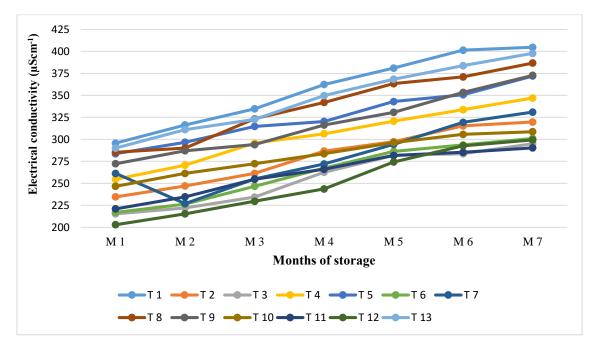
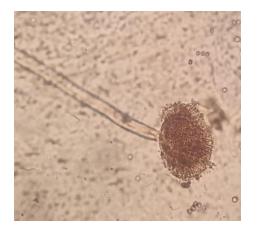


Fig. 12. Influence of nano grade ZnO and TiO₂on electrical conductivity after seven months of storage





Aspergillus flavus





Aspergillus niger

PLATE 6: Microorganisms observed after seven months of storage

FUTURE LINE OF WORK

- 1. Field performance of seeds wet dressed with nanoparticles is to be ascertained inorder to study its performance on growth parameters.
- 2. The present study has to be confined to a single variety. Hence it would be advisable to repeat the experiment in other varieties.
- 3. The study can be extended to ascertain the use of nanoparticle against pest population during storage for effective recommendations.
- 4. The impact of nanoparticle treatments on biochemical parameters like lipid peroxidation and dehydrogenase activity can be assessed.

<u>Summary</u>

6. SUMMARY

The study "Seed invigoration with nanoparticles for seed yield and quality in chilli (*Capsicum annuum* L.)" was conducted in the Department of Seed Science and Technology, College of Horticulture, Vellanikkara, Kerala Agricultural University, Thrissur. The study was conducted using chilli cv Anugraha. Seed invigoration was done with nano and normal grade ZnO and TiO₂.

6.1 Impact of nano and normal grade ZnO and TiO2 on seed yield

- The seeds treated with normal and nano grades of ZnO and TiO₂ performed better than untreated seeds. Significant variations was observed for traits such as plant spread, plant height, days to harvest, fruit length, number of fruits per plant, fruit weight at maturity, fruit yield, number of seeds per fruit, seed yield per plant and 100 seed weight.
- Nano ZnO@ 1300 mg kg⁻¹ of seed (T₄) performed superior for plant spread, plant height, number of fruits per plant, fruit length, fruit yield and seed yield per plant.
- Normal grade ZnO treatments was effective in improving number of seeds per fruit (ZnO @ 900 mg kg⁻¹ of seed).
- Significant effects were recorded when seeds were treated with normal grade TiO₂ @ 500 mg kg⁻¹ of seed (T₁₁) for fruit weight at maturity.
- In case of days to harvest, treatments nano ZnO @ 500 mg kg⁻¹ of seed (T₂), nano TiO₂ @ 900 mg kg⁻¹ of seed (T₉), nano TiO₂ @ 1300 mg kg⁻¹ of seed (T₁₀), ZnO @ 1300 mg kg⁻¹ of seed (T₇) and control recorded the least number of days.
- 100 seed weight of 0.55g was recorded in treatments like TiO₂ @900 mg kg⁻¹ of seed (T₉), TiO₂ @ 500 mg kg⁻¹ of seed (T₁₁), nano ZnO @ 1300 mg kg⁻¹ of seed (T₄), nanoTiO₂ @1300 mg kg⁻¹ of seed (T₁₀) and ZnO @ 900 mg kg⁻¹ of seed (T₆).

In general, nano grade seed treatments performed better than normal grade treatments in case of ZnO and for TiO₂, both normal and nano grade treatments performed well.

Impact of nano ZnO and TiO₂ on seed storage

- Seeds wet dressed with nano ZnO and TiO₂ were dried and packed in 700 gauge polythene bags and stored for 7 months. The treatments showed significant variations for germination, shoot length, root length, seedling dry weight, vigour indices and electrical conductivity of seed leachate over the period of storage.
- A declining trend was observed for germination, seedling length, dry weight and vigour indices while electrical conductivity and seed infection increased throughout the storage period.
- Nano seed treatments ZnO@100 mg kg⁻¹ of seed (T₂), ZnO@250 mg kg⁻¹ of seed (T₃), ZnO@500 mg kg⁻¹ of seed (T₄), ZnO@1000 mg kg⁻¹ of seed (T₆), TiO₂@500 mg kg⁻¹ of seed (T₁₀), TiO₂@750 mg kg⁻¹ of seed (T₁₁) and TiO₂@1000 mg kg⁻¹ of seed (T₁₂) retained germination above 60 % (IMSCS) after seven months of storage and nano ZnO @ 250 mg kg⁻¹ of seed (T₃) recorded highest value (65.66).
- Vigour index-I was found to be high in ZnO @ 1000mg kg⁻¹ of seeds (T₆) (925.88) followed by nano TiO₂ @ 500 mg kg⁻¹ of seed (866.93) and vigour index II in TiO₂ @1000 mg kg⁻¹ of seed (T₁₂) (1471.81) followed by ZnO @ 250 mg kg⁻¹ of seed (1457.58) at the end of storage.
- In case of root length, higher values were recorded by ZnO @750 mg kg⁻¹ of seed (9.37 cm) and shoot length by ZnO @ 1000 mg kg⁻¹ of seeds (5.57 cm).
- Dry matter production was high in nano ZnO @ 1000 mg kg⁻¹ of seed (T₆) (22.41mg).
- Seed treatments at nano ZnO @ 250 mg kg⁻¹ of seed (295 μScm⁻¹) followed by nano TiO₂ @ 750 mg kg⁻¹ of seed (290.33 μScm⁻¹) were able to maintain lower electrical conductivity of seeds.

Seed infection in agar plate method was noticed in treatments nanoZnO

 (a) 100 mg kg⁻¹ of seeds (T₂) (13.3%) and nano TiO₂ (a) 250 mg kg⁻¹ of seeds (T₉) (10%) at the end of storage. *Aspergillus niger* and *Aspergillus flavus* were the seed micrflora observed.

Seed treatment with nano ZnO at 1000 mg kg⁻¹ of seed was found to be effective in retaining seed longevity.

<u>References</u>

REFERENCES

- Abdul-Baki, A.A., and Anderson, J. D. 1973. Vigor determination in soybean seed by multiple criteria. *Crop Sci.* 13(6): 630-633.
- Adhikari, T., Kundu, S., and Rao, A. S. 2016. Zinc delivery to plants through seed coating with nano-zinc oxide particles. *J. Plant Nutr.* 39(1): 136–146.
- Afrayeem, S. M. and Chaurasia, A. K. 2017. Effect of zinc oxide nanoparticles on seed germination and seed vigour in chilli (*Capsicum annuum* L.). *Int. J. Pharmacol. Phytochem.* 6: 1564-1566.
- Agarwal, S. and Rathore, P. 2014. Nanotechnology pros and cons to agriculture: a review. *Int. J. Curr. Microbiol. App. Sci.* 3(3): 43-55.
- Argawy, E., Rahhal, M. M. H., El-Korany, A., Elshabrawy, E. M., and Eltahan, R. M.
 2017. Efficacy of some nanoparticles to control damping-off and root rot of sugar beet in El-Behiera governorate. *Asian J. Plant Pathol.* 11: 35-47.
- Alam, M. Z., Hamim, I., Ali, M. A., and Ashrafuzzaman, M. 2014. Effect of seed treatment on seedling health of chili. J. Environ. Sci. Nat. Resour. 7(1): 177-181.
- Anandaraj and Natarajan, N. 2017. Effect of nanoparticles for seed quality enhancement in onion [Allium cepa (Linn) cv. CO (On)] Int. J. Curr. Microbiol. App. Sci. 6(11): 3714-3724.
- Arnon, D. I. and Stout, P. R. 1939. The essentiality of certain elements in minute quantity for plants with special reference to copper. *Plant Physiol.* 14: 371-375.
- Athmaja, S., Francies, R. M., Bastian, D., George, S. T., and Cherian, K. A. 2019. Impact of pre-storage seed invigoration in ash gourd (*Benincasa hispida* (Thunb.) Cogn.). Veg. Sci. 45(2): 297-299.

- Avinash, C. P., Sharda, S. S., and Raghavendra, S. Y. 2010. Application of ZnO nanoparticles in influencing the growth rate of *Cicer arietinum*. J. Exp. Nanoscience 5(6): 488-497.
- Baddar, Z. E. and Unrine, J. M. 2018. Functionalized-ZnO-nanoparticle seed treatments to enhance growth and Zn content of wheat (*Triticum aestivum*) seedlings, *Agric. Food Chem.* 66(46): 12166–12178.
- Bagawade, J. A. and Jagtap, S. S. 2020. Effect of ZnO nanoparticles on chlorophyll content of wheat plants (*Triticum Aestivum* L.) *Int. J. Eng. Adv. Technol.* 9(4): 28-30.
- Bailly, C. 2004. Active oxygen species and antioxidants in seed biology. Seed Sci. 14: 93-107.
- Bala, R., Kalia, A., and Dhaliwal, S. S. 2019. Evaluation of Efficacy of ZnO Nanoparticles as Remedial Zinc Nanofertilizer for Rice. J. Soil. Sci. Plant. Nut. 1-10.
- Balogun, O. S., Odeyemi, G. A., and Fawole, O. B. 2005. Evaluation of the pathogenic effect of some fungal isolates on fruits and seedlings of pepper (*Capsicum* spp) J. Agric. Res. Dev. 4(2): 159-169.
- Barua, H., Rahman, M. M., and Masud, M. M. 2009. Effect of storage containers environment at different storage period on the quality of chilli seed. *Int. J. Sustain. Crop Prod.* 4(4): 28-32.
- Boonyanitipong, L. P., Kositsup, B., Kumar, P., Baruah, S., and Dutta, J. 2011. Toxicity of ZnO and TiO₂ nanoparticles on germinating rice seed. *Int. J. Biosci. Biochem. Bioinforma.* 1(4):282-285.
- Buchanan, B. B., Gruissem, W., and Jones, R. L. 2000. Biochemistry and molecular biology of plants (1st Ed.). American Society of Plant Physiologists, Rockville. 1222p.
- Burman, U., Saini, M., and Kumar, P. 2013. Effect of zinc oxide nanoparticles on growth and antioxidant system of chickpea seedlings. *Toxicol. Environ. Chem.*

95(4): 605–612.

- Chauhan R.T., Patel, P.R., and Thumar, V.M. 2018. Occurrence of seed borne pathogens in Chilli (*Capsicum frutescence* L.) cv. GVC 111 in vitro. Int. J. Chem. Stud. 6(2): 374-1376.
- Chigoziri, E. and Ekefan, E. J. 2013. Seed-borne fungi of chilli pepper (*Capsicum frutescens*) from pepper producing areas of Benue state, Nigeria. *Agric. Biol. J. Am.* 4: 370-374.
- Chiub, K. Y., Wang, C. S., and Sung, J. M. 1995. Lipid peroxidation and peroxidescavenging enzymes associated with accelerated aging and hydration of watermelon seeds differing in ploidy. *Physiol. Plant.* 94: 441-446.
- Clément, L., Hurel, C., Marmier, N. 2013. Toxicity of TiO₂ nanoparticles to cladocerans, algae, rotifers and plants - Effects of size and crystalline structure. *Chemosphere* 90(3): 1083-1090.
- Debnath, K., Das, A., Das, B., and Karfoma, J. 2020. TiO₂ nanoparticles enhancing germination, growth and yield of rice. *Int. Res. J. Pure Appl. Chem.* 25-30.
- Davarpanah, S., Tehranifar, A., Davarynejad, G., Aran, M., Abadía, J., and Khorassani,
 R. 2017 Effects of foliar nano-nitrogen and urea fertilizers on the physical and
 chemical properties of pomegranate (Punica granatum cv.Ardestani) fruits.
 Hort. Sci. 52: 288–294.
- Elizabath, A., Bahadur, V., Misra, P., Prasad, V. M., and Thomas, T. 2017. Effect of different concentrations of iron oxide and zinc oxide nanoparticles on growth and yield of carrot (*Daucus carota* L.). J. Pharmacognosy Phytochem. 6(4): 1266-1269.
- Farooq, M., Basra, S. M. A., Wahid, A., Khaliq, A., and Kobayashi, N. 2009. Rice seed invigoration. In: Lichtfouse, E, (ed.), *Sustainable Agriculture Reviews*. Proceedings of a conference. Springer, Netherlands. pp. 137-175.

Feizi, H., Kamali, M., Jafari, L., and Moghaddam, P. R. 2013. Phytotoxicity and

stimulatory impacts of nanosized and bulk titanium dioxide on fennel (*Foeniculum vulgare* Mill). *Chemosphere* 91(4): 506-511.

- Feizi, H., Moghaddam, P. R., Shahtahmassebi, N., and Fotova, A. 2012. Impact of bulk and nanosized titanium dioxide (TiO₂) on wheat seed germination and seedling growth. *Biol. Trace Elem. Res.* 146: 101–106.
- Frazer, L. 2001. Titanium dioxide: environmental white knight?. Environ. Health Perspect. 109(4): A174-A177.
- Gao, F., Hong, F., Liu, C., Zheng, L., Su, M., Wu, X., Yang, F., Wu, C., and Yang, P.
 2006. Mechanism of nano-anatase TiO₂ on promoting photosynthetic carbon reaction of spinach. *Biol. Trace Element Res.* 111: 239-253.
- García-López, J., Zavala-García, F., Olivares-Sáenz, E., Lira-Saldívar, R. H., Barriga-Castro, E. D., Ruiz-Torres, N. A., Ramos-Cortez, E., Vázquez-Alvarado, R., and Niño-Medina, G. 2018. Zinc oxide nanoparticles boosts phenolic compounds and antioxidant activity of *Capsicum annuum* L. during germination. *Agronomy* 8(10): 215.
- Gayathri, S. 2019. Seed invigoration with inorganic nanoparticles in chillies (*Capsicum annuum* L.). M.Sc.(Ag) thesis, Kerala Agricultural University, Thrissur, 146p.
- Gomez, K. A. and Gomez, A. A. 1976. Statistical Procedures for Agricultural Research (2nd Ed.). John Wiley and Sons, New York, 680p.
- Grajkowski, J. and Ochmian, I. 2007. Influence of three biostimulants on yielding and fruit quality of three primocane raspberry cultivars. *Acta. Sci. Pol. Hortorum Cult.* 6: 29-36.
- Grusak, M. L. 2002. Enhancing mineral content in plant food product. *J. Am. Coll. Nutr.* 21: 178-183.
- Gyasi, E., Kwoseh, C., and Moses, E. 2020. Identification of seed-borne fungi of farmer-saved seeds of pepper and their control with some selected botanicals. *Ghana J. Agric. Sci.* 55(1): 43 - 53.

- Haghighi, M., Heidarian, S., and Teixeira-da-Silva, J. A. 2014. The effect of titanium amendment in N-withholding nutrient solution on physiological and photosynthesis attributes and micronutrient uptake of tomato. *Biol. Trace. Elem. Res.* 150: 381–390.
- Hajra, A. and Mondal, N. K. 2017. Effects of ZnO and TiO₂ nanoparticles on germination, biochemical and morphoanatomical attributes of *Cicer arietinum* L. *Energy, Ecol. Environ.* 2(4): 277-288.
- Hatami, M., Ghorbanpour, M., Salehiarjomand, H. 2014. Nanoanatase TiO₂ modulates the germination behavior and seedling vigority of some commercially important medicinal and aromatic plants. J. Biol. Environ. Sci. 8: 53–59.
- He, L., Liu, Y., Mustapha, A., and Lin, M. 2011. Antifungal activity of zinc oxide nanoparticles against *Botrytis cinerea* and *Penicillium expansum*, *Microbiol. Res.* 166(3): 207-215.
- Hong, F. S., Yang, F., Ma, Z. N., Zhou, J., Liu, C., Wu, C., and Yang, P. 2005. Influences of nano-TiO₂ on the chloroplast ageing of spinach under light. *Biol. Trace Elem. Res.* 104(3): 249–260.
- Hruby, M., Cigler, P., and Kuzel, S. 2002. Contribution to understanding the mechanism of titanium action in plant. *J. Plant Nutr.* 25: 577–598.
- ISTA [International Seed Testing Association] 1985. International rules for seed testing. *Seed Sci. Technol.* 13: 299-355.
- ISTA [International Seed Testing Association] 1999. International rules for seed testing. *Seed Sci. Technol.* 27: 1-340.
- Jaberzadeh, A., Moaveni, P., Moghadam, H. R. T., and Zahedi, H. 2013. Influence of bulk and nanoparticles titanium foliar application on some agronomic traits, seed gluten and starch contents of wheat subjected to water deficit stress. *Not. Bot. Horti. Agrobo.* 41(1): 201-207.

Jalill, A. and Yousef, A. M. 2015. Comparison the phytotoxicity of TiO2 nanoparticles

with bulk particles on amber 33 variety of rice (*Oryza sativa*) in vitro. *Sch. Acad. J. Biosci.* 3(3): 254-262.

- Janmohammadi, M., Amanzadeh, T., Sabaghnia, N., and Dashti, S. 2016. Impact of foliar application of nano micronutrient fertilizers and titanium dioxide nanoparticles on the growth and yield components of barley under supplemental irrigation. Acta. Agriculturae. Slovenica. 107: 265 – 276.
- Jayarambabu, N., Kumari, B. S., Rao, K. V., and Prabhu, Y. T. 2014. Germination and growth characteristics of mungbean seeds (*Vigna radiata* L.) affected by synthesized zinc oxide nanoparticles. *Int. J. Curr. Eng. Technol.* 4(5): 3411-3416.
- Jo, Y. K. and Kim, B. H. 2009. Antifungal activity of silver ions and nanoparticles on phytopathogenic fungi. *Plant Dis.* 93: 1037–1043.
- Jogi, M. G., Padule, D. N., and Kamdi, S. R. 2010. Detection of seed mycoflora of chilli and its impact on seed germination and seedling vigour. *Int. J. Plant Sci.* 5(2): 502-504.
- Justic, O. L. and Bass, L. N. 1978. Principles and Practices of Seed Storage. USDA Agricultural Handbook, No. 506 Washington, pp. 53-60.
- Kavitha, S. 2002. Seed hardening and pelleting for maximizing the productivity of blackgram (*Vigna mungo* (L.) Hepper) cv. Vamban-3 under rainfed conditions. M.Sc.(Agri.) Thesis, Tamil Nadu Agricultural University, Coimbatore.
- Khan, M. M., Hendry, G. A. F., Atherton, N. M., and Vertucci-Walters, C. W. 1996. Free radical accumulation and lipid peroxidation in testas of rapidly aged soybean seeds: A light promoted process. *Seed Sci. Res.* 6: 101-107.
- Khanm, H., Vaishnavi, B. A., Namratha, M. R., and Shankar, A. G. 2017. Nano zinc oxide boosting growth and yield in tomato: the rise of "nano fertilizer era". *Int. J. Agric. Sci.Res.* 7(3): 197-206.

- Khater, M. S. 2015. Effect of titanium nanoparticles (TiO₂) on growth, yield and chemical constituents of coriander plants. *Arab J. Nucl. Sci. Appl.* 48(4): 187-194.
- Kheyri, N., Norouzi, H. A., Mobasser, H. R., and Torabi, B. 2019. Effects of silicon and zinc nanoparticles on growth, yield, and biochemical characteristics of rice. *Crop Ecol. Physiol.* 111(6): 1-7.
- Khot, L. R., Sankaran, S., Maja, J. M., Ehsani, R., and Schuster, E. W. 2012. Application of nanomaterial in agricultural production and protection: a review. *Crop Prot.* 64-70.
- Korishettar, P., Vasudevan, S. N., Shakuntala, N. M., Doddagoudar, S. R., Hiregoudar, S., and Kisan, B., 2016. Seed polymer coating with Zn and Fe nanoparticles: An innovative seed quality enhancement technique in pigeonpea. J. Appl. Nat. Sci. 8(1): 445-450.
- Kumar, G.D., Raja, K. Natarajan, N., Govindaraju, K., and Subramanian, K. S. 2019 Invigouration treatment of metal and metal oxide nanoparticles for improving the seed quality of aged chilli seeds (*Capsicum annum* L.) *Mater. Chem. Phys.* 1-16
- Kumar, S. 2012. Cultural approaches for plant disease management; research & reviews. J. Agric. Sci. Technol. 1(2): 12-21.
- Laware, S. L. and Raskar, S. 2014. Effect of titanium dioxide nanoparticles on hydrolytic and antioxidant enzymes during seed germination in onion. *Int. J. Curr. Microbiol. App. Sci.* 3(7): 749-760.
- Liscano, J. F., Wilson, C. E., Norman, R. J., and Slaton, N. A. 2000. Zinc availability to rice from seven granular fertilizers. *AAES. Res. Bulletin.* 963: 1–31.
- López-Moreno, M. L., de la Rosa, G., Hernandez-Viezcas, J.A., Castillo- Michael, H., Botez, C.E., Peralta-Videa, J.R., and Gardea-Torresdey, J.L. 2010. Evidence of the differential biotransformation and genotoxicity of ZnO and CeO2

nanoparticles on soybean (*Glycine max*) plants. *Environ. Sci. Technol.* 44: 7315–7320.

- Lu, C. M., Zhang, C. Y., Wen, J. Q., Wu, G. R., and Tao, M. X. 2002. Research of the effect of nanometer materials on germination and growth enhancement of *Glycine max* and its mechanism. *Soybean Sci.* 21: 168-172.
- Machenahalli, S., Nargund, V. B., and Patil, S. 2014. Quick detection and diagnosis of chilli fruit rot pathogens. *Int. J. Plant Res.* 27(3):403-406.
- Mahajan, P., Dhoke, S. K., Khanna, A. S., and Tarafdar, J. C. 2011. Effect of nano-zno on growth of mung bean (*Vigna radiata*) and chickpea (*Cicer arietinum*) seedlings using plant agar method. *Appl. Bio. Res.* 13 (2): 54-61.
- Mahdieh, M., Sangi, M. R., Bamdad, F., and Ghanem, A. 2018. Effect of seed and foliar application of nano-zinc oxide, zinc chelate, and zinc sulphate rates on yield and growth of pinto bean (*Phaseolus vulgaris*) cultivars. J. Plant Nutr. 1-12.
- Mahmoodzadeh, H., Nabavi, M., and Kashefi, H. 2013. Effect of nanoscale titanium dioxide particles on the germination and growth of canola (*Brassica napus*).J. Ornam. Hortic. Plants 3(1): 25-32.
- Mahmoud, A.W.M., Abdelaziz, S.M., El-Mogy, M.M., and Abdeldaym, E.A.2019. Effect of foliar ZnO and FeO nanoparticles application on growth and nutritional quality of red radish and assessment of their accumulation on human health. *Agric*. 65(1).16–29.
- Maity, A., Natarajan, N., Pastor, M., Vijay, D., Gupta, C. K., and Wasnik, V. K. 2018.
 Nanoparticles influence seed germination traits and seed pathogen infection rate in forage sorghum (*Sorghum bicolour*) and cowpea (*Vigna unguiculata*). *Indian J. Exp. Biol.* 56: 363-372.
- Marschner, H. 1995. *Mineral Nutrition of Higher Plants* (2nd Ed.). Academic Press, London. 889p.

Mattiello, A. and Marchio, L. 2017. Application of nanotechnology in agriculture:

assessment of tio₂ nanoparticle effects on barley. 29-39.

- Meena, D. S., Jayadeva, H. M., Gautam, C., and Meena, H. M. 2017. Effects of nano zinc oxide (ZnO) particles on germination of maize (*Zea mays* L.) seeds. *Int. J. Plant Soil Sci.* 16(1): 1-5.
- Mittler, R., Vanderauwera, S., Gollery, M., and Van Breusegem, F. 2004. Reactive oxygen gene network of plants. *Trends Plant Sci.* 9: 490–498.
- Moaveni, P., Talebi, A., Farahani, A., and Maroufi, K. 2011. Study of nano particles TiO₂ spraying on some yield components in barley (*Hordem vulgare* L.). *Intl. Conf. Environ. Agri. Eng.* 115–119.
- Mortvedt, J. J. 1992. Crop response to level of water -soluble zinc in granular zinc fertilizers. *Fertil. Res.* 33: 249-255.
- Murali, M. R., Shashidhara, S. D., and Vyakatanahal, B. S. 2002. Ivestigation on seed viability in blackgram and greengram *J. Res. Angrau.* 30(1): 34-39.
- Nagendra, M. S., Bastian, D., Francies, R. M., and Rajendra, A. A. 2017. Effect of sowing time on fruit and seed yield in oriental pickling melon (Cucumis melo var. conomon). Int. J. Chem. Stud. 5(4): 1910-1912.
- Narendhran, S., Rajiv, P., and Sivaraj, R. 2016. Influence of zinc oxide nanoparticles on growth of *Sesamum indicum* L. in zinc deficient soil. *Int. J. Pharm. Pharm. Sci.* 8(3): 365-371.
- Navya, P. 2016. Halogenation for improvement of storage life in chilli seeds. M.Sc.(Ag) thesis, Kerala Agricultural University, Thrissur, 116p.

Neergaard, P. 1979. Seed Pathology: Vol 1. The Macmillan Press Ltd, London, 746p.

NHB [National Horticultural Board]. 2019. NHB homepage. Available: http://www.**nhb**.gov.in.

Owolade, O. F. and Ogunleti, D. O. 2008. Effects of titanium dioxide on the diseases,

development and yield of edible cowpea. J. Plant Prot. Res. 329-335.

- Perez-de-Luque, A. and Rubiales, D. 2009. Nanotechnology for parasitic plant control. *Pest Manage. Sci.* 65: 540-545.
- Pokhrel, L. R. and Dubey, B. 2013. Evaluation of developmental responses of two crop plants exposed to silver and zinc oxide nanoparticles. *Sci. Total Environ.* 452: 321-332.
- Poornima, R. and Koti, R. V. 2019. Effect of nano zinc oxide on growth, yield and grain zinc content of sorghum (Sorghum bicolor). J. Pharmacognosy Phytochem. 8(4): 727-731.
- Potarzycki, J. and Grzebisz, W. 2009. Effect of zinc foliar application on grain yield of maize and its yielding components. *Plant Soil Environ*. 55(12): 519-527.
- Prasad, T. N. V. K. V., Sudhakar, P., Sreenivasulu, Y., Latha, P., Munaswamy, V., Reddy, K. R., Sreeprasad, T. S., Sajanlal, P. R., and Pradeep, T. 2012. Effect of nanoscale zinc oxide particles on the germination, growth and yield of peanut. J. Plant Nutr. 905-927.
- Priya, B., Srinivasarao, M., Satyanarayana, N. H., Mukherjee, S., Das, B., and Sarkar, K. K. 2016. Effect of metal based nanoparticles (ZnO and TiO₂) on germination and growth of cowpea seedling. In: Proceedings of National Conference on Harmony with Nature in Context of Resource Conservation and Climate Change, 22 – 24 Oct 2016, Hazaribag. Vinoba Bhave University, Hazaribag, Jharkhand, pp. 359-365.
- Raju, B. B. and Rai, P. K. 2017. Studies on effect of polymer seed coating, nanoparticles and hydro priming on seedling characters of pigeonpea (*Cajanus cajan* L.) seed. J. Pharmacognosy Phytochem. 6(4): 140-145.
- Raliya, R., Nair, R., Chavalmane, S., Wei-Ning, W., and Biswas, P. 2015. Mechanistic evaluation of translocation and physiological impact of titanium dioxide and zinc oxide nanoparticles on the tomato (*Solanum lycopersicum* L.) plant. *R*.

Soc. Chem. 1584-1594.

- Ram, N., Verloo, M., and Cottenie, A. 1983. Response of bean to foliar spray of titanium. *Plant Soil* 73(2): 285-290.
- Ratajczak, E. and Pukacka, S. 2005. Decrease in beech (*Fagus sylvatica*) seed viability caused by temperature and humidity conditions as related to membrane damage and lipid composition. *Acta. Physiol. Plant.* 27: 3–12.
- Rezaei, I. F., Moaveni, P., and Mozafari, H. 2015. Effect of different concentrations and time of nano TiO₂ spraying on quantitative and qualitative yield of soybean (*Glycine max* L.) at Shahr-e-Qods. *Biol. Forum Int. J.* 7(1): 957-964.
- Rizwan, M., Ali, S., Ali, B., Adrees, M., Arshad, M., Hussain, A., Rehman, M. Z., and Waris, A. A. 2018. Zinc and iron oxide nanoparticles improved the plant growth and reduced the oxidative stress and cadmium concentration in wheat. *Chemosphere* 214: 269-277.
- Sadak, M. S. and Bakry, B. A. 2020. Zinc-oxide and nano ZnO oxide effects on growth, some biochemical aspects, yield quantity, and quality of flax (*Linum uitatissimum* L.) in absence and presence of compost under sandy soil. Bull. Natl. Res. Cent. 1-12.
- Salama, H. M. H. 2012. Effects of silver nanoparticles in some crop plants, common bean (*Phaseolus vulgaris* L.) and corn (*Zea mays* L.). *Int. Res. J. Biotech.* 3(10): 190-197.
- Sandhya, R. 2016. Seed treatment with botanicals to enhance seedling vigour in chilli (*Capsicum annuum* L.). M.Sc.(Ag) thesis, Kerala Agricultural University, Thrissur, 104p.
- Segatto, C., Ternus, R., Junges, M., de Mello, J. M. M., da Luz, G. L., Riella, H. G., Silva, L. L., Lajús, C. R., and Fiori, M. A. 2018. Adsorption and incorporation of the zinc oxide nanoparticles in seeds of corn: germination performance and antimicrobial protection. *Int. J. Adv. Eng. Res. Sci.* 5(5): 277-295.

- Sharfun-Nahar, S. N., Mushtaq, M., and Pathan, I. H. 2004. Seed-borne mycoflora of *Capsicum annuum* imported from India. *Pak. J. Bot.* 36(1): 191-198.
- Shyla, K. K. and Natarajan, N. 2014. Customizing zinc oxide, silver and titanium dioxide nanoparticles for enhancing groundnut seed quality. *Indian J. Sci. Technol.* 7(9): 1376-1381.
- Singh, G. and Gill, A. 1994. Evaluation of soybean genotypes for seed storability. Seed Res. 22:137-140.
- Subbaiah, L. V. 2014. Effect of nanoscale zinc oxide particles on the yield and yield attributes of maize. M.Sc. (Ag) thesis, Acharya N. G. Ranga Agricultural University, Guntur. 102p.
- Subbaiah, L.V., Prasad, T. N. V. K. V., Krishna, T. G., Sudhakar, P., Reddy, B. R., and Pradeep, T. 2016. Novel effects of nanoparticulate delivery of zinc on growth, productivity, and zinc biofortification in maize (*Zea mays L.*). J. Agric. Food Chem. 64: 3778–3788.
- Sujatha, K. and Srimathi, P. 2006. Chemical priming for improved seed yield and quality in blackgram (*Vigna mungo* (L.) Hepper) var. Co 5. *Plant Arch.* 6(1): 177-180.
- Suppan, S. 2013. *Nanomaterials in Soil: Our Future Food Chain?* Institute for Agriculture and Trade Policy, Minnesota, United States, 17p.
- Tarafdar, J. C., Raliya, R., Mahawar, H., and Rathore, I. 2014. Development of zinc nanofertilizer to enhance crop production in pearl millet (*Pennisetum americanum*), Agric. Res. 3(3): 257–262.
- Tekrony, D. M. and Egli, D. B. 1977. Relationship between laboratory indices of soybean seed vigour and field performance. *Crop Sci.* 17: 573 577.
- Tiwari, P. K. 2017. Effect of zinc oxide nanoparticles on germination, growth and yield of maize (*Zea mays* L.). M.Sc.(Ag) thesis, Anand Agricultural University, Gujarat, 116p.

- Vazin, F. 2012. Effect of zinc sulphate on quantitative and qualitative characteristics of corn (*Zea mays*) in drought stress. Cercetari Agronomice in Moldova. 45(3): 15-24.
- Vijayalakshmi, V., Ramamoorthy, K., and Natarajan, N. 2018. TiO₂ nano particles on extending seed vigour and viability of naturally aged maize (*Zea mays* L.) seeds. J. Pharmacognosy Phytochem. 7(1): 2221-2224.
- Wigginton, N. S., Haus K. L., and Hochella, M. F. Jr. 2007. Aquatic environmental nanoparticles. J. Env. Monit. 9: 1306-1316.
- Yusefi- Tanha, E., Fallah, S., Rostamnejadi, A., and Pokhrel, L.R. 2020. Zinc oxide nanoparticles (ZnONPs) as a novel nanofertilizer: Influence on seed yield and antioxidant defense system in soil grown soybean (Glycine max cv. Kowsar). Sci. Total, Env. 738: 1-13.
- Zagzog, O. A., Gad, M. M., and Hafez, N. K. 2017. Effect of nano-chitosan on vegetative growth, fruiting and resistance of malformation of mango. *Trends Hortic. Res.* 7: 11-18.
- Zheng, L., Hong, F., Lu, S., and Liu, C. 2005. Effect of nano-TiO₂ on strength of naturally aged seeds and growth of spinach. *Biol. Trace Elem. Res.* 104(1): 83-91.
- Ziaeyan, A. H. and Rajaie, M. 2009. Combined effect of zinc and boron on yield and nutrients accumulation in corn. *Int. J. Plant Prod.* 3(3): 35-44.

SEED INVIGORATION WITH NANOPARTICLES FOR SEED YIELD AND QUALITY IN CHILLI

(Capsicum annuum L.)

by

RIYA MARY MATHEW

(2018 - 11 - 154)

ABSTRACT OF THE THESIS

Submitted in partial fulfilment of the requirements for the degree of

Master of Science in Agriculture

Faculty of Agriculture Kerala Agricultural University



DEPARTMENT OF SEED SCIENCE AND TECHNOLOGY COLLEGE OF HORTICULTURE VELLANIKKARA, THRISSUR - 680 656 KERALA, INDIA 2020

ABSTRACT

The study "Seed invigoration with nanoparticles for seed yield and quality in chilli" was conducted in the Department of Seed Science and Technology, College of Horticulture, Vellanikkara during May 2019 with the objective of standardizing the optimum seed treatment dose required for increasing yield, improving quality and prolonging seed longevity. Normal grade and nano grade zinc oxide (ZnO) and titanium dioxide (TiO₂) were used for seed treatment on chilli variety Anugraha.

The study consisted of two experiments. In the first experiment, chilli seeds dry dressed with 500, 900 and 1300 mg kg⁻¹ of normal and nano grade ZnO and TiO₂ along with control (thirteen treatments) were raised in a Randomized block design with three replications. While all treated seeds performed better than control (untreated seeds) treatments with nanoparticles performed better than the normal grade particles. Highly significant variations was observed for traits such as plant spread, plant height, days to harvest, fruit length, number of fruits per plant, fruit weight at maturity, fruit yield, number of seeds per fruit, seed yield per plant and 100 seed weight.

Seed treatments with nano ZnO @ 1300 mg kg⁻¹ of seed (T₄) performed superior for plant spread (60.1 cm), plant height (67.30 cm), number of fruits per plant (122), fruit length (7.40cm), fruit weight (3.46g), fruit yield (422.70 g) and seed yield per plant (41.14 g). Normal grade ZnO treatments was effective in improving number of seeds per fruit (62) (ZnO @ 900 mg kg⁻¹ of seed). Significant effects were recorded when seeds were treated with normal grade TiO₂ @ 500 mg kg⁻¹ of seed (T₁₁) for fruit weight at maturity (3.53 g).

In the second experiment the effect of wet seed treatment with nanoparticles on seed storability was assessed. The experiment was laid out in Completely Randomized Design with thirteen treatments in three replications. Treatments include nano ZnO and TiO₂ at 100,250,500 and 100 mg kg⁻¹. Seed quality parameters like germination, seedling length, dry weight and vigour indices showed a declining trend throughout the storage period while electrical conductivity and seed infection increased. Treatments ZnO@100 mg kg⁻¹ of seed (T₂), ZnO@250 mg kg⁻¹ of seed (T₃), ZnO@500 mg kg⁻¹ of seed (T₄), ZnO@1000 mg kg⁻¹ of seed (T₆), TiO₂@500 mg kg⁻¹ of seed (T₁₀),

TiO₂@750 mg kg⁻¹ of seed (T₁₁) and TiO₂@1000 mg kg⁻¹ of seed (T₁₂) retained germination above 60 % (IMSCS) at the end of seven months of storage. ZnO @ 250 mg kg⁻¹ of seed (T₃) retained the highest germination per cent of 65.66 %.

Vigour index-I was found to be high in ZnO @ 1000mg kg⁻¹ of seeds (T₆) (925.88) and vigour index II in TiO₂ @1000 mg kg⁻¹ of seed (T₁₂) (1471.81) at the end of storage. The electrical conductivity of seed leachate obtained from nano ZnO @ 250 mg kg⁻¹ of seed and nano TiO₂ @ 750 mg kg⁻¹ of seed recorded the lowest values (295 μ Scm⁻¹and 290.33 μ Scm⁻¹). Pathogen infection in seeds were found to be lower in treated seeds than in untreated control. Infection was recorded in nano ZnO @ 100 mg kg⁻¹ of seeds (T₂) (13.3%) and nano TiO₂ @ 250 mg kg⁻¹ of seeds (T₉) (10%) treatments. The seed storage fungi observed were *Aspergillus niger* and *Aspergillus flavus*.

Seed treatments with inorganic nanoparticles are effective in improving field performance and as well as retaining seed quality in storage. Nano ZnO at 1300 mg kg⁻¹ of seeds was the best treatment followed by nano TiO₂ at 900mg kg⁻¹ of seeds in improving yield in chilli. Treatments nano ZnO at 250 and 1000 mg kg⁻¹ of seed may be used as seed treatments to enhance seed longevity in chilli.

Appendix-I

| Months | Temperature (⁰ C) | | Relative | Rainfall | Rainy | Mean | Wind |
|----------------|-------------------------------|---------|----------|----------|-------|-----------|--------|
| | Mean | Mean | Humidity | (mm) | days | sunshine | speed |
| | Maximum | minimum | (%) | | | hours | (kmph) |
| | | | | | | (hrs/day) | |
| June 2019 | 32.2 | 23.5 | 83 | 324.4 | 15 | 3.7 | 1.7 |
| July 2019 | 30.4 | 22.8 | 85 | 654.4 | 21 | 2.6 | 1.7 |
| August 2019 | 29.5 | 21.9 | 89 | 977.5 | 24 | 1.5 | 1.5 |
| September 2019 | 31.2 | 22 | 85 | 419.0 | 19 | 3.3 | 1.4 |
| October 2019 | 32.4 | 21.4 | 80 | 418.4 | 16 | 5.5 | 1.8 |
| November 2019 | 32.9 | 21.7 | 71 | 205.0 | 5 | 7.5 | 4 |
| December 2019 | 32.3 | 22.1 | 63 | 4.4 | 1 | 6.7 | 8.7 |
| January 2020 | 34.1 | 22.4 | 60 | 0 | 0 | 9.4 | 5.9 |
| February 2020 | 35.5 | 23.2 | 54 | 0 | 0 | 9.5 | 5.3 |
| March 2020 | 36.4 | 24.4 | 65 | 33.4 | 2 | 8.5 | 2.8 |

Monthly meteorological data from June 2019 to March 2020